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TRICKLE-BED BIOFILTER**

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K. Thomas Klasson, Brian H. Davison, John W. Barton, and Jennifer E. Jacobs

Chemical Technology Division
Oak Ridge National Laboratory¹
Oak Ridge, Tennessee 37831-6044

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Removal of Chlorinated and Non-Chlorinated Alkanes in a Trickle-Bed Biofilter

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K. Thomas Klasson, Brian H. Davison, John W. Barton, and Jennifer E. Jacobs

Oak Ridge National Laboratory (managed by Lockheed Martin Energy Research Corp. for the U.S. Department of Energy under contract DE-AC05-96OR22464), Oak Ridge, TN 37831-6044

ABSTRACT

Increasing restrictions in emissions from a variety of industrial settings demand low cost removal of dilute contaminants in air. Many of these contaminants such as volatile organic components (VOCs) and sulfur compounds are biodegradable and can be removed from air streams via biofiltration. The simplest form of biofiltration consists of compost-based systems. More advanced systems designed for unique contaminants are biofilters with bioactive structured packing operating in trickle-bed mode. These advanced systems rely on a microbial consortium capable of degrading the contaminants of concern and the consortium usually is isolated or enriched from a more complex microbial mixture.

This paper describes the use of a trickle-bed reactor seeded with a microbial consortium enriched from a methanotrophic culture. The microbial consortium has been found to degrade chlorinated alkanes as the sole carbon source. Degradation rates of alkane mixtures are presented for the trickle-bed as well as results from batch cultures experiments designed to study degradation of various chlorinated and non-chlorinated VOCs.

INTRODUCTION

Increasing restrictions in emissions from a variety of industrial settings demand low cost removal of dilute contaminants in air. Many of these contaminants, such as volatile organic components (VOCs) and sulfur compounds, are biodegradable and can be removed from air streams via biofiltration.¹ The simplest form of biofiltration consists of compost- or soil-based systems. More advanced designs for unique contaminants are biofilters with a bioactive, structured packing operating in a trickle-bed mode.² These advanced systems rely on microbial consortia capable of degrading the contaminants of concern and the cultures are usually isolated or enriched from a more complex microbial mixture.

METHODS

The mixed culture used in the study has been described elsewhere;^{3,4} it can be characterized as an aerobic alkanotrophic culture enriched from a methanotrophic consortium; the improved culture has lost its ability to consume methane. The culture was maintained on salts medium (Medium 1, Table 1). The gas phase typically consisted of different alkanes or chlorinated alkanes in air (details in figure captions). Glass vials (various volumes) used for the batch studies were sealed with butyl-rubber stoppers and aluminum crimp-seals. Sterile controls were prepared in an identical fashion before sterilization at 121°C for 30 min. Blank controls had viable microorganisms but no gaseous carbon source. The vials were shaken in a rotary shaker

incubator at about 125 rpm at 30°C. The trickle-bed reactor was 50 cm long and had an internal diameter of 5 cm. It contained a structured polyethylene packing (Koch Industries) to maximize surface area, minimize pressure drop, and enhance mass transfer. Gas and liquid were fed to the top of the reactor. A schematic of the system may be seen in Figure 1.

Gas samples (0.1 to 0.3 mL) were injected directly into a Hewlett-Packard gas chromatograph (Model 5890) equipped with a flame ionization detector and an Alltech AT-624 capillary column (60m × 0.53mm ID). For gas phase oxygen, a thermocouple detector and a Hewlett-Packard HP-PLOT (Molecular Sieve 5A) capillary column (30m × 0.53mm ID) were utilized. Cell density was measured at 600 nm on a Milton Roy Spectronic 21D spectrometer and compared to a previously created calibration curve. Chloride concentration was measured using a Orion chloride-sensitive electrode.

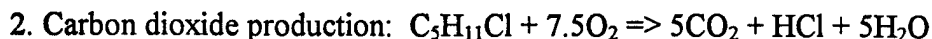
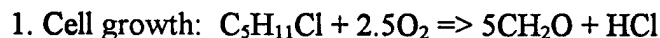
RESULTS

The consumption of straight and branched alkanes simultaneously is not new. Babu and Brown reported that a strain of *Nocardia paraffinicum* consumed propane and isobutane in a gas mixture.⁵ In their studies, they noted that either substrate was consumed by itself, regardless of prior substrate preference; e.g., propane-grown cells readily consumed isobutane. However, when the gases were added in mixtures, significant inhibition was noted. Isobutane and propane was also degraded in a soil bioreactor by Kampbell et al.⁶ Togna and Singh noted that isopentane was consumed by a mixed culture as a sole carbon source in a trickle-bed study.⁷ Numerous other publications have reported microbial n-alkane consumption as well.^{8,9,10,11}

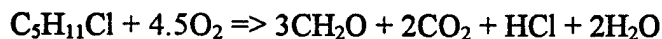
In Figure 2, the simultaneous consumption of C₂–C₅ alkanes by the culture is displayed. As is noted, methane is not consumed even after prolonged exposure once all other gases were depleted. Pentane and butane were consumed at almost identical rates; propane and ethane consumption was slower. The lack of methane consumption is not new. Takahashi et al. isolated and identified *Pseudomonas butanovora* as a bacterium capable of utilizing C₂–C₉ n-alkanes as sole carbon source (the medium contained 0.1 g/L yeast extract).¹² The organism would not grow on methane. Patel et al. isolated 16 new strains of bacteria capable of growth on C₂–C₄ alkanes but none could consume methane.¹¹ Perry, on the other hand, reported that the bacterial strain JOB5 was capable of growth on C₁–C₈ alkanes.⁸ Interestingly enough, the capability of consuming C₁–C₃ alkanes depended on the growth history of the culture; e.g., cells grown on short chain alkanes could consume longer chain alkanes, but the reverse was not true. Brisbane and Ladd also noted simultaneous conversion of methane and other alkanes.¹³ They speculated that enriched microbial cultures from soil contained at least two types of organisms; one that consumed methane and co-consumed ethane, another that consumed propane.

To study the release of chloride via the degradation of chlorinated organics and also to elucidate overall reaction sequence, a set of experiments was conducted by adding different amounts of 1-chloropentane as the sole carbon source to otherwise identical microcosms. The cell growth on chloropentane is shown in Figure 3. As is noted in the figure, cell growth was inhibited at chloropentane levels of 41 µmol. Also note the lack of growth in the absence of chloropentane. At the completion of the experiment, the microcosms were sampled for residual chloropentane (qualitative) and oxygen (quantitative); chloride and pH were also measured. The results are displayed in Figures 4, 5, and 6.

To interpret the results, we speculate that there are two predominant reactions for chloropentane conversion:



where CH_2O refers to the simplified molecular structure of cells. Based on the results presented in Figures 4 to 6, the following overall reaction can be derived.



indicating that 60% of available carbon was incorporated into cell mass.

The performance of the consortium may be compared with Janssen et al. who describes the organism *Xanthobacter autotrophicus* GJ10 as being an organism capable of growth on 1-chloropropane; however, this bacterium lacks capability of growth on pentane or hexane.¹⁴ The culture in our consortium grow on both these substrates. The proposed degradative pathway for *X. autotrophicus* goes through the alcohol, aldehyde, and carboxylic acid. Another organism, *Ancylobacter aquaticus*, has been described by van den Vijngaard.¹⁵ This organism grows on 1,2-dichlorethane (as does *X. autotrophicus*) but cannot support growth on longer-chain chloroalkanes. The current consortium may reveal other than these organisms when subjected to rigorous isolation protocol (ongoing).

The conversion of straight and branched alkanes in a trickle-bed reactor is shown in Figure 7. This experiment was conducted with the same culture previously described. It is clear that the conversion was more efficient at 30°C than at 15°C. Further information on the column performance has been published elsewhere.¹⁶ Future column experiments will test the degradation of chloropentane.

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Table 1. Media compositions used in experiments.

Component	Medium 1 (mg/L)	Medium 2 (mg/L)
$\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$		85
CaCl_2	200	
$\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$		0.2
$\text{CoCl}_2 \cdot \text{H}_2\text{O}$	0.01	
$\text{CuCl}_2 \cdot \text{H}_2\text{O}$		0.01
$\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$	0.05	
$\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$		1.5
FeCl_3	27	
H_3BO_4		0.3
K_2HPO_4	560	1500
KH_2PO_4	440	1200
KNO_3	1000	500
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	1000	500
$\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$		0.03
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	0.01	
$\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$		0.03
NaSeO_3		0.01
NH_4Cl	100	
$(\text{NH}_4)_2\text{SO}_4$		1250
$\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$		0.02
$\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$	0.07	
$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$		0.1

Figure 1. Schematic of trickle-bed reactor used for continuous flow studies. Experiments were conducted with Medium 1 (Table 1). Inlet gas feed consisted of 0.5% n-pentane and isobutane in air.

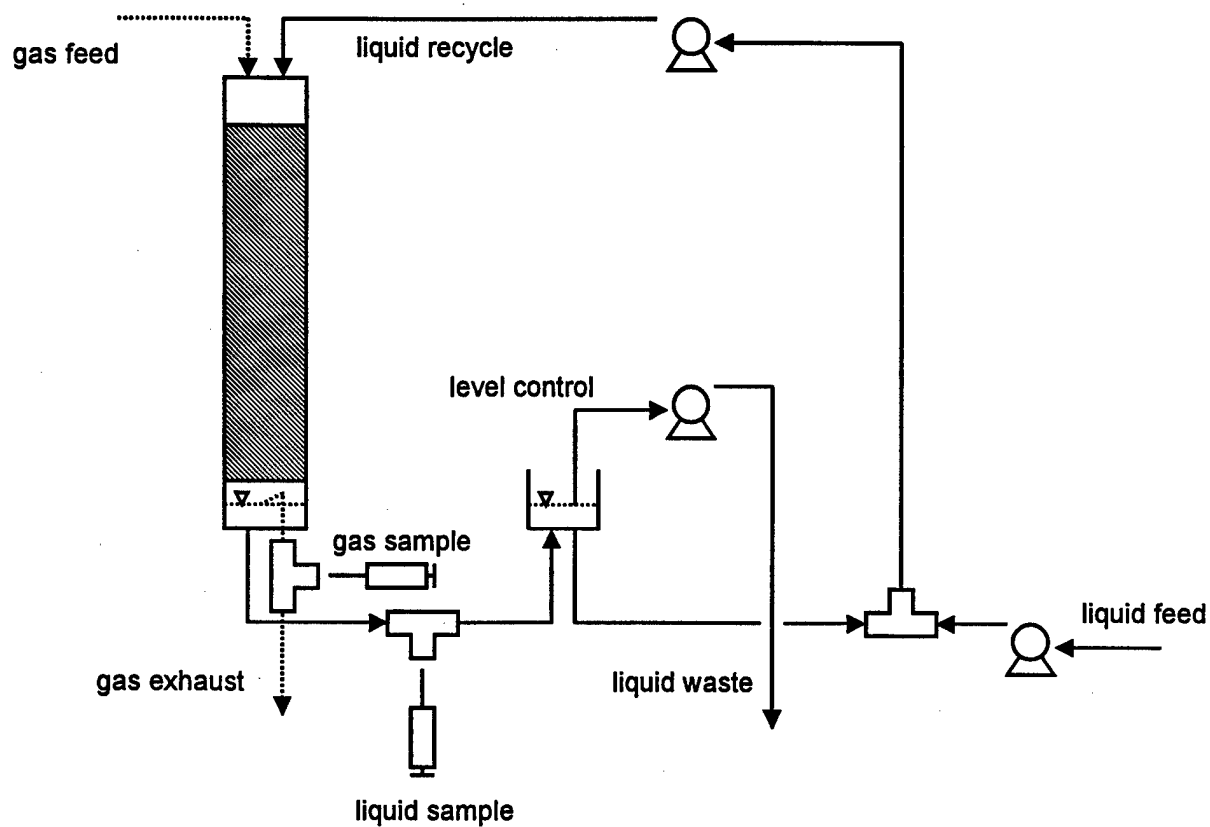


Figure 2. Consumption of C_1 – C_5 alkanes by the mixed culture. These studies were conducted in 157-mL serum bottles with a liquid volume of 50 mL (Medium 1, Table 1). The initial concentration of each gas was approximately 0.85% in air.

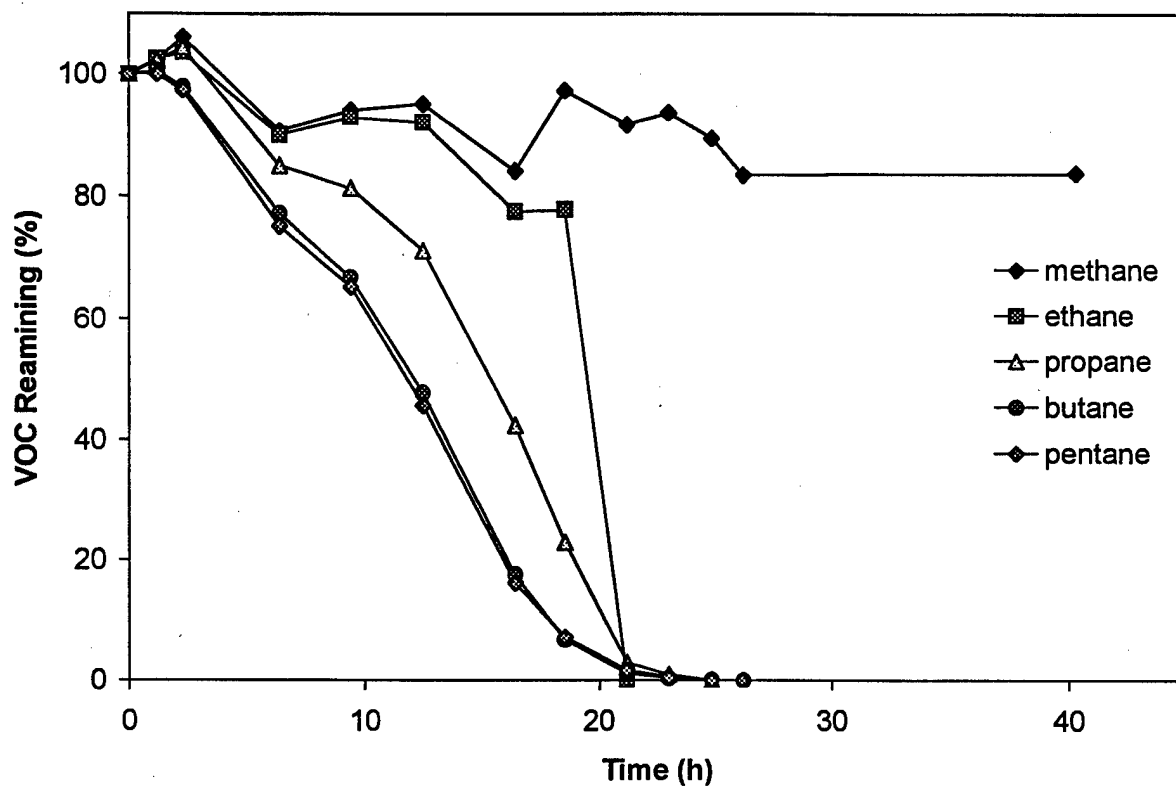


Figure 3. Growth of cell culture on various amounts of chloropentane. These studies were conducted in 26-mL serum test tubes with a liquid volume of 5.5 mL (Medium 2, Table 1). Various amounts of chloropentane were added to each microcosm.

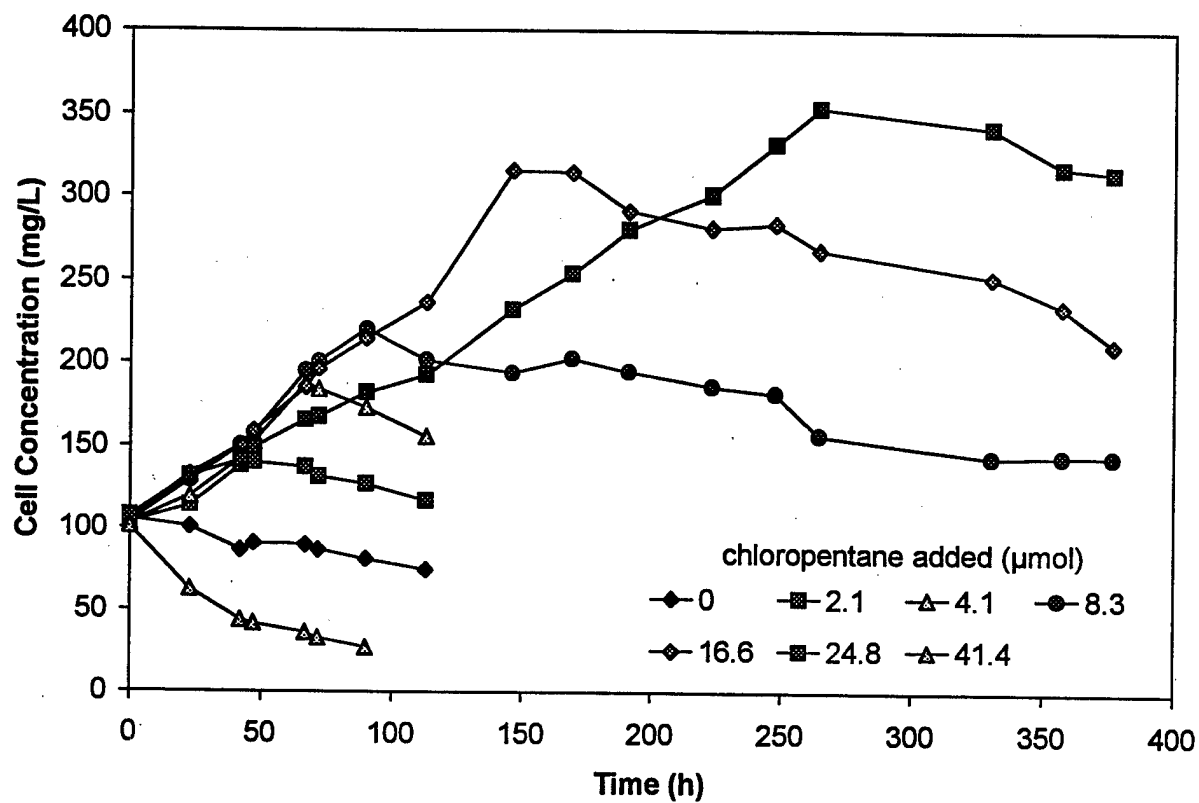


Figure 4. Cell mass produced in experiment set described in Figure 3 caption.

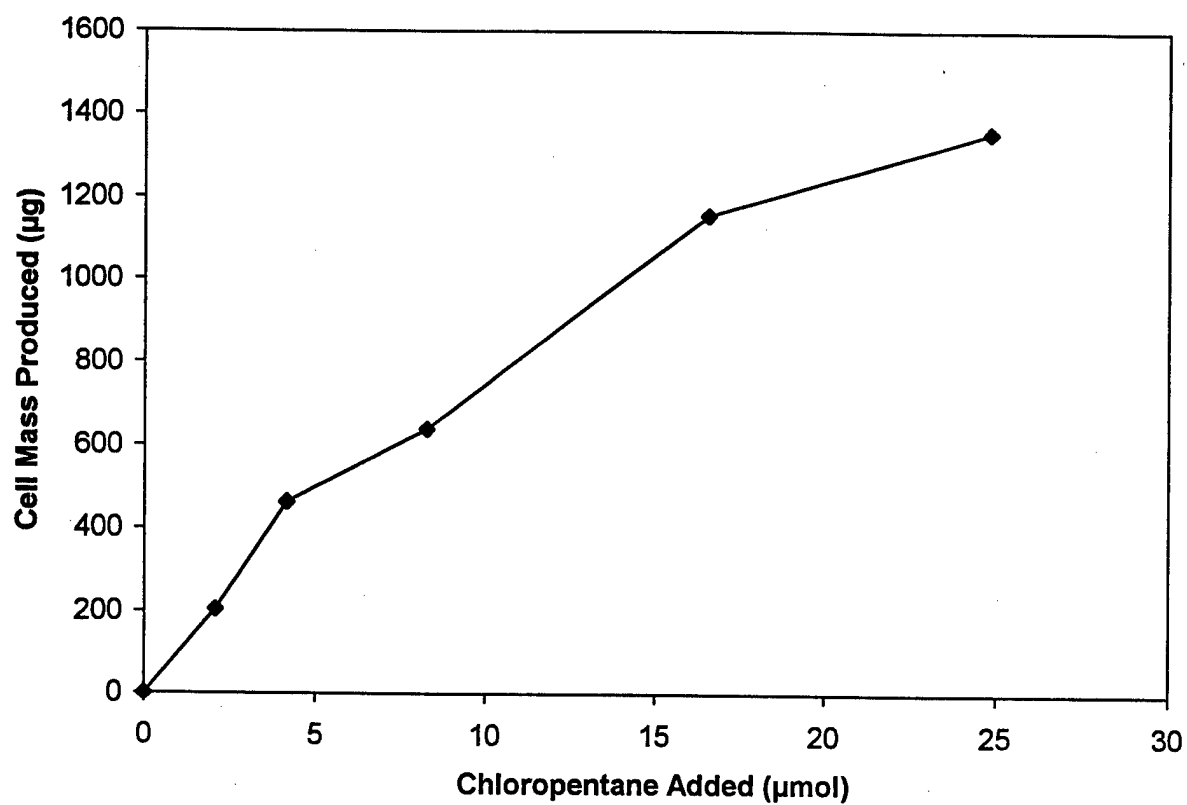


Figure 5. Chloride release in experiment set described in Figure 3 caption.

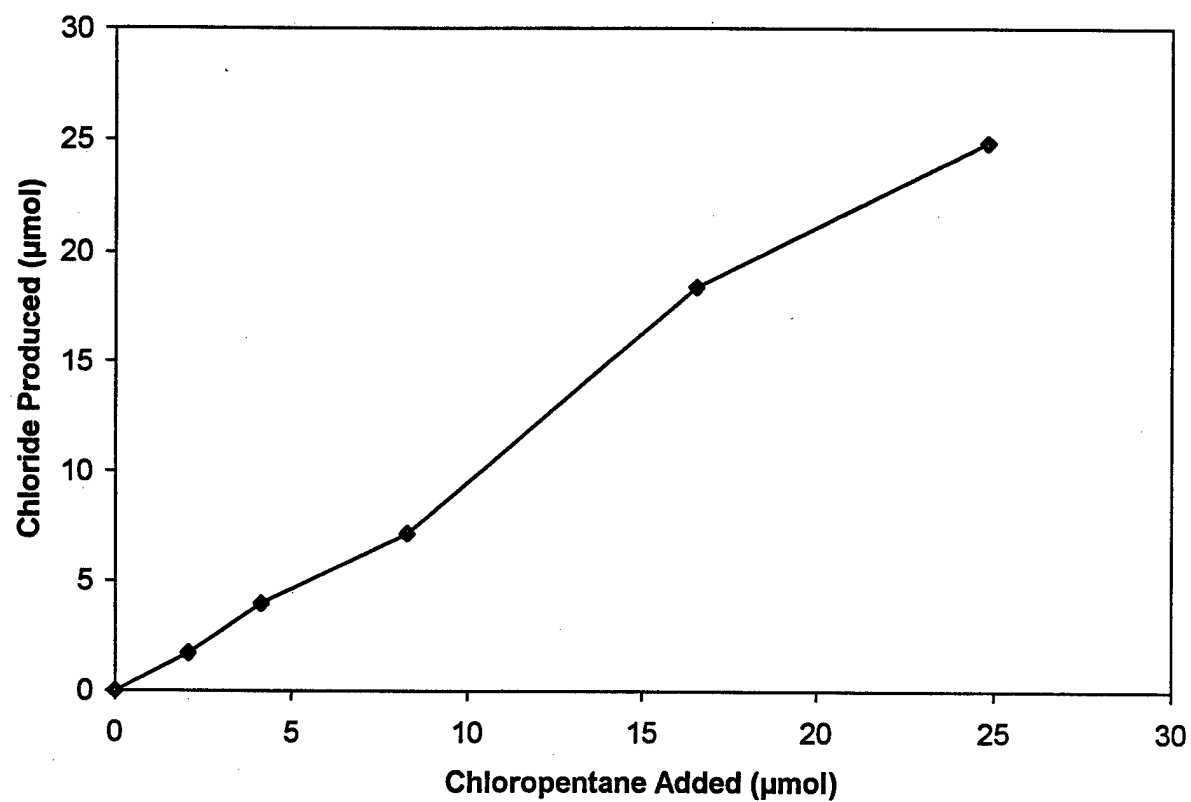


Figure 6. Residual oxygen in experiment set described in Figure 3 caption.

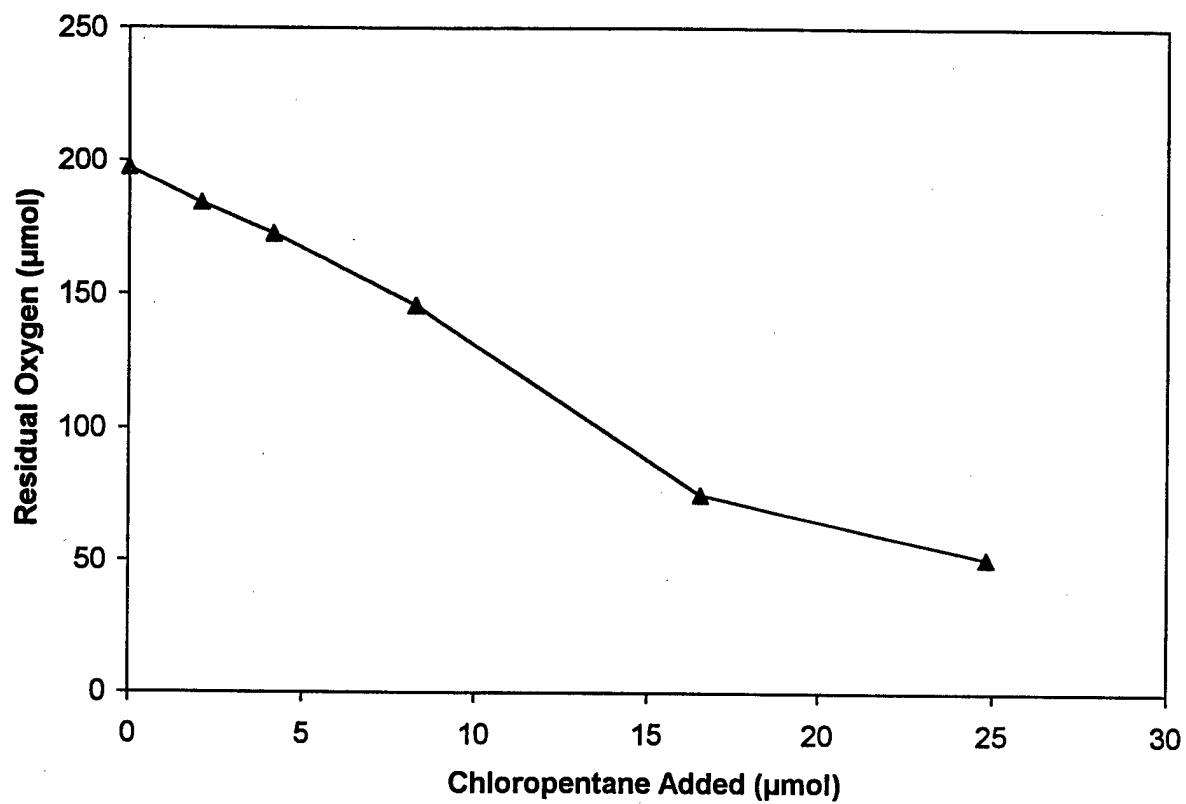
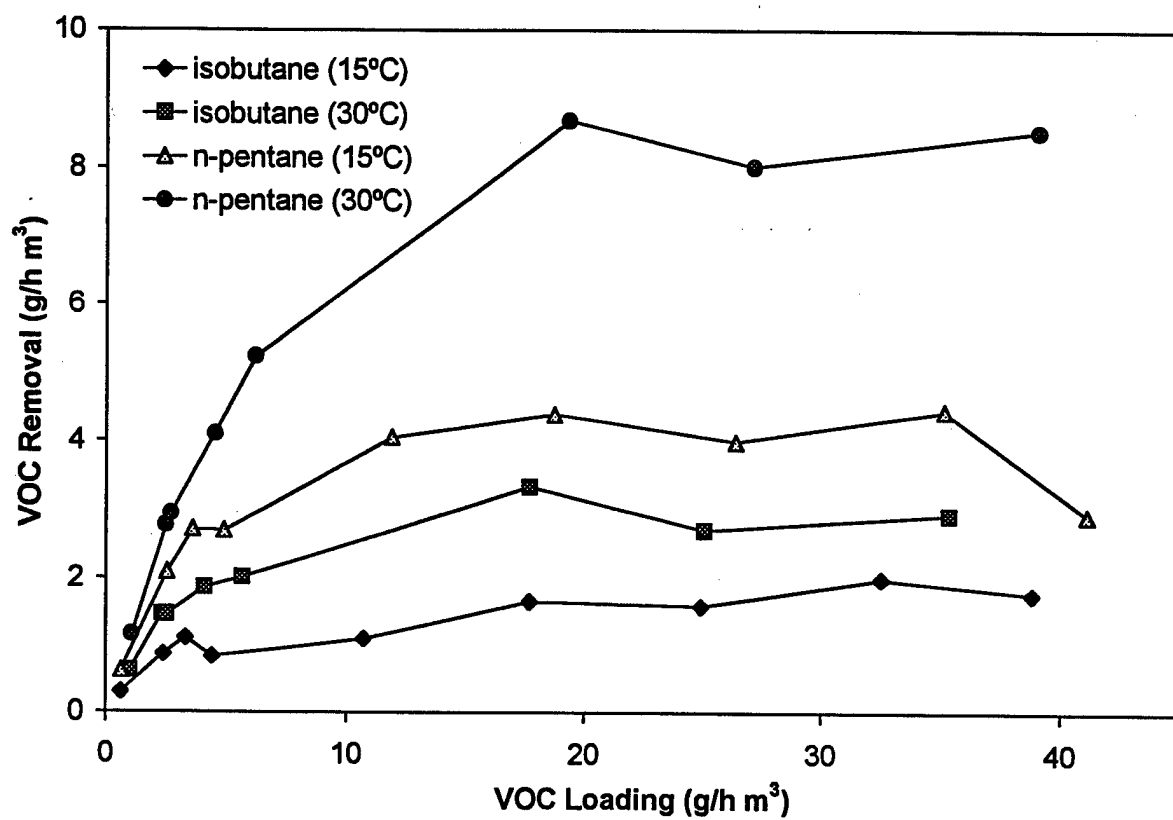


Figure 7. VOC removal efficiency in a continuous flow trickle bed reactor for various temperatures.



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