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Quality-Assurance Plan and Field Methods for Quality-of-Water Activities, U.S. Geological Survey, Idaho National Engineering Laboratory, Idaho

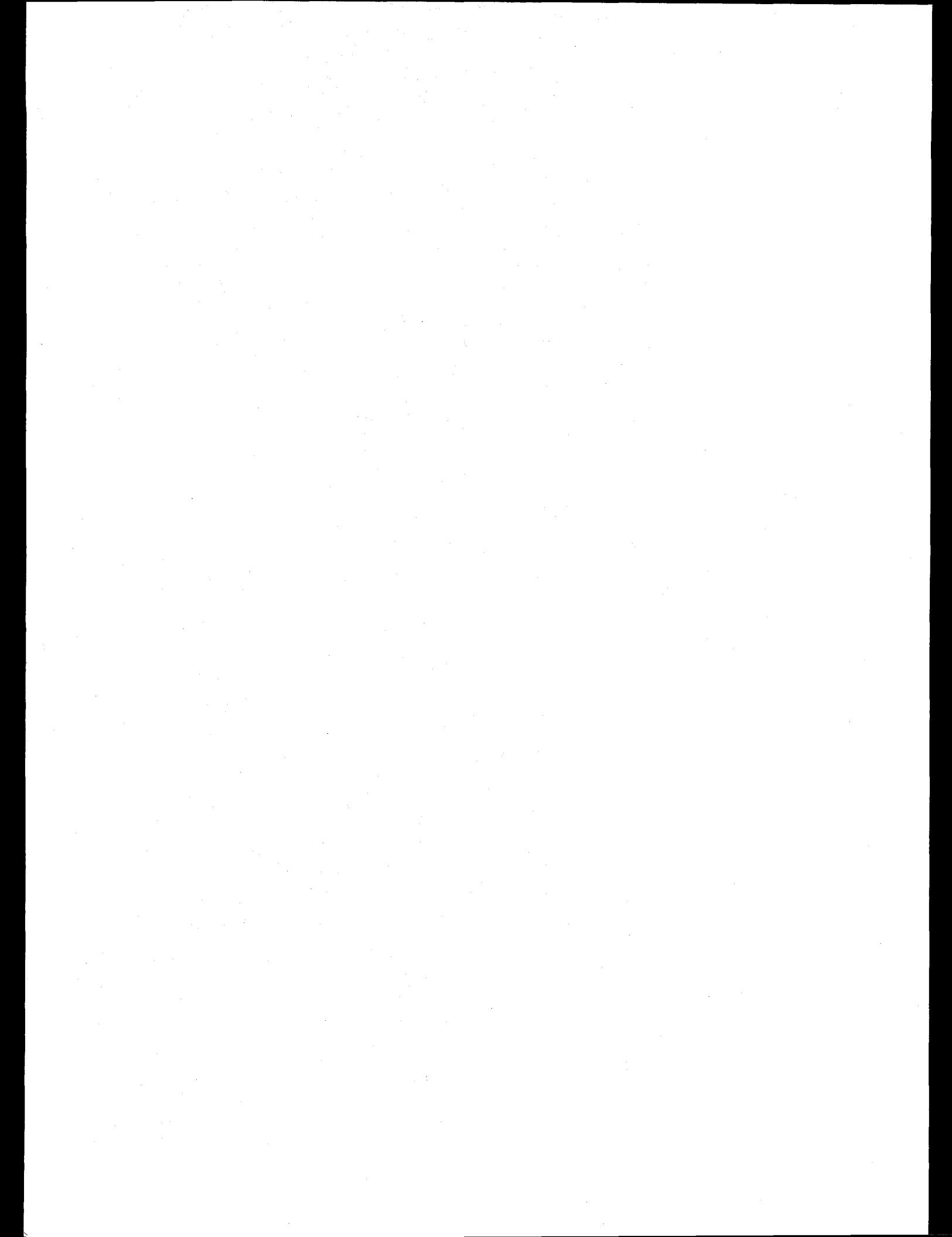
**U.S. GEOLOGICAL SURVEY
Open-File Report 96-615**

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Quality-Assurance Plan and Field Methods for Quality-of-Water Activities, U.S. Geological Survey, Idaho National Engineering Laboratory, Idaho

By Larry J. Mann

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October 1996



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by Larry J. Mann

INTRODUCTION

Water-quality activities at the Idaho National Engineering Laboratory (INEL) Project Office are part of the U.S. Geological Survey's (USGS) Water Resources Division (WRD) mission of appraising the quantity and quality of the Nation's water resources. The activities are conducted in cooperation with the U.S. Department of Energy's (DOE) Idaho Operations Office and the U.S. Environmental Protection Agency, Region 10. Results of the water-quality investigations are presented in various USGS publications or in refereed scientific journals. The results of the studies are highly regarded and are used with confidence by researchers, regulatory and managerial agencies, and interested civic groups.

In its broadest sense, quality assurance refers to doing the job right, the first time. It includes the functions of planning for products, review and acceptance of the products, and an audit designed to evaluate the system that produces the product. Quality assurance and quality control differ in that: quality control ensures that things are done correctly given the "state-of-the-art" technology; and quality assurance ensures that quality control is maintained within specified limits.

Purpose of and Responsibility for Maintaining the Quality-Assurance Plan

The purpose of the Quality Assurance Plan (QAP) for water-quality activities performed by

the INEL Project Office is to maintain and improve the quality of technical products, and to provide a formal standardization, documentation, and review of the activities that lead to these products. The principles of this plan are as follows:

1. Water-quality programs will be planned in a competent manner and activities will be monitored for compliance with stated objectives and approaches. The objectives and approaches are defined in an annual project work plan.
2. Field, laboratory, and office activities will be performed in a conscientious and professional manner in accordance with specified WRD practices and procedures by qualified and experienced employees who are well trained and supervised. If, or when, WRD practices and procedures are inadequate, data will be collected in a manner that its quality will be documented.
3. All water-quality activities will be reviewed for completeness, reliability, credibility, and conformance to specified standards and guidelines.
4. A record of actions will be kept to document the activity and the assigned responsibility.
5. Remedial action will be taken to correct activities that are deficient.

The overall responsibility for maintaining this QAP belongs to the Chief of the INEL Project Office and the Quality-Assurance Officer. The principal investigators for geochemistry and radiochemistry and the lead personnel for the water-quality monitoring network, however, are directly responsible for the day-to-day maintenance of the

QAP. The QAP will be formally revised and reprinted every 2 to 4 years; changes that take place in the interim, however, will be communicated by memoranda to project office personnel on an as-needed basis.

Scope

The QAP for water-quality activities at the INEL defines procedures and tasks performed by project-office personnel that ensure the reliability of water-quality data. Virtually all of the principles of the plan have been in effect during past and current operations, but the QAP provides a method to formalize and communicate the plan to all employees of the project office and to users of the hydrologic data and interpretive reports. A comprehensive list of references that contains guidelines used in data collection is given in the section entitled "Selected References." Tasks not described by the references owing to field conditions are detailed in the following sections.

Information on water-quality sampling schedules, data quality objectives, and water-quality field equipment are included in attachments 1 to 4.

Monitoring Networks

The water-quality monitoring network for the INEL consists of about 180 sites and include production wells, wells dedicated to water-quality monitoring, and surface-water sites. The network originally was established to document the distribution and concentration of radionuclides and industrial chemicals contained in wastewater discharged at the INEL either to the Snake River Plain aquifer or to the overlying perched ground-water zones. Disposal has taken place through deep disposal wells and shallow infiltration ponds. Recently, wells have been drilled to ascertain whether leachates from landfills have migrated to the aquifer. Additional monitoring sites will be selected if and when they are needed to better document the migration of solutes.

The frequency of sampling wells in the INEL network varies depending on the distance a particular well is from a disposal site, the historic

concentration of the radioactive or chemical waste in the ground water, and the location of the well relative to other wells. In general, water samples routinely are collected at 3- to 6-month intervals. The wells and streams at which water samples are collected, the method and frequency of sample collection, and the constituents routinely analyzed for are shown on attachment 1. In addition to the routine sampling, some wells periodically may be sampled for other constituents, including, chlorine-36, iodine-129, trace metals, and purgeable organic compounds. The frequency of sampling ranges from monthly to biannually depending on whether the constituent(s) of interest has been detected during previous sampling campaigns.

In addition to the INEL water-quality monitoring network, 50 wells and 19 springs between the southern boundary of the INEL and the Hagerman area are sampled at 1- to 3-year intervals. This off-site water-quality network was established to monitor natural contaminants and manmade pollutants in the aquifer that potentially could migrate from the INEL to hydraulically downgradient populated and agricultural areas.

Data-Quality Objectives

Data-quality objectives are qualitative and quantitative criteria that describe the data needed by managers or regulators to support environmental decisions and actions or by scientists to study natural or induced chemical processes in the Snake River Plain aquifer. The first steps of the scientific method are somewhat analogous to and are supported by data-quality objectives. Identifying problems is followed by hypothesizing solutions. Unbiased and thorough scientific experiments are proposed and then conducted, analyzed, and reported in the literature for peer review and use by others.

Data-quality objectives for the water-sample analyses by the USGS's National Water Quality Laboratory (NWQL) are included in attachment 2; objectives for radionuclides in water sample analyzed by and for the U.S. Department of Energy's Radiological and Environmental Sciences Laboratory (RESL) are in attachment 3.

METHODS

Sample containers, sample-preservation methods, field equipment, and well-head decontamination and sample-collection procedures are integral and crucial steps in assuring that data-quality objectives are achieved at the field level. Equally important are the analytical methods, and quality-control and quality-assurance activities exercised by the laboratories that analyze the samples.

Sample Containers and Preservation Methods

Sample containers and preservation methods

differ depending on the chemistry of the constituents being analyzed. Samples analyzed by the NWQL are containerized and preserved in accordance with laboratory requirements specified by Timme (1995). Containers and chemical preservatives are supplied by the NWQL, where they undergo a rigorous quality control to ensure that they are free of contamination (Pritt, 1989, p. 75). Samples analyzed by the RESL are containerized and preserved in accordance with requirements specified by the laboratory's Analytical Chemistry Measurements Team; changes in procedures are documented in writing. Containers and preservatives for selected constituents are summarized on table 1.

Table 1. Containers and preservatives used for water samples, Idaho National Engineering Laboratory and vicinity

[Abbreviations: L, liter; mL, milliliter; HNO₃, nitric acid; H₂SO₄, sulfuric acid; K₂Cr₂O₇, potassium dichromate; °C, degrees Celsius. Analyzing laboratory: NWQL—U.S. Geological Survey's National Water Quality Laboratory; RESL—U.S. Department of Energy's Radiological and Environmental Sciences Laboratory]

Type of constituent	Container		Preservative		Other treatment	Analyzing laboratory
	Type	Volume	Type	Volume		
Anions, dissolved	Polyethylene	250 mL	None	None	Filter	NWQL
Cations, dissolved	Polyethylene, acid-rinsed	500 mL	HNO ₃	2 mL	Filter	NWQL
Cations, total	Polyethylene, acid-rinsed	500 mL	HNO ₃	2 mL	None	NWQL
Metals, dissolved	Polyethylene, acid-rinsed	250 mL	HNO ₃	1 mL	Filter	NWQL
Metals, total	Polyethylene, acid-rinsed	250 mL	HNO ₃	1 mL	None	NWQL
Mercury, dissolved	Glass, acid-rinsed	250 mL	K ₂ Cr ₂ O ₇ /HNO ₃	10 mL	Filter	NWQL
Mercury, total	Glass, acid-rinsed	250 mL	K ₂ Cr ₂ O ₇ /HNO ₃	10 mL	None	NWQL
Chromium, total	Polyethylene, acid-rinsed	250 mL	HNO ₃	1 mL	None	NWQL
Chromium, dissolved and hexavalent	Polyethylene, acid-rinsed	500 mL	HNO ₃	2 mL	Filter	NWQL
Nutrients, dissolved	Polyethylene, brown	125 mL	None	None	Filter, chill 4°C	NWQL
	Polyethylene, brown	125 mL	H ₂ SO ₄	1 mL	Filter, chill 4°C	NWQL
Nutrients, total	Polyethylene, brown	125 mL	H ₂ SO ₄	1 mL	Chill 4°C	NWQL
Purgeable organic compounds	Glass, baked	40 mL	None	None	Chill 4°C	NWQL
Semivolatile organic compounds	Glass, baked	1 L	None	None	Chill 4°C	NWQL

Table 1. Containers and preservatives used for water samples, Idaho National Engineering Laboratory and vicinity—Continued

Type of constituent	Container		Preservative		Other treatment	Analyzing laboratory
	Type	Volume	Type	Volume		
Dissolved organic carbon	Glass, baked	125 mL	None	None	Silver filter, chill 4°C	NWQL
Total organic carbon	Glass, baked	125 mL	None	None	Chill 4°C	NWQL
Gross alpha and beta, dissolved	Polyethylene, acid-rinsed	1 L	HNO ₃	4 mL	Filter	NWQL
	Polyethylene, acid-rinsed	500 mL	HNO ₃	2 mL	None	RESL
Pesticides	Glass, baked	1 L	None	None	Chill 4°C	NWQL
Tritium	Polyethylene	125 mL	None	None	None	NWQL
	Polyethylene	1L	None	None	None	NWQL
	Polyethylene	125 mL	None	None	None	RESL
Strontium-90	Polyethylene, acid-rinsed	500 mL	None	None	None	RESL
	Polyethylene, acid-rinsed	1 L	HNO ₃	4 mL	Filter	NWQL
	Polyethylene, acid-rinsed	500 mL	HNO ₃	2 mL	None	RESL
Gamma spectroscopy	Polyethylene, acid-rinsed	7 L	HNO ₃	4 mL	Filter	NWQL
	Polyethylene, acid-rinsed	500 mL	HNO ₃	2 mL	None	RESL
Transuranics	Polyethylene, acid-rinsed	1 L	HNO ₃	4 mL	None	RESL

Field Equipment

Analytical and other associated equipment used in the field include: pH and specific conductance meters; thermometers; titrators for dissolved oxygen and alkalinity; a peristaltic pump; an in-line disposable filter capsule with a 0.45-micron filter; and associated glassware. The analytical equipment is housed and usually operated in mobile field laboratories. The purpose of the mobile laboratories is threefold: (1) they provide a relatively clean area to measure field parameters while minimizing the potential for contamination or degradation of the samples from the wind, dust, rain, snow and sunlight; (2) they are used as storage for sample and shipping con-

ainers, chemical reagents and preservatives, analytical instrumentation, and deionized water used for decontaminating equipment in the field; and (3) they provide a place where samples can be containerized, preserved and placed in shipping containers within minutes after withdrawal from a well or stream.

Instruments used to measure field water-quality parameters, such as pH and specific conductance, are maintained and calibrated in the field or in the laboratory in accordance with procedures specified by the instrument manufacturer. Instrument calibration is checked and, if necessary, instruments are recalibrated at each sampling site; calibration methods are recorded in a

field logbook (see figure 4 in section on sample collection.) A logbook that documents changes to equipment—for example, modifications to pH and conductivity meters—is kept in the office. An inventory of field equipment is given in attachment 4.

Decontamination Procedures at the Well Head

Wells that are equipped with dedicated submersible or line-shaft turbine pumps do not require decontamination except for the equipment that is attached to the discharge pipe to accommodate the collection of a water sample. Additionally, at least three wellbore volumes of water are pumped from the well to remove stagnant water and to rinse and equilibrate the pump and delivery line. Production wells generally have a spigot at or near the well head; decontamination consists of thoroughly rinsing the spigot with pumped ground water to remove foreign materials.

Sample collection is facilitated and excess water is diverted away from the well head by fitting wells equipped with dedicated pumps with a portable discharge pipe about 2 ft long. The discharge pipe has a 1.5-in. I.D. (inside diameter) and is equipped with a gate valve to control the flow rate. A T-joint is inserted into the pipe between the well head and the control valve and a series of nipples, a valve to control the flow rate of the sampling port, and connectors are attached to the T-joint to reduce the diameter so that a 1/4-in. I.D. delivery line can be attached as a sampling point. The line is made up with two 1/4-in. I.D. nipples connected with a 90-degree elbow to facilitate sample collection. All fittings and pipes are stainless steel and are rinsed with deionized water before installation at the well head. Subsequent flushing with several hundred to thousands of gallons of purged well water further reduces the possibility of cross-contamination with water from previously sampled wells.

At wells that are not equipped with dedicated pumps, one of two methods is used to collect water samples depending on the amount of water in the well and depth to water. A generator-powered portable pump is used to collect samples

from wells at which the depth to water is less than 200 ft. The portable pump and attendant hose routinely are decontaminated by flushing many cycles of water and detergent through them and then rinsing the equipment with about 30 gallons of tap water and about 25 gallons of deionized water; 1.5 gallons of water is enough to flush the pump and hose one time. Before the pump is installed in a well, it is thoroughly flushed by pumping deionized water through pump and discharge hose. Samples of the deionized-water rinse periodically are collected and analyzed to document whether the portable pumps are contaminated by constituents of interest.

For wells without dedicated pumps and at which the depth to water exceeds 200 ft, and for wells with only a few feet of water in the wellbore, a bailer is used for collecting water samples. The bailer and that part of the bailer line that enters the well are washed with hot water and detergent and rinsed with deionized water prior to use; samples of the rinse periodically are collected and analyzed to document whether the equipment is contaminated by constituents of interest. At some wells, bailers are dedicated to and stored in the well casing. This eliminates the possibility of cross-contamination of samples from different wells.

Sample Collection

At wells equipped with a dedicated pump or at which the portable pump is used to ensure that water representative of the Snake River Plain aquifer or perched ground-water zone is sampled, a volume of water equivalent to a minimum of 3 wellbore volumes is pumped prior to collecting the samples; at many wells, 5 to 10 wellbore volumes are pumped. The diameter of the wellbore, rather than the volume of the casing is used to calculate the minimum volume because of the potentially large difference between the two. In addition, temperature, specific conductance, and pH are monitored periodically during pumping using methods described by Wood (1981) and Hardy and others (1989). Field measurements made immediately prior to sample collection are used to represent those for the sample. When

these measurements stabilize, indicating probable hydraulic and chemical stability, a water sample is collected using the following steps:

1. The field person responsible for collecting the water sample wears disposable vinyl gloves and stands in a position where neither the collector nor the sample can become contaminated.
2. The outside of the sample delivery line is thoroughly rinsed with water pumped from the well.
3. If appropriate, sample containers and filtration equipment are thoroughly rinsed with water pumped from the well or surface-water site before being used. A new, disposable capsule filter with a 0.45-micron membrane filter is used at each site. One liter of deionized water or water from the well is used to rinse the capsule filter prior to sample collection to remove any surfactants that are adhered to the filters. The capsule filter is inverted during rinsing to remove any trapped air bubbles.
4. For ground-water samples from wells equipped with dedicated pumps, the capsule filter is connected to the sample port with pre-cleaned Tygon tubing; unfiltered samples are collected directly from the sample port. For surface-water samples and bailer samples, a grab sample is collected in a pre-cleaned container and the inlet tubing of a peristaltic pump is placed into the container to supply sample water to the capsule filter. Unfiltered samples are collected by submersing the sample container into the surface water-body or drawing water from a pre-cleaned container.
5. Samples are capped loosely and moved into the mobile field laboratory where they are uncapped and preserved as described in Table 1.
6. The bottles are capped and the caps are sealed with laboratory film. The bottles then are labeled (see fig. 1 for example of label). An alternate method for labeling sample containers is to record information directly on the sample container using a permanent marker. Recording the information both on a label and directly on the bottle is the preferable option.

LOCATION=	1 OF 1
STA NAME=	SPEC COND=
DATE=	TIME=
SAMP SIZE=	pH=
DISCHARGE	TREATMENT=
W TEMP=	A TEMP=
SAMPLE TYPE=	SCHEDULE=

Figure 1.—Example of label attached to each sample bottle.

7. Field measurements are made again after samples are collected. If the temperature differs by more than 0.5 °C, the pH differs by more than 0.1 units, or the conductance differs by more than 5 percent, the measurements are verified and a second set of samples are collected.
8. A laboratory request schedule is completed for use by each laboratory to which the sample(s) will be sent for analysis (see figs. 2 and 3 for examples).
9. The water samples are chilled if necessary, and stored in the field laboratory until they can be transferred to a secured storage area. Samples are transported to the analyzing laboratory as soon as reasonably possible. Samples sent to the NWQL for analysis are transported in a sealed ice chest by a contract carrier; overnight delivery is stipulated for water samples for analyses of nutrients and purgeable organic compounds. Samples sent to the RESL for analysis are hand carried to the laboratory.
10. All equipment is decontaminated with deionized water and, if necessary, organic-free water.

Some wells completed in the perched-water zones do not contain or produce enough water to be sampled with a portable pump. For these wells, either a 1,000-mL Teflon bailer, a 1,000-mL brass thief sampler rigged as a bailer, or a 200-mL galvanized bailer normally is used for sample collection. The well is bailed until enough water is collected for all the samples required or until the well is bailed dry.



ID F-54841A
(Rev. 02-90)
Ref. ID 5484.1 RESI

Routine _____ Urgent _____

Non-Routine _____ Date Needed _____

Collected By _____ Date Sent _____

Organization _____

UNITED STATES DEPARTMENT OF ENERGY
IDAHO OPERATIONS OFFICE
RADIOLoGICAL AND ENVIRONMENTAL SCIENCES LABORATORY
SAMPLE RECORD SHEET

Onsite _____
Offsite _____

Serial No. _____

Samples Received: _____

Analysis Completed: _____

Notified: _____ Date: _____

Analyzed By: _____

Approved By: _____

Anal. for _____

Inst. used _____

Quant. used _____

Date _____

cntd. _____

Count time _____

Gross count _____

Bkgd. _____

Net count _____

Result \pm 1S.O (attached) _____

Sample _____

Sample Description _____

No. _____ Date _____ Hour _____

Figure 3.—Sample record sheet for the Radiological and Environmental Sciences Laboratory.

When the bailer is retrieved, its contents either are placed directly in bottles for raw samples or in a pre-cleaned container as described in step 4. Field measurements are made on excess water from the bailer or in the pre-cleaned container. After the sample bottle is filled with either raw or filtered water, samples are preserved appropriately and labeled, stored, and shipped as described in steps 6-9.

Some perched-water wells contain enough water to use a portable pump, but have such low recovery rates that the wells pump dry before 2 or 3 wellbore volumes can be pumped. For these wells, a sample is collected when temperature, pH, and specific conductance measurements stabilize. If the well pumps dry before measurements stabilize, the field person waits for the well to recover to collect a pumped or bailed sample, or, if the well does not recover, a grab sample is collected from the pre-rinsed bucket into which water is discharged while pumping. Exceptions such as these to usual sample-collection procedures are described in the field logbook.

An added precaution is used at wells that are sampled with a portable submersible pump or bailer. The concentrations of most contaminants are greatest in wells nearest disposal sites and the concentrations decrease with increasing distance from the disposal sites. Therefore, when conditions permit, the most distant wells are sampled first. Wells closest to sources of contamination generally have the greatest contaminant concentrations and are sampled last. This process of sampling minimizes the potential for measurable amounts of cross contamination between wells because of the portable equipment.

Conditions at the well during sample collection are recorded in a bound field logbook (fig. 4) and a chain-of-custody record (fig. 5) is used to track samples from the time of collection until delivery to the RESL or until mailing to the NWQL. These records are available for inspection at the USGS Project Office at the INEL.

QUALITY ASSURANCE

The USGS's Quality-Assurance Program at the INEL Project Office incorporates the previously described methods with several other elements: (1) analytical methods used by the laboratories; (2) quality-control samples; (3) review of analytical results of chemical constituents provided by the laboratories; (4) audits of performance in the field and in the laboratory; (5) corrective actions to resolve problems with field and laboratory methods; and (6) reporting of data. These elements effectively are performed to assure the following: (1) reliability of the water-quality data; (2) compatibility of the data with data collected by other organizations at the INEL; and (3) the data meet the programmatic needs of the DOE and its contractors and the scientific and regulatory communities.

Analytical Methods and Quality-Control Samples

A detailed description of internal quality control and of the overall quality-assurance practices used by the NWQL is provided in reports by Friedman and Erdmann (1982) and Pritt and Raese (1992); quality-control practices at the laboratory are described by Jones (1987); and quality-assurance data for routine water analyses are presented in a report by Maloney and others (1993). Additional quality assurance instituted by the INEL Project Office includes collection and analysis of the following: (1) duplicate samples—two or more samples collected concurrently or sequentially and sent to different laboratories; (2) blind replicates—duplicate samples with different sample identification numbers submitted to a laboratory; (3) blank samples—samples of deionized water sent to a laboratory and identified as routine samples; (4) equipment blanks—rinse collected during decontamination procedures; (5) splits—large sample volumes divided into two or more equal volumes and sent to different laboratories for analysis; (6) trip blanks—laboratory supplied samples of boiled deionized water that travel with water samples from time of collection to time of analysis; and (7) spiked samples—samples to which a known concentration of a

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Figure 5.—Chain-of-custody record.

constituent is added. Analytical methods used by the NWQL are described by Barnett and Mallory (1981), Faires (1992), Fishman (1993), Fishman and Friedman (1989), Goerlitz and Brown (1972), Rose and Schroeder (1995), Skougstad and others (1979), Thatcher and others (1977), and Wershaw and others (1987). The type of analysis and analytical procedure are specified on the NWQL services request form (fig. 2).

A discussion of procedures used by the RESL for the analysis of radionuclides in water is provided in reports by Bodnar and Percival [eds.] (1982) and U.S. Department of Energy (1995). Additional quality assurance implemented by the INEL Project Office for samples sent to the RESL is consistent with procedures used for samples sent to the NWQL. The type of analysis to be performed on a water sample is specified on the RESL sample record sheet (fig. 3).

In general, about 10 percent of the samples collected are dedicated to quality assurance. That is, for every 10 samples submitted to one of the laboratories for analysis, at least one is a blind replicate, a blank, a split, or another type of quality-assurance sample. For samples which are to be analyzed for non-routine constituents, 15 to 20 percent of the samples are dedicated to quality assurance.

Review of Analyses

After the analytical results are obtained from the analyzing laboratory, the concentration of each constituent of interest is reviewed by personnel at the INEL Project Office for consistency, precision, and accuracy. Factors considered during the review are:

1. The historical concentration of the solute at the site where the sample was collected;
2. The concentration of the solute in replicate, split, blank, or other quality-assurance samples;
3. The concentration of the solute in nearby wells that obtain water from the same aquifer or perched-water zone;

4. A review of waste-disposal records and changes in disposal techniques, land use, and recharge that may influence the concentration of a solute(s);
5. Cation-anion balance of analyses for which common ions are analyzed; and
6. Other accepted tests for accuracy of analytical results, when appropriate (Hem, 1985, p. 163-165).

Constituents for which previous analyses have been made are reviewed for consistency with items 1, 2, and 3. If a constituent exceeds or is less than the historical trend, if it differs markedly from the concentration in water from nearby wells, or if an initial analysis for a solute exceeds 80 percent of the maximum contaminant level (MCL) for that constituent set by the Environmental Protection Agency, either a re-analysis by the laboratory is requested or a second sample is collected and analyzed to verify the concentration of the solute in the water. If resampling is necessary, replicates generally are collected to evaluate laboratory precision. Constituents for which MCLs have been proposed or established are shown on tables 2-7.

If analytical results indicate that concentrations in samples from one site vary by more than 50 percent for no obvious reason, the results are evaluated by replicate sampling. If the analytical results for the replicates do not agree, the analyzing laboratory is contacted to resolve the problem.

Performance Audits

Performance audits are conducted routinely at three levels: (1) at the field level, (2) at the laboratory level, and (3) through National Field Quality Assurance Tests. At the field level, the Project Chief or a designee routinely accompanies the field personnel to a selected number of sites to ascertain whether proper field techniques are used to collect and preserve the samples and to ensure safety procedures. Replicate and split samples are used to evaluate the precision of the field laboratory methods; spikes and reference samples are used to measure accuracy.

Table 2. Maximum contaminant levels for types of radioactivity and selected radionuclides in drinking water

[The maximum contaminant levels were established pursuant to the recommendations of the U.S. Environmental Protection Agency (1994a) for community water systems and are for comparison purposes only. The maximum contaminant level given for gross alpha-particle radioactivity includes radium-226 but excludes radon and uranium. The maximum contaminant level given for gross beta-particle and gamma radioactivity excludes radioactivity from natural sources and should be used only for comparison. Maximum contaminant level values listed for strontium-90 and tritium are average annual concentrations assumed to produce a total body or organ dose of 4 mrem/year (millirem per year) of beta-particle radiation. Abbreviations: pCi/L—picocuries per liter; $\mu\text{g}/\text{L}$ —microgram per liter; mrem—millirem]

Type of radioactivity or radionuclide	Maximum contaminant level
Gross alpha-particle radioactivity	15 pCi/L
Gross beta-particle and gamma radioactivity	4 mrem/year
Radium-226 plus radium-228	5 pCi/L
Radium-226	20 pCi/L
Radium-228	20 pCi/L
Radon-222	300 pCi/L
Strontium-90	8 pCi/L
Total uranium	20 $\mu\text{g}/\text{L}$
Tritium	20,000 pCi/L

The INEL Project Office participates in the National Field Quality Assurance Program established by the USGS to evaluate the accuracy of water-quality field measurements. Quality-assurance samples are sent to field personnel for testing. The results are sent back to the water quality service unit for evaluation. If field personnel or equipment do not pass the test, corrective action is taken. The program is described in detail by Erdmann and Thomas (1985).

In addition to the routine performance audits, water-quality activities at the INEL Project Office periodically are monitored and reviewed by other USGS personnel: the Water-Quality Specialists for the Idaho District, Boise, Idaho; personnel at the Office of the Regional Hydrologist, Western Region, Menlo Park, Calif.; and personnel at the Office of Water Quality at Headquarters, Reston, Va. Reviews by personnel at the Idaho District take place at 1- to 2-year intervals and by the Western Region Office and Office of Water Quality at 2- to 3-year intervals. The reviews are summarized in writing and distributed to the Project Office, Regional Office, and the Office of Water Quality. If deficiencies are documented, a written reply outlining corrective action is required of the Project Office.

Corrective Actions

If the performance audits indicate inconsistencies or inadequacies in field methods or in analytical results by the laboratories, the problems are documented and the field personnel or laboratories are notified in writing of the inconsistencies or inadequacies. Training is provided to the field personnel as needed and the frequency of performance audits is increased until the performance is judged by the INEL Project Office Chief as suitable and consistent with written guidelines.

Inconsistencies and inadequacies in laboratory analyses are discussed with or submitted in writing to the appropriate laboratory director, and who is responsible for initiating the appropriate action to resolve the problem. To evaluate whether appropriate actions are taken, the frequency and numbers of replicate, blank, split, other quality assurance samples are increased until it is demonstrated that problems in the laboratory methods are resolved.

If project-office personnel discover a problem with sampling procedures, equipment calibration, or data review analysis and interpretation that cannot be resolved at the project level, the Idaho District Water-Quality Specialist is notified of the problem. If he or she cannot resolve the problem in consultation with the Regional Water-Quality

Table 3. Maximum contaminant levels and minimum reporting levels of selected trace elements in drinking water

[The maximum contaminant levels are for total measurements and were established pursuant to the recommendations of the U.S. Environmental Protection Agency (1994a, p. 753; 1994b) for community water systems and are for comparison purposes only. Secondary maximum contaminant levels—in brackets—are from U.S. Environmental Protection Agency (1994a, p. 836). Minimum reporting levels for the U.S. Geological Survey's National Water Quality Laboratory are from Timme (1995); concentrations for constituents for which two minimum reporting levels are shown, can be determined by two analytical methods. Units are in micrograms per liter ($\mu\text{g/L}$). Symbols: -- , maximum contaminant level has not been established; $*$, lead has an action level of 15 $\mu\text{g/L}$]

Constituent	Maximum contaminant level	Minimum reporting level
Aluminum	[50 to 200]	1, 10
Arsenic	50	1
Barium	2,000	1
Beryllium	4	.5, 1
Cadmium	5	1
Chromium, dissolved	--	5
Chromium, hexavalent	--	1
Chromium, total	100	1
Cobalt	--	1, 3
Copper	[1,000]	1, 10
Iron	[300]	3
Lead	*	1
Lithium	--	4
Manganese	[50]	1
Mercury	2	.1
Molybdenum	--	10
Nickel	100	1, 10
Selenium	50	1
Silver	[100]	1
Strontium	--	.5
Vanadium	--	6
Zinc	[5,000]	1, 3

Table 4. Maximum contaminant levels and minimum reporting levels of selected nutrients, dissolved organic carbon, and anionic surfactants in drinking water

[The maximum contaminant levels were established pursuant to the recommendations of the U.S. Environmental Protection Agency (1994a, p. 753) for community water systems and are included for comparison purposes only. The secondary maximum contaminant level—in brackets—is from U.S. Environmental Protection Agency (1994a, p. 836). Minimum reporting levels are from Timme (1995). Units are milligrams per liter. Symbol: -- , maximum contaminant level has not been established or proposed]

Constituent	Maximum contaminant level	Minimum reporting level
Ammonia (as nitrogen)	--	0.015
Nitrite (as nitrogen)	1	.01
Nitrite plus nitrate (as nitrogen)	10	.05
Orthophosphate (as phosphorus)	--	.01
Dissolved organic carbon	--	.1
Anionic surfactants	[0.5]	.02

Table 5. Maximum contaminant levels and minimum reporting levels of selected insecticides and gross polychlorinated compounds in drinking water

[The maximum contaminant levels, in micrograms per liter, were established pursuant to the recommendations of the U.S. Environmental Protection Agency (1994b) for community water systems and are included only for comparison purposes. Proposed maximum contaminant levels—shown in parentheses—are from U.S. Environmental Protection Agency (1994b). Minimum reporting levels are from Timme (1995). Symbols: .., indicates that a maximum contaminant level has not been established or proposed for that compound. Abbreviation: µg/L, microgram per liter]

Carbamate insecticides: minimum reporting level is 0.5 µg/L

Insecticide	Maximum contaminant level	Insecticide	Maximum contaminant level
Aldicarb	7	3-Hydroxycarbofuran	..
Aldicarb sulfone	7	Methomyl	..
Aldicarb sulfoxide	7	1-Naphthol	..
Carbaryl (Sevin)	..	Oxamyl	200
Carbofuran	40	Propham	..

Organophosphorus insecticides: minimum reporting level is 0.01 µg/L

Insecticide	Maximum contaminant level	Insecticide	Maximum contaminant level
Chlorpyrifos; Dursban	..	Methyl parathion	..
Diazinon	..	Methyl trithion	..
Disulfoton	..	Parathion	..
Ethion	..	Phorate	..
Fonofos	..	Trithion	..
Malathion	..		

Organochlorine insecticides: minimum reporting level is 0.01 µg/L except for chlordane and perthane (0.1 µg/L), and toxaphene (1.0 µg/L)

Insecticide	Maximum contaminant level	Insecticide	Maximum contaminant level
Aldrin	..	Heptachlor	0.4
Chlordane	2	Heptachlor epoxide	.2
DDD	..	Lindane	.2
DDE	..	Methoxychlor	40
DDT	..	Mirex	..
Dieldrin	..	Perthane	..
Endosulfan	..	Toxaphene	3
Endrin	2		

Gross polychlorinated compounds, minimum reporting level is 0.1 µg/L

Compound	Maximum contaminant level
Gross polychlorinated biphenyls (PCB)	0.5
Gross polychlorinated naphthalenes (PCN)	..

Table 6. Maximum contaminant levels and minimum reporting levels of chlorophenoxy-acid herbicides and other herbicides in drinking water

[Analyses were performed by the U.S. Geological Survey's National Water-Quality Laboratory. The maximum contaminant levels were established pursuant to the recommendations of the U.S. Environmental Protection Agency (1994b) for community water systems and are included for comparison purposes only. Minimum reporting levels are from Timme (1995). Units are in micrograms per liter. Symbol: .., maximum contaminant level has not been established or proposed. Abbreviations: MCL, maximum contaminant level; MRL, minimum reporting level]

Chlorophenoxy-acid herbicides

Herbicide	MCL	MRL	Herbicide	MCL	MRL
2,4-D	70	0.01	Silvex	50	0.01
2,4- <i>DP</i>	..	.01	2,4,5-T	..	.01

Other herbicides

Herbicide	MCL	MRL	Herbicide	MCL	MRL
Alachlor	2	0.009	Napropamide	..	0.01
Atrazine	3	.017	Pebulate	..	.009
Atrazine, Desethyl-	..	.005	Pendimethalin	..	.018
Benfluralin	..	.013	Prometon	..	.008
Butylate	..	.008	Pronamide	..	.009
Cyanazine	..	.013	Propachlor	..	.015
DCPA (Dacthal)	..	.004	Propanil	..	.016
Diethylaniline	..	.006	Simazine	4	.008
EPTC (Eptam)	..	.005	Tebuthiuron	..	.015
Ethalfluralin	..	.013	Terbacil	..	.03
Linuron	..	.039	Thiobencarb	..	.008
Metolachlor	..	.009	Triallate	..	.008
Metribuzin	..	.012	Trifluralin	..	.012
Molinate	..	.007			

Table 7. Maximum contaminant levels and minimum reporting levels of selected purgeable organic compounds in drinking water

[Analyses were performed by the U.S. Geological Survey's National Water-Quality Laboratory using an analytical method equivalent to U.S. Environmental Protection Agency method 524.2. Maximum contaminant levels were established pursuant to the recommendation of the U.S. Environmental Protection Agency (1994b) for community water systems and are included for comparison purposes only. Minimum reporting levels are from Timme (1995). Units are in micrograms per liter ($\mu\text{g/L}$). Symbols: -- , maximum contaminant level has not been established or proposed; $*$, total trihalomethanes—which include bromoform, chlorodibromomethane, chloroform, and dichlorobromomethane—in community water systems serving 10,000 or more persons cannot exceed 100 $\mu\text{g/L}$ (U.S. Environmental Protection Agency, 1994b, p. 2). Abbreviations: MCL, maximum contaminant level; MRL, minimum reporting level]

Compound	MCL	MRL	Compound	MCL	MRL
Acrolein	--	20	1,2-Dichloropropane	5	0.2
Acrylonitrile	--	20	1,3-Dichloropropane	--	.2
Benzene	5	.2	2,2-Dichloropropane	--	.2
Bromobenzene	--	.2	Cis-1,3-Dichloropropene	--	.2
Bromochloromethane	--	.2	Trans-1,3-Dichloropropene	--	.2
Bromoform	*	.2	1,1-Dichloropropene	--	.2
Bromomethane	--	.2	Ethylbenzene	700	.2
n-Butylbenzene	--	.2	Hexachlorobutadiene	--	.2
Sec-butylbenzene	--	.2	Isopropylbenzene	--	.2
Tert-butylbenzene	--	.2	p-Isopropyltoluene	--	.2
Carbon tetrachloride	5	.2	Methylene chloride	5	.2
Chlorobenzene	100	.2	Methyltertbutylether	--	.2
Chlorodibromomethane	*	.2	Naphthalene	--	.2
Chloroethane	--	.2	n-Propylbenzene	--	.2
2-Chloroethylvinylether	--	1	Styrene	100	.2
Chloroform	*	.2	1,1,1,2-Tetrachloroethane	--	.2
Chloromethane	--	.2	1,1,2,2-Tetrachloroethane	--	.2
2-Chlorotoluene	--	.2	Tetrachloroethene	5	.2
4-Chlorotoluene	--	.2	Toluene	1,000	.2
1,2-Dibromo-3-chloropropane	.2	1	1,2,3-Trichlorobenzene	--	.2
1,2-Dibromoethane	.05	.2	1,2,4-Trichlorobenzene	70	.2
Dibromomethane	--	.2	1,1,1-Trichloroethane	200	.2
1,2-Dichlorobenzene	600	.2	1,1,2-Trichloroethane	5	.2
1,3-Dichlorobenzene	600	.2	Trichloroethene	5	.2
1,4-Dichlorobenzene	75	.2	Trichlorofluoromethane	--	.2
Dichlorobromomethane	*	.2	1,2,3-Trichloropropane	--	.2
Dichlorodifluoromethane	--	.2	Trichlorotrifluoroethane	--	.2
1,1-Dichloroethane	--	.2	1,2,4-Trimethylbenzene	--	.2
1,2-Dichloroethane	5	.2	1,3,5-Trimethylbenzene	--	.2
Cis-1,2-Dichloroethene	--	.2	Vinyl chloride	2	.2
1,1-Dichloroethene	7	.2	Xylenes, total ortho, meta, and para	10,000	.2
1,2-Transdichloroethene	100	.2			

Specialist, the problem may be referred to the Office of Water Quality or National Research Program, where research hydrologists and chemists will aid in resolving the problem.

Reporting of Data

Most data collected by the USGS are published in data reports and used in interpretive reports. Water-quality information, subsequent to its review, is entered into a local data base and periodically merged with a nationally accessible data base. Data that suggests that there could be a human health or environmental problem are provided to managerial agencies such as the DOE and to regulatory agencies, such as the State of Idaho's Department of Health and Welfare and the U.S. Environmental Protection Agency, Region 10. After data have been reviewed and verified by resampling if necessary, they are available to the general public upon request.

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ATTACHMENTS

Attachment 1—Field schedule showing well and pump information and sampling schedules for selected wells and streamflow sites.

Attachment 1

Date	Local site identifier	Method of sampling	Well depth (feet)	Analysis type (see code)			RESL	NWQL	Field value.
				Jan.	Apr.	July			
	ANP #6	Pump 40 gpm	12	305		14			
	ANP #9	Pump 20 gpm	12	322	34		35		
	Arbor Test	Pump 20 gpm	16	790		2	2		
	AREA II	Pump 14 gpm	16	877		14			
	Atomic City*	Spigot	8	639	1		9		
	Badging Facility	Pump 35 gpm	8	644		14			
	BLR (near Mackay)	Surface water			4		4		
	BLR (near Arco)	Surface water			4		4		
	BLR (INEL Div.)	Surface water			4		4		
	BLR (Dairy Farm)	Surface water			4		4		
	Birch Creek*	Surface water			1		1		
	CFA #1*	Pump 1,000 gpm	16	685	11	13	11	14	
	CFA #2*	Pump 1,400 gpm	20	681	11	13	11	14	
	CFA LF 2-10	Pump 6.6 gpm	6	765	26	26	26	27	
	CFA LF 3-9	Pump 5.7 gpm	4	500			23		
	CFA LF 3-11	Pump 7 gpm	4	492			23		
	CPP #1	Pump 3,000 gpm	20	585	28		29		
	CPP #2	Pump 3,000 gpm	16	605	17		43		
	CPP #4	Pump 400 gpm	16	700	17		43		
	CPP #5	Pump 200 gpm	16	721	17		43		
	CWP 1	Bail @ 65 feet	10	66.0			6		
	CWP 2	Bail @ 52 feet	10	52.5			6		

Attachment 1

Date	Time	Local site identifier	Method of sampling	Well depth (feet)	Analysis type (see code)			RESL	NWQL	Field value.
					Jan.	Apr.	July	Oct.		
									SH or lab code	Temp
		CWP 3	Bail @60 feet	10	60.5			6		
		CWP 4	Bail @60 feet	10	61.0			6		
		CWP 5	Bail @53 feet	10	53.5			6		
		CWP 6	Bail @52 feet	10	52.5			6		
		CWP 7	Bail @53 feet	10	53.5			6		
		CWP 8	Bail @65 feet	10	66.0			6		
		EBR-I	Pump 1000 gpm	12	1.075			24		
		Fire Station #2	Pump 400 gpm	16	518			2		
		Highway #3*	Spigot	8	750			24		
		IET Disp	Pump 40 gpm	20	324			14		
		Leo Rogers #1	Pump 20 gpm	20	702			15		
		Little Lost River	Surface water					1		
		MTR Test	Pump 26 gpm	8	588			11		
		Mud Lake*	Surface water					1		
		NRF 6***	Pump 2.5 gpm	8	417					
		NRF 7***	Pump 2.5 gpm	10	417					
		NRF 8***	Pump 22 gpm	8	423					
		NRF 9***	Pump 30 gpm	8	422					
		NRF 10***	Pump 30 gpm	8	427					
		NRF 11***	Pump 28 gpm	8	417					
		NRF 12***	Pump 30 gpm	8	421					
		NRF 13***	Pump 1 gpm	8	425					
		NPR Test	Pump 26 gpm	6	599			24		
									38	

Attachment 1

Date	Time	Local site identifier	Method of sampling	Well depth (feet)	Analysis type (see code)			RESL	NWQL	Field value.
					Jan.	Apr.	July			
		PSTF	Pump 44 gpm	16	322		34	35		
		P&W#2*	Pump 35 gpm	10	386		24	25		
		PW-1	Grundfos @105 feet; 1.5 gpm	10	117	3	5	3	13	
		PW-2	Bail @ 115 feet	10	131		5	13		
		PW-3	Bail @121 feet	10	125		5	13		
		PW-4	Pump 6 gpm	10	136	3	5	3	13	
		PW-5	Pump 8 gpm	10	124		5	13		
		PW-6	Bail @125 feet	10	125	3	5	3	13	
		PW-7	Bail @220 feet	10	237		7	17		
		PW-8	Pump 8 gpm	10	166	6	7	6	17	
		PW-9	Pump 5 gpm	10	200	6	7	6	17	
		RWMC M3S	Pump 4.5 gpm	6	633			23		
		RWMC M7S	Pump 4.5 gpm	6	638			23		
		RWMC Production**	Pump 200 gpm	16	683	39	21	39	22	
		Site #4	Pump 500 gpm	15	496		2		11	
		Site #9	Pump 25 gpm	10	1,057			14		
		Site 14*	Pump 40 gpm	8>377 10<377 12<340	717		24		25	
		Site #17	Pump 25 gpm	20	600				14	
		Site #19	Pump 25 gpm	10>576 18<576	865				11	
		SPERT 1	Pump 400 gpm	24	653				10	

Attachment 1

Date	File	Local site identifier	Method of sampling	Hole diameter (inches) (feet)	Analysis type (see code)			RESL	NWQL	Field value.
					Jan.	Apr.	July			
		SWP 8	Bail from bottom of well	2	28			16		
		SWP 13	Bail from bottom of well	2	27			16		
		TAN Expl. (no name 1)	Pump 42 gpm	12	550		34		35	
		TRA #1	Pump 3,400 gpm	20	600			11		
		TRA #3	Pump 3,800 gpm	20	602			11		
		TRA #4	Pump 2,000 gpm	20	975			11		
		TRA A-13	Bail from bottom of well	2	59		7		18	
		TRA A-77	Bail from bottom of well	2	33		7		18	
		TRA Disp.	Pump 25 gpm	12	1,267		7		19	
		W.S. for INEL-1	Pump 12 gpm	8	595		2		11	
		USGS #1	Pump 16 gpm	6	636		24		25	
		USGS #2	Pump 16 gpm	5	704			14		
		USGS #4	Pump 40 gpm	6	553		24		25	
		USGS #5	Pump 5 gpm	6	500		24		38	
		USGS #6	Pump 25 gpm	6	620			14		
		USGS #7	Pump 45 gpm	4 >760 6 <760	1,200		34		35	
		USGS #8*	Pump 16 gpm	6	812		24		25	
		USGS #9	Pump 19 gpm	8	655		24		25	
		USGS #11*	Pump 20 gpm	12	704		24		25	
		USGS #12**	Pump 32 gpm	10 >387 12 <387	560		30		31	
		USGS #14*	Pump 16 gpm	6	751		4		4	

Attachment 1

Date	Time	Local site identifier	Method of sampling	Analysis type (see code)				NWQL	Field value.
				Well depth (feet)	Jan.	Apr.	July		
		USGS #15	Pump 30 gpm	10 >480 16 <480	610			14	
		USGS #17	Pump 32 gpm	6 >416 8 <416	498		24	25	
		USGS #18	Pump 25 gpm	4	329		14		
		USGS #19*	Pump 33 gpm	6	405		24	25	
		USGS #20	Pump 25 gpm	6	676		3	14	
		USGS #22	Pump 2.5 gpm	6	657			9	
		USGS #23	Pump 25 gpm	6	467		24	25	
		USGS #26	Pump 40 gpm	8	266		34	35	
		USGS #27*	Pump 20 gpm	8	312		24	25	
		USGS #29	Pump 32 gpm	6	422			14	
		USGS #31	Pump 40 gpm	8 >306 10 <306	428			14	
		USGS #32	Pump 28 gpm	6 >323 10 <323	392			14	
		USGS #34	Pump 25 gpm	10 >499 13 <499	700		28	29	
		USGS #35	Pump 25 gpm	7	578			3	14
		USGS #36	Pump 25 gpm	6	567		3	3	14
		USGS #37	Pump 25 gpm	8	573		3	20	
		USGS #38	Pump 6 gpm	6 >505 8 <505	729		28	29	
		USGS #39	Pump 25 gpm	6 >507 8 <507	572	3	3	3	14

Attachment 1

Date	Time	Local site identifier	Method of sampling	Well depth (feet)	Analysis type (see code)				RESL	NWQL	Field value.	
					Jan.	Apr.	July	Oct.				
		USGS #40	Pump 8 gpm	6	483	8	8	8	20			
		USGS #41	Pump 25 gpm	6	674		3					
		USGS #42	Pump 25 gpm	6	678		3					
		USGS #43	Pump 6 gpm	6	676		5					
		USGS #44	Pump 24 gpm	6	650		5					
		USGS #45	Pump 24 gpm	6	651		3					
		USGS #46	Pump 24 gpm	6	651		5					
		USGS #47	Pump 8 gpm	6	652		5					
		USGS #48	Pump 24 gpm	6	750		3					
		USGS #50	Pump 0.5 gpm	6	405		5					
		USGS #51	Pump 5 gpm	6	659		3					
		USGS #52	Pump 25 gpm	6	650		3					
		USGS #53	Pump 6 gpm	6	90		7					
		USGS #54	Pump 6 gpm	6	91	6	7	6				
		USGS #55	Pump 6 gpm	6	79		7					
		USGS #56	Pump 1 gpm	6	80		7					
		USGS #57	Pump 25 gpm	6	732	3	3	3				
		USGS #58	Pump 26 gpm	6	503		7					
		USGS #59	Pump 3 gpm	6	657		3					
		USGS #60	Pump 6 gpm	6	117		7					
		USGS #61	Pump 6 gpm	10	123		7					
		USGS #62	Pump 5 gpm	8	165		7					
		USGS #63	Pump 5 gpm	10	97		7					

Attachment 1

Date	Time	Local site identifier	Method of sampling	Hole diameter (inches)	Analysis type (see code)			RESL	NWQL	Field value.
					Well depth (feet)	Jan.	Apr.	July	Oct.	
		USGS #65*	Pump 7 gpm	6	498	7	36	7	37	
		USGS #66	Bail @ 214 feet	6	475				17	
		USGS #67	Pump 8 gpm	6	698		3		14	
		USGS #68	Pump 1 gpm	10	128		7		17	
		USGS #69	Pump 5 gpm	10	115			17		
		USGS #70	Pump 6 gpm	8	100		7		17	
		USGS #71	Pump <1 gpm	8	184		7		17	
		USGS #72	Pump 1 gpm	6	177			17		
		USGS #73	Grundfos @100 feet; 1.5 gpm	6	127		7		17	
		USGS #74	Bail @ 190 feet	6	192		7		17	
		USGS #76	Pump 25 gpm	6	718		7		19	
		USGS #77	Pump 25 gpm	6	610		28		29	
		USGS #78	Bail	7	204			17		
		USGS #79	Pump 25 gpm	6	702		2		11	
		USGS #82	Pump 25 gpm	8	700	3	3	3	14	
		USGS #83	Pump 26 gpm	6	752		24		38	
		USGS #84	Pump 5 gpm	6	505			36	37	
		USGS #85*	Pump 23 gpm	6	637		3		14	
		USGS #86	Pump 19 gpm	8	691		24		25	
		USGS #87*	Pump 2 gpm	6	673	21	32	21	33	
		USGS #88	Pump 1 gpm	6	662	21	21	21	22	
		USGS #89	Pump 5 gpm	6	646	3	21	3	22	

Attachment 1

Date	Time	Local site identifier	Method of sampling	Well depth (feet)	Analysis type (see code)			RESL	NWQL	Field value.
					Jan.	Apr.	July	Oct.		
									SH or Temp	pH
		USGS #90	Pump 4 gpm	6	626	21	21	22		
		USGS #92	Bail @213 feet	6	214	21				8
		USGS #97***	Pump 27 gpm	4	510		36			37
		USGS #98***	Pump 18 gpm	6	505		36			37
		USGS #99***	Pump 25 gpm	6	450		2			11
		USGS #100*	Pump 10 gpm	6	750		2			11
		USGS #101	Pump 13 gpm	6	865		24			25
		USGS #102***	Pump 29 gpm	6	445			14		
		USGS #103*	Pump 21 gpm	8	760	1	24	1		25
		USGS #104*	Pump 21 gpm	8	700	1	1	1		10
		USGS #105	Pump 19 gpm	8	800		24			25
		USGS #106	Pump 22 gpm	8	760		1			9
		USGS #107	Pump 25 gpm	8	690		24			38
		USGS #108*	Pump 20 gpm	8	760		24			25
		USGS #109	Pump 16 gpm	6	800		24			25
		USGS #110A	Pump 24 gpm	10	644		24			25
		USGS #111	Pump 15 gpm	8	595		3			14
		USGS #112*	Pump 25 gpm	8	563		3			14
		USGS #113	Pump 25 gpm	6	564		3			16
		USGS #114	Pump 10 gpm	6	562		3			14
		USGS #115*	Pump 5 gpm	6	581		3			14
		USGS #116	Pump 20 gpm	6	580		3			14
		USGS #117	Pump 10 gpm	8	655	3	21	3		22

Attachment 1

Local site identifier	Method of sampling	Well depth (feet)	Analysis type (see code)			RESL	NWQL	Field value.
			Jan.	Apr.	July			
USGS #119	Pump 3 gpm	8	705	3	21	3	22	
USGS #120*	Pump 21 gpm	8	705	21	32	21	33	
USGS #121	Pump 8 gpm	8	475		3		14	
USGS #123	Pump 3 gpm	8	481		3		14	
USGS #124	Pump 18 gpm	4	800		1		10	
USGS #125	Pump 20 gpm	10	760		24		25	

*Well is sampled with someone from the State of Idaho's INEL Oversight Program

**Well is sampled monthly for organics - SH1380

***Well is sampled quarterly for the NRF study--Dec., Mar., June, Sept.

Attachment 1Codes for types of analyses

1. ^3H , Cl^-
2. ^3H , Cl^- , Cr
3. ^3H , ^{90}Sr , Cl^-
4. ^3H , Cl^- , α , β , γ Spec
5. ^3H , ^{90}Sr , γ Spec, Cl
6. ^3H , ^{90}Sr , Cl^- , Cr, SO_4^-
7. ^3H , ^{90}Sr , γ Spec, Cl^- , Cr
8. ^3H , ^{90}Sr , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^-
9. ^3H , Cl^- , Na^+
10. ^3H , Cl^- , Na^+ , NO_3^-
11. ^3H , Cl^- , Cr, Na^+
12. ^3H , Cl^- , Cr, Na^+ , NO_3^- , SO_4^-
13. ^3H , ^{90}Sr , Cl^- , Na^+ , SO_4^-
14. ^3H , ^{90}Sr , Cl^- , Na^+ , NO_3^-
15. ^3H , α , β , γ Spec, Cl^- , Na^+
16. ^3H , ^{90}Sr , γ Spec, Cl^- , Na^+ , NO_3^- , SO_4^-
17. ^3H , ^{90}Sr , Cl^- , Cr, Na^+ , SO_4^-
18. ^3H , ^{90}Sr , γ Spec, Cl^- , Cr, Na^+ , SO_4^-
19. ^3H , ^{90}Sr , γ Spec, Cl^- , Cr, Na^+ , NO_3^- , SO_4^-
20. ^3H , ^{90}Sr , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , NO_3^- , SO_4^-
21. ^3H , ^{90}Sr , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , POC's
22. ^3H , ^{90}Sr , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , NO_3^- , POC's, SO_4^-
23. ^3H , ^{90}Sr , Cl, NO_3^-
24. ^3H , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^-
25. ^3H , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^- , TOC
26. ^3H , ^{90}Sr , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^-
27. ^3H , ^{90}Sr , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^- , TOC
28. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , SO_4^- , F, ^{129}I , POC's
29. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , SO_4^- , F, ^{129}I , POC's, TOC
30. ^3H , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^- , SO_4^- , POC's
31. ^3H , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^- , SO_4^- , POC's, TOC
32. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , POC's
33. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , POC's, TOC, SO_4^-
34. ^3H , ^{90}Sr , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^- , POC's, Sb, Ar, Cr, Pb, Hg, Ni, Tl, Zn
35. ^3H , ^{90}Sr , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^- , POC's, TOC, Sb, Ar, Cr, Pb, Hg, Ni, Tl, Zn
36. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , SO_4^- , POC's, Al, Ar, Ba, Cd, Pb, Mn, Ni, Hg, Se, Ag, Zn
37. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , SO_4^- , POC's, TOC, Al, Ar, Ba, Cd, Pb, Mn, Ni, Hg, Se, Ag, Zn
38. ^3H , α , β , γ Spec, Cl^- , Na^+ , Cr, NO_3^- , TOC, POC's
39. ^3H , ^{90}Sr , Cl^- , POC'S
40. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , POC's, SH1043 metals
41. ^3H , ^{90}Sr , α , β , γ Spec, ^{241}Am , ^{238}Pu , $^{239,240}\text{Pu}$, Cl^- , Na^+ , Cr, NO_3^- , POC's, SH1043 metals, TOC
42. POC's
43. ^3H , ^{90}Sr , Cl^- , Cr, Na^+ , NO_3^-

Attachment 1

Constituent and type of sample treatment

Constituent	Lab	Bottle type	Description
³ H	RESL	500 mL (Apr.); 125 mL (Jan., Jul., Oct.)	Raw water, unacidified
⁹⁰ Sr	RESL	500 mL	Raw water, preserved with 2 mL HNO ₃
⁹⁰ Sr, Y Spec	RESL	500 mL	Raw water, preserved with 2 mL HNO ₃
α , β	RESL	500 mL	Raw water, preserved with 2 mL HNO ₃
²⁴¹ Am, ²³⁸ Pu, ^{239,240} Pu,	RESL	500 mL	Raw water, preserved with 2 mL HNO ₃
⁹⁰ Sr, Y Spec, ²⁴¹ Am, ²³⁸ Pu, ^{239,240} Pu	RESL	1 L	Raw water, preserved with 4 mL HNO ₃
Y Spec	RESL	500 mL	Raw water, preserved with 2 mL HNO ₃
Cl ⁻ **	NWQL	250 mL; LC 1571	Filtered, unacidified
Cr*	NWQL	250 mL; LC 722	Filtered, preserved with 1 mL HNO ₃
Na ⁺ *	NWQL	250 mL; LC 675	Filtered, preserved with 1 mL HNO ₃
NO ₃ ⁻	NWQL	125 mL; SH101	Filtered, chilled, brown poly
POC's	NWQL	(3) 40 mL GCV-SH1380	Raw water, chilled, unacidified
SO ₄ ²⁻ **	NWQL	250 mL; LC 1572	Filtered, unacidified
F**	NWQL	250 mL; LC 31	Filtered, unacidified
TOC	NWQL	125 mL; LC 114	Raw water, chilled, unacidified, glass amber
Sb, Cr, Pb, Ni, Zn Ar, Tl	NWQL	250 mL; SH 1050 250 mL; LC 112 and LC 492	Filtered, preserved with 1 mL HNO ₃ Raw water, unacidified
Hg	NWQL	250 mL; LC 226	Filtered, preserved with 10 mL potassium dichromate
Al, Ba, Cd, Cr, Pb, Mn, Ni, Ag, Zn, Ar, Se	NWQL	250 mL; SH 1050 250 mL; LC 112 and LC 87	Filtered, preserved with 1 mL HNO ₃ Raw water, unacidified
¹²⁹ I	CL	1 L	Raw water, chilled, unacidified, polyseal cap
³⁶ Cl	CL	1 L RU	Raw water, unacidified, polyseal cap
SH 1043 metals	NWQL	250 mL; SH 1043 125 mL; SH 1043	Filtered, preserved with 1 mL HNO ₃ Raw water, unacidified

*Analysis can be requested from the same bottle.

**Analysis can be requested from the same bottle

CL - contract laboratory

Attachment 2—DATA-QUALITY OBJECTIVES FOR WATER SAMPLES ANALYZED BY THE NATIONAL WATER QUALITY LABORATORY

Constituent	Minimum reporting level (micrograms per liter)	Precision (\pm percent)	Accuracy* (percent)	Lab code/ schedule
I. Purgeable organic compounds	1	30	70-130	SH 1380
II. Base/neutral extractable compounds, total recoverable	10	30	70-130	SH 1385
III. Inorganic compounds: (dissolved)				
1. Aluminum	10	10	90-110	SH 1050
2. Antimony	1	10	90-110	SH 1050
3. Arsenic	1	10	90-110	LC 112
4. Barium	1	10	90-110	SH 1050
5. Beryllium	5	10	90-110	SH 1043
6. Cadmium	1	10	90-110	SH 121
7. Calcium	20	10	90-110	SH 1043
8. Chromium	5	10	90-110	LC 722
9. Hexavalent chromium	1	10	90-110	LC 16
10. Cobalt	3	10	90-110	SH 1043
11. Copper	10	10	90-110	SH 1043
12. Iron	3	10	90-110	SH 1043
13. Lead	10	10	90-110	SH 1050
14. Magnesium	10	10	90-110	SH 121
15. Manganese	1	10	90-110	SH 1050
16. Nickel	10	10	90-110	SH 1050
17. Potassium	100	10	90-110	SH 121
18. Selenium	1	10	90-110	LC 87
19. Silica	10	10	90-110	SH 121
20. Silver	1	10	90-110	SH 1050
21. Sodium	100	10	90-110	LC 59
22. Thallium	0.5	10	90-110	LC 492
23. Vanadium	6	10	90-110	SH 1043
24. Zinc	3	10	90-110	SH 1050
25. Mercury	0.1	10	90-110	LC 226
26. Sulfate	100	10	90-110	LC 1572
27. Chloride	1,000	10	90-110	LC 1571
28. Nitrate (as N)	50	10	90-110	SH 101
29. Phosphate	10	10	90-110	SH 101
30. Ammonia (as N)	15	40	60-140	SH 101
IV. Organic compounds				
1. Polychlorinated biphenyls (PCB's)	1	40	60-140	SH 1324 (PCB only)
2. Chlorinated phenols (PCP's), includes tetra-, tri, and dichlorophenyls	30	40	60-140	SH 1385

*Coefficient of variance measured by replicate analysis; precision at 10% level.

Attachment 3—DATA-QUALITY OBJECTIVES FOR RADIONUCLIDES IN WATER SAMPLES ANALYZED BY THE RADIOLOGICAL AND ENVIRONMENTAL SCIENCES LABORATORY

For each radionuclide concentration, an associated analytical uncertainty, s , is calculated such that there is a 67-percent probability that the true concentration of a radionuclide in a sample is in the range of the reported concentration plus or minus the analytical uncertainty. For example, given an analytical result of 1.0 ± 0.2 pCi/L (picocuries per liter), there is a 67 percent probability that the true concentration is in the range of 0.8 to 1.2 pCi/L. Some laboratories report the analytical uncertainty as $2s$, at which there is a 95-percent probability that the true concentration is in the range of 0.6 to 1.4 pCi/L. Therefore, unlike analyses for most inorganic or organic constituents, the analytical uncertainty is specified for each analysis for a specified radionuclide. The following guidelines for interpreting analytical results are based on an extension of the method described by Currie (1968).

In the analysis for a selected radionuclide, laboratory measurements are made on a target sample and a prepared blank. Instrument signals for the sample and the blank vary randomly. Therefore, it is essential to distinguish between two key aspects of the problem of detection: (1) the instrument signal for the sample must be greater than the signal observed for the blank to make the decision that a selected radionuclide was detected; and (2) an estimation must be made of the minimum radionuclide concentration that will yield a sufficiently large observed signal to make the correct decision for detection or nondetection of that radionuclide most of the time. The first aspect of the problem is a qualitative decision based on an observed signal and a definite criterion for detection. The second aspect of the problem is an intuitive estimation of the detection capabilities of a given measurement process.

In the laboratory, instrument signals must exceed a critical level to make the qualitative decision whether a selected radionuclide was detected. Radionuclide concentrations that equal $1.6s$ meet this criterion; at $1.6s$, there is a 95-percent probability that the correct decision—not detected—will be made. Given a large number of samples, up to 5 percent of the samples with true concentrations greater than or equal to $1.6s$, which were concluded as being detected, might not contain the selected radionuclide. These measurements are referred to as false positives and are errors of the first kind in hypothesis testing.

Once the critical level of $1.6s$ has been defined, the minimum detectable concentration may be established. Radionuclide concentrations that equal $3s$ represent a measurement of the minimum detectable concentration. For true concentrations of $3s$ or greater, there is a 95-percent-or-more probability of correctly concluding that a selected radionuclide was detected in a sample. Given a large number of samples, up to 5 percent of the samples with true concentrations greater than or equal to $3s$, which were concluded as being nondetected, could contain the selected radionuclide at the minimum detectable concentration. These measurements are referred to as false negatives and are errors of the second kind in hypothesis testing.

True radionuclide concentrations between $1.6s$ and $3s$ have larger errors of the second kind. That is, there is a greater-than-5-percent probability of false negative results for samples with true concentrations between $1.6s$ and $3s$, and although the selected radionuclide might not have been detected, such nondetection may not be reliable; at $1.6s$, the probability of a false negative is about 50 percent.

These guidelines are based on counting statistics alone and do not include systematic or random errors inherent in laboratory procedures. The values $1.6s$ and $3s$ vary slightly with background or blank counts and with the number of gross counts for individual analyses and for different selected radionuclides. The use of the critical level and minimum detectable concentration aid in the interpretation of analytical results and do not represent absolute concentrations of radioactivity which may or may not have been

detected. The minimum detectable concentration should not be confused with the detection limit, which is based on instrument sensitivity, sample volumes, analytical procedures and counting times used in the laboratory.

Bodnar and Percival (1982) summarized detection limits normally available from the RESL. Special arrangements can be made to achieve smaller detection limits for selected constituents. For example, by using a five-fold counting time for tritium in water, that is, increasing the counting time from 20 to 100 minutes, the detection limit can be reduced from 0.5 to 0.2 pCi/L.

Detection limits for selected types of radioactivity and nuclides as a function of sample size and detection method are shown on table 1; the limits are intended as guides to order-of-magnitude sensitivities and, in practice, can easily change by a factor of two or more even for the conditions specified.

Table 1—Detection limits for selected types of radioactivity and nuclides

Type of radioactivity or nuclide	Sample material	Size of sample (milliliter)	Counting time (minutes)	Detection method or instrument	Detection limit (picocuries per milliliter)
Gross alpha	Water	100	60	Scintillation	3X10 ⁻³
Gross beta	Water	250	20	Low bkgd counter	5X10 ⁻³
	Water	100	20	Low bkgd counter	4X10 ⁻³
	Water	5	20	Low bkgd counter	0.1
Po-210	Water	100	60	Scintillation	1X10 ⁻³
Sr-90	Water	400	50	Low bkgd counter	5X10 ⁻³
Th-230	Water	500	60	Scintillation	1X10 ⁻³
Tritium	Water	10	20	Liquid scintillation	0.5
U-234	Water	1,000	1,000	Alpha spectrometry	4X10 ⁻⁵
Th-230	Water	1,000	1,000	Alpha spectrometry	4X10 ⁻⁵
Pu-238	Water	1,000	1,000	Alpha spectrometry	2X10 ⁻⁵
Am-241	Water	1,000	1,000	Alpha spectrometry	3X10 ⁻⁵
Np-239	Water	400	60	Ge(Li)	0.4
Tc-99	Water	400	60	Ge(Li)	0.7
Te-132	Water	400	60	Ge(Li)	6X10 ⁻²
Pb-212	Water	400	60	Ge(Li)	0.1
Se-75	Water	400	60	Ge(Li)	8X10 ⁻²
Sb-125	Water	400	60	Ge(Li)	0.2
Ru-103	Water	400	60	Ge(Li)	1X10 ⁻²
Tl-108	Water	400	60	Ge(Li)	0.2
Sb-124	Water	400	60	Ge(Li)	0.1
Ta-182	Water	400	60	Ge(Li)	0.2
Co-60	Water	400	60	Ge(Li)	6X10 ⁻²
Na-22	Water	400	60	Ge(Li)	9X10 ⁻²
K-40	Water	400	60	Ge(Li)	1
La-140	Water	400	60	Ge(Li)	7X10 ⁻²
Co-56	Water	400	60	Ge(Li)	5X10 ⁻²
Ce-144	Water	400	60	Ge(Li)	0.4
Ce-141	Water	400	60	Ge(Li)	9X10 ⁻²
Cr-51	Water	400	60	Ge(Li)	0.6
I-131	Water	400	60	Ge(Li)	6X10 ⁻²
Ba-140	Water	400	60	Ge(Li)	0.2
Ru-106	Water	400	60	Ge(Li)	0.5
Cs-137	Water	400	60	Ge(Li)	6X10 ⁻²
Bi-212	Water	400	60	Ge(Li)	1.0
Nb-95	Water	400	60	Ge(Li)	6X10 ⁻²
Cs-134	Water	400	60	Ge(Li)	6X10 ⁻²
Mo-99	Water	400	60	Ge(Li)	5X10 ⁻²
Hg-203	Water	400	60	Ge(Li)	6X10 ⁻²

Table 1—Detection limits for selected types of radioactivity and nuclides—continued

Type of radioactivity or nuclide	Sample material	Size of sample (milliliter)	Counting time (minutes)	Detection method or instrument	Detection limit (picocuries per milliliter)
Kr-85	Water	400	60	Ge(Li)	21
Bi-214	Water	400	60	Ge(Li)	0.4
Zr-95	Water	400	60	Ge(Li)	9X10 ⁻²
Co-58	Water	400	60	Ge(Li)	6X10 ⁻²
Mn-54	Water	400	60	Ge(Li)	5X10 ⁻²
Ag-110m	Water	400	60	Ge(Li)	7X10 ⁻²
Ac-228	Water	400	60	Ge(Li)	0.2
Fe-59	Water	400	60	Ge(Li)	0.1
Zn-65	Water	400	60	Ge(Li)	0.1



Attachment 4—INVENTORY OF WATER-QUALITY FIELD EQUIPMENT

Type of meter	Model	Manufacturer	Serial number
pH	SA250	Orion	7567
pH	SA250	Orion	4154
pH	250A	Orion	064290
pH	250A	Orion	004385
pH	250A	Orion	005808
Sp. Cond.	122	Orion	0905040
Sp. Cond.	122	Orion	0905045
Sp. Cond.	122	Orion	42402093
Sp. Cond.	122	Orion	42556041