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Informal Report

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**THE PHOTOSYNTHETIC ACCLIMATION OF *LOLIUM PERENNE*  
IN RESPONSE TO THREE YEARS GROWTH IN A FREE-AIR CO<sub>2</sub>  
ENRICHMENT (FACE) SYSTEM**

**Graham J. Hymus**

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by

Graham J. Hymus  
Department of Biology  
University of Essex  
and  
University of Writtle College  
United Kingdom

August 1996

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**PERENNE IN RESPONSE TO THREE YEARS GROWTH IN A**  
**FREE-AIR CO<sub>2</sub> ENRICHMENT (FACE) SYSTEM.**

BY GRAHAM J. HYMUS

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Dept of Biology, University of Essex; and Writtle College.

## CONTENTS.

Abstract.	1
Abbreviations and symbols.	2
<u>1. Introduction.</u>	
1:1. Climate change.	3
1:2. Theoretical crop responses.	3
1:3. Acclimation.	4
1:4. A/Ci response.	6
1:5. <i>Lolium perenne</i> .	7
<u>2. Materials and methods.</u>	
2:1. The site.	11
2:2. Materials.	12
2:3. Methods.	14
2:3:1. Diurnal measurements.	14
2:3:2. A/Ci determination.	15
2:3:3. Diurnal A/Ci determination.	16
2:3:4. Manipulation experiment.	17
2:3:5. Canopy effects on A/Ci response.	18
<u>3. Results.</u>	
3:1. Diurnal Measurements.	19
3:2. A/Ci results.	29
3:3. Manipulation experiment results.	33
3:4. Canopy experiment results.	35
3:5. Diurnal A/Ci results.	38
<u>4. Discussion.</u>	41
<u>5. Conclusion.</u>	49
Appendix.	52
Literature cited.	53

## ABSTRACT.

Pure stands of Ryegrass (*Lolium perenne* L. cv Bastion) were in their third year of growth in the field, exposed to either ambient ( $355 \mu\text{mol mol}^{-1}$ ), or elevated ( $600 \mu\text{mol mol}^{-1}$ ) atmospheric  $\text{CO}_2$  concentration. A Free-Air  $\text{CO}_2$  Enrichment (FACE) system was used to maintain the elevated  $\text{CO}_2$  concentration whilst limiting experimental constraints on the field conditions. The theoretically predicted increase in the net rates of  $\text{CO}_2$  uptake per unit leaf area ( $A \mu\text{mol mol}^{-1}$ ) as a consequence, primarily, of the suppression of photorespiration by  $\text{CO}_2$  a competitive inhibitor of RubP oxygenation by Rubisco, was observed for the *Lolium perenne* studied. Also observed was a general decline in leaf evapotranspiration (E) consistent with observations of increased water use efficiency of crops grown in elevated  $\text{CO}_2$ . Enhancement of leaf A in the FACE grown *L. perenne* ranged from 26.51% to 44.95% over the course of a diurnal set of measurements. Whilst reductions in leaf E reached a maximum of 16.61% over the same diurnal course of measurements. The increase in A was reconciled with an absence of the commonly observed decline in  $V_{c \text{ max}}$  as a measure of the maximum *in vivo* carboxylation capacity of the primary carboxylating enzyme Rubisco and  $J_{\text{max}}$  a measure of the maximum rate of electron transport. The manipulation of the source sink balance of the crop, stage of canopy regrowth or height in the canopy had no effect on the observation of a lack of response. The findings of this study will be interpreted with respect to the long term implications of  $\text{C}_3$  crops being able to adapt physiologically to maximise the potential benefits conferred by growth in elevated  $\text{CO}_2$ .

**Key words:** *Lolium perenne*, elevated CO<sub>2</sub>, photosynthesis, acclimation, Free-Air CO<sub>2</sub> Enrichment.

**Abbreviations and symbols:**

**A** = Net rate of CO<sub>2</sub> uptake per unit area ( $\mu\text{mol mol}^{-1}$ ); **Ca** = Atmospheric CO<sub>2</sub> concentration ( $\mu\text{mol CO}_2 / \text{mol air}$ ); **Ci** = Intercellular CO<sub>2</sub> concentration ( $\mu\text{mol CO}_2 / \text{mol air}$ ); **E** = H<sub>2</sub>O transpiration rate ( $\text{mmol m}^{-2} \text{s}^{-1}$ ); **FACE** = Free-Air CO<sub>2</sub> Enrichment ; **J<sub>max</sub>** = Maximum light saturated rate of electron transport ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ); **Pi** = Inorganic phosphate; **PPFD** = Photosynthetic photon flux density ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ); **Rubisco** = Ribulose-1,5 biphosphate Carboxylase-Oxygenase; **RubP** = Ribulose-1,5 biphosphate; **V<sub>cmax</sub>** = Maximum rate of carboxylation ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ).

## **1. INTRODUCTION.**

### **1:1. Climate change.**

The exponential increase in the atmospheric carbon dioxide concentration ( $C_a$ ), from pre-industrial revolution levels of  $275 \mu\text{mol mol}^{-1}$ , to the current level of  $345 \mu\text{mol mol}^{-1}$  (Lindzen 1993) has been facilitated primarily by anthropogenic emissions of  $\text{CO}_2$ . Namely, the burning of fossil fuels (5.4 Gt per annum) and deforestation (1.6 Gt per annum) (Idso 1989, Post *et al* 1990). Projections suggest that  $C_a$  will reach 530 and  $700 \mu\text{mol mol}^{-1}$  by 2050 and 2100 respectively (IPCC business as usual scenario) (Watson *et al* 1990). A concurrent rise in temperature of  $3^\circ\text{C}$  and  $4^\circ\text{C}$  by 2050 and 2100 is also predicted (Watson *et al* 1990).

The implications for crop plants, of such increases in atmospheric  $\text{CO}_2$  concentration. Concurrent with increases in temperature are considerable. With new balances between respiration (carbon release) and photosynthesis (carbon sequestration) dependent on the interactive nature of the increasing  $C_a$  and temperature (Long 1991). It is envisaged that fundamental to the crop plants performance will be the way in which its biochemical systems, geared towards coping with an atmosphere low in  $\text{CO}_2$  can adapt (acclimate) to a higher  $C_a$  under existing genetic feedback mechanisms so that limiting resources like nitrogen are not wasted (Woodrow 1994).

### **1:2. Theoretical crop responses.**

In the short term, increases in average crop yields for  $C_3$  crops, of which Ryegrass is one, are expected to be in the region of 34% (Kimball 1983). These increases are largely attributable to substantial increases in the net rate of  $\text{CO}_2$  uptake per unit area (A) ( $\mu\text{mol mol}^{-1}$ ) (Long 1991). In turn being as a result of the suppression

of RubP oxygenation by Rubisco, by  $\text{CO}_2$  a competitive inhibitor (Webber *et al* 1994). Photorespiration is responsible for a decrease in A of 20% - 50% in temperate climates (Long 1985). In addition, provided RubP is in excess, there will be an expected increase in carboxylation velocity as present  $C_a$  does not saturate Rubisco (Stitt 1991).

In intact leaves, photosynthesis is usually limited by one of three processes.

- 1) The capacity of Rubisco to consume RubP in  $\text{CO}_2$  fixation.
- 2) The capacity of the thylakoid membranes to supply ATP and NADPH for RubP regeneration.
- 3) The capacity of starch and sucrose to utilise triose phosphates and subsequently regenerate inorganic orthophosphate (Pi) for photosynthesis. (Sage *et al* 1994).

### **1:3. Acclimation.**

With an elevated  $C_a$  the balance of control exerted on the plant by 1-3 above, appears to be upset and a new equilibrium sought at a new photosynthetic capacity (Stitt 1991). The primary cause of acclimation is deemed to be the inability of the plant to utilise the excess carbohydrate produced due to an imbalance between excess sources and insufficient sinks for carbohydrate (Arp 1991, Long & Drake 1992).

Whilst the extreme consequences of this imbalance are the accumulation of carbohydrate, in the form of starch, in the chloroplast both damaging (Cave *et al* 1981) and changing its structure (Sionit *et al* 1984), these now seem to be rare events. More common appears to be the inability of the plant to utilise sucrose produced which will result in the reduction of inorganic phosphate (Pi) flux from the cytosol to the chloroplast and the inhibition of ATP and NADPH production and subsequently the rate of RubP regeneration. In addition evidence is pointing to the repression of specific

genes via the accumulation of soluble carbohydrates leading to decreased levels of photosynthetic proteins (Webber *et al.* 1994).

The degree to which these theoretical and often observed initial stimulatory responses to elevated  $C_a$  will be maintained longer term is of paramount importance. Evidence increasingly suggests that physiological adjustment (Oechel & Strain 1985) and acclimation of the photosynthetic apparatus (Sage *et al.* 1989, Beresford 1990) lead to downregulation of photosynthesis in  $C_3$  crops in response to prolonged growth in elevated  $C_a$  (Delucia *et al.* 1985, Cure & Acock 1986). Whilst acclimation is a commonly observed phenomenon of crop plants grown in controlled environments the relevance of these results to those that occur in the field are routinely questioned (Fischer & Aguilar 1976, Lawlor & Mitchell 1991). Indeed there is evidence that the downregulation of photosynthesis in controlled environments is an artefact of the constraints of the experimental system, particularly pot size (Arp 1991) and coupling of environmental growth parameters (Lawlor & Mitchell 1991).

The observed artefacts or "chamber effects" of plant growth in environments that are controlled to some degree commonly include changes in wind velocity, humidity, temperature, light quality or intensity and soil variables (Lewin *et al.* 1994). In an attempt to negate such alterations in the microclimate around the plants compared to the natural environment. A Free-Air (chamberless) carbon dioxide enrichment (FACE) system has been developed by Brookhaven National Laboratory (BNL) to provide controlled fumigation conditions over a large area of field grown

plants throughout the plants phenological development whilst minimising the aforementioned "chamber effects" (Lewin *et al* 1994).

As yet the few field relative to controlled environment experiments and the restricted number of crops studied, are quoted as reasons that conclusive hypotheses regarding field responses to elevated CO<sub>2</sub> cannot be formulated (Rozema *et al* 1993). Indeed results range from acclimation being achieved within three years on Arctic Tussock Grass (Oechel & Strain 1984), to the lack of downward acclimation observed over a four year study on a C<sub>3</sub> sedge (Arp and Drake 1991), whilst acclimation observed late in one growing season on cotton was attributed to earlier senescence (Radkin 1987). To a conclusion that there was a lack of acclimation in field grown crops in natural environments (Arp 1991, Sage *et al* 1994).

#### 1:4. A / Ci response.

The major aid to quantifying the acclimatory response of crops to growth in elevated C<sub>a</sub> is the construction of an A (Leaf net CO<sub>2</sub> uptake, Assimilation), Ci (Internal CO<sub>2</sub> concentration of the leaf) response curve. Derived from the equations of Farquhar *et al* (1980) the A / Ci response provides an *in vivo* measure of.

- 1) The maximum capacity of the leaf to regenerate RubP, a plateau or shallow slope at high Ci where photosynthesis is limited by the rate of regeneration of RubP.
- 2) A measure of the carboxylation efficiency of the leaf, an initial slope at low Ci determined by Rubisco activity, which will be RubP saturated.

Acclimation, defined for the purpose of this study as the downregulation of photosynthetic capacity, is characterised by a decreased carboxylation efficiency, commonly caused by a reduction in the amount / activity of Rubisco (Long & Drake

1992) and can be determined from changes in the initial slope of the  $A / C_i$  response curve. In practice upgrading of the photosynthetic capacity of a plant is also acclimation and highlights the great inter and intra species differences in photosynthetic response to elevated  $C_a$ .

The response of perennial systems to elevated  $C_a$  will be of particular importance in an elevated  $C_a$  world. Ecosystems where enhanced production will be expected to result from increased productivity and an extended growing season into winter months, as a consequence of concurrent rises in  $C_a$  and temperature (Ryle *et al* 1992). When considering perennial crops additional factors will determine their response to elevated  $C_a$ . In particular the allocation of photosynthates to roots and shoots, a trade off which represents the choice of maximising above ground growth in one year and possibly compromising growth in the next, or investing in a larger root system and underground carbohydrate storage for enhanced growth in subsequent growth periods. Increased tillering one year may compromise nitrogen sequestration in the soil in the next year.

#### **1:5. *Lolium perenne*.**

Given these considerations a study of Perennial Ryegrass, *Lolium perenne* was undertaken to ascertain its photosynthetic response to prolonged growth of three years in elevated  $C_a$  in a FACE system (Lewin *et al* 1994). The responses observed could then be reconciled with generalised response of  $C_3$  crops and prior work on Ryegrasses in a variety of experimental designs. A number of facts justify the time devoted to studying this crop. As the most useful and widely spread pasture grasses in many parts of the world several species of *Lolium* are of economic importance (Langer & Hill 1991). At present and for the foreseeable future, Ryegrasses will provide

the mainstay of lowland pasture production in temperate western Europe (Ryle *et al* 1992). Since reserves for the expansion of grassland are limited and since farmland tends to be reducing in industrialised areas emphasis must be placed on the monitoring of changes in grassland productivity and predict future changes. Adaption of management techniques to a higher CO<sub>2</sub> atmosphere may be required (Nijs *et al* 1988) particularly under low input conditions.

Recent experiments suggest that the responses of *Lolium perenne* to elevated C<sub>a</sub> are consistent with those of other C<sub>3</sub> crops, with increases in A at both the single leaf (Nijs *et al* 1989, Ryle *et al* 1992, Gay & Hauck 1993) and canopy (Ryle *et al* 1992, Nijs & Impens 1993). Dry matter production also exhibited expected increases (Nijs *et al* 1988, Ryle *et al* 1992). Acclimatory modification of the photosynthetic apparatus also mirrors generalised C<sub>3</sub> responses, V<sub>c,max</sub> and J<sub>max</sub> values as *in vivo* measurements of the maximum velocity of carboxylation and maximum light saturated rate of electron transport respectively have been shown to decline in response to exposure to elevated C<sub>a</sub> (Moon *et al* 1990). Contention at present seems to surround the response of *Lolium* to elevated C<sub>a</sub> at low light intensities. Evidence tends to suggest that at lower light intensities leaf A decreases for *Lolium perenne* (Nijs *et al* 1989, Ryle *et al* 1992, Gay & Hauck 1993). Such results are inconsistent with the predictions of Long and Drake (1992) who predict that A should increase in elevated C<sub>a</sub> even at low light intensities, and the work of Bryant (1994) where enhanced leaf A was achieved even at low light intensities. Consensus on this area is needed to be able to better determine whether the extension of the growing season for *Lolium perenne* into winter months will be a result of elevated C<sub>a</sub>.

The implications of using different management strategies is mimicked in the F.A.C.E experiment by varying nitrogen treatments and cutting regimes. Again the effects of nitrogen availability on the response of crops to elevated  $C_a$  is in dispute. It is argued that a shortage of nitrogen to facilitate the growth of new sinks for carbohydrate assimilation will enhance downregulation (Wong 1979, Larigauderie 1988, Vessey *et al* 1990). In contrast, it is claimed that a loss of 30 % of Rubisco could be sustained by a plant before its photosynthetic performance is reduced (Long & Drake 1992), ultimately allowing the re-apportioning of nitrogen from Rubisco and mitigating the effects of nitrogen deficits (Sinclair 1992). Manipulation of the source sink balance by cutting should alleviate any photosynthetic downregulation and allow the higher carbon storage in the roots of Ryegrass stands to facilitate speedy development of a new canopy (Nijs & Impens 1993).

This study will, therefore attempt to address a number of key questions and evaluate whether or not the results validate or refute previous findings obtained in a number of experimental environments, for *Lolium perenne*.

- 1) Does prolonged growth in elevated  $C_a$  stimulate  $A$  throughout the course of a day and at different stages of growth. As inferred from diurnal course's of measurement.
- 2) Does prolonged growth in elevated  $C_a$  induce an acclimatory response in the photosynthetic apparatus in the field, as inferred from the construction of the  $A/C_i$  response curve. Three experiments have been designed to allow acclimation to be studied .1) Measurements will be taken at varying times through the regrowth cycle in all plots, as well as, 2) on plots of *L.perenne* cut to two different heights within each

low nitrogen plot. This will allow the effects of changing source sink balances to be studied at a single point in time. Removing the effects of variable climate conditions from the equation. The low nitrogen plots only have been chosen as it is believed that acclimation is most likely to be apparent in these plots.3) The interaction between leaf age, shading and acclimation will be addressed by taking measurements on the same leaves at two points through the regrowth cycle.

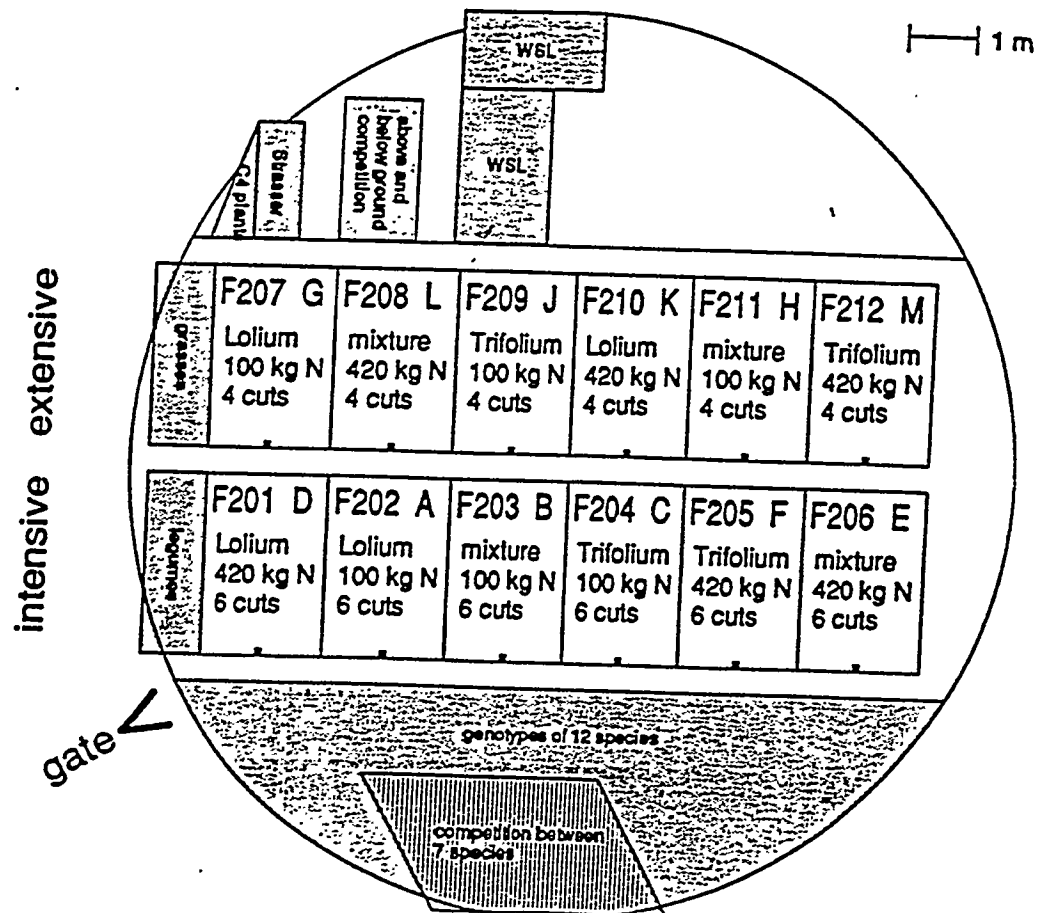
3) What are the implications for 1 and 2 above of variations in cutting regimes and nitrogen treatments, as methods of altering the source sink balance.

## 2. Materials and Methods.

### 2:1 The Site.

The ETHZ Eshikon research site utilises the Free Air Carbon Dioxide Enrichment (F.A.C.E.) system developed by Brookhaven National Laboratory. (Hendrey *et al* 1993, Lewin *et al* 1994). The FACE system at Eshikon consists of a 14 m diameter toroidal plenum below the ground. 32 vertical vent pipes protrude from below ground from the plenum at regular intervals. From these pipes the release of CO<sub>2</sub> is controlled by a central computer and governed by wind speed, wind direction and air temperature to maintain an elevated C<sub>a</sub> of 600 μmol mol<sup>-1</sup> within the FACE rings. There are six rings paired within three blocks, three control at ambient C<sub>a</sub> (355 μmol mol<sup>-1</sup>) and three FACE at elevated C<sub>a</sub> (600 μmol mol<sup>-1</sup>). Figure 1 details the layout of one of the six rings.

FIGURE 1.



Fumigation of the rings commenced in March this year and is scheduled to end in November and is switched on when leaf temperatures exceed 8°C. Elevated  $C_a$  was maintained within 10% of the target  $CO_2$  concentrations (540-660  $\mu\text{mol mol}^{-1}$ ) for 89 - 94% of the time, measured over one minute integrals during 1994 (Nagy *et al* 1995).

Perennial Ryegrass *Lolium perenne* L (cv Bastion) is in its third year of growth in pure stands in each of the six rings. Variation in growing conditions is achieved by the interaction of three main variables.

- 1)  $CO_2$  level control (355  $\mu\text{mol mol}^{-1}$ ) or FACE (600  $\mu\text{mol mol}^{-1}$ )
- 2) Level of Nitrogen application (140 or 540 kg / N ha), in the form of  $NH_4 NO_3$ .
- 3) Cutting regime ( 8 cuts during the growing season). Every 4 weeks commencing 24.3.95 and ending 2.11.95.

Additional variants influencing growth of the crop are provided by fluctuations in atmospheric and meteorological conditions.

## 2:2. Materials.

Data collected consisted exclusively of leaf gas analysis measurements, obtained non destructively, from which the main variables  $A$  and  $C_i$  were derived based on the equations of Von Caemmerer & Farquar (1981) and Harley (1992) to standardise to 25 °C. Photosynthesis was measured using an Open Combined Infra-Red Gas Analysis System (CIRAS 1 model 1.4, PP Systems ,Hitchin , Herts UK). in conjunction with a Parkinson Leaf Cuvette (Model 1.1, PP Systems, Hitchin, Herts UK). Weekly calibration of the CIRAS was made for  $CO_2$  at 600  $\mu\text{mol mol}^{-1}$  and water

vapour at 10 mb. CO<sub>2</sub> calibration was made against a gravimetric standard CO<sub>2</sub> concentration (Carbagas Zurich), whilst a known humidity was generated with a water vapour generator (Type WE 600, Analytical Development Co, Hoddesden UK).

Whilst leaf gas exchange measurements were always made with the upper leaf surface displayed in the top of the leaf chamber, the type of leaf chosen and the position along the leaf at which measurements were taken differed and depended on the experiment being conducted. For most of the sampling period the plants were in the vegetative stage of growth, it was only towards the end of the cutting period that some entered the reproductive stage of growth. Care was taken not to sample these plants where substantial non photosynthesising sinks had developed. The selection and measurement criteria will therefore be discussed at the appropriate place. Fundamental to analysis was the estimation of leaf area in the circular leaf cuvette of 1.8 cm diameter. This was achieved by multiplying the width of the leaf by the diameter of the chamber and assuming a rectangle was formed. This assumption results in the overestimation of leaf area to a tiny degree. Which was considered negligible when compared to the ability of the CIRAS to only accept leaf areas calculated to 1 mm<sup>2</sup> accuracy.

For the construction of A/Ci curves saturating photosynthetic photon flux densities were required, so a quartz iodide lamp was attached to the upper surface of the leaf chamber to provide a constant light saturating photon flux density of 750 μmol m<sup>-2</sup> s<sup>-1</sup>.

### 2:3. Methods.

#### 2:3:1. Diurnal Measurements.

Gas analysis measurements were performed every two hours in each of the six rings to determine the diurnal course of, photosynthetic leaf CO<sub>2</sub> uptake per unit area (A). This was done in 2 hour runs between 6.00 and 18.00 hrs for both the low and high nitrogen *Lolium* plots for the 8 cut regime on the 24<sup>th</sup> May and the 20<sup>th</sup> June, whilst on the 9<sup>th</sup> July the last run started at 20.00 hrs. For both nitrogen treatments in each ring three leaves were sampled every two hours. Measurements were made at the growth C<sub>a</sub> (either 355 or 600 μmol mol<sup>-1</sup>) and in the climatic conditions prevailing at the time, the only restriction on measurements was enforced by rain, yet days with a minimum of cloud cover were considered preferable.

Leaves chosen for the measurements were consistently the youngest fully expanded leaf, namely those leaves nearest to the top of the canopy bearing a ligule. Measurements were made half way along these leaves with the leaf cuvette always held horizontally parallel to the ground. Table 1 details the days when measurements were taken whilst the rings were sampled in the following order, C2, F1, C1, F2, F3, C3. See Appendix 1 for ring locations.

**TABLE 1.** Dates of diurnal measurements, nitrogen treatments sampled and the canopy regrowth status.

	DATE.	NITROGEN	DAYS FROM CUT
DIURNAL 1	24 <sup>th</sup> MAY 1995	HIGH & LOW	9 AFTER
DIURNAL 2	20 <sup>th</sup> JUNE 1995	HIGH & LOW	8 AFTER
DIURNAL 3	9 <sup>th</sup> JULY 1995	HIGH & LOW	1 BEFORE

A two way anova with replication was used to statistically test the results obtained during the midday hours of peak insolation and temperature, between 11.00 hrs and 15.00 hrs. Compared with the results gained earlier in the morning and in the evening.

### 2:3:2. A / C<sub>i</sub> determination.

The construction of light saturated leaf CO<sub>2</sub> uptake per unit area ( $A_{sat}$ ) to intercellular CO<sub>2</sub> concentration ( $C_i$ ) plots was made by gas analysis measurements. For both the low and high nitrogen treatments of the eight cut regime A /  $C_i$  curves were constructed for two leaves in each treatment in all six rings from a sequence of six  $C_a$  levels.

For ambient plots the sequence was : 355, 250, 150, 50, 600, 900  $\mu\text{mol mol}^{-1}$ .

For F.A.C.E plots the sequence was : 600, 355, 250, 150, 50, 900  $\mu\text{mol mol}^{-1}$ .

Adherence to these sequences was designed to negate any problems associated with the activation state of Rubisco being raised above naturally occurring levels.

Measurements were taken at  $C_a$  concentrations within + or - 40  $\mu\text{mol mol}^{-1}$  of the above stated values except at the growth  $C_a$  when a divergence of only + or - 15  $\mu\text{mol mol}^{-1}$  was permitted. For all measurements photosynthetic photon flux densities that were light saturating were used and measurements were concluded before 15.00 hrs to reduce the possibility of feedback inhibition of photosynthesis by carbohydrate accumulation and cytosolic inorganic phosphate limitation. The criteria for leaf sampling was the same as that already detailed for the Diurnal measurements (p 14).

Four sets of measurements were obtained in total, one set was made either side of the

cuts on the 12<sup>th</sup> June and the 10<sup>th</sup> July. This enabled analysis before and after the two cuts as well as during the regrowth period between the cuts.

From the plots sampled, values of  $V_{c,max}$  and  $J_{max}$  were fitted. These values represent *in vivo* quantifications of the maximum rate of carboxylation ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) and hence Rubisco activity ( $V_{c,max}$ ) and the maximum light saturated rate of electron transport ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) or the maximum capacity for RubP regeneration ( $J_{max}$ ). These values were achieved by fitting the data of McMurtrie & Wang (1993) to the  $A_{sat}$  values achieved.

Three way analysis of variance was performed on the  $V_{c,max}$  and  $J_{max}$  values obtained from each individual A/Ci curve obtained to highlight any significant differences in those values caused by CO<sub>2</sub>, nitrogen and the date (before or after cutting).

**Dates of experiments: 22/23 May, 6/11 June, 19/21 June, & 1 July.**

### **2:3:3. Diurnal A / C<sub>i</sub> determination.**

An attempt to detail the diurnal nature of fluctuating  $V_{c,max}$  and  $J_{max}$  values was made by constructing A / C<sub>i</sub> curves for two rings, one F.A.C.E and one ambient in the time window between 15.00 and 17.00 hrs when feedback inhibition of photosynthesis would be expected to exert an influence on the  $V_{c,max}$  and  $J_{max}$  values achieved. The values gained from the two rings sampled in the afternoon were compared to the values achieved in the morning in the same rings and on the same two tagged leaves in both the low and high nitrogen 8 cut regime plots. Leaf selection and the sequences of

$C_e$  exposure levels to construct the  $A / C_i$  responses have already been detailed (pgs 14 & 15).

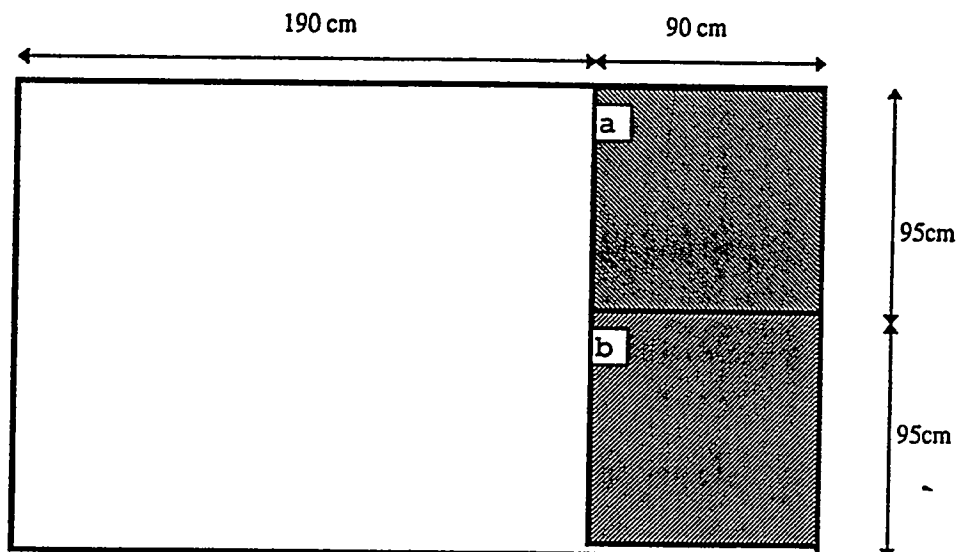
Dates of experiments: 23 May, & 26 June.

#### 2:3:4. Manipulation Experiment.

The size of the area cut provided a central area 30 by 35 cm's for sampling where border and shade effects would be at a minimum. (Figure 2).

After allowing a regrowth period of between 3 and 5 days measurements were taken on two leaves in the cut and uncut section of each of the six rings (NB for the measurements on the 6<sup>th</sup> of June only four rings were sampled due to constraints imposed in the other two rings by other experiments, they were C2, F2, C3, F3).

**FIGURE 1.** Low nitrogen 8 cut *L.perenne* plot. Shaded areas denote areas cut in manipulation experiment. Area a cut on 2<sup>nd</sup> June, area b cut on 30<sup>th</sup> June.



The leaves chosen for the measurements were chosen in a way that aimed to ensure uniformity of age. This involved taking measurements on leaves that were not fully developed, (None were in the cut section) 5 cm's from the leaf juncture with the stems high in the canopy as possible for the uncut plot.  $A / C_i$  curves were then constructed in the aforementioned manner. An assumption that rates of leaf expansion were uniform was fundamental to sampling leaves of the same age, in reality rates of expansion may differ.

**Dates of experiments: 8 June & 5 July.**

#### **2:3:5. Canopy Effects on $A / C_i$ response.**

In an effort to assess the implications of shading and leaf age on the  $A/C_i$  response of *Lolium perenne*,  $A/C_i$  curves were constructed at two time periods during canopy regrowth, firstly eight and then fourteen days into the regrowth period. The leaves used for the initial  $A/C_i$  measurements were tagged to allow recognition and re-measurement on the two subsequent measurement days, Whilst also ensuring that the leaves sampled were of the same age.

**Dates of experiment: 19/21 June & 27/28 June.**

### 3. Results

#### 3.1. Diurnal Measurements.

Studies of the diurnal nature of CO<sub>2</sub> and water vapour exchange undertaken on three separate days throughout June and July show substantial increases in A and slightly more variable, yet generally decreases in E for *L.perenne* grown at elevated C<sub>a</sub> with respect to that grown at ambient C<sub>a</sub> (Table 3 , & Figs 3-8).

**TABLE 3.** Relative enhancement (%) of A and E for FACE grown *L.perenne* over controls, over a diurnal course of measurements. On three separate days and two nitrogen treatments.

DATE	Days from cut	A % Enhancement		E % Enhancement	
		HIGH N	LOW N	HIGH N	LOW N
24.5.95	+9	30.51	39.35	-13.40	-15.14
20.6.95	+8	26.51	38.14	-8.84	-10.03
9.7.95	-1	33.05	44.95	+4.98	-16.01

A stimulation in A was recorded in the elevated plots during every two hour period and in both nitrogen treatments over the course of the measurement days. This stimulation was irrespective of the period into regrowth of the crop. Expressed as a percentage over the whole study period this stimulation in A ranged from 26.51 % to 44.95 %.

Greatest total enhancement was shown to correspond to periods of the highest Photosynthetic photon flux densities (PPFD) and higher temperatures. These conditions occurred between 11:00 hrs and 15:00 hrs, a period of time when there was a significant difference between A in the FACE and control plots (Table 4). During the

**FIGURE 3.** Diurnal Assimilation and Evapotranspiration rates for Control and FACE grown *L.perenne*. Low nitrogen treatment 23.5.95, 9 Days after the cut.

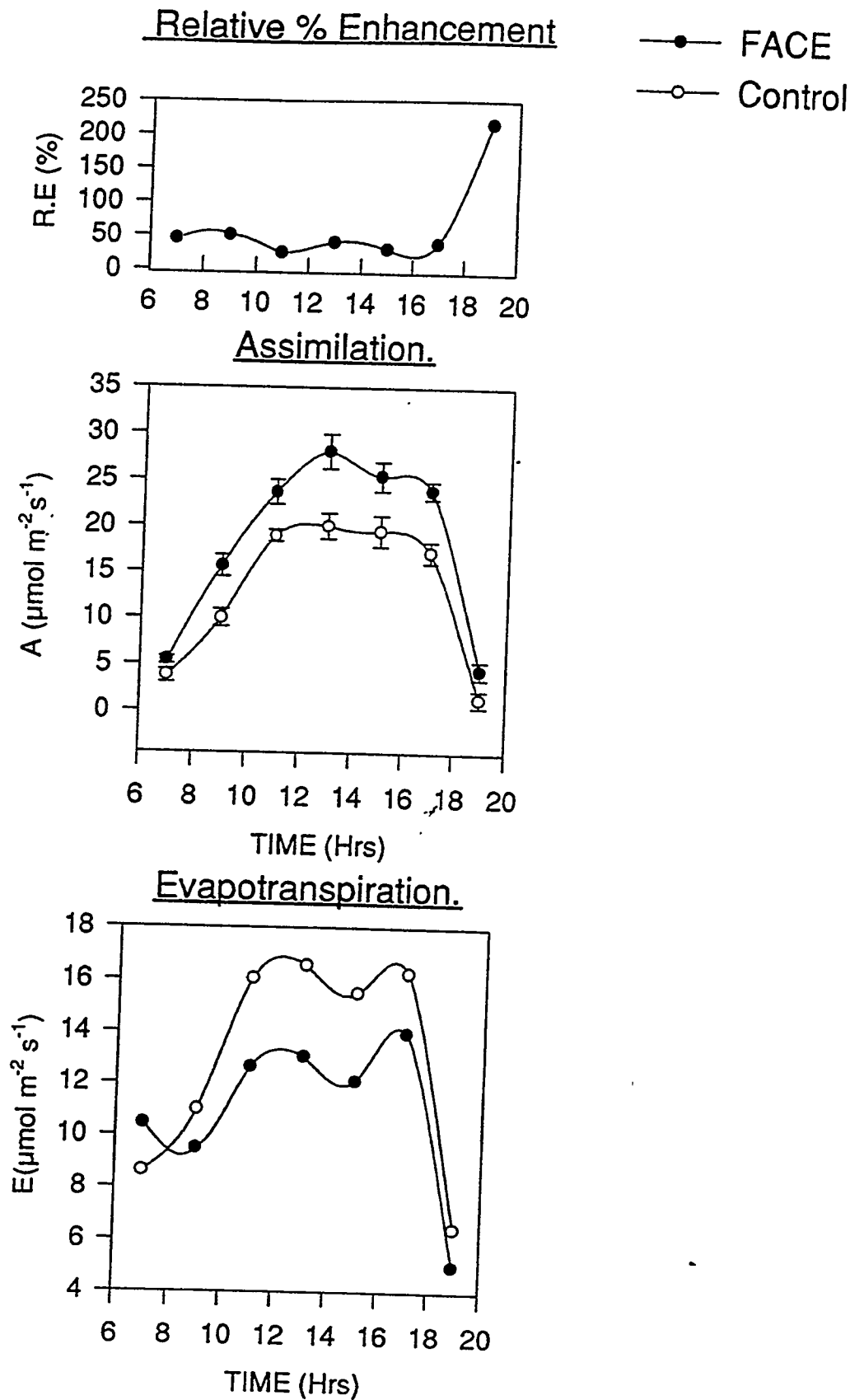
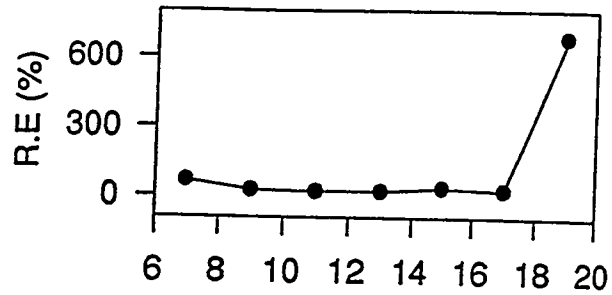
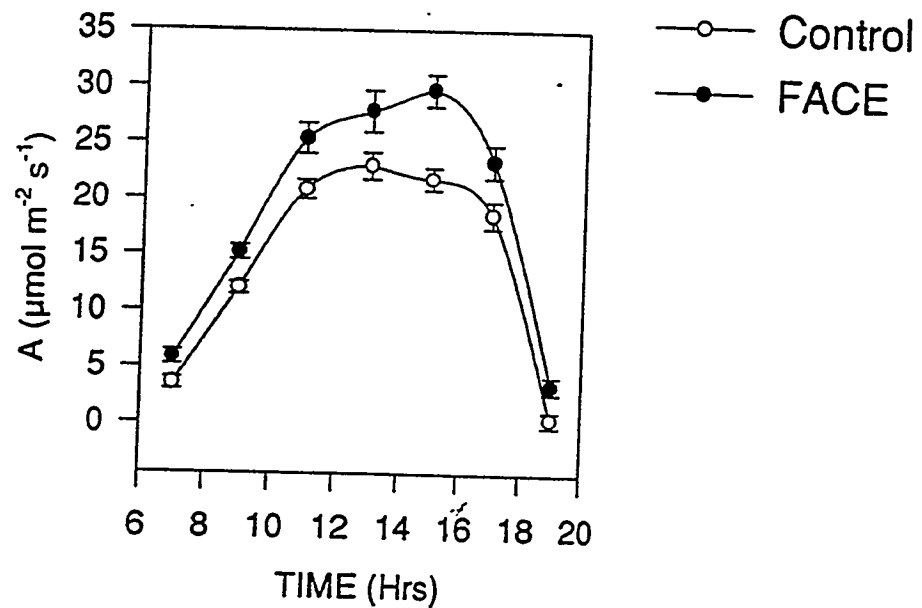


FIGURE 4. Diurnal Assimilation and Evapotranspiration rates for Control and FACE grown *L. perenne*. High nitrogen treatment 23.5.95, 9 Days after the cut.

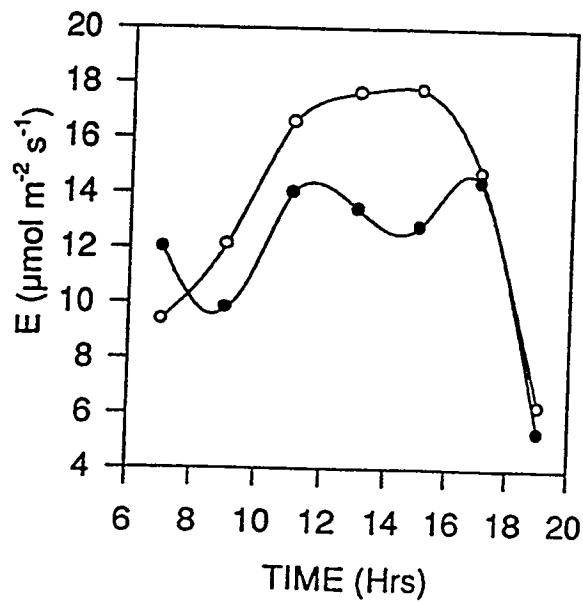
**Relative % Enhancement of A.**



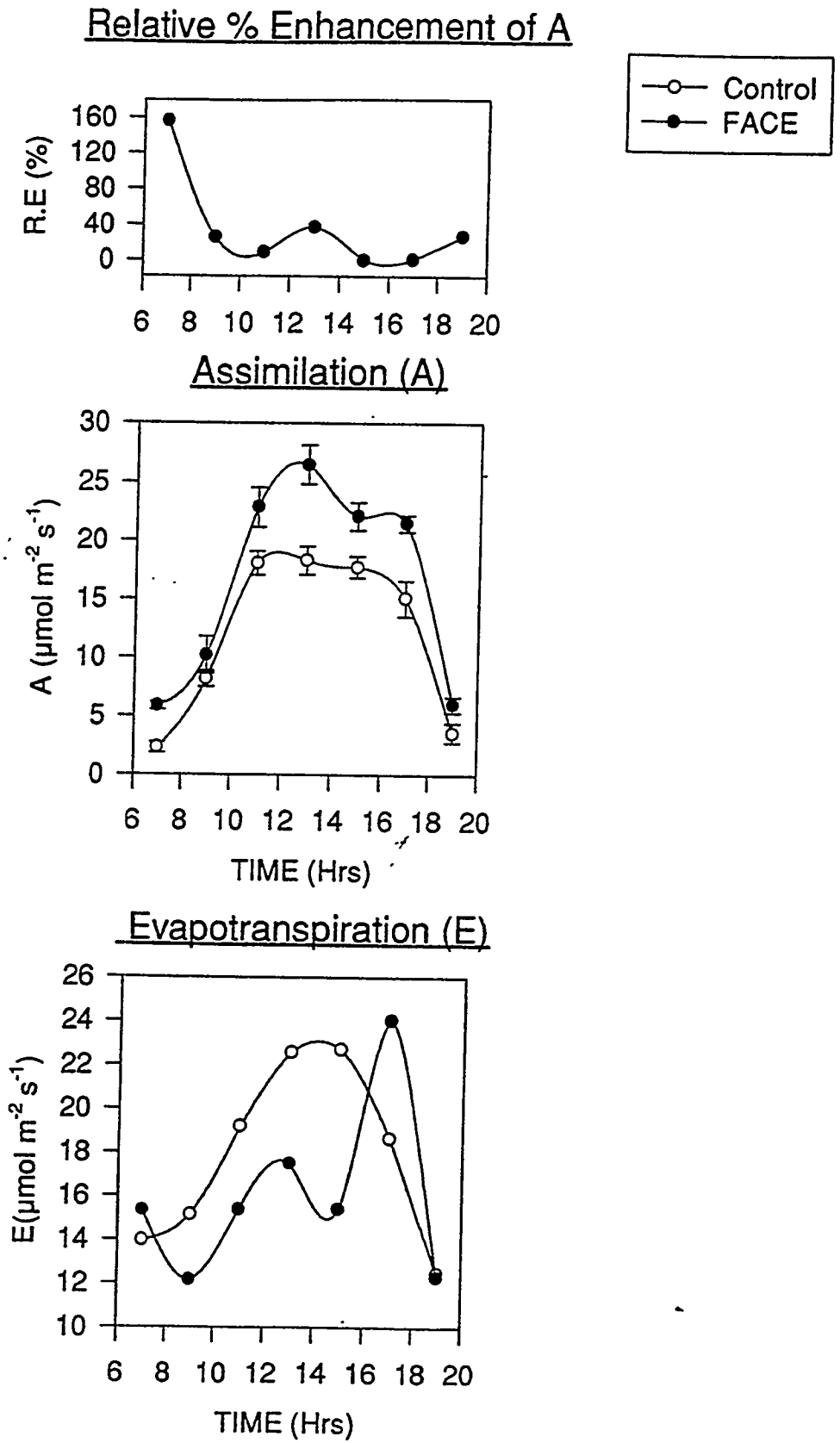
**Assimilation (A).**



**Evapotranspiration (E).**

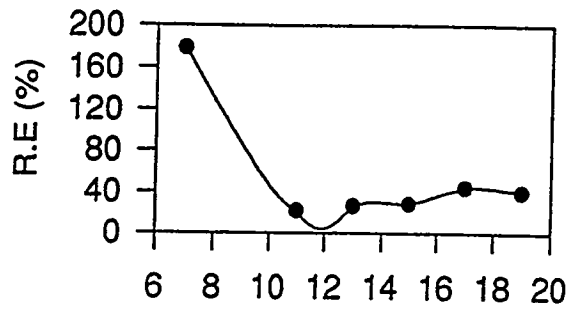


**FIGURE 5.** Diurnal Assimilation and Evapotranspiration rates for Control and FACE grown *L.perenne*. Low nitrogen treatment. 20.6.95 8 Days after cut.

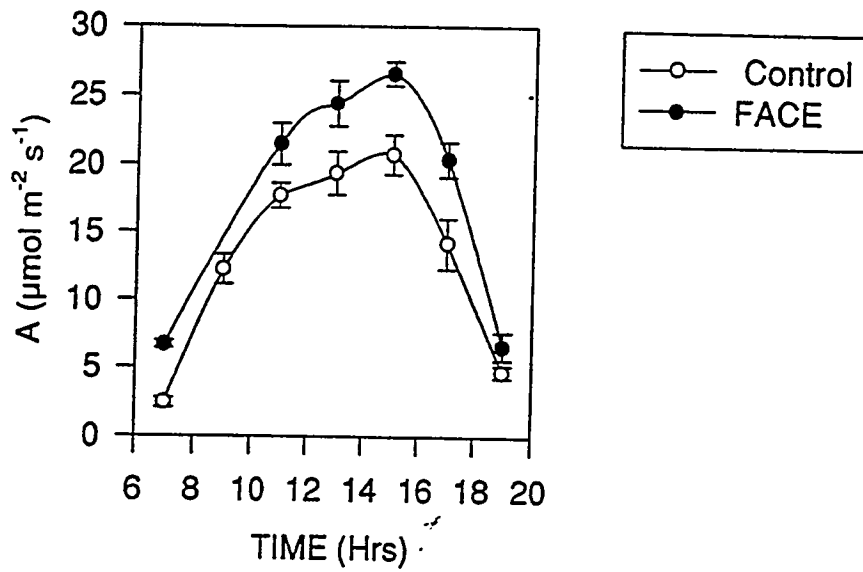


**FIGURE 6.** Diurnal Assimilation and Evapotranspiration rates for Control and FACE grown *L.perenne*. High nitrogen treatment. 20.6.95 8 Days after cut.

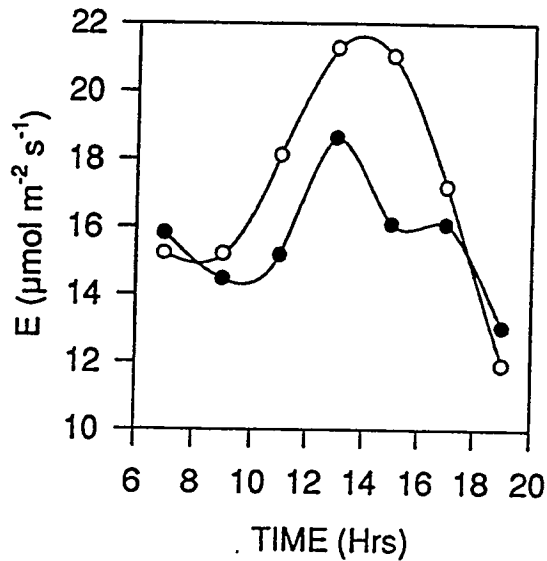
Relative % Enhancement of A.



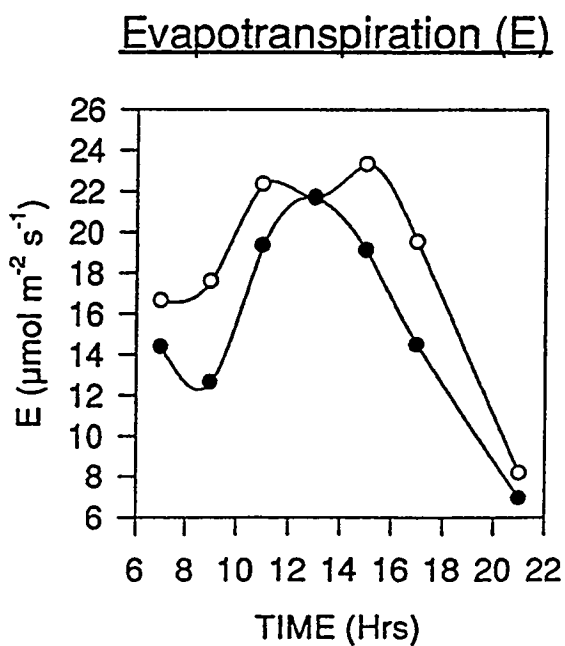
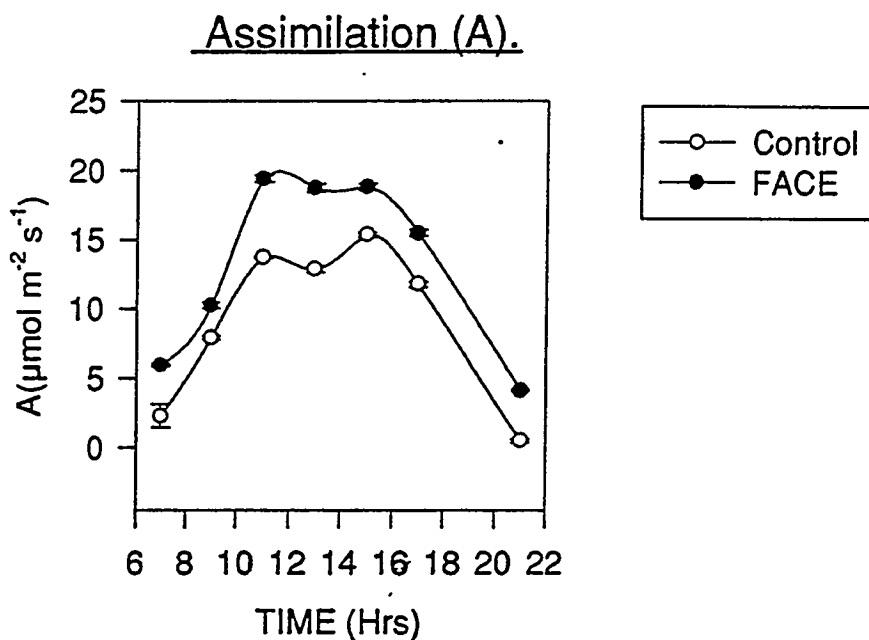
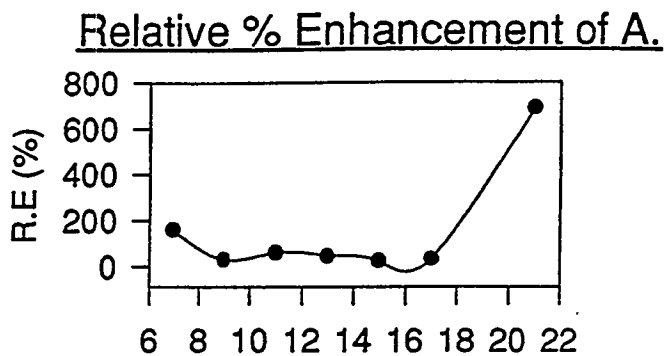
Assimilation (A)



Evapotranspiration (E).

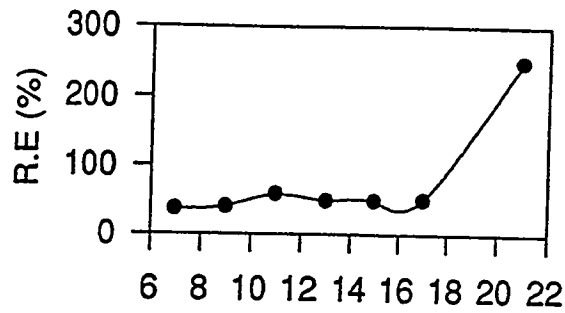


**FIGURE 7.** Diurnal Assimilation and Evapotranspiration rates for Control and FACE grown *L. perenne*. Low nitrogen treatment. 9.7.95. 1 Day before cut.

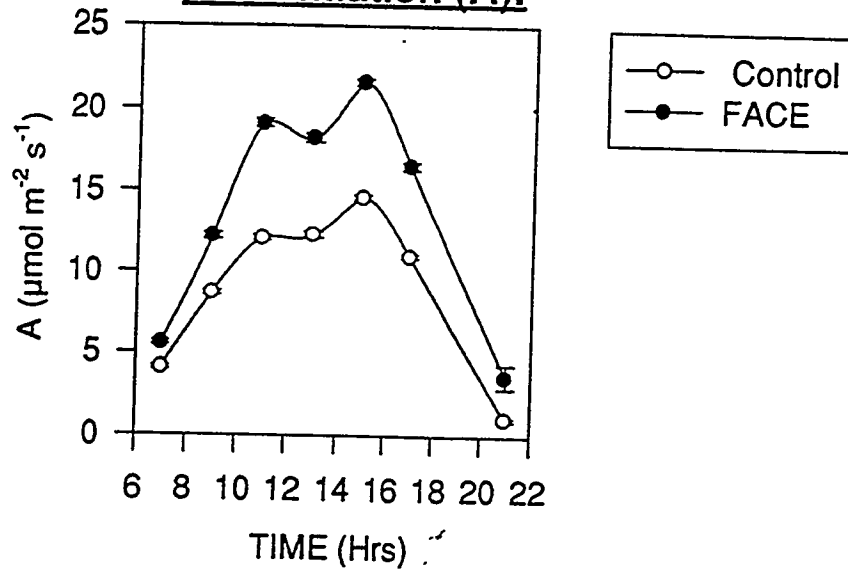


**FIGURE 8.** Diurnal Assimilation and Evapotranspiration rates for Control and FACE grown *L. perenne*. High nitrogen treatment. 9.7.95. 1 Day before cut.

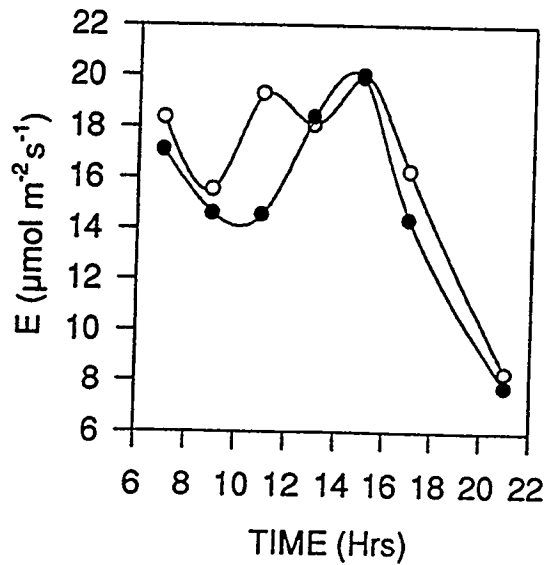
**Relative % Enhancement of A.**



**Assimilation (A).**



**Evapotranspiration (E).**



hours of measurement preceding and following this time period when temperatures and PPFD were lower, a stimulation of A was still evidenced in the FACE plots on each occasion yet the stimulation was not significant (Table 5).

**TABLE 4.** Significant levels of variation in A due to CO<sub>2</sub> and Nitrogen between 11:00 & 15:00

VARIABLES	25 <sup>th</sup> MAY 1995	20 <sup>th</sup> JUNE 1995	9 <sup>th</sup> JULY 1995
Nitrogen	P=0.052, F=33.45	P=0.50, F=0.51	P=0.84, F=0.044
CO <sub>2</sub>	P<0.05, F=5.20	P<0.05, F=23.47	P<0.05, F=59.32
Interaction	P=0.982, F=0.001	P=0.695, F=0.166	P=0.31, F=1.86
n	3	3	3

**TABLE 5.** Significant levels of variation in A due to CO<sub>2</sub> and Nitrogen before 11:00 and after 15:00

VARIABLES	25 <sup>th</sup> MAY 1995	20 <sup>th</sup> JUNE 1995	9 <sup>th</sup> JULY 1995
Nitrogen	P=0.874, F=0.026	P=0.986, F=0.0003	P=0.831, F=0.047
CO <sub>2</sub>	P=0.363, F=0.893	P=0.384, F=0.815	P=0.224, F=1.643
Interaction	P=0.853, F=0.036	P=0.915, F=0.012	P=0.991, F=0.0001
n	4	4	4

During neither of the two time periods on any of the dates was there a significant effect of nitrogen treatment on the stimulation of A in the FACE plots (Tables 4 & 5). Indeed in both the FACE and control plots there was little difference between peak A values and mean diurnal A values between nitrogen treatments. Not

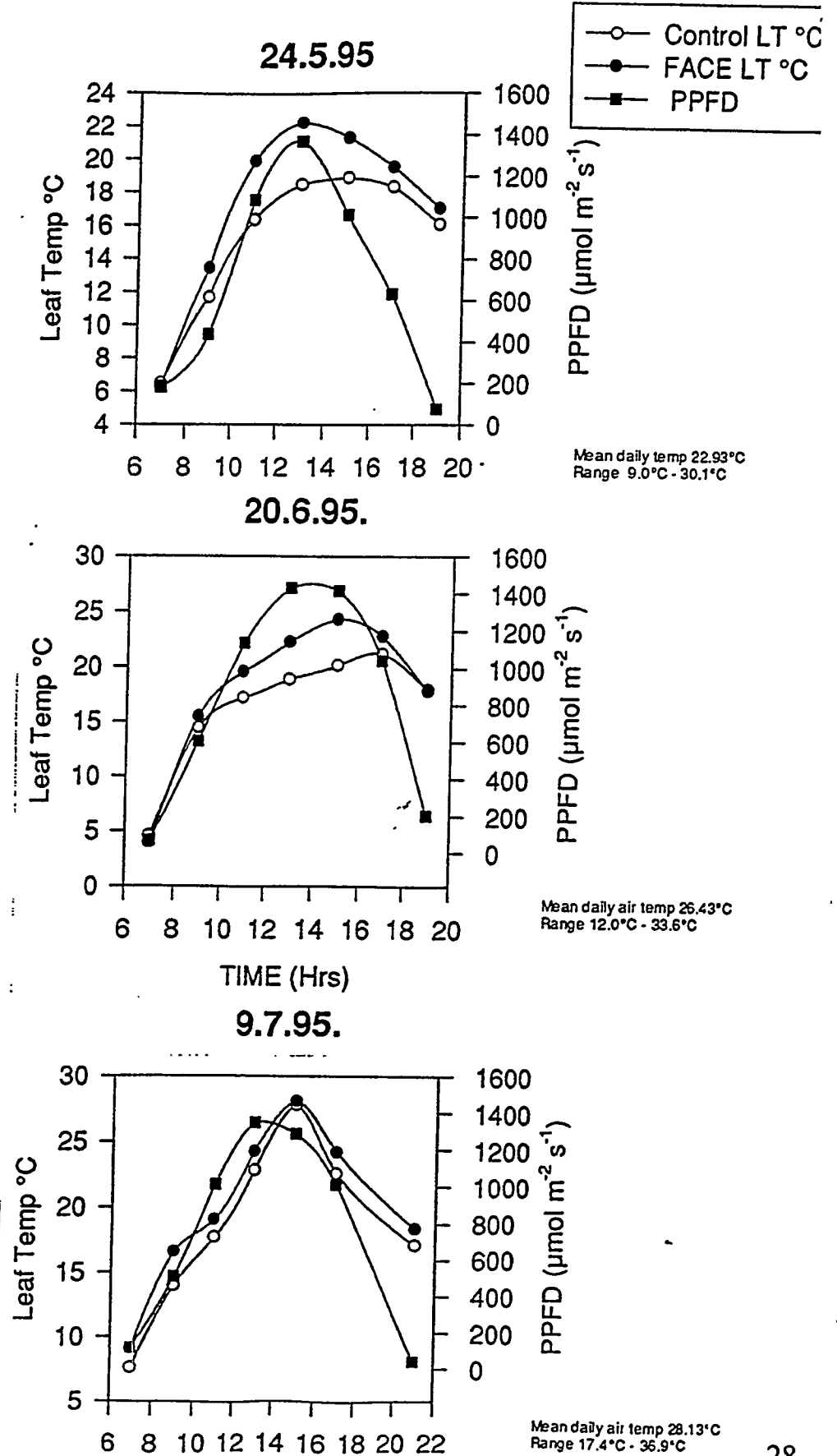
withstanding this fact on all three measurement days the greatest percentage enhancement of A was between the low nitrogen control and FACE plots (Table 3 Figs 3, 5 & 7), the largest being 44.95% on the 9<sup>th</sup> July. This day also saw the highest percentage enhancement between the high nitrogen FACE and control plots, 33.05 %. The 9<sup>th</sup> July was 27 days into the regrowth period, a time when some degree of sink limitation to photosynthesis would be expected compared to the previous two measurement days when the swards were less than 10 days into their regrowth. Whilst the percentage enhancement was greatest for both nitrogen treatments on the 9<sup>th</sup> July the actual values of A obtained were substantially lower than on the other two dates. The average diurnal value of A gained on the 9<sup>th</sup> July was 26.6% lower and 17.89 % lower than the values observed in the control and FACE plots respectively on the 20<sup>th</sup> June and 32.8 % and 36.4 % lower than the values gained on the 23<sup>rd</sup> May in the control and FACE plots.

As well as being further into the regrowth period the 9<sup>th</sup> July experienced the highest mean daily and maximum air and leaf temperatures, as well as PPFD comparable to the other two measurement days. (Figs 9) As a consequence it is likely that photoinhibition and carbohydrate feedback were limiting photosynthesis to some degree as well as longer term sink limitation.

With the exception of the low nitrogen treatment on the 9<sup>th</sup> July, *L.perenne* grown in the FACE plots experienced reduced rates of evapotranspiration expressed as a percentage over the course of the day (Table 3). These reductions ranged from 8.84 to 16.01% and were uniformly higher in the low nitrogen treatments, whilst also highest between 11:00 and 15:00 hrs when insolation and temperature were at there

**FIGURE 9.** Microclimate data for the three diurnal measurement days.

Control and FACE leaf temperatures and PPFD.



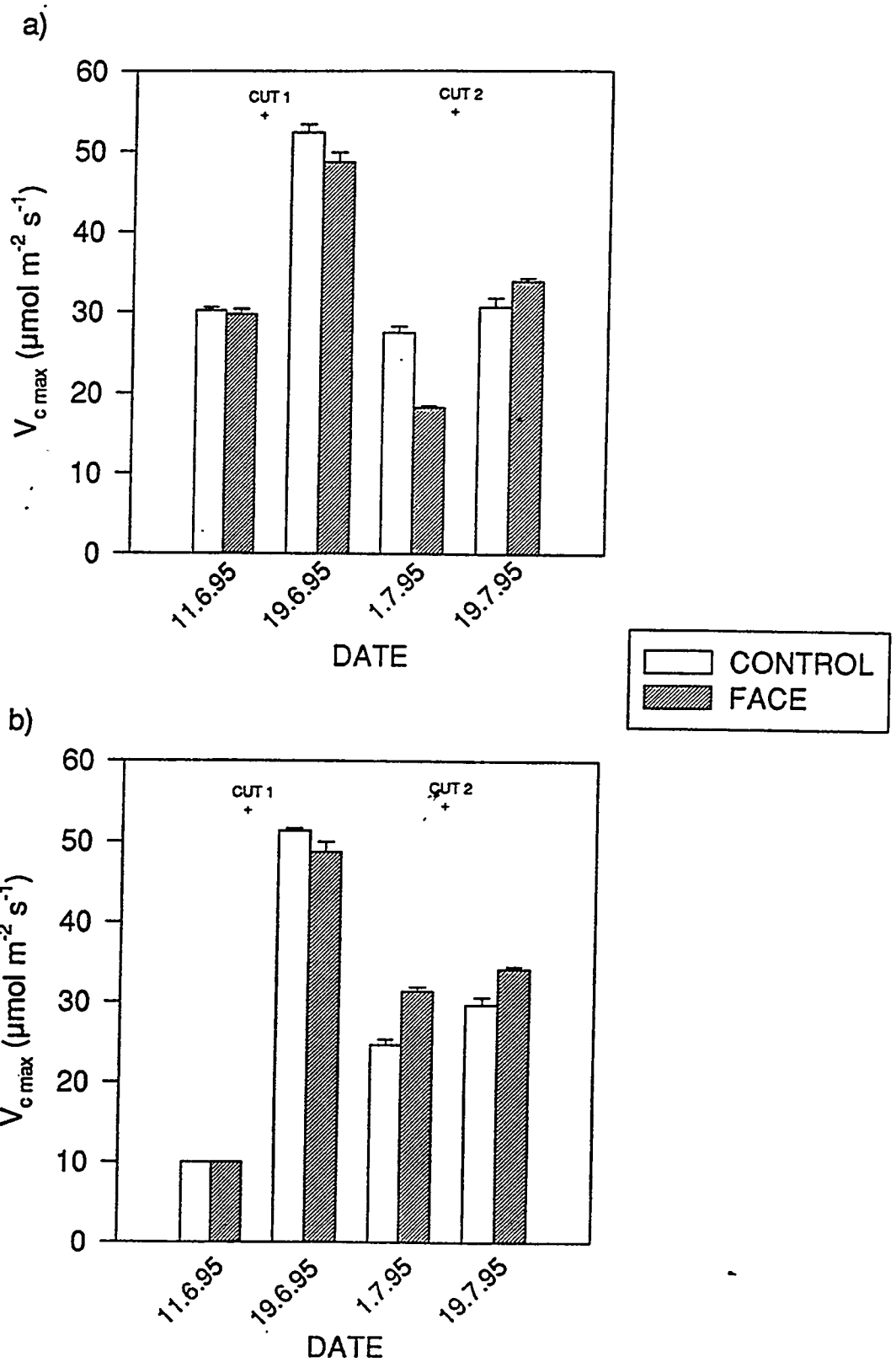
highest levels, in both nitrogen treatments (Figs 3-8). A time period when growth in elevated  $C_a$  seemed to confer a clear advantage in terms of water conservation. Only on the 9<sup>th</sup> July was this not the case when the aforementioned weather conditions and possibly supraoptimal temperatures, seemed to have induced stomatal closure in the Control plots (Figs 7 & 8) reducing midday E to levels of the FACE grown plants.

A manifestation of these reduced rates of evapotranspiration from the FACE grown *L. perenne* is a consistent increase in the leaf temperatures of the FACE plots over the whole measurement period on each of the measurement days. This is attributable to the reduced rate of latent heat loss through evaporation for the FACE leaves and is again greatest during the 11:00 to 15:00 hrs time period. Except on the 9<sup>th</sup> July when the aforementioned midday early afternoon stomatal closure has reduced latent heat loss in the Control leaves (Fig 9).

### 3:2. A/Ci Results.

Values of  $V_{c,max}$  as indications of Rubisco activity and carboxylation efficiency were shown to increase significantly after a cut on two separate occasions in both control and FACE rings at both low and high nitrogen treatments (Table 6, Fig 10). Whilst the differences in  $V_{c,max}$  were significant between the days before and after both cuts, the increase was far smaller numerically after the second cut in both control and FACE low and high nitrogen treatments (Fig 10). The significance of the  $V_{c,max}$  values before and after the cut is not mirrored by any significant differences in  $V_{c,max}$  attributable to the effects of  $CO_2$  treatment (TABLE 6). Only with respect to nitrogen treatments for measurements around the first cut is another significant difference in  $V_{c,max}$  values observed (Table 6).

FIGURE 10. Variations in  $V_c$  max , Pre, Post and Intra cuts for Control and FACE grown *L.perenne*, eight cut regime.  
a) Low Nitrogen, b) High Nitrogen.



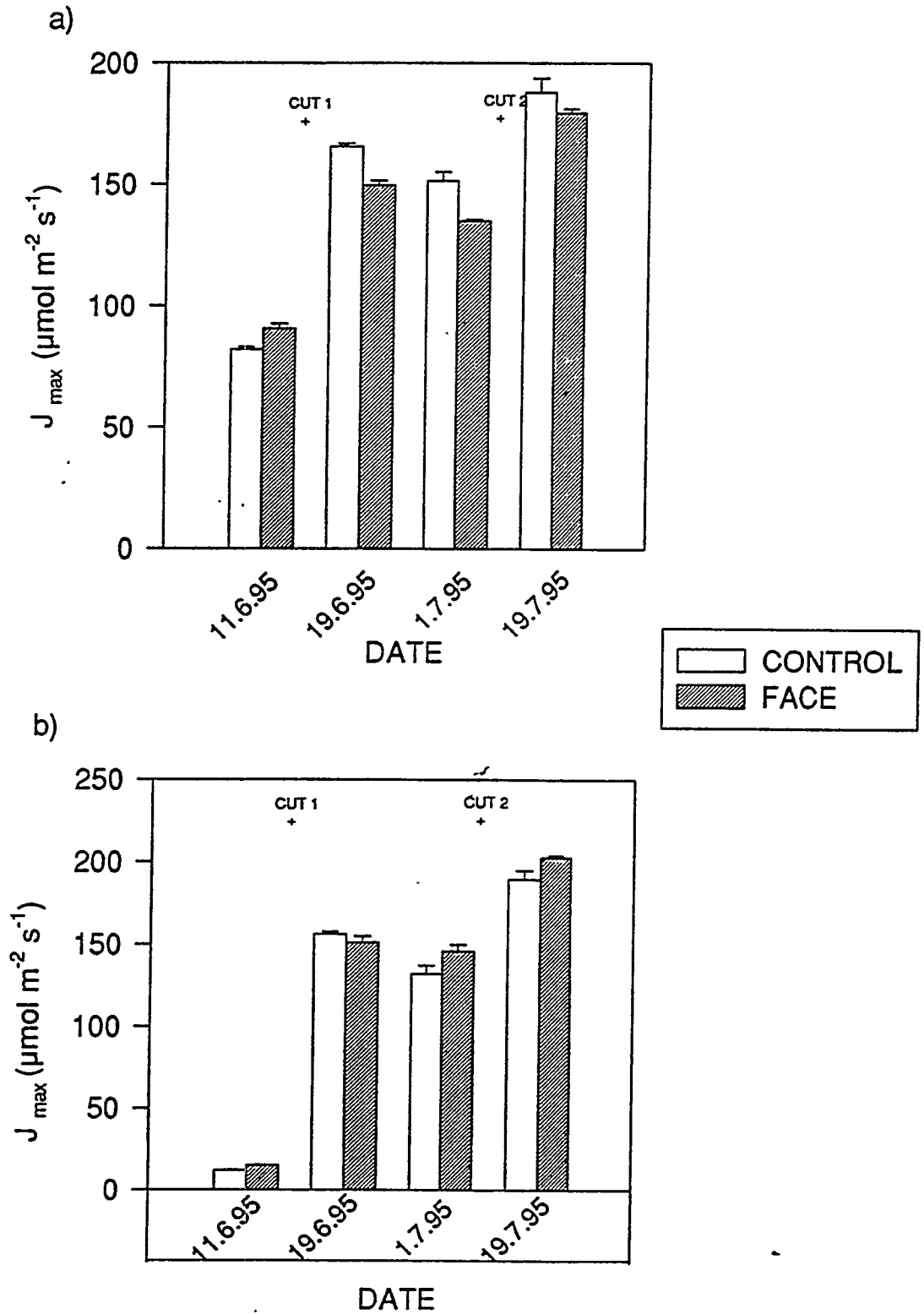
**TABLE 6.** Significant variation in  $V_{c\max}$  and  $J_{\max}$  values due to CO<sub>2</sub> nitrogen treatment and measurement day pre or post cut.

VARIABLES	CUT 1 $V_{c\max}$	CUT 2 $V_{c\max}$	CUT 1 $J_{\max}$	CUT 2 $J_{\max}$
Day	P<0.05, F=90.2	P<0.05, F=11.2	P<0.05, F=337	P<0.05, F=14
CO <sub>2</sub>	P=0.53, F=0.40	P=0.42, F=0.67	P=0.96, F=0.00	P=0.99, F=0.0
Day*CO <sub>2</sub>	P=0.65, F=0.21	P=0.33, F=1.01	P=0.43, F=0.63	P=0.9, F=0.02
Nitrogen	P<0.05, F=7.06	P=0.21, F=1.63	P<0.05, F=21.7	P=0.85, F=0.04
Day*Nitrogen	P<0.05, F=9.23	P=0.21, F=1.62	P<0.05, F=40.6	P=0.52, F=0.4
CO <sub>2</sub> *Nitrogen	P=9.0, F=0.01	P=0.43, F=4.39	P=0.28, F=1.20	P=0.29, F=1.1
Day*CO <sub>2</sub> *N	P=0.97, F=0.00	P=0.07, F=3.56	P=0.21, F=1.63	P=0.86, F=0.03
n	6	4	6	6

The effect of cutting is just as pronounced for  $J_{\max}$  which represents the maximum rate of electron transport and as such RubP regeneration. Again there was a highly significant increase in  $J_{\max}$  values after each cut in both control and FACE plots and in both nitrogen treatments (Table 6, Fig 11). Again the increase was greatest after the first cut. Whilst there was no significant difference in  $V_{c\max}$  values attributable to CO<sub>2</sub> treatment. There was again a significant effect of nitrogen before and after the first cut.

Whilst CO<sub>2</sub> treatments confer no significant differences in either  $V_{c\max}$  or  $J_{\max}$ , there is a significant effect of nitrogen before and after the first cuts for both  $V_{c\max}$  and  $J_{\max}$ . The effect of nitrogen treatment appears enhanced when studying the regrowth period between the two cuts, from 19<sup>th</sup> June to 1<sup>st</sup> July. In the low nitrogen treatment

FIGURE 11. Variations in  $J_{max}$ , Pre, Post and Intra cuts for Control and FACE grown *L.perenne*, eight cut regime. a) Low Nitrogen, b) High Nitrogen.



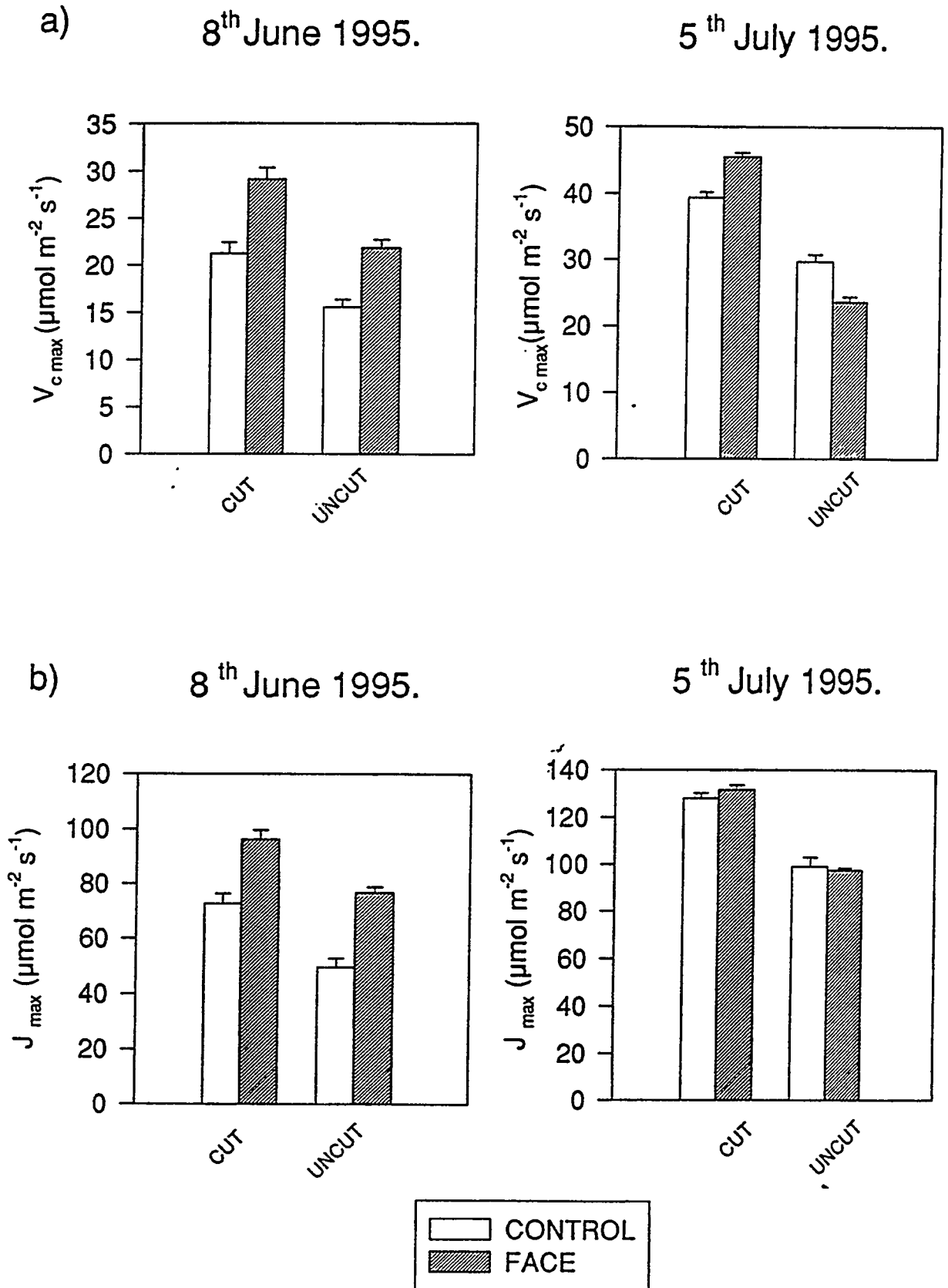
both  $V_{c,max}$  and  $J_{max}$  values are clearly lower in the face rings than the control rings. Immediately after the first cut  $V_{c,max}$  is 7% lower in the FACE rings than the control ring, prior to the second cut this figure rises to 33.58%. The same reduction is evidenced in the low nitrogen treatment for  $J_{max}$  values where the initial reduction in the FACE plots of 9.62% rises to 10.87% prior to the second cut (Fig 11). This effect is not observed in the high nitrogen treatments where  $V_{c,max}$  and  $J_{max}$  increase in the FACE plots relative to the control plots through the regrowth period.

### 3:3. Manipulation Experiment results.

The results of the manipulation experiment seem to confirm the findings of the A / Ci experiments. For both  $V_{c,max}$  and  $J_{max}$  values there was a significant difference in the values obtained in the uncut and cut plots during the second experiment on the 5<sup>th</sup> July 1995 when all six rings were sampled. With a significant enhancement of both sets of values occurring in both FACE and control plots in the cut areas. (Table 7, Fig 12). Also in agreement with the A / Ci findings is the lack of significant effect of  $CO_2$  treatment on the results obtained on either day (Table 7, Fig 12).

Analysis of the relative enhancements of  $V_{c,max}$  and  $J_{max}$  between the two cutting treatments on both experimental dates also showed a lack of significant  $CO_2$  effect (Table 8). In fact, the results clearly show that the enhancement of  $V_{c,max}$  and  $J_{max}$  in the cut FACE plots over the control plots is mirrored in the uncut plots where in experiment 1 the enhancement remains and in experiment 2 where the reductions are not significant (Fig 12). Given that this experiment was conducted solely in the low nitrogen plots it would be here where acclimatory downregulation of photosynthesis in

**FIGURE 12.** Variations in a)  $V_c$  max & b)  $J$  max for *L. perenne* measured on the same day, for a cut plot 5 days into regrowth and an uncut plot 22 days into regrowth. In low nitrogen Control and FACE plots.



response to sink saturation would be expected to be most evident.

**TABLE 7.** Significance levels of variation in  $V_{c,max}$  and  $J_{max}$  due to  $CO_2$  and cutting on two days. Experiment 1, 8<sup>th</sup> June 1995. Experiment 2, 5<sup>th</sup> July 1995.

VARIABLES	$V_{c,max}$ Expt 1	$V_{c,max}$ Expt 2	$J_{max}$ Expt 1	$J_{max}$ Expt 2
$CO_2$	P=0.14, F=2.57	P=0.98, F=0.00	P=0.07, F=4.08	P=0.94, F=0.01
Cut/Uncut	P=1.33, F=2.59	P<0.05, F=14.3	P=0.12, F=2.87	P<0.05, F=6.8
Interaction	P=0.91, F=0.01	P=0.16, F=2.16	P=0.89, F=0.02	P=0.84, F=0.04
n	4	6	4	6

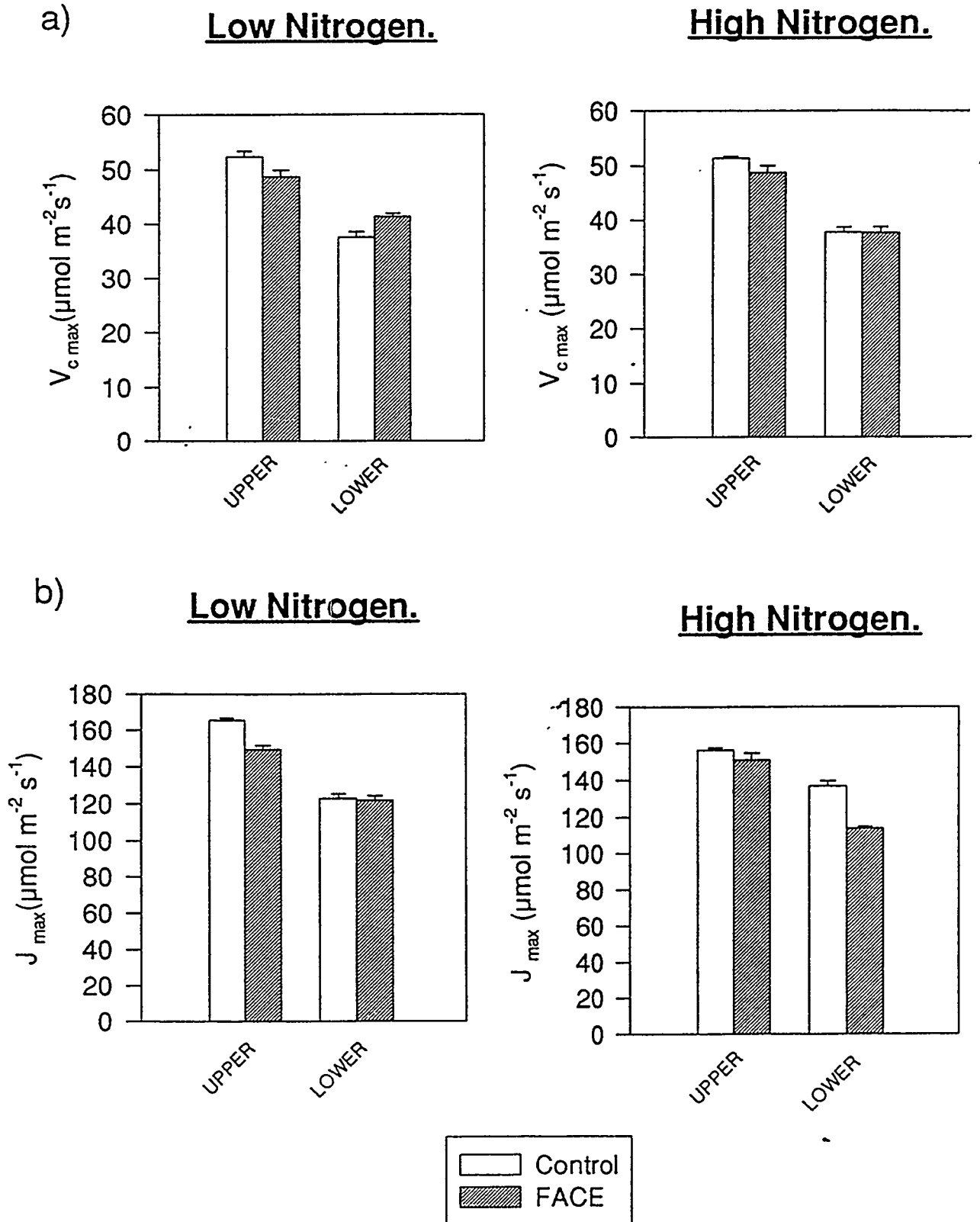
**TABLE 8.** Significant levels of variation in the relative enhancement of  $V_{c,max}$  and  $J_{max}$  due to  $CO_2$  and cutting on two days (As in Table 7).

VARIABLE	$V_{c,max}$ EXPT 1	$V_{c,max}$ EXPT 2	$J_{max}$ EXPT 1	$J_{max}$ EXPT 2
$CO_2$	P=0.74, F=0.07	P=0.18, F=2.19	P=0.43, F=0.72	P=0.55, F=0.39
n	4	6	4	6

### 3:4. Canopy Experiment Results.

As would be expected there was a significant downregulation of both  $V_{c,max}$  and  $J_{max}$  values in both the control and FACE plots when the leaves were older and positioned lower in the canopy, measured on the 27<sup>th</sup> June 1995, in contrast to the initial measurements taken on the 19<sup>th</sup> June 1995.. This effect was observed in both the low and high nitrogen treatments. (Table 9, Fig 13). The results do not show any significant influence of  $CO_2$  treatment (Table 9). Further analysis of the relative declines in  $V_{c,max}$  and  $J_{max}$  of the leaves on the second measurements day compared to

**FIGURE 13.** Variations in a)  $V_c$  max & b)  $J$  max measured on two occasions on the same leaves. When the leaves were in the upper canopy 7 days into regrowth. Then 8 days later when the same leaves are in the lower canopy.



the first again shows no significant influence of CO<sub>2</sub> treatment or nitrogen treatment on the values obtained. (Table 10).

**TABLE 9.** Significant variations in  $V_{c_{max}}$  and  $J_{max}$  due to CO<sub>2</sub> and leaf age / canopy position for two nitrogen treatments.

VARIABLES	$V_{c_{max}}$ Low N	$J_{max}$ Low N	$V_{c_{max}}$ High N	$J_{max}$ High N
CO <sub>2</sub>	P=0.98, F=0.00	P=0.44; F=0.63	P=0.76, F=0.09	P=0.27, F=1.3
Leaf Age	P<0.05, F=5.4	P<0.05, F=10.9	P<0.05, F=7.6	P<0.05, F=5.2
Interaction	P=0.45, F=0.59	P=0.49, F=0.49	P=0.78, F=0.08	P=0.48, F=0.5
n	6	6	6	6

**TABLE 10.** Significant variations in the relative percentage decline of  $V_{c_{max}}$  and  $J_{max}$  due to CO<sub>2</sub> and nitrogen for the same leaves through canopy regrowth.

VARIABLES	$V_{c_{max}}$	$J_{max}$
CO <sub>2</sub>	P=0.28, F=1.26	P=0.83, F=0.05
Nitrogen	P=0.94, F=0.01	P=0.51, F=0.46
Interaction	P=0.98, F=0.00	P=0.52, F=0.42
n	6	6

The only indications of an interactive effect of CO<sub>2</sub> and nitrogen treatments are when the relative percentage declines in the  $V_{c_{max}}$  and  $J_{max}$  values are calculated based on average figures from the leaves sampled. Then we see higher percentage decreases in the FACE treatments than the control treatments for each combination of variables except high nitrogen  $J_{max}$  values. This analysis also suggests that for the FACE treatments downregulation is higher in the high nitrogen treatment. (Table 11).

**TABLE 11.** Percentage declines in average  $V_{c\max}$  and  $J_{\max}$  values for two  $CO_2$  and two nitrogen treatments.

	$V_{c\max}$ FACE	$V_{c\max}$ Control	$J_{\max}$ FACE	$J_{\max}$ Control
Low Nitrogen	15.25	28.32	18.55	25.80
High Nitrogen	22.0	26.48	24.63	12.50

### 3:5. Diurnal A/Ci results.

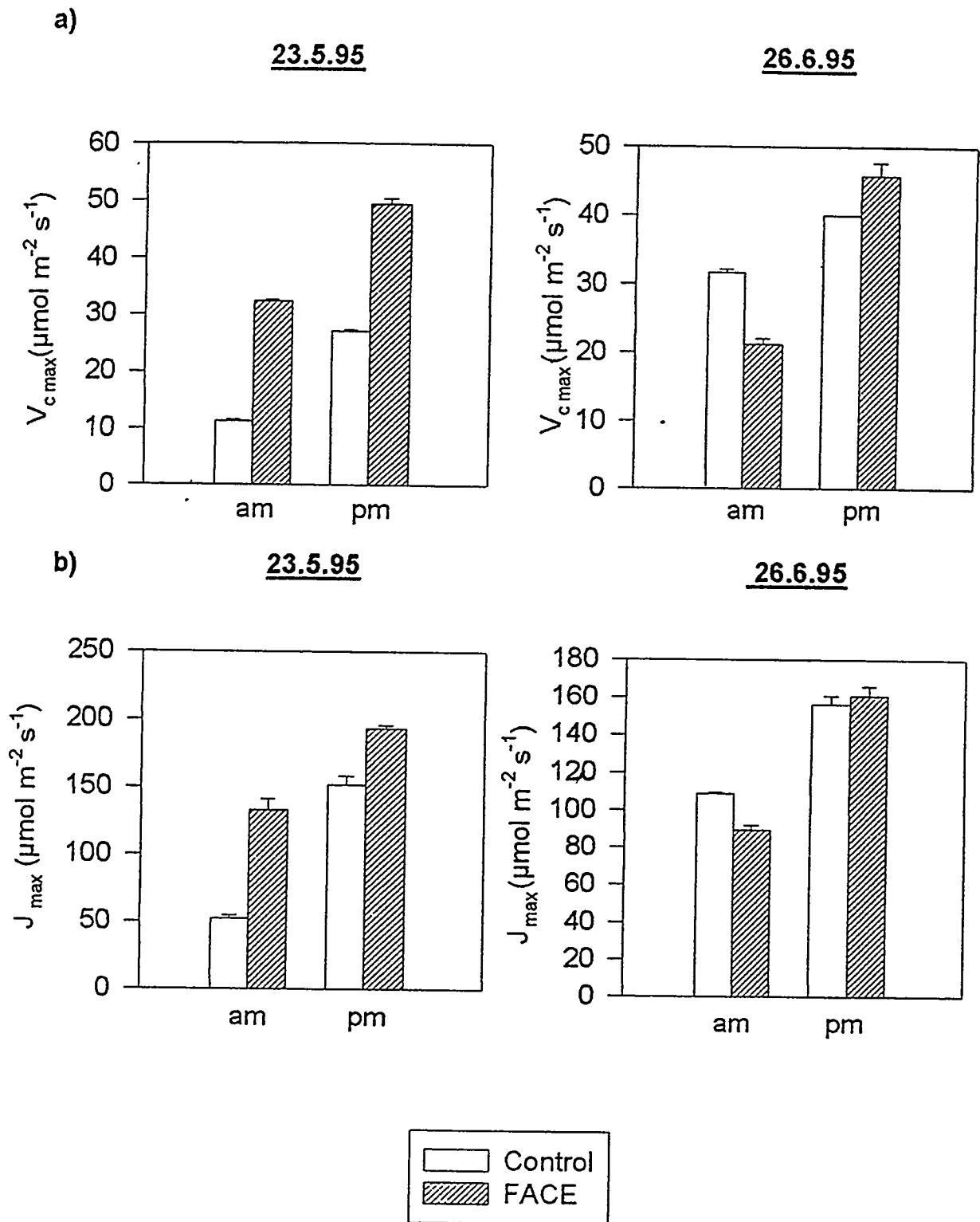
The results of this experiment show a significant difference in both  $V_{c\max}$  and  $J_{\max}$  values obtained before 08:00 hrs and after 15:00 hrs on both the experiment days (23<sup>rd</sup> May & 26<sup>th</sup> June). On both days there was a significant increase in the values obtained for both control and FACE plots (Table 12, Fig 14)

**TABLE 12.** Significant levels of variation in  $V_{c\max}$  and  $J_{\max}$  between two time periods. Before 08:00 and after 15:00 hrs on two separate measurement days .

Variables	$V_{c\max}$ 23.5.95	$J_{\max}$ 23.5.95	$V_{c\max}$ 26.6.95	$J_{\max}$ 26.6.95
$CO_2$	P=0.62,F=0.26	P=0.64,F=0.23	P<0.05,F=95.6	P<0.05,F=16.36
Time	P<0.05,F=12.5	P<0.05,F=15.65	P<0.05,F=55.3	P<0.05,F=27.91
Interaction	P=0.11,F=3.05	P=0.43,F=0.66	P=0.76,F=0.09	P=0.22,F=1.69
n	4	4	4	4

The results also showed a significant effect of  $CO_2$  on the 26<sup>th</sup> June with a significant increase in  $V_{c\max}$  and  $J_{\max}$  in the FACE plots over the increase in the control plots (Table 12). Indeed the enhancement of the average  $V_{c\max}$  and  $J_{\max}$  values in the control plots on the 26<sup>th</sup> June was 26.44% and 43.28% respectively. In the FACE plots the enhancement was 116.71% for  $V_{c\max}$  and 80.23 % for  $J_{\max}$  . On the 23<sup>rd</sup> May the

**FIGURE 14.** Variations in a)  $V_{c\max}$  and b)  $J_{\max}$  measured on the same leaves at two times of the day. Before 08:00 hrs (am) and after 15:00 hrs (pm).



enhancement was greater in the control plots,  $V_{c\ max}$  values increased by 139 % in the afternoon in the control plots whilst  $J_{\ max}$  increased by 190%. Corresponding increases in the FACE plots were 53.1 % for  $V_{c\ max}$  and 45.17% for  $J_{\ max}$ .

#### 4. Discussion.

The enhancement of A observed during the diurnal set of experiments on *L.perenne* grown in elevated  $C_a$ , ranged from 26.51% to 44.95% over the course of a diurnal set of measurements. Such findings are in agreement with previous work on *L.perenne* single leaves (Nijs *et al* 1989, Ryle *et al* 1992, Gay & Hauck 1993). They also correspond with the normal and expected pattern of  $C_3$  crops photosynthesising in elevated  $C_a$  (Kimbal 1983, Long 1985). Indeed the "Clear and Unequivocal" enhancement of A described by Ryle *et al* (1992) could equally well describe this studies findings.

Over the course of the three day's measurements the largest and significant increases in A between control and FACE rings corresponded to the period of the day when temperatures were at their highest. Whilst in ambient  $C_a$  there is a reduction in carboxylation relative to oxygenation as temperature rises, as a consequence of the decreased specificity of Rubisco for  $CO_2$  and the reduced solubility of  $CO_2$  (Jordan & Ogren 1984). These effects are offset in elevated  $C_a$ . As the key effect of increased  $C_a$  is increased competitive inhibition of RubP oxygenation (Long 1991, Kirshbaum 1994), it follows that proportionate increases in photosynthesis resulting from elevated  $C_a$  will rise with temperature. This study confirms these predictions and is in agreement with other findings demonstrating this principle (Sage & Sharkey 1987). The increasing burden of oxygenation in plants grown in ambient  $C_a$  is not mirrored in FACE grown plants whose temperature optimum of photosynthesis also shifts upwards (Sage & Sharkey 1987).

A related discovery of this study is the consistent increase in the temperature of the leaves of the FACE grown *L. perenne* over and above those grown in the control plots. This effect has been predicted by Long (1985) and is attributed to a reduction in evaporative latent heat loss from leaves and plants grown in elevated  $C_a$ . A consequence of commonly observed decreases in stomatal conductance (Valle *et al* 1985, Idso *et al* 1987). This effect was observed previously by Bryant (1994) for *L. perenne*, and would be expected to confer the aforementioned benefits for plants photosynthesising in elevated  $C_a$  at higher temperatures even if the air temperature does not increase.

Whilst not significant this study observed consistent enhancement of A during periods of the day when PPFD and temperature were lower. Namely the beginning and end of the sunlight hours of the day, mirroring findings the previous year on the same crop (Bryant 1994). This finding is in agreement with the general predictions of Long & Drake (1992) that stimulation of photosynthesis will occur even at lower light intensities, but is in contrast to previous observations on *L. perenne* (Nijs *et al* 1989, Ryle *et al* 1992). Nijs *et al* (1989) site the increased structural components associated with growth in elevated  $C_a$  as a disadvantage at lower irradiance but of far greater benefit at higher irradiance.

This study revealed no significant differences in A as a result of differing nitrogen treatment levels or nitrogen  $CO_2$  interaction. With little differences in maximum A values achieved between FACE and control plots on any of the days.

Trends observed over the three day's measurements suggest that percentage enhancement of  $A$  in FACE plots over control plots was consistently greater in the low nitrogen treatment. Such a finding is juxtaposed to the theory that nitrogen supply, if sub-optimal, will be insufficient to maintain plant growth at a level where it can utilise the additional carbohydrate produced by plant growing in elevated  $C_a$  (Wong 1979, Larigauderie *et al* 1988, Vessey *et al* 1990). The end result being sink limitation and carbohydrate accumulation leading to feedback inhibition of photosynthesis (Mauney *et al* 1979, Clough *et al* 1981, Ehret & Jolliffe 1985). These results tend to support the claims of Sinclair (1992) that a far more complex interactive relationship exists between nitrogen fertilisation and  $CO_2$ . The absence of any downregulation of  $A$  in the low nitrogen plots has been observed previously by Sage *et al* (1990). Analysis of the  $A/C_i$  response curve has revealed that at elevated  $C_a$ , where Rubisco levels are in excess of those required to maintain photosynthetic performance, up to 30 % could be re-apportioned without any reduction in photosynthetic capacity (Long & Drake 1992). These results contrast those on the same crop the previous year when the enhancement of  $A$  was greater in the high nitrogen treatment (Bryant 1994). It was speculated last year that this was due to the denser high nitrogen canopy not reaching its full photosynthetic potential. For the measurements taken this year on the 24<sup>th</sup> May and 20<sup>th</sup> June when the canopies were only 9 and 8 days respectively into regrowth such a proposal would not seem valid. At low light levels or low temperature any depression of photosynthetic performance resulting from sub-optimal nutrient supply would be expected to be reduced (Bunce 1992).

These results evidence a clear decline in A as regrowth continues (9<sup>th</sup> July) with maximum values of A obtained noticeably lower than the A values attained on the measurement days immediately after the cut (24<sup>th</sup> May & 20<sup>th</sup> June). These findings are consistent with the findings that the maximum rate of individual leaf photosynthesis of successive youngest fully expanded leaves of *L. perenne* declines as the vegetative sward develops (Leafe *et al* 1976, Sheehy 1977). Whilst supporting the notion of Nijs & Impens (1993) that the effective use of cutting as a management tool could yield higher increases in seasonal production in conditions of elevated C<sub>a</sub> when the Ryegrass stands would be in the younger stages of growth, when response to CO<sub>2</sub> is largest. At the same time the enhancement of A in the FACE plots in both nitrogen treatments was higher than the previous two days. Current thought would lead us to believe that the further into the regrowth period the crop is, the greater will be the effects of photosynthetic disruptions as a consequence of carbohydrate feedback inhibition in the FACE plots. Closer inspection of the results points to a far greater downregulation of A in the control plots being the cause of this enhancement not an increase in A in the FACE plots. The downregulation of A between the 24<sup>th</sup> May and the 9<sup>th</sup> July in the control plots was 26.6 % compared to 17.89% in the FACE plots. Compared to the 20<sup>th</sup> June the downregulation in the control plots was 32.8% whilst in the FACE plots it was 26.4 %. This greater downregulation in the control plots is a likely consequence of the supra-optimal temperature experienced on the 9<sup>th</sup> July and the midday stomatal closure more evident in the control plots on that day.

The increased water use efficiency of plants grown in elevated CO<sub>2</sub> would seemingly explain this phenomenon. Evapotranspiration was consistently, yet not

exclusively lower in the FACE plots than the levels in the control plots on the three measurement days. Proposals that plant water use efficiency commonly increases with elevated  $C_a$  (Morison 1993), largely as a result of reductions in stomatal aperture (Morison 1987), would seemingly explain the lower rates of evapotranspiration observed in this study for *L. perenne* grown in elevated  $C_a$ . Reduced daytime evapotranspiration is but one factor affecting the water use efficiency of a crop, water use efficiency that has been observed to increase in near unity to increasing  $C_a$  in *L. perenne* (Nijs *et al* 1989).

The increased temperature optimum for photosynthesis and improved water use efficiency predicted and observed for  $C_3$  crops such as *L. perenne* would seemingly equip such crops well for predicted atmospheric changes. Unfortunately any further terrestrial or atmospheric feedback effects associated with increased temperature and atmospheric  $CO_2$  increase could negate such benefits.

Given that the *L. perenne* crop studied was in its third year of growth in elevated  $C_a$  the results obtained provide a firm basis for determining the extent to which the long term acclimatory response (defined on pg 6) will negate the substantial short term increases in photosynthesis by  $C_3$  plants growing in elevated  $C_a$ . From the results obtained by this study it would have to be concluded that there is little evidence to suggest that after three years of growth in elevated  $C_a$  of  $600 \mu\text{mol mol}^{-1} CO_2$  that the photosynthetic capacity of plants grown in elevated  $C_a$  has downregulated with respect to plants grown in ambient  $C_a$ . Such a conclusion would appear to be in agreement with the findings of Arp (1991) who suggested that downregulation of the

photosynthetic capacity of crops in response to growth in elevated  $C_a$  was primarily an artefact of experimental constraints. It is also in agreement with Sage (1994) who reviewed field studies since 1986 to conclude that there was a lack of acclimation of field grown crops to elevated  $C_a$ . At the same time this study is at odds with previous work on *L. perenne* which showed acclimation in response to long term growth in elevated  $C_a$  (Moon *et al* 1990, Ryle *et al* 1992), and indeed with work on the same crop 1 year previously that showed "Clear evidence of acclimation" (Bryant 1994). The controlled environments in which the experiments of Ryle *et al* (1992) and Moon *et al* (1990) were conducted may explain the apparent contradiction of the results.

The results of the 4 A /  $C_i$  experiments conducted either side of the two cuts showed no significant differences in values of  $V_{c \max}$  and  $J_{\max}$  obtained as a result of growth in different  $CO_2$  treatments. In contrast the advent of cutting greatly relieved the pre-cut depression of both  $V_{c \max}$  and  $J_{\max}$  values, significantly increasing them for both  $CO_2$  treatments. Depression would have been brought on by a combination of increasing leaf age and sink limitation. This was relieved by cutting and countered by the post cut need to ensure quick and effective re-establishment of the canopy.

Whilst there was not a significant  $CO_2$  effect there was a substantial downregulation of  $V_{c \max}$  and to a lesser extent  $J_{\max}$  values in the low nitrogen treatments on the 1<sup>st</sup> July for the FACE rings with respect to the control rings. Such a discovery if it were significant would be consistent with the aforementioned theory (pg 9) that when nitrogen supply or translocation are insufficient to support the growth rate of the plant, leaf nitrogen amounts start to fall below those levels associated with

more optimal supplies of nitrogen with a corresponding decrease in photosynthetic capacity (Evans 1989).

These findings appear to be enforced by the results of the manipulation experiment conducted in the low nitrogen plots only, where theory would suggest that acclimation will be most extreme. Values of  $V_{c \max}$  and  $J_{\max}$  obtained in both the cut and uncut plot were at no time significantly different as a response to growth in different  $CO_2$  treatments or as a result of an interaction between  $CO_2$  and the different cut status's of the plots. This conclusion was reached based on the actual values of  $V_{c \max}$  and  $J_{\max}$  obtained and on analysis of the relative enhancement of the two sets of values between the cut and uncut plots. At the same time significant differences in  $V_{c \max}$  and  $J_{\max}$  were obtained as a consequence of the manipulation of the source sink balance. As would be expected and was evidenced in the previously described A / Ci experiments. The removal of sink limitation in the cut sections of the plots used where measurements were taken 5 days into regrowth exhibited significantly higher values of  $V_{c \max}$  and  $J_{\max}$  than the areas of the plot that were 23 days into the regrowth cycle, on the 5<sup>th</sup> July when all six rings were sampled. On the 8<sup>th</sup> June when only four rings were sampled the results did not achieve significance.

The canopy experiment allowed the way in which leaf ageing and shading affect acclimation to be studied. As canopy regrowth advances the initially measured young leaves at the top of the canopy will age and become shaded to some extent as they migrate down through the canopy profile. As well as the photosynthetic performance of individual leaves of *L.perenne* declining with age (Leafe *et al* 1976, Sheehy 1977),

where light is limiting the photosynthetic capacity of the shaded leaves photosynthesis will be most limited by the supply of light dependent ATP and NADPH for RubP regeneration . Whilst Rubisco levels or activity will be in excess. A situation that mirrors that in elevated CO<sub>2</sub> . It has been proposed by Evans (1989) that such an excess of Rubisco in a shaded lower canopy will be countered by plants, by adapting and re-apportioning more leaf nitrogen into the thylakoids and chlorophyll away from Rubisco. Whilst the results of the canopy experiment show significant declines in both V<sub>c max</sub> and J<sub>max</sub> of single leaves as canopy regrowth advances there was again no significant effect of CO<sub>2</sub> treatment on the values obtained. Whilst this would seem to mirror the findings so far that the FACE grown *L. perenne* is exhibiting no signs of acclimation, for this experiment we must consider the structure of the ryegrass canopy. It has been described how the combination of the low leaf area index in the top strata of the canopy and the vertical distribution of the youngest leaves (extinction co-eff (k) = 0.5) means that photosynthetically active radiation can penetrate deeply into the crop stand and canopy photosynthesis can be spread over a large area of leaf (Hay & Walker 1989). This would limit the reductions in V<sub>c max</sub> and J<sub>max</sub> values in both the FACE and control plots as shading would not be expected to be such a consideration. If we also consider the fact that the results show higher downregulation of V<sub>c max</sub> and J<sub>max</sub> in the low nitrogen plots for the control grown plants whilst down regulation is higher in the high nitrogen treatment in the FACE plots. We can speculate that in the control plots leaf age is more of a determining factor than shading that would be expected to be minimal in the low density low nitrogen canopy. Whilst the higher downregulation in the high nitrogen treatments of the FACE plots supports the view that the interactive effects of elevated CO<sub>2</sub>, shade and high nitrogen have resulted in

excess Rubisco levels that is re-apportioned, or its activity reduced as the canopy develops.

The final experiment designed to test for acclimation in the FACE plots was the diurnal  $A / C_i$  experiment. For the two experiments conducted on the 23<sup>rd</sup> May and the 26<sup>th</sup> June the expected downregulation of the  $V_{c \max}$  and  $J_{\max}$  values in response to carbohydrate feedback between 15:00 hrs and 17:00 hrs did not occur. In fact there was a statistically significant enhancement of  $V_{c \max}$  and  $J_{\max}$  on both measurement days in the afternoon measurement period. This increase was higher in the FACE plots for both sets of values on both days, significantly so for the measurements on the 23<sup>rd</sup> May. It must be borne in mind that as well as feedback inhibition of photosynthesis the plants used would potentially have been subject to some magnitude of photoinhibition. A phenomenon that plants grown in elevated  $C_a$  potentially have some protection against possibly giving some explanation for the greater enhancement of  $V_{c \max}$  and  $J_{\max}$  observed in the FACE plots. Whilst there appears to have been no evidence of carbohydrate feedback inhibition on these two measurement days exposure of the leaf in the leaf chamber to a very low  $O_2$  concentration of 1% would evidence inorganic phosphate (Pi) limitation as a horizontal line on the  $A / C_i$  response curve at higher  $C_i$  values.

## **5. Conclusion.**

It is clearly possible to reconcile the substantial stimulation of  $A$  during the diurnal sets of measurements for *L.perenne* grown in the FACE rings with the apparent

lack of acclimation of the photosynthetic apparatus of *L. perenne* to the long term exposure to elevated  $C_a$ . Studies have suggested that typically plants will undergo a succession of adaptive responses of the photosynthetic mechanism to growth in elevated  $C_a$ . Initially there will be a downregulation of the non limiting processes at high  $C_a$ , namely RubP carboxylation, to create a balance with the limiting process RubP regeneration (Long & Drake 1992). Next it is proposed that these resources released by this downregulation, namely leaf nitrogen, will be re-apportioned to the limiting processes of photosynthesis at elevated  $C_a$  (RubP regeneration). The period of time this takes to occur is of paramount importance for perennial crops.

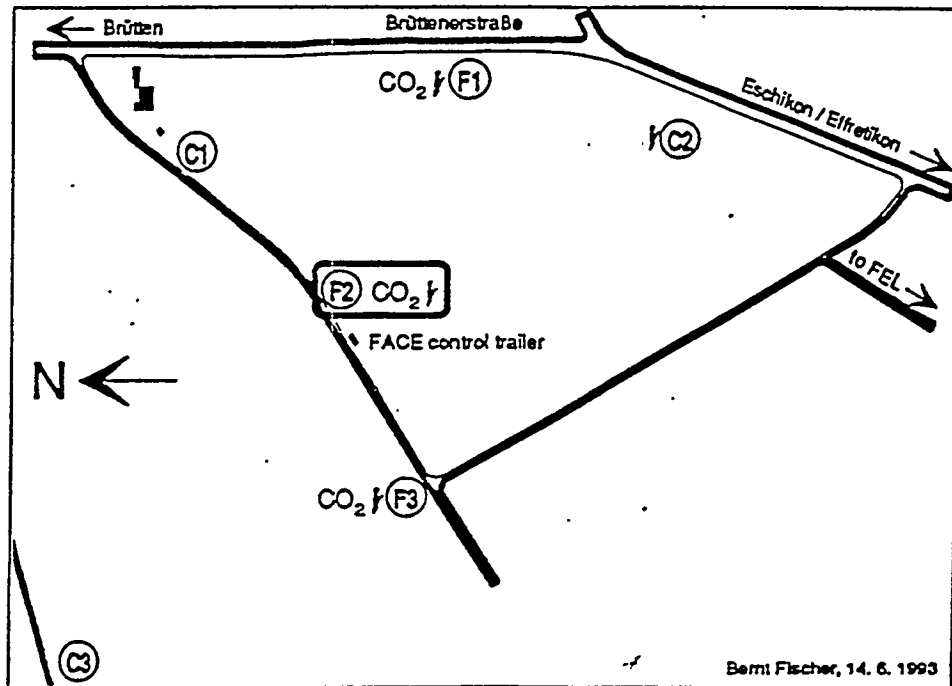
Whilst more work is needed to firmly establish whether such a complete acclimation of the photosynthetic apparatus occurs. It could be suggested from this data that after three years of growth in elevated  $CO_2$  that the crop of *L. perenne* studied is approaching if not at the stage where this re-apportioning of the resources of the non limiting processes to the limiting processes has been achieved. The fact that this study has observed no significant downregulation of  $J_{max}$  values, in FACE grown *L. perenne*, with respect to control plots, would support such a claim that the initially limiting capacity for RubP regeneration has now been upgraded. Fundamental to such a transformation would be the ability of the crop to overcome long term sink limitation. The restriction of tillering ability of *L. perenne* at high light intensities (low red : far red radiation ratio) (Deregibus *et al* 1983) and the fact that *L. perenne* is known to partition relatively more of additional photosynthate produced in elevated  $C_a$  into storage rather than growth of new sinks (Wardlaw 1982), have been cited as factors limiting the ability of *L. perenne* to generate new sinks. What may be more pertinent is

the fact that after three years of growth in elevated  $C_a$ , *L. perenne* may have generated a root system large enough that it does not experience prolonged sink limitation. This area of study revealing root responses to elevated  $C_a$  of *L. perenne* would perhaps enforce or dispel such a claim.

Whatever the causes, this study has evidenced a prolonged, considerable enhancement of photosynthesis and a lack of acclimatory downregulation of photosynthetic capacity of *L. perenne* grown in elevated  $C_a$ . These factors suggest that over the longer term, in a future high  $CO_2$  world, seasonal enhancement in productivity could be attained, as could extensions of the growing season for *Lolium perenne*.

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APPENDIX I.



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