Characterization and Reactivity of Organic Monolayers on Gold and Platinum Surfaces

by

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Characterization and reactivity of organic monolayers on gold and platinum surfaces

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The focus of this study is to understand how the immobilization, dielectric, orientation, composition, coverage, and structure of self-assembled organic monolayers on metal surfaces affects the surface reactivities and properties of these films in order to facilitate the construction of desired films.

Two model systems used: tiols at gold and aromatic acids at platinum. Both methyl- and carboxylic acid-terminated thiolate monolayers were formed at evaporated gold films by the self-assembly technique via the thiol moiety of thiols. The phenylcarboxylic acid and pyridylcarboxylic acid monolayers were formed at platinum electrodes cleaned electrochemically in 1 N H₂SO₄ aqueous solution.

Several surface analysis technologies (including contact angle measurement, electrochemistry, ellipsometry, infrared reflection absorption spectroscopy (IRRAS), and x-ray photospectroscopy) have been used to study the formation, composition, structure, surface properties and reactivity of self-assembled organic monolayers on gold and platinum surfaces.

IRRAS, contact angle measurements, and an electrochemistry methods were used to determine the surface pK_a of phenylcarboxylic acids and pyridylcarboxylic acids monolayers on platinum.

These techniques were also used to determine the orientation of polymethylene chain axis and the carboxylic follow the structural evolution of the chains and end group of the thiolate monolayers during formation.

In addition, IRRAS was used to asses the carboxylic acid group in terms of its possible existence as the non-hydrogen-bonded species, the hydrogen-bonded dimeric group, and the hydrogen-bonded polymeric group. The existence of these different forms of the end group were also followed as a function of coverage and as well as their influence a reactivity as a function of solution pH.

IRRAS and contact angle measurements were used to calculate the rate constant of the esterification of carboxylic acid-terminated monolayers on gold.

DEDICATION

This thesis is dedicated to the love from my mother Wei Liu and my father Yu-Wey Wu who passed away on December 24, 1993.

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GENERAL INTRODUCTION

Dissertation Organization

This dissertation consists of five research papers followed by a general summary and prospective.

Overview

The attractive properties of self-assembled monolayers including simple fabrication, stability, well-defined structures, and flexible synthetic modification have attracted enormous worldwide attention for the last decade. This interest is due largely from their potential and sometimes demonstrated use as models of organic interfaces.

The use of self-assembled monolayers is primarily controlled by their surface properties and reactivities which are dictated by the coverage, structure, dielectire, spatial orientation, and composition of these films. Therefore, it is essential to fully understand how these factors affect the surface properties and reactivities of these films. However, there is little work has been done in this regard. The main goal of this study is to probe these relationships between these factors and the surface properties of these films to construct desired materials more effectively.

Although it has been pointed out that the adsorption of thiolate monolayers on Au is a two-step process using the electrochemical quartz micro balance experiment and contact angle measurement, little work has been done concerning the details of this fundamental formation process.

In Paper I, the pH dependence of the electron transfer of ferricyanide at phenylcarboxylic acids and pyridylcarboxylic acids adsorbed at Pt electrodes is used to determine the surface pK_a of the coatings. The results of this measurement, which relies on the change in the potential at the electrode reaction plane via adsorbate deprotonation, are compared to those from contact angle titrations. Unexpected

differences in the reactivity of the adsorbate (as opposed to their solution analogs) are discussed.

In Paper II, the formation of alkane and acid-terminated thiolate monolayers on gold is examined using infrared reflection absorption spectroscopy (IRRAS). The discovery of agostic bonds at 2903 cm⁻¹ due to the interaction between methylene groups and a bare Au surface based on the shift in energy of the $\nu_a(\text{CH}_2)$ from 2926 cm⁻¹ to 2903 cm⁻¹ and the absence of the agostic bond at 2903 cm⁻¹ on the Au surface which was pretreated with a sulfur monolayer is reported. The change in the orientation of the polymethylene chain as a function of coverage, which is based on the fact that the presence of the $\nu_s(\text{CH}_2)$ is at the expense of the amount of $\nu_a(\text{CH}_2)$, is described.

In Paper III, the coverage dependence of the amount of the non-hydrogen-bonded, the hydrogen-bonded dimeric, and the hydrogen-bonded polymeric C=O groups of a 16-mercaptohexadecanoic acid $(HO_2C(CH_2)_{15}SH)$ on Au monolayer, which is evidenced by the fact that decrease in peak area of the hydrogen-bonded dimeric $\nu(C=O)$ is almost equal to the total increase in peak area of the other two peaks, is studied with ellipsometry, IRRAS, contact angle measurements techniques.

In Paper IV, the surface pK_a values of the non-hydrogen-bonded, the hydrogen-bonded dimeric, and the hydrogen-bonded polymeric C=O groups of several carboxylic acid-terminated thiolate monolayers on Au are examined with IRRAS as a function of solution pH and calculated based on the pH where 50 % of the decreases in the peak areas of the corresponding ν (C=O) peaks.

In Paper V, the extent of esterification, θ , of 11-mercaptoundecanoic acid on Au monolayers was examined and calculated based on the decrease in the IRRAS peak area of the $\nu(C=0)$ mode of the carboxylic acid or the increase in that of the $\nu(C=0)$ mode of the ester and on the increase in contact angles. The coverage-dependent rate constant of the surface esterification reaction is discussed.

PAPER I

ELECTRON TRANSFER OF FERRICYANIDE AT ORGANIC CARBOXYLIC ACIDS ON PLATINUM SURFACES

INTRODUCTION

Enormous attention has been paid to molecular films because they have many very promising properties for fundamental and technological applications ¹⁻⁴. The component, structure and orientation of these films can affect their properties. In the densely-packed monolayer, each molecule experiences not only a highly concentrated but also well-organized environment. It is important to understand how these factors affect the properties of these films. In many instances, the reactivities of the immobilized species are markedly different from those of their solution analogs. The main focus of this study is to see how the pK_a values of carboxylic acids tethered to Pt electrodes via aromatic moieties are affected upon immobilization. The other focus is to understand how the potential at the electrode double layer is changed due to the deprotonation of the carboxylic acid-terminated monolayer.

The packing density, structure, and orientation of a variety of aromatic species formed as monolayers at Pt have been extensively characterized by Hubbard and co-workers⁵⁻¹⁹ and other groups²⁰⁻²⁹. All these properties are a function of the solution concentration and depend on the nature of molecules as well. The orientation of the aromatic ring of some aromatic molecules can be parallel or vertical to the Pt surfaces at low or high concentration, respectively⁶⁻⁹. Some aromatic molecules can only form flat orientation¹⁸. In this study, there are two different kinds of carboxylic acids studied: phenylcarboxylic acids and pyridylcarboxylic acids. The aromatic ring of phenylcarboxylic acids are oriented parallel to the Pt surface.

On the other hand, pyridylcarboxylic acids form stable monolayers on Pt surface and orient differently from phenylcarboxylic acids³⁰. Based on Auger electron spectroscopy (AES) data, the tilt angles of the aromatic rings of the nicotinic acid and isonicotinic acid on Pt were calculated to be 19° and 15° off the surface normal, respectively³⁰. The observed packing for the NA/Pt and INA/Pt were calculated to be 3.8 x 10⁻¹⁰ mol/cm² and 4.2 x 10⁻¹⁰ mol/cm², respectively³⁰. This provides a very good opportunity to see how the orientation and component affect the surface properties of monolayers.

In this paper, we report the results of a characterization of carboxylic acid functionalized monolayers (i.e. phenylcarboxylic acids:

 $C_6H_5(CH_2)_nCOOH$, n=0 to 3; pyridylcarboxylic acids: $C_5NH_4(CH_2)_nCOOH$, n=0,1) at Pt using tow techniques: electrochemistry, and contact angle measurements (CA). The electrochemical measurements relies on the influence of the change of the charge at the electrode reaction plane as a function of the pH-dependent ionization of the surface bond carboxylic acid groups. These changes are followed through measurements of the apparent heterogeneous electron-transfer rate constant of the solution-bases redox couple $Fe(CN)_6^{-4/-3}$. For comparison purposes, the ionization of these layers were monitored using CA. The effects of immobilization on reactivity are discussed.

EXPERIMENTAL SECTION

Electrode preparation

The 0.15-cm-diameter platinum electrodes (Bioanalytical System, West Lafayette, IN) and a 2" x 1" Pt foil (Engelhard Minerals & Chem. Corp., Carteret, NJ) were smoothed to a mirror-like finish using successively fine grades of wet emery paper, followed by slurries of 1.0,0.3 and 0.05-mm diamond polishing compound (Buehler, Lake Forest, IL). Between each step, the electrodes were rinsed thoroughly with deionized water. The finial pretreatment step consisted of scanning the applied voltage at 100 mV/s repetitively between +1.2 and -0.2 V until a steady-state voltammetric response that exhibited the characteristic hydrogen adsorption/desorption and oxide formation/desorption fine structure was obtained (see Figure 1). Electrodes so prepared will be referred to simply as an uncoated platinum electrodes. An Ag/AgCl (sat'd) reference electrode and a Pt auxiliary electrode were used. All the potentials are reported with respect to this silver reference electrode.

The active surface of the platinum electrode was determined by the charge consumed in the hydrogen atom underpotential deposition region 11 . The voltammetric response of an electrode coated with a layer of adsorbed iodine provided the base line for charge integration 11 . A value of 210 mC/cm^2 was used as representative for the monolayer coverage of hydrogen at polycrystalline platinum 11 . This analysis yielded a roughness factor, which is given as the active surface area divided by the geometric surface area, of 1.3 ± 0.1 . The uncertainty of the roughness factor is based on repetitive testing of five separate electrodes using the described polishing and electrochemical pretreatment methods.

Monolayer preparation

The electrodes were coated with each of the aromatic moieties by terminating a voltammetric scan in 1.0 M H₂SO₄ aqueous solution in the region of oxide formation. The electrode was disconnected from the potentiostat, and the electrolytic solution changed to a known volume of 1.0 M NaF solution (pH 3.0). After reconnection to the potentiostat, the voltage was stepped to

0.0 V. A small volume of concentrated adsorbate was then added to the solution with a micropipette to yield the targeted concentration. Film formation was essentially complete within 5 min after injection, as indicated by loss of the hydrogen underpotential deposition waves, longer adsorption times were used for convenience. Over a ten minute time period, and uncoated platinum platinum electrode exhibited only a marginal loss (< 5 %) of in the area of the electrode active toward hydrogen adsorption.

Instrumentation and measurements

Electrochemistry

A conventional three electrode electrochemical cell with a Luggin-Harber capillary and a Cypress system CY-1 computer controlled potentiostat (Cypress Systems Inc., Lawrence, KS) were used in all electrochemical experiments.

The heterogeneous electron-transfer rate constant, k_{app} , of ferriferrocyanide at the monolayer-coated electrodes was determined by cyclic voltammetry according to the method of Nicholson³³. The working solution were 1.0 mM ferrocyanide with 0.50 M K_2SO_4 as the supporting electrolyte.

For the determinations, the following constants were used for the redox couple: $D_{ox} = 7.63 \times 10^{-6} \text{ cm}^2/\text{s}^{34}$, $D_{red} = 6.32 \times 10^{-6} \text{ cm}^2/\text{s}^{34}$, $\alpha = 0.5$, and n = 1.0. With these constants and the separation of the cathodic and anodic peak current potentials, k_{app} was calculated as $k_{app} = 2.78 \times 10^{-2} \text{ v y}^{1/2}$ (cm/s), where v is the scan rate and ψ was taken from a working curve³⁵ or from a numerical data³⁶.

Contact angles

Advancing contact angles (θ_a) were measured in air with 0.100 N of HF/NaF aqueous buffer solutions as probe liquids by using a Rame-Hart Model 100-00 115 goniometer. For these measurements, a 2.00 mL droplet was formed on the substrate with the needle of the syringe in the droplet. The value of θ_a was determined as the volume of the droplet was slowly increased. The standard deviation of these data is ~10 %, a reflection of the relatively small values of the contact angle.

Chemicals and reagents

All the organic compounds including benzoic acid (BA), 2-phenylacetic acid (PAA), 3-phenylpropanoic acid (PPA), 4-phenylbutric acid (PBA), isonicotinic acid (INA), nicotinic acid (NA), pyridylacetic acid (PYAA) (Aldrich, Milwaukee, WI) and inorganic chemicals (Fisher Scientific, Pittsburgh, PA) were reagent grades and were used without further purification. All solutions were prepared daily with purified water from a Mili-Q system (Millipore, Bedford, MA). The pH was adjusted by adding appropriate amount of H₂SO₄ or KOH. Solution were deoxygenated by purging with high purity argon gas for 20 min prior to each electrochemistry experiment and were kept under an argon atmosphere during the experiments.

RESULTS AND DISCUSSION

Characterization of monolayers

Figure 1 shows cyclic voltammograms for an electrochemically pretreated platinum electrode (solid curve) and for platinum electrodes coated by immersion of a platinum electrode in solution of different concentration of BA (dashed curves). The voltammograms were obtained by sweeping the voltage at 100 mV/s from + 0.2 V to a lower limit of -0.2 V, to an upper limit of +1.5 V, and then returning to the initial voltage. The curve for the uncoated electrode exhibits both the hydrogen adsorption and desorption waves and the oxide formation and reduction waves characteristic of a platinum surface free of strongly bond organic adsorbates¹¹. The presence of adsorbed BA is evident by the loss of the hydrogen adsorption/desorption waves as well as by the anodic current that flows at large positive applied voltages (~+ 1.3 V). The adsorbed layer blocks access to sites for hydrogen atom adsorption, and hinders the oxidation of the platinum surface. The broad anodic wave corresponds to oxidation of the surface-bonded layer. For all the acids, the charge under this anodic wave increases with increases in the concentration of the aromatic acids as shown in Figure 1.

The values for the oxidation charge were determined by integrating the charge passed at the coated electrode, from which the value over the same potential region of an uncoated electrode was subtracted. The oxidation charge density Q_{ox} is calculated by dividing the value of the oxidation charge by the real electrode surface area which is equal to the geometric area of the electrode multiplied by the roughness factor (1.3). The dependencies of Q_{ox} on the concentration of each of the aromatic acids are shown in **Figure 2**. In each instance, the value of Q_{ox} increases as the adsorbate precursor concentration increases, reaching limiting values between 0.05 and 1.0 mM depending on the acids as shown in **Table 1**.

The apparent surface coverage for BA and the other adsorbed layers were estimated for the limiting values based the charge density, Q_{ox} , consumed during the anodic sweep in the voltammotry experiments. The values of Q_{ox} for BA, PAA, PPA, and PBA at saturation coverages on platinum are 8.2×10^4 , 9.5×10^4 , 9.7×10^4 , and 7.9×10^4 C/cm²,

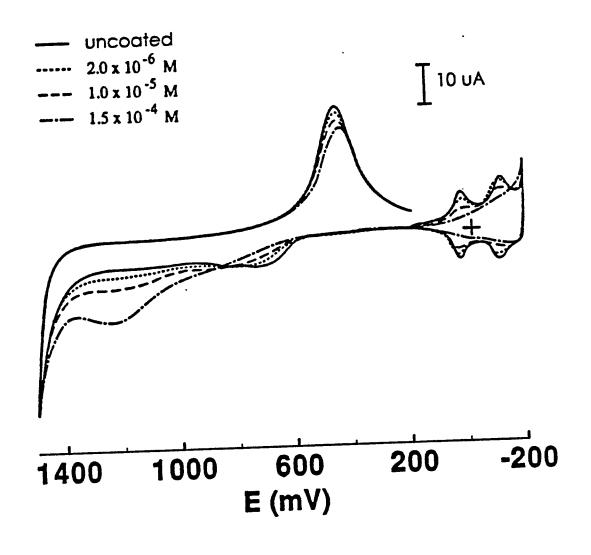


Figure 1. Cyclic voltammograms of a clean Pt electrode (solid curve) and the BA/Pt electrodes adsorbed in 2.0 x 10⁻⁶ M, 1.00 x 10⁻⁵ M, and 1.50 x 10⁻⁴ M BA solution for 5 min (dashed curves) in 1.0 N H₂SO₄ aqueous solution at 100 mV/s scan rate.

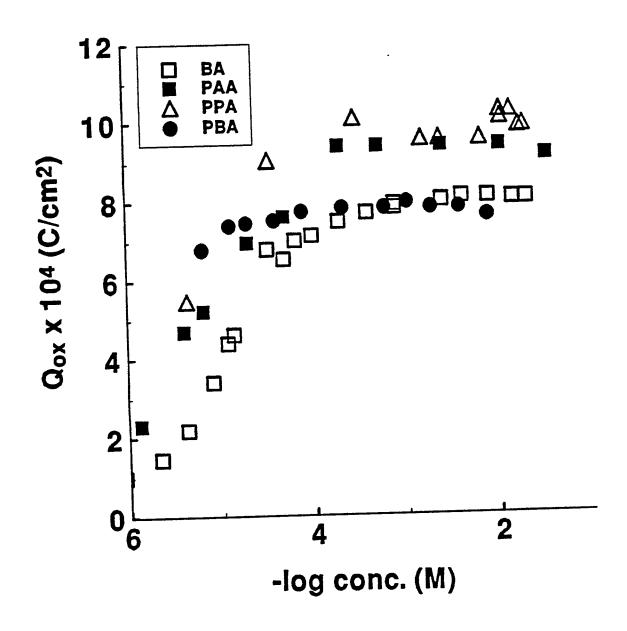


Figure 2. The Q_{ox} of BA, PAA, PPA, and PBA on platinum electrode as a function of -log concentration.

Table 1. Oxidative stripping charge density (Q_{ox}) , and saturation surface coverage estimates experimental (Γ_{ox}) and theoretical (Γ_{calc}) at limiting concentrations of aromatic acid layers at platinum electrodes.

adsorbate	$\frac{Q_{ox} \times 10^4}{(C/cm^2)}$	n_{ox}	$\Gamma_{\rm ox} \times 10^{10}$ (mol/cm ²)	$\Gamma_{\rm calc} \times 10^{10}$ (mol/cm ²)
precursor	(C/GIII)		(IIIOI/CIII)	(111011 02111)
BA	8.2 ± 10	30	2.8	2.9
PAA	9.5 ± 10	36	2.8	2.8
PPA	9.7 ± 10	42	2.4	2.5
PBA.	7.9 ± 10	48	1.7	2.3

respectively with an error bar of $0.1 \times 10^{-4} \text{ C/cm}^2$. Saturation coverages, Γ_{ox} , from these measurements are calculated as

$$\Gamma_{\rm ox} = Q_{\rm ox}/n_{\rm ox} F \tag{1}$$

where n_{ox} is the number of electrons required to oxidize the adsorbed layer exhaustively to CO₂ (e.g., C₆H₅CO₂H + 12 H₂O -> 7 CO₂ + 30 H⁺ + 30 e⁻), F is the Faraday¹⁷. In addition to the exhaustive electrolysis to CO₂, there are two further assumptions: 1) the adsorbate orientation and/or mode of attachment is constant over the concentration range studied; and 2) the composition of the adsorbate is the same. The limitation of this approach have been examined in depth by Hubbard and co-workers¹⁹; our intent, however, is to use this as a qualitative indicator of coverage and orientation. The errors in measurement arise from the assumption that oxidation of adsorbed species always proceeds completely to CO₂. Complete conversion of adsorbed aromatics occurs only for flat-adsorbed molecules. In other words, this method cannot be applied to the edge-adsorbed molecules because these adsorbed molecules cannot be oxidized exhaustively to CO₂. However, this method can be used to differentiate a flat-adsorbed molecule from an edge-adsorbed molecule.

The values of Γ_{ox} for BA, PAA, PPA, and PBA are 2.8×10^{-10} , 2.8×10^{-10} , 2.4×10^{-10} , and 1.7×10^{-10} mol/cm², respectively. The results of the coverage analyses are listed in **Table 1** along with coverage calculated for the closest-packed model structures for each adsorbate, Γ_{calc} . The values for Γ_{calc} were determined based on a fully extended alkyl chain with the plane of the aromatic ring oriented parallel to the Pt surface, i.e. in the $\eta 6$ orientation 6,37,38 . The Γ for the closest-packed model structures with the plane of the aromatic ring oriented vertical to the Pt surface is not used because its value (~7.0 x 10 $^{-10}$ mol/cm²) is ~3 times larger than the experimental values. From **Table 1**, the values of Γ_{ox} for BA, PAA, and PPA are in good agreement with the corresponding Γ_{calc} values. This suggests that the BA, PAA, and PPA form closest-packed monolayers with the aromatic rings oriented parallel to the Pt surface. The Γ_{ox} value for PBA is, however, much less than the corresponding Γ_{calc} value, precluding an orientation analysis.

The difference in the Γ_{ox} and Γ_{calc} values for PBA is possibly due to the incomplete oxidation of long alkyl chain¹⁹. For the purposes of our discussion, we will nevertheless assume that the aromatic ring for PBA is oriented parallel to the Pt surface.

On the other hand, pyridylcarboxylic acid monolayers on platinum electrodes are oriented with the aromatic ring almost vertical to the Pt surface. As shown by Hubbard and co-workers, rings with a vertical orientation cannot be fully oxidized electrochemically before the occurrence of the strong oxygen evolution wave. This prohibits the same coverage analysis for all pyridylcarboxylic acids on platinum electrodes. Therefore, the packing densities of the INA of $(4.2 \times 10^{-10} \text{ mol/cm}^2)$ and NA $(3.8 \times 10^{-10} \text{ mol/cm}^2)$ monolayers on platinum electrodes calculated by Hubbard and co-workers³⁰ were used in this study.

Electrochemical characterization of acid-base reactivity

To probe the acid-base chemistry of these monolayers, the pH dependence of the apparent heterogeneous rate constant (k_{app}) for ferricyanide redox couple $(Fe(CN)_6^{-4/-3})$ in solution pH was monitored. The underlying basis of this approach relies on the effect of the build up of a negative charge at the electrode surface as the carboxylic acid ionized as the solution pH increases. Values of k_{app} was calculated based on the difference in potential of the anodic and cathodic peak currents from cyclic voltammograms 33,35,36 . (see experimental section) Figure 3 presents a portion of such data at an INA/Pt, which is summarized in Table 2. As expected, the peak separation increases as the solution pH increases, indicative of a decrease in k_{app} .

Plots of k_{appa} for the redox couple at BA/Pt, PAA/Pt, PPA/Pt and PAA/Pt electrodes as a function of solution pH are shown in Figure 4. At low pH, the values of k_{app} at BA/Pt, PAA/Pt, and PPA/Pt are almost the same but that at PBA/Pt is slightly smaller. This indicates that the thicknesses of BA/Pt, PAA/Pt, and PPA/Pt monolayers are similar. In contrast, the lower value of k_{app} for the PBA/Pt monolayer is indicative of a slightly thicker film as dictated by the electron tunneling i.e. the greater the thickness of the adsorbed monolayer the larger the decrease in k_{app} . In other words, the structural

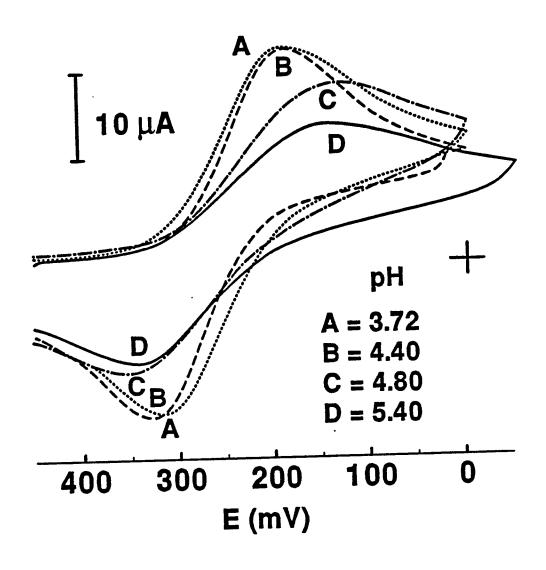


Figure 3. Cyclic voltammograms of 1.0 mM ferricyanide at a INA/Pt electrode in 1.0 N K₂SO₄ solution at pH 3.7 (curve A), pH 4.4 (curve B), pH 4.8 (curve C), and pH 5.4 (curve D).

Table 2. The peak separation (ΔE_{peak}) and k_{app} for ferricyanide at INA/Pt as a function of solution pH and scan rate (ν).

ΔE_{peak} (mV)	pH=2.20	pH=3.72	pH=4.40	pH=5.40
v(V/s) = 5	102	113	137	223
v(V/s) = 3	81	94	117	177
v(V/s) = 2	79	84	110	162
v(V/s) = 1	77	79	96	135
k _{app} (cm/s)	5.2	3.0	2.0	0.85

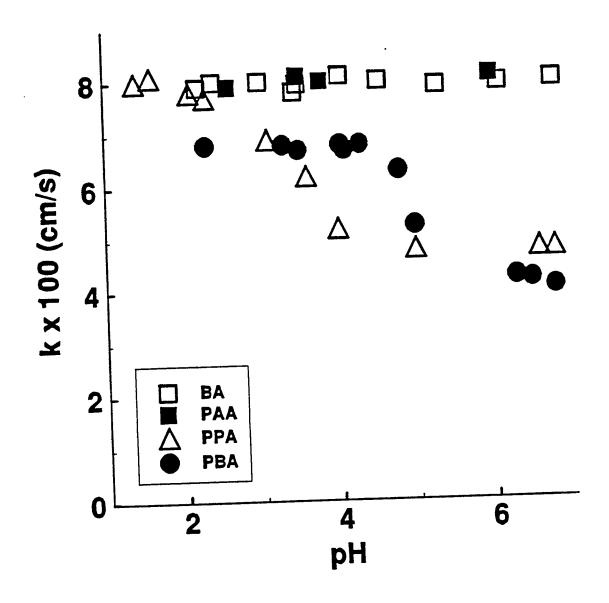


Figure 4. The k_{app} of ferricyanide at the BA/Pt, PAA/Pt, PPA/Pt, and PBA/Pt electrodes as a function of solution pH.

differences between the BA/Pt, PAA/Pt, PPA/Pt and the PBA/Pt monolayers led to differences in the plane of closest approach for ferricyanide. This is consistent with our interpretation of coverage analysis data as discussed early. The values of kapp of ferricyanide at both BA/Pt and PAA/Pt are constant within the pH window studied while those at PAA/Pt and PBA/Pt decrease with increasing solution pH. The k_{app} values at control monolayers (i.e. these without an acidic functional group, like pyridine and benzene) exhibit no observable dependence on pH as shown in Figure 5. Importantly, the transition in the k_{app} values spans ~3 decade in hydrogen ion concentration, diagnostic of the deprotonation of an acidic functional group. Since it has been demonstrated that the carboxylic acid groups of benzoic acid can form a carboxylate-Pt bond at these electrode³⁰ and effectively lose any acid-base chemistry at this site, we should lose acid-base reactivity. Since the $k_{\mbox{\tiny app}}$ of ferricyanide at BA/Pt and PAA/Pt are the same as that at protonated PPA/Pt, it is unlikely that both BA/Pt and PAA/Pt exist in their basic forms at low solution pH. In other words, if both BA/Pt and PAA/Pt monolayers were strong acids because of immobilization, then k_{app} would be lower than observed at low pH (see below). More evidence and more discussion will be provided later in support our conclusion and make one believe that the carbonyl of both BA/Pt and PAA/Pt monolayers are inactive.

Plots of k_{app} of ferricyanide at INA/Pt, NA/Pt, and PYAA/Pt electrodes as a function of solution pH are shown in **Figure 6**. In each plot, there is a titration curve centered at around pH 4. Since PYAA/Pt should have the largest film thickness, ferricyanide should as before have the smallest k_{app} at PYAA/Pt. The thicknesses differences of the INA and NA monolayers are probably too small to influence these data to an observable extent³⁹. This curve has a very similar shape to the one found by using IRRAS (see below). We therefore attribute the decreases in k_{app} to the deprotonation process.

The k_{app} values of ferricyanide are smaller at pyridylcarboxylic acids monolayers than that at PYR/Pt because the thicknesses of the foamier are thicker than that of the latter which does not have the carboxylic acid group at all as shown in **Figure 4.** Notice, the k_{app} values of ferricyanide at PYR/Pt is

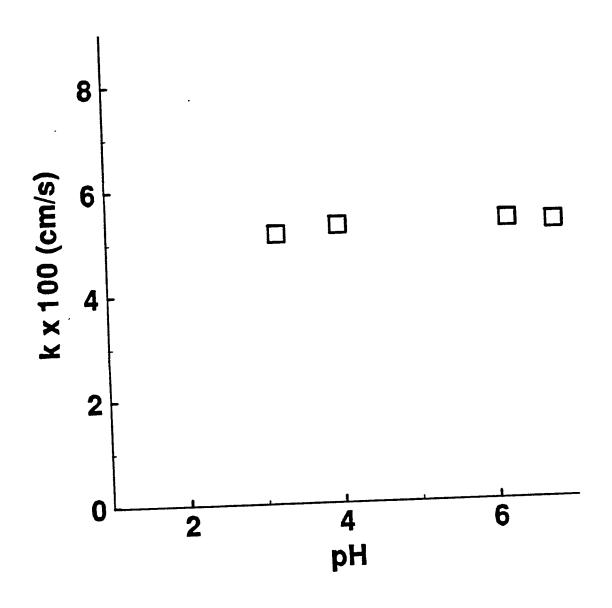


Figure 5. The k_{app} of ferricyanide at the BEN/Pt electrode as a function of solution pH.

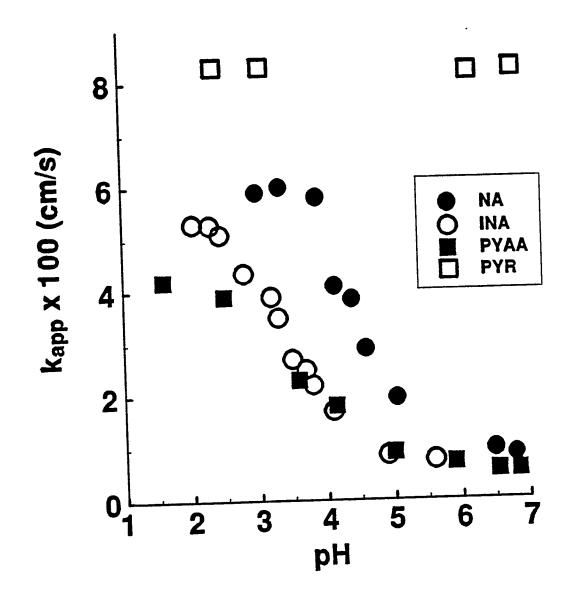


Figure 6. The k_{app} of ferricyanide at the NA/Pt, INA/Pt, PYAA/Pt, and PYR/Pt electrodes as a function of solution pH.

independent with solution pH.

The k_{app} values of ferricyanide are smaller at pyridylcarboxylic acids monolayers than that at phenylcarboxylic acids monolayers because the thickness of the former is thicker than the that of the latter as a result of different orientation as shown in Figures 4 and 6.

The Δ k_{app} is greater at phenylcarboxylic acids monolayers than that at phenylcarboxylic acids monolayers mainly because the latter has lower packing density of carboxylic acid groups and has inactive carboxylic acid groups as well.

The basis of the observed data are caused by a change in the potential at the reaction plane which can cause a change in k_{app} . For acid-terminated monolayers, the potential at the reaction plane becomes negative upon deprotonation. The electrostatic repulsion between ferricyanide and the acid-terminated surface upon deprotonation discourage the electron transfer rate of ferricyanide. This decrease in k_{app} is a consequence of the Frumkin effect⁴⁰⁻⁵² through a change in the potential in the reaction plane and is examined in detail in the next section.

Potential change at the reaction plane

The potential at the reaction plane, ϕ_r , in a heterogeneous electron transfer process will decrease as a result of the increase in the amount of negative charges from the carboxylate group generated by the deprotonation of the carboxylic acid group. The decrease in ϕ_r causes the decrease in the rate constant of ferricyanide which has negative charge as well. The change in ϕ_r can be calculated based on the following equation from Frumkin³¹

$$\Delta \ln(k_{app}) = (\alpha - z)(F/RT)(\Delta \phi_r)$$
 (2)

where α is the electron transfer coefficient, z is the apparent charge on the redox species, and F, R, and T have their conventional meanings. A value of 2 instead of -3 is used for z because of the almost complete association between K⁺ and Fe(CN) $_6^{4/3}$. A value of 0.5 is used for α^{53} . From the data in **Table 3**, these changes in ϕ_r for BA/Pt, PAA/Pt, PPA/Pt, PBA/Pt, NA/Pt,

Table 3. The ratio of k of ferricyanide at pH 2 (k_{app2}) to that at pH6 (k_{app6}), the potential change at reaction plane ($\Delta \phi_r$) and the theoretical potential change ($\Delta \phi_{theo}$) due to deprotonation, and the ratio of these two values for the following monolayers on platinum electrodes: BA/Pt, PAA/Pt, PPA/Pt, PBA/Pt, INA/Pt, and NA/Pt.

monolayers	k _{app2} /k _{app6}	$\Delta \phi_r$ (V)	$\Delta \phi_{\text{theo}} (V)$	$\Delta \phi_{ m I} / \Delta \phi_{ m theo}$
BA/Pt	1.00	0.00	-0.875	0.0000
PAA/Pt	1.00	0.00	-0.854	0.0000
PPA/Pt	1.67	-0.00525	-0.742	0.0071
PBA/Pt	1.61	-0.00500	-0.717	0.0070
INA/Pt	7.8	-0.0211	-1.76	0.0124
NA/Pt	6.9	-0.0198	-1.59	0.0123

INA/Pt and PYAA/Pt from low to high pH were calculated to be 0.00 mV, 0.00 mV, -5.25 mV, -5.00 mV, -19.8 mV, -21.11 mV and -20.0 mV, respectively (see **Table 3**).

The theoretical $\Delta \phi_{\Gamma}$ values can be calculated using the following equation

$$\Delta \phi_{\text{theo}} = \chi \Gamma F/c \tag{3}$$

where χ , Γ , and c are the molar fraction of carboxylate, the packing density, and the capacitance (a value of 23 μ F/cm² for all except a value of 31 μ F/cm² for BA/Pt, PPA, and PPA/Pt^{31,54}), respectively. Since there is no coverage data available for PYAA/Pt, there is no way of calculating the theoretical change in ϕ r for PYAA/Pt. The theoretical change in ϕ r for BA/Pt, PAA/Pt, PPA/Pt, PBA/Pt, INA/Pt, and NA/Pt were calculated to be -0.875 V, -0.854 V, -0.742 V, -0.717 V, -1.76 V, and -1.59 V, respectively. Therefore, the observed $\Delta \phi$ values for PPA/Pt, PBA/Pt, INA/Pt and NA/Pt are significantly smaller than theoretical prediction. (see **Table 3**)

Other than the difference in the packing density, the difference in $\Delta \phi_{\Gamma}$ mainly is a result of the difference in orientation of monolayers. The carboxylic acid groups of both BA/Pt and PAA/Pt are so close to the Pt surface that all the acid groups form carboxylate-Pt bonds which do not have any acid-base functionality at all. The carboxylic acid groups of PPA/Pt and PBA/Pt are not quite far away from the Pt surface so that part of the them are bonded to Pt surface and the rest are in acidic form. To the extreme, the carboxylic acid groups of INA/Pt, NA/Pt, and PYAA/Pt are completely away from Pt surface and therefore all of them are in acidic form. The results clearly demonstrates that the orientation of the monolayers can influence the properties of the films substantially.

The k_{app} of ferricyanide as a function of solution pH for PPA/Pt, PBA/Pt, INA/Pt, and NA/Pt can be fitted with the same percentage of the theoretical $\Delta \phi_{\Gamma}$ values well as shown in **Figures 7-10**. With only ~1 % of the theoretical value, the $\Delta \phi_{\Gamma}$ is most likely due to the screening effect of the

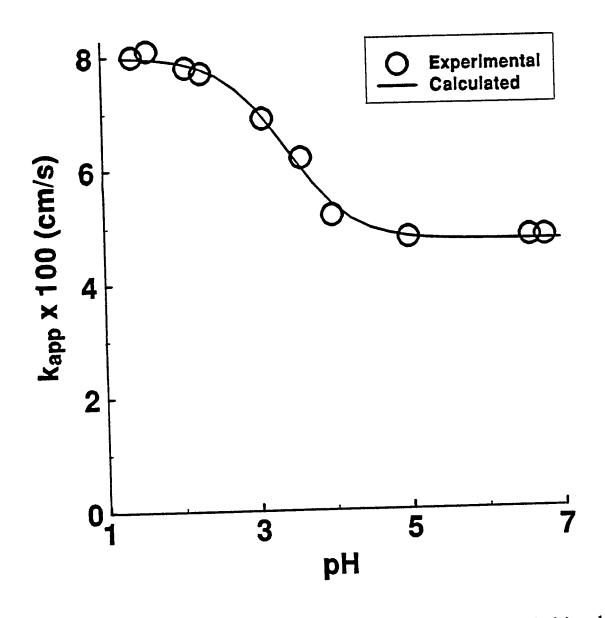


Figure 7. The k_{app} of ferricyanide at the PPA/Pt electrode (open circle) and calculated k_{app} using 0.71 % of the theoretical $\Delta \phi$ value (solid line) as a function of solution pH.

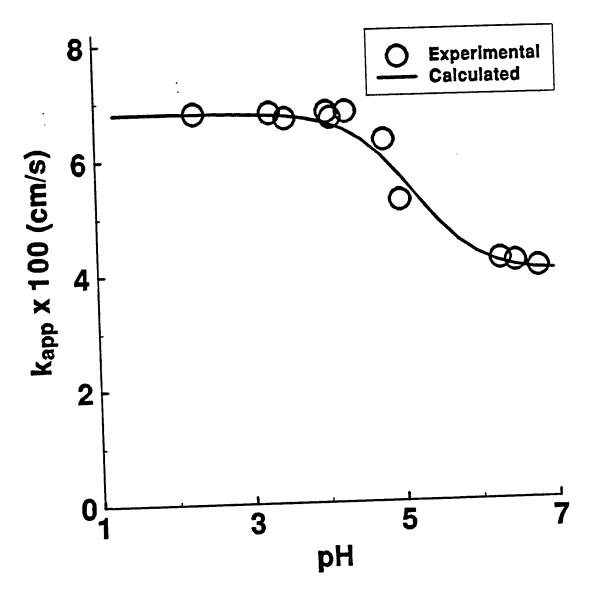


Figure 8. The k_{app} of ferricyanide at the PBA/Pt electrode (open circle) and calculated k_{app} using 0.70 % of the theoretical $\Delta \phi$ value (solid line) as a function of solution pH.

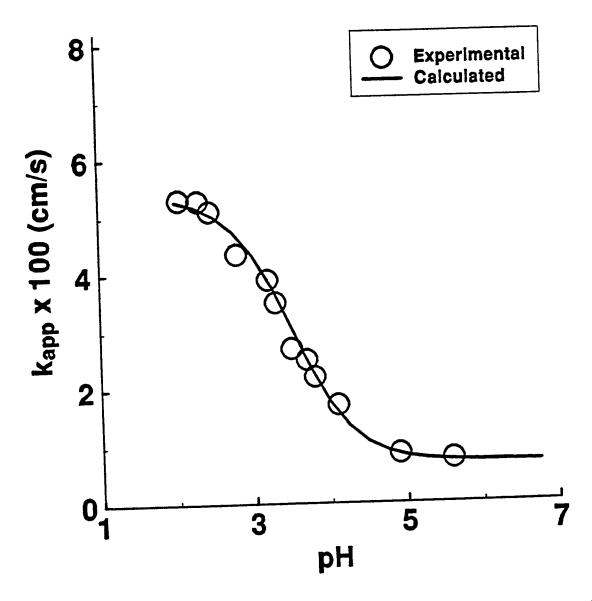


Figure 9. The k_{app} of ferricyanide at the INA/Pt electrode (open circle) and calculated k_{app} using 1.2% of the theoretical $\Delta \phi$ value (solid line) as a function of solution pH.

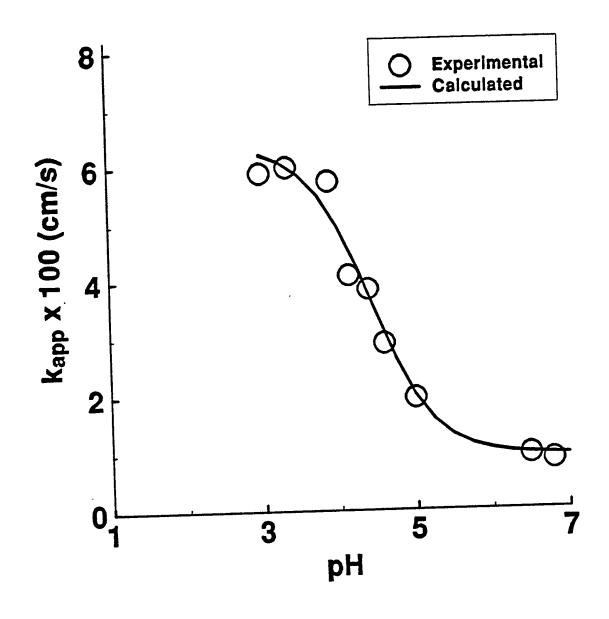


Figure 10. The k_{app} of ferricyanide at the NA/Pt electrode (open circle) and calculated k_{app} using 1.2% of the theoretical $\Delta \phi$ value (solid line) as a function of solution pH.

counter ions at the reaction plane because of the COO group^{51,55} i.e. most of the negative charges at the electrode are screened by K⁺ and H₂O.

Determination of pKa using electrochemistry methods

Since the decrease in k_{app} is associated with deprotonation process, it is possible to calculate the pK_a for these monolayers on Pt from the k_{app} vs. pH plot. The pK_a values for PPA/Pt, PBA/Pt, INA/Pt, NA/Pt and PAA/Pt were calculated based on this plot by using equation (1) at 50 % of theoretical $\Delta \phi_T$ values to be 3.5 ± 0.3 , 5.3 ± 0.3 , 3.9 ± 0.3 , 4.8 ± 0.3 , and 4.3 ± 0.4 , respectively. To make sure that this decrease in k_{app} is only due to deprotonation process, the same kind of experiments were studied at PYR/Pt electrodes and the result is shown in **Figure 6**. Constant k_{app} values of these non-acid-terminated monolayers confirm the previous conclusion. These pK_a values are slightly higher than that of the corresponding precursors in water except PPA/Pt. The decrease in pK_a for PPA/Pt is most likely due to the electron withdrawing effect of the aromatic ring when coupled to Pt.

Determination of pKa using IRRAS

The IRRAS spectra in **Figure 11** are the low energy spectra of INA/Pt as a function of solution pH. Obviously, the peak intensity of $\nu(C=O)$ decreases with increasing solution pH while that of $\nu_s(COO-)$ peak at 1389 cm⁻¹ and the $\nu_a(COO-)$ at 1520 cm⁻¹ increase with increase in solution pH. Notice that the $\nu(C-O)$ at 1181 cm⁻¹ decreases with increase in solution pH.

The complete disappearance of the peak at 1181 cm⁻¹ at high solution pH proves that this peak is the ν (C-O) mode only without anything else as cited in literature³⁰.

The peak areas of $\nu(C=O)$, $\nu_s(COO)$, and $\nu(C=O)$ are shown in Figure 12. The peak area of non-hydrogen-bonded $\nu(C=O)$ starts to decrease at very low pH and has a transition of ~ 3 pH units. The peak area of the hydrogen-bonded dimeric C=O starts to decrease at pH ~ 3.5 with a transition range of 6 pH units. The pK_a values calculated from the reflection

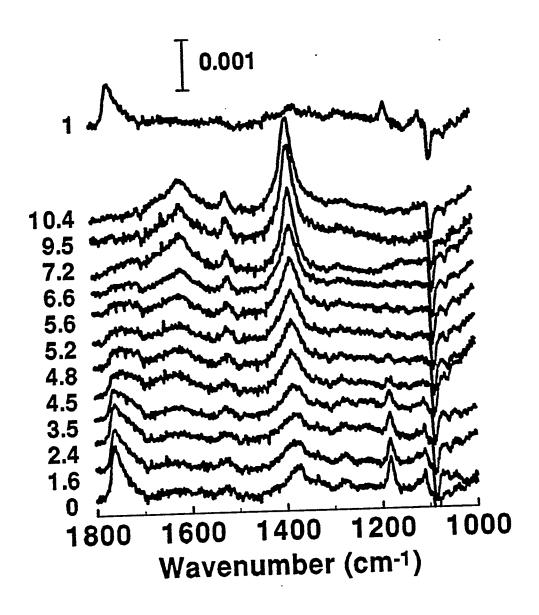


Figure 11. IRRAS spectra in the low energy region for INA/Pt at 80° degree of incident angle from the surface as a function of solution pH.

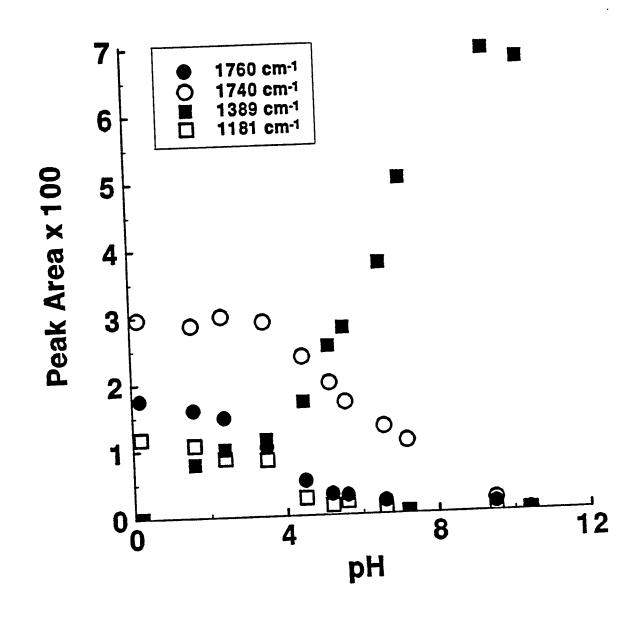


Figure 12. The IRRAS peaks at 1760 cm⁻¹, 1740 cm⁻¹, 1389 cm⁻¹, and 1181 cm⁻¹ of INA/Pt at 80° degree of incident angle from the surface as a function of solution pH.

point of the curves for non-hydrogen-bonded and hydrogen-bonded dimeric C=O groups are 3.8 and 6.0, respectively. The pK_a value of non-hydrogen-bonded C=O is almost the same as that of the adsorbate precursor in water (3.44)⁵⁶. On the other hand, the pK_a of hydrogen-bonded dimeric C=O group is higher than that of the adsorbate precursor in water by 2.56 pH unites. The calculated pK_a value for NA/Pt is listed in **Table 4** for comparison as well.

Interestingly, the change in the peak area of $\nu(\text{C-O})$ not only has a reflection point the same as the pKa of non-hydrogen-bonded C=O group but also has the same trend as non-hydrogen-bonded C=O group. This suggests that the $\nu(\text{C-O})$ mode observed at 1181 cm⁻¹ is associated with non-hydrogen-bonded C=O group rather the hydrogen-bonded dimeric C=O group. This implies that the spatial orientation of non-hydrogen-bonded C=O group is quite different from that of hydrogen-bonded dimeric C=O group. It is strange that there is not much of the amount of COO- until pH 4. In fact the peak area of the COO- increases from ~0.01 at pH 4 to ~0.07 at pH 10. This suggests that the orientation of the $\nu_s(\text{COO-})$ dipole from the non-hydrogen-bonded C=O group is more parallel to the Pt surface than that from the hydrogen-bonded diemric C=O group.

Determination of pKa using contact angle measurements

Contact angle measurements have long been used to probe wettability⁵⁷. Recently, this method was used to "titrate" acidity of polymers⁵⁸ and carboxylic acid monolayers⁵⁹ structure and metal oxides⁶⁰. The basis of a contact angle measurement is based on the Young equation⁶¹.

$$\gamma_{\rm SV} - \gamma_{\rm SL} = \gamma_{\rm LV} \cos \theta \tag{3}$$

where γ_{SV} and γ_{LV} are the respective solid and liquid surface tensions in equilibrium with the saturated liquid vapor, γ_{SL} is the surface tension between solid and liquid, and θ is the contact angle between liquid and solid. Since changes in both γ_{SV} and γ_{LV} with pH are relatively small, it follows that decreases in θ with pH reflect an increase in γ_{SL}^{60} . This is the

Table 4. The pK_a values of BA/Pt, PAA/Pt, PPA/Pt, PBA/Pt, INA/Pt, NA/Pt and PAA/Pt monolayers based on IRRAS, electrochemistry (ECHEM) and contact angle (CA) measurements.

pKa	IRRASª	ECHEM ^a	CAª	aqueous solution value
BA/Pt	ъ	none	c	4.19
PAA/Pt	ъ	none	c	4.9
PPA/Pt	ъ	3.5	С	4.9
PBA/Pt	ъ	5.3	С	5.0
INA/Pt	3.8,6.0	3.9	4.0	3.75^{d}
NA/Pt	3.9,6.0	4.8	4.7	3.44 ^d
PAA/Pt	b	4.3	4.6	3.88 ^d

^aThe standard deviation for IRRAS, ECHEM, and CA data are 0.3, 0.3, and 0.4 pK units, respectively.

^bNo detectable spectroscopic signal.

^cThe contact angle at low pH is low for titration.

^d Refer to reference 56.

basis of the "contact angle titration" measurement.

The following presents results from using this technique to characterize three of our reactive monolayers samples (i.e. NA/Pt, INA/Pt, and PAA/Pt) and a control sample (PYR/Pt); the low surface tension of other monolayers precludes this characterization. As with the IRRAS data, these measurements were performed for methods verification. The contact angles of water at NA/Pt, INA/Pt, PAA/Pt, and PYR/Pt as a function of pH are shown in Figures 13-15. As can been seen, the contact angles of water at NA/Pt, INA/Pt, and PAA/Pt decrease upon increases in pH. This is due to progressive ionization of the carboxylic acid functionalities, which more and more amount of the carboxylate groups which have higher surface energy than that of carbonyl groups upon deprotonation 58,59. As expected, PYR/Pt does not show any change as shown in Figure 16.

It is possible to calculate pK_a of these acid on Pt surface based on the relationship between $cos\theta_a$ and solution pH. The pK_a values of INA/Pt, NA/Pt and PAA/Pt surface, which were calculated at 50 % of the total change in $cos\theta_a$, were found to be 4.0 ± 0.4 , 4.7 ± 0.4 , and 4.6 ± 0.4 , respectively. (see **Table 4**)

Comparison of pKa values

The pK_a values of NA/Pt, INA/Pt and PAA/Pt have been calculated by using IRRAS, cyclic voltammetry and contact angle measurements and are listed in **Table 4**. Although the standard deviations of these values are a little large because of experiment difficulties, it is possible to make some comparisons from these values.

Owing to too weak IRRAS signals of PAA/Pt, there is no pK_a value available for PAA/Pt from IRRAS data. The pK_a values of pyridylcarboxylic acids in water are calculated values⁵⁷ instead of the dissociation constants because these acids are zwitterionic. From IRRAS data, both NA/Pt and INA/Pt monolayers have two pK_a values. The pK_a values for non-hydrogen-bonded C=O group are almost the same as that in water. On the other hand, the pK_a values for dimeric hydrogen-bonded C=O group for NA/Pt and INA/Pt are higher than that in water by ~2.5 pH

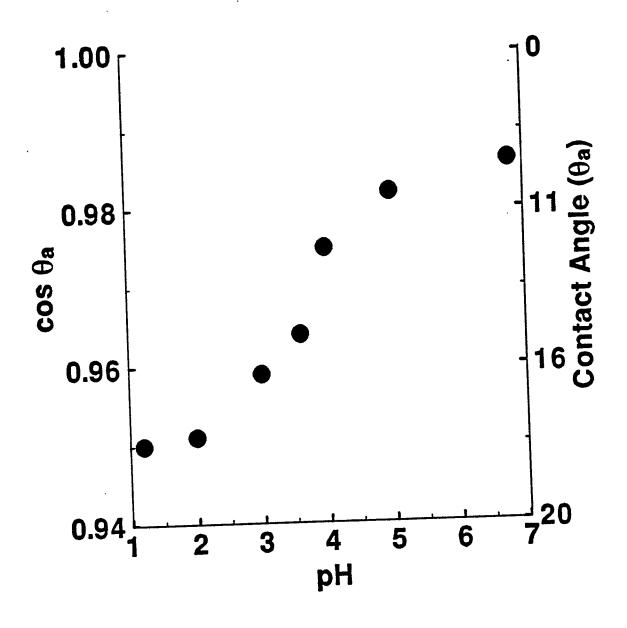


Figure 13. The $\cos\theta_a$ of water on the INA/Pt surface as a function of solution pH.

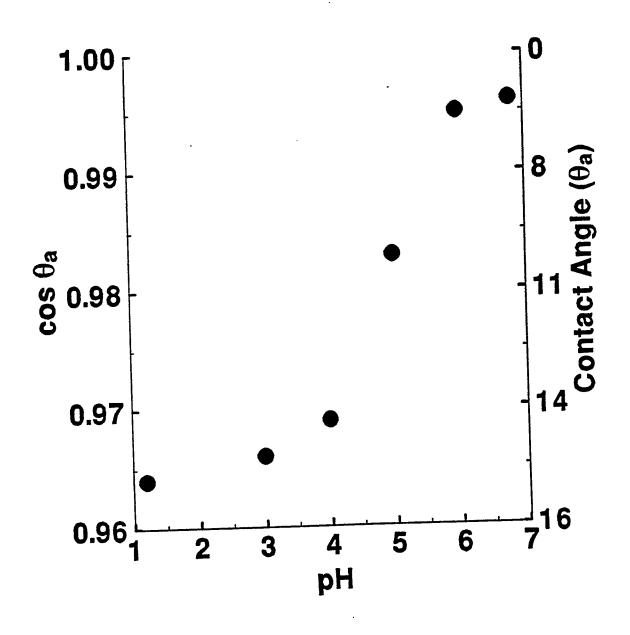


Figure 14. The $cos\theta_a$ of water on the NA/Pt as a function of solution pH.

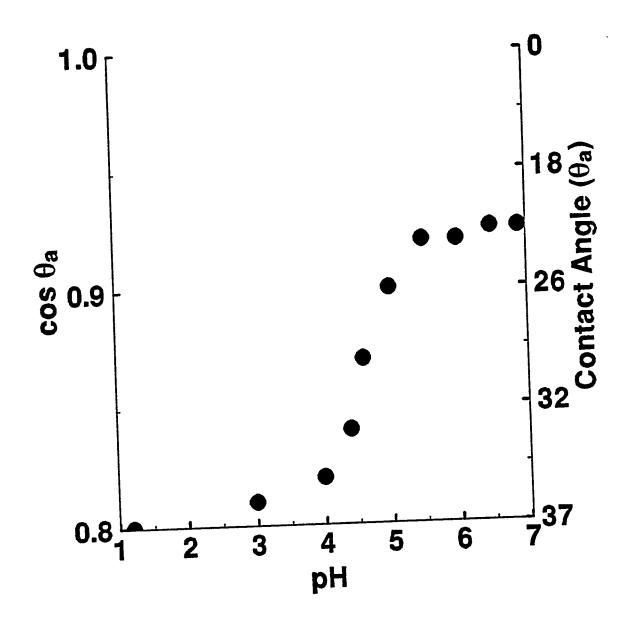


Figure 15. The $\cos\theta_a$ of water on the PYAA/Pt as a function of solution pH.

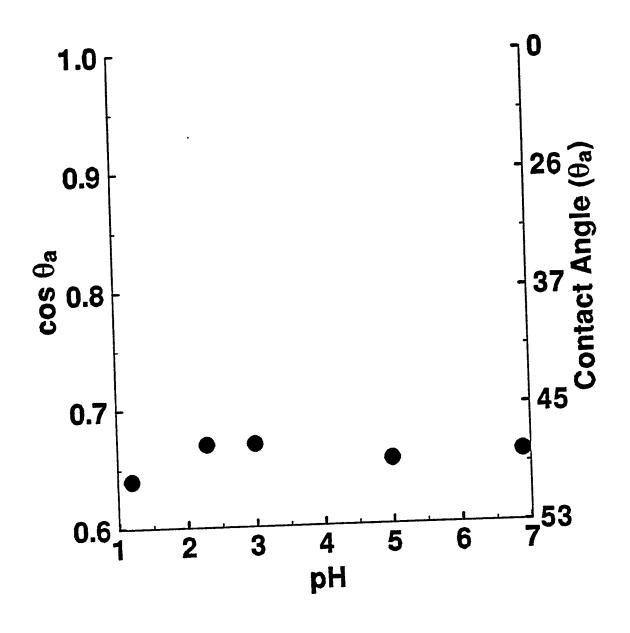


Figure 16. The $\cos\theta_a$ of water on the PYR/Pt as a function of solution pH.

units. This strongly suggests that the decrease in the acidity of both INA/Pt and NA/Pt is primly due to the presence of the hydrogen bonding but not the low dielectric environment.

Both electrochemistry and contact angle measurements can only provide the overall acidity and give almost the same pK_a values for INA/Pt, NA/Pt, and PAA/Pt. All these INA/Pt, NA/Pt, and PAA/Pt monolayers have higher pK_a values than that of corresponding precursor in water by ~0.2, ~1.3, ~0.6 pH units, respectively. In other words, deprotonation process of these pyridylcarboxylic acid on Pt surfaces is more difficult than in water due to the existence of hydrogen bonding.

CONCLUSIONS

Self-assembly monolayers of short methylene phenylcarboxylic acids on Pt can form the closest-packed structure with the rings and methylene groups oriented parallel to the Pt surface that can be exhaustively oxidized to CO₂ and H₂O electrochemically. Unlike phenylcarboxylic acids, pyridylcarboxylic acids are bonded to Pt surface through nitrogen rather the aromatic rings and have the ring oriented almost perpendicular to the surface. This makes pyridylcarboxylic acid monolayers on Pt more difficult to be oxidized electrochemically than aromatic acid monolayers on Pt.

Deprotonation of all these acids on Pt electrodes causes 0 % to 40 % and 85% decrease in the heterogeneous electron transfer rate of ferricyanide at phenylcarboxylic acids coated Pt electrodes and pyridylcarboxylic acids coated Pt electrodes, respectively. The changes in the potential at the reaction plane are about 0 mV to -5 mV and -20 mV for phenylcarboxylic acid coated Pt electrodes and pyridylcarboxylic acids coated Pt electrodes, respectively. The observed $\Delta \phi_r$ values due to deprotonation for PPA/Pt and PBA/Pt are only 0.7 % of the theoretical values and that for INA/Pt and NA/Pt are only 1.2 % due to the screening effect of the electrolyte and water. No change in ϕ_r was observed for BA/Pt and PAA/Pt electrodes because both BA and PAA lose the carboxylic acid groups upon immobilization onto Pt surfaces.

INA, NA, PYAA become weaker acids mainly because of the formation of hydrogen bond while PPA becomes a stronger acid because of the electron withdrawing nature of the aromatic ring.

REFERENCES

- (1) Kajzar, F.; Messier, J.; Zyss, J.; Ledoux, I. Opt. Commun., 1983, 45, 13.
- (2) Carter, G. M.; Chen, Y. J.; Tripathy, S. K. Opt. Engl. 1985, 24, 609.
- (3) Mathur, S.C.; Scheinbeim, J. I.; Newman, B. A. J. Appl. Phys. 1984, 56, 2419.
- (4) Murray, R. W. Annu. Rev. Mater. Sci. 1984, 14, 145.
- (5) Stern, D. A.; Wellner, E.; Salaita, G. N.; Laguren-Davidson, L.; Lu. F.; Batina, N. Frank, D. G.; Zapien, D. C.; Walton, N.; Hubbard, A. T. J. Am. Chem. Soc. 1988, 110, 4885.
- (6) Soriaga, M. P.; Hubbard, A. T. J. Am. Chem. Soc. 1982, 104, 2735.
- (7) Soriaga, M. P.; Wilson, P. H.; Hubbard, A. T.; Benton, C. S. J. *Electroanal. Chem.* **1982**, *142*, 317.
- (8) Soriaga, M. P.; Hubbard, A. T. J. Am. Chem. Soc. 1982, 104, 3937.
- (9) Hubbare, A. T.; Stickney, J. L.; Soriaga, M. P.; Chia, V. K. F.; Rosasco, S. D.; Schardt, B. C.; Solomun, T.; Song, D.; White, J. H.; Wieckowski, A. J. Electroanal. Chem. 1984, 168, 43.
- (10) Lai, C, N.; Hubbard, A. T. Inorg. Chem. 1972, 11, 2081.
- (11) Ishikawa, R. M.; Katekaru, J.; Hubbard, A. T. J. Electroanal. Chem. 1978, 86, 271.
- (12) Katekaru, J.; Hershberger, J.; Garwood, G. A. Jr.; Hubbard, A. T. Surface Sci. 1982, 121, L524.
- (13) Lumsden, J. B.; Garwood, G. A.; Hubbard, A. T. J. Am. Chem. Soc. 1982, 104, 3937.
- (14) Soriaga, M. P.; Hubbard, A. T. J. Am. Chem. Soc. 1982, 104, 2742.
- (15) Gui, J. Y.; Kahn, B. E.; Laguren-Davidson, L.; Lin, C.-H.; Lu, F.; Salaita, G. N.; Stern, D. A.; Hubbard, A. T. Langmuir 1989, 5, 819.
- (16) White, J. H.; Soriaga, M. P.; Hubbard, A. T. J. Electroanal. Chem. 1984, 177, 89.
- (17) Soriaga, M. P.; Stickney, J. L.; Hubbard, A. T. J. Electroanal. Chem. 1983, 144, 207.
- (18) Stern, D. A.; Salaita, G. N.; Lu, F.; McCargar, J. W.; Batina, N.; Frank, D. G.; Laguren-Davidson, L.; Lin, C.-H.; Walton, N.; Gui, J. Y.; Hubbard, A. T. Langmuir 1988, 4, 711.

- (19) Soriaga, M. P.; Hubbard, A. T. J. Electroanal. Chem. 1984, 167, 79.
- (20) Sasaki, K.; Kunai, A.; Harada, J.; Nakaboni, S. *Electrochim. Acta*, 1983, 28, 671.
- (21) Vasiliev, Yu. B.; Bagotzky, V. S.; Khazova, O. A.; Krasnova, N. N.; Sergeeva, T. A.; Bogdanovich, V. B. *Electrochim. Acta* 1981, 26, 545.
- (22) Vasiliev, Yu. B.; Bagotzky, V. S.; Khazova, Cherny, V. V.; Meretsky, L. M. J. Electroanal. Chem. 1979, 98, 253.
- (23) Mirkind, L. A.; Fioshin, M. Y.; Dubinin, A. G. Elektrokhimiya 1973, 9, 257.
- (24) Mirkind, L. A.; Kazarinov, A. N.; Dubinin, A. G.: Fioshin, M. Y. Elektrokhimiya 1973, 9, 1067.
- (25) Mirkind, L. A.; Kazarinov, A. N.; Dubinin, A. G.: Fioshin, M. Y. Elektrokhimiya 1973, 9, 1069.
- (26) Gileadi, E.; Duic, L.; Bockris, J.O'M. *Electrochim. Acta* 1968, 13, 1915.
- (27) Bockris, J.O'M.; Green, M.; Swinkels, D. A. J. J. Electrochem. Soc. 1964, 111, 743.
- (28) Horanyi, G.; Nagy, E. J. Electroanal. Chem. 1973, 43, 441.
- (29) Horanyi, G.; Andreev, V. N. J. Electroanal. Chem. 1982, 18, 1209.
- (30) Stern, D. A.; Laguren-Davidson, Laarni; Frank, D. G.; Gui, J. Y.; Lin, C.-H.; Lu, F.; Salaita, G. N.; Walton, N.; Zapien, D. C.; Hubbard, A. T. J. Am. Chem. Soc. 1989, 111, 877.
- (31) Fawcett, W. R. J. Chem. Phys. 1974, 61, 3842.
- (32) Parson, R. J. Electroanal. Chem. 1969, 21, 35.
- (33) Nicolson, R. S. Anal. Chem., 1965, 37(11), 667.
- (34) Heyrovsky, J.; Kuta, J. *Principle of Polarography*, Academic Press, New York, **1966**, p. 106.
- (35) Nicolson, R. S. Anal. Chem., 1964, 37, 1351.
- (36) Perone, S. P. Anal. Chem., 1966, 38, 1158.
- (37) Stickney, J. L.; Soriaga, M. P.; Hubbard, A. T.; Anderson, S. E. J. Electroanal. Chem. 1981, 125, 73.
- (38) Pauling, L. C. The Nature of the Chemical Bond, 3rd ed.; New York: Cornell University Press, 1960.

- (39) Lane, R. F.; Hubbard, A. T. J. Phys. Chem. 1973, 77, 1411.
- (40) Frumkin, A. N. Z. Physik. Chem. 1933, 164, 121.
- (41) Frumkin, A. N.; Florianovich, G. M. Zh. Fiz. Khim. 1955, 29, 1827.
- (42) Frumkin, A. N. Z. Elektrochem. 1955, 59, 807.
- (43) Frumkin, A. N. Advan. Electrochem. Eng. 1961, 1, 65.
- (44) Parsons, R. Advan. Electrochem. Eng. 1961, 1, 1.
- (45) Grahame, D. C. J. Electrochem. Soc. 1951, 98, 343.
- (46) Grahame, D. C. J. Am. Chem. Soc. 1958, 80, 4201.
- (47) Grahame, D. C.; Parsons, R. J. Am. Chem. Soc. 1961, 83, 1291.
- (48) Lawrence, J.; Parsons, R.; Payne, R. J. Electroanal. Chem. 1968, 16, 193.
- (49) Schimid, R. W.; Reilley, C. N. J. Am. Chem. Soc. 1958, 80, 2087.
- (50) Deakin, M. R.; Stutts, K. J.; Wightman, R. M. J. Electroanal. Chem. 1985, 182, 113.
- (51) Weaver, M. J.; Satterberg, T. L. J. Phys. Chem. 1977, 81, 1772.
- (52) Hu, I.-F.; Karweik, D. H.; Kuwana, T. J. Electroanal. Chem. 1985, 188, 59.
- (53) Scherer, G.; Willig, F. J. Electroanal. Chem. 1977, 85,77.
- (54) Randin, J. P.; Yeager, E. J. Electroanal. Chem. 1975, 58, 313.
- (55) Bard, A. J.; Faulkner, L. R. Electrochemical Methods Fundamentals and Application, New York: Wiley, 1980; pp513-516.
- (56) Blanch, J. H. J. Chem. Soc. (B), 1966, 937.
- (57) Kaelble, D. H. *Physical Chemistry of Adhesion*, New York: Wiley, 1971.
- (58) Holmes-Farlsy, S. R.; Whitesides, G. M. Langmuir, 1987, 3, 62.
- (59) Bain, C. D.; Whitesides, G. M. Langmuir 1989, 5, 1370.
- (60) Chau, L.-K.; Porter, M. D. J. Colloid and Interface Sci. 1991, 145, 283.
- (61) Adamson, A. W. Physical Chemistry of Surface, 4th ed.; Wiley: New York, 1982, p338;

PAPER II

INFRARED REFLECTION ABSORPTION SPECTROSCOPIC STUDY OF THE TEMPORAL EVOLUTION OF THE STRUCTURE OF ALKANE THIOLATE MONOLAYERS ON GOLD

INTRODUCTION

The high stability, well-ordered, densely-packed structure, simple fabrication process, and very flexible modification of the self-assembled monolayers on gold have attracted worldwide attention. As a consequence, the study of the properties and construction of the self-assembled thiol monolayers on gold has grown enormously in recent years¹⁻³.

Porter and co-workers used infrared reflection absorption spectroscopy (IRRAS), ellipsometry and contact angle measurements to demonstrate that the tilt angle is 30° with twist angle of 50° and there is a odd-even chain-length-dependent orientation of the methyl group which provides the best evidence of a conserved Au-S-C bond angle of the thiolate on Au monolayers⁴⁻⁶. Monolayers have a well ordered pattern of $\sqrt{3} \times \sqrt{3} \text{ R30°}$ with 5.0 Å separation to the nearest neighbor from STM images^{7,8}. The packing density calculated from the electrochemical reduction desorption data of these is 8.0 × 10^{-10} mol/cm² on Au/mica annealed at 300° C assuming roughness factor of 1.2°. This supports the $\sqrt{3} \times \sqrt{3} \text{ R30°}$ structure with 30° tilt angle from the surface normal. The loss of the mercapto hydrogen and formation of a gold alkanethiolate was concluded⁹.

Nuzzo and co-workers have used low-energy electron diffraction (LEED) and concluded that these monolayers have a $\sqrt{3}$ x $\sqrt{3}$ R30° with a 4.99 Å value for the distance between the nearest neighbor structure¹⁰. These molecular architecture is adopted independent of the chain length or chain-terminating group and the tilt angle is ~30° with twist angle of 52° which implies that the gold surface-sulfur-carbon bond angle is near 120° .¹¹

Whitesides and co-worker have shown that the distance between the nearest neighbor is 4.97 ± 0.05 Å by transmission electron diffraction spectroscopy¹². Bain and Whitesides have studied the wetting properties of a series of ethers of the form $HS(CH_2)_{16}O(CH_2)_nCH_3$, where n varied from 0 to 5. They found that when n was >1 the surface properties of these material were identical to those of the pure n-alkane thiol monolayers¹³. This has demonstrated that these monolayers are sufficiently densely packed so that the

surface properties measured are only those of the top layer but not those of the underlying polymethylene chains.

Pemberton and co-worker have used surface Raman scattering spectroscopy (SERS) and shown that the H in H-S bond is gone when the molecule adsorbed, the C-S bond is largely parallel to the surface at Au. The S-C bond at 630 cm⁻¹ is gauche mode , 700 cm⁻¹ is trans mode. Gauche mode decreases with immersion time but small amount of gauche still exists due to defects in the film at grain boundaries on the polycrystalline Au surface. They calculated the tilt angle of 30° and twist angle of ca. 45°. 14

Grunze and co-workers have calculated the tilt angle of these monolayers is ~35° by second harmonic generation spectroscopy¹⁵. and nearedge X-ray absorption fine structure spectroscopy (NEXAFS)^{16,17}.

Shimazu and co-workers have used the electrochemical quartz micro balance and demonstrated that the number of adsorbed thiol molecules, 2.6×10^{14} molecules/cm², estimated from the frequency change, suggests the formation of a monolayer of this thiol. This number agrees well with that calculated from the electrochemical charge associated with oxidation/reduction of the monolayer, 2.4×10^{14} molecules/m² for ferrocenylundecanethiol¹⁸.

Klein and co-workers have used molecular dynamics calculations to demonstrate that these monolayers form a hugely corrugated yet dense surface phase with the chain cant toward the next-nearest neighbor ^{19,20}. The monolayers must be densely packed at 300° K with few gauche defects. The defects that exist are concentrated near the chain terminal ²¹.

Ulman and co-workers have used molecular dynamics calculation and suggested that the tilt angle should be 30° for n-alkane thiol monolayers on Au^{22} .

Chidsey and co-workers have used helium diffraction technique and shown that the difference between the azimuths of maximum intensity for the (-1, -1) and (n, 0) diffraction peaks is 30° , as would be expected for oriented domains of hexagonally packed adsorbates. The resulting lattice has a distance between the nearest neighbor of 5.01 ± 0.02 Å. Shorter chains give the same lattice constant but weaker diffraction, and the domains appear to be much less well oriented.²³.

Although there are a few studies on the formation of these monolayers with a finding of a two-step processes including a Langmuirian kinetic model $^{24-27}$, little work has been done concerning the details of this very fundamental adsorption process. In this study, the formation and the orientation of a 16-mercaptohexadecanoic acid $(HO_2C(CH_2)_{15}SH)$ and a n-octadecanethiol monolayers on Au were characterized using IRRAS as a function of immersion time. The discovery of agostic bonds at 2903 cm⁻¹ due to the interaction between methylene groups and a bare Au surface based on the shift in energy of the $\nu_a(CH_2)$ and the absence of the agostic bond at 2903 cm⁻¹ on the Au surface which was pretreated with a sulfur monolayer is reported as well.

EXPERIMENTAL SECTION

Substrate preparation

Electrodes were prepared by the resistive evaporation of 15 nm of chromium, followed by 300 nm of gold, onto 3 inch by 1 inch glass substrates. The temperature during the evaporation, measured at the plate supporting the substrates, increased to ~50° C as a result of radiative heating by the evaporation source. The evaporation rates were 0.2 and 0.5 nm/s for chromium and gold, respectively. The pressure in a cryopumped E360A Edwards Coating System during evaporation was <9x10⁻⁵ Pa. After ~45 min cooling time, the evaporator was back-filled with purified N₂. The substrates were then removed, analyzed by optical ellipsometry for the determination of the optical functions of the uncoated substrates, and immersed in the thiol-containing solution.

The surfaces of the uncoated gold electrodes have been characterized previously at both a macroscopic and microscopic level using scanning tunneling microscopy⁷ and electrochemical measurements of surface texture and surface area ⁹. Macroscopically, the STM images show that the electrode surface is composed of atomically-terraced crystallites ~25 nm in diameter. The electrodes have a roughness factor of ~1.3, which is given as the surface area determined from the oxidative desorption of iodine⁹ divided by the geometric electrode area.

Monolayer preparation

Thiolate monolayers were prepared by their spontaneous adsorption onto the evaporated gold from 1.00×10^{-6} M solutions in absolute ethanol for different periods of immersion time and rinsed successively with absolute ethanol and hexane, and dried under a stream of prepurified N_2 .

Ellipsometric measurements of film thickness

The thicknesses of the monolayers were determined by optical ellipsometry at 632.8 nm in two steps with a computer-interfaced Gaertner Model L-116B ellipsometer. The angle of incidence was 70° from the surface normal. Upon removal from the evaporation chamber, the analyzer and

polarizer angles for a reflected light beam from each uncoated substrate were measured on at least three different spots. The average complex refractive index for each substrate was then calculated with a two phase parallel layer model from classical electromagnetic theory²⁸. After monolayer formation, each sample was again analyzed and the film thickness calculated from a three-phase parallel layer model, using the average complex refractive index of the individual sample and a real refractive index of 1.45 for the film. A value of 1.45, which is representative of the adsorbate precursors, facilitates comparison with thickness data that has been reported for a variety of monolayers. The influence of several refinements on this treatment of ellipsometric data have been recently discussed²⁹.

Infrared spectroscopy

Infrared reflection absorption spectra (IRRAS) were acquired with a Nicolet 740 spectrometer. Spectra of the monolayers were obtained in a single reflection mode by using p-polarized light incident at 82° from the surface normal and a liquid nitrogen cooled HgCdTe detector. A home-built sample holder was used to position the substrates reproducibly in the sample chamber of the spectrometer³⁰. The spectrometer was purged with boil-off from liquid N₂. Spectra were obtained by referencing 1024 sample scans to 1024 background scans at 2 cm⁻¹ resolution (zero filled) with Happ-Genzel apodization. All spectra are reported as -log(R/R_o), where R is the reflectance of the coated substrate and R_o is that of a reference Au substrate. The reference substrates were prepared by immersion of an uncoated gold substrate into ~1 mM ethanolic solution of octadecanethiol-d₃₇. for 24 hours.

Chemicals and reagents

The 16-mercaptohexadecanoic acid (HO₂C(CH₂)₁₅SH) (MHA) was synthesized in this group and characterized with IR and NMR. All other reagents including n-octadecanethiol (ODT) were obtained from Aldrich and used as received. Aqueous solutions were freshly prepared from water purified by passage through a Milli-Q purification system (Millipore Corp., Bedford, MA).

RESULTS AND DISCUSSION

IRRAS spectra as a function of immersion time MHA /Au

At low coverage, MHA/Au has only one observable $\nu_a(CH_2)$ mode at 2903 cm⁻¹ with no $\nu_s(CH_2)$ observed as shown in Figure 1. The absence of the $\nu_a(CH_{3,ip})$, $\nu_s(CH_3,FR_1)$, and $\nu_s(CH_3,FR_2)$ modes at 2965 cm⁻¹, 2936 cm⁻¹, and cm⁻¹, respectively, confirms the formation of the acid-terminated monolayer. The absence of the $\nu_s(CH_2)$ and the presence of the $\nu_a(CH_2)$ strongly suggests that all the polymethylene groups must lie on the Au surface in order to have the $\nu_s(CH_2)$ dipole parallel to the Au surface and the $\nu_a(CH_2)$ perpendicular to the Au surface at the same time. The peak at 2903 cm⁻¹ increases with increases in coverage and starts to decrease at high coverage where the $\nu_a(CH_2)$ and $\nu_s(CH_2)$ modes start to appear at 2926 cm⁻¹ and 2860 cm⁻¹, respectively. These two $\nu_s(CH_2)$ peaks increase in peak area to a certain point where the peak at 2903 cm⁻¹ totally disappear and decrease in peak area with decreases in peak frequency.

Figure 2 shows the IRRAS spectra in the low energy region of a MHA/Au as a function of immersion time. Peaks found at 1380 cm⁻¹, 1396 cm⁻¹, 1436 cm⁻¹, 1460 cm⁻¹ are ω (CH₂), δ (α -CH₂), ι (C-O),and δ (CH₂), respectively. All the IRRAS modes assignment is listed in **Table 1**. The intensity of the δ (α -CH₂) and ι (C-O) peaks decrease with increasing the coverage while that of ω (CH₂) and δ (CH₂) increase with increasing the coverage.

ODT/Au

The same trend was found in the case of ODT/Au as shown in Figures 3 and 4: it has only one observable $v_a(CH_2)$ mode at 2903 cm⁻¹ with no $v_s(CH_2)$ at low coverage and the intensity of the $\delta(\alpha$ -CH₂) peak decreases with increasing the coverage while that of $\omega(CH_2)$ increases with increasing the coverage.

In Figure 3, the peak at 2965 cm⁻¹, 2936 cm⁻¹, and 2878 cm⁻¹ are assigned as the $\nu_a(\text{CH}_{3,ip})$, $\nu_s(\text{CH}_3,\text{FR}_1)$, and $\nu_s(\text{CH}_3,\text{FR}_2)$ modes, respectively.

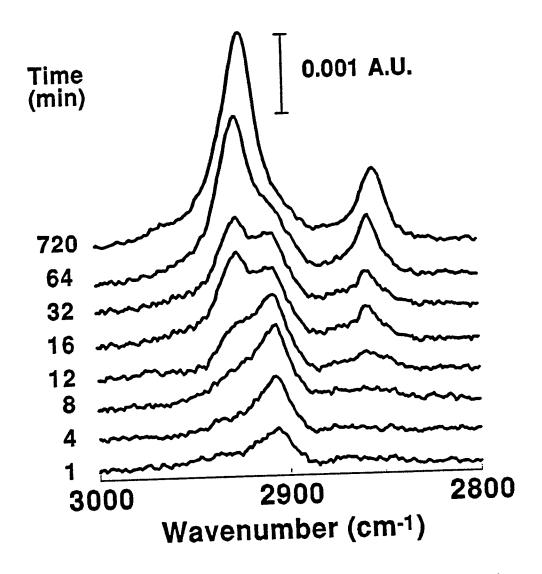


Figure 1. The IRRAS spectra in the high energy region of MHA on Au as a function of immersion time.

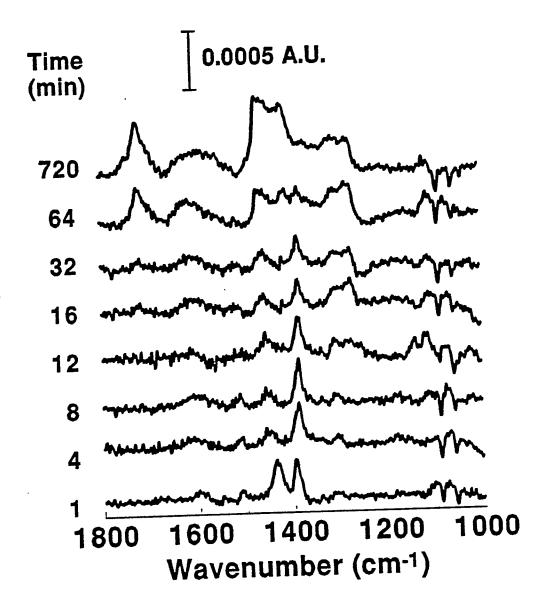


Figure 2. The IRRAS spectra in the low energy region of MHA on Au as a function of immersion time.

Table 1. Peak positions, mode assignments, and transition dipole moment directions of carboxylic acid- and methyl-terminated alkanethiolate monolayers at gold.

	HO ₂ C(CH ₂) ₁₅ S/Au		CH ₃ (CH ₂) ₁₇ S/A		.u 	
Mode assignment	obs.	calc.	obs.	calc.	direction of transition dipole ^a	
$\nu_{\rm a}({ m CH_{3,ip}})$			2964		to C-CH₃ bond in C-C-C chain plane	
$\nu_{\rm a}({ m CH_{3,op}})$			b	2954	to C-CH ₃ bond	
$v_{\rm s}({\rm CH_3,FR_1})^{\rm c}$			2936		to C-CH ₃ bond	
$\nu_{\rm a}({ m CH_2})$	2919 ^d 2903 ^e	2920	2919 ^d 2903 ^e	2920		
$\nu_{\rm s}({ m CH_3,FR_2})^{\rm c}$			2878		to C-CH3 bond	
$\nu_{\rm S}({ m CH_2})$	2850 ^j	2850	2850 ^j	2851	to C-C-C plane bisceting H-C-H	
v(C=O)	1743	1699			to C=O bond	
((0 0)	1717,168	0				
δ(CH ₂)	1460	1472	1468	1467	in HCH plane, bisecting HCH	
ν(C-O)+	1436	1431			to C-O bond ^f	
δ(C-O-H) ^g						
$\delta_a(\mathrm{CH_3})$			1439	1450	$h \stackrel{1}{\downarrow}$ to C- CH ₃ bond	
$\delta(\alpha - CH_2)$	1396	1410	1394		1	
$\delta_s(\mathrm{CH_3})$,	1375	5h to C-CH3 bond	
$\omega(CH_2)$	1382	1382	1380	1380	to axis of chain	
and twists	1170 -	*	1344	1170)	

Table 1. (continued)

^aOrientation of transition dipoles for CH modes assuming all-trans conformation (Snyder, R. G. J. Chem. Phys. 1965, 42, 1744). ^bThe $v_a(CH_{3,op})$ mode was not observed in the monolayer spectrum. ^cThe fermi-resonance couplet, $\nu_{\rm S}({\rm CH_3,FR_1})$ and $\nu_{\rm S}({\rm CH_3,FR_2})$, is designated by the subscripts 1 and 2, which refer to the higher and lower energy components, respectively. ^dThe peak frequency of this band shifts from ~2926 cm⁻¹ to 2919 cm⁻¹ as increasing immersion time. eThis $\nu_a(CH_2)$ is the agostic C-H...Au bond which has a direction of dipole perpendicular to the surface and will disappears when the coverage is high enough. The direction of the ν (C-O) is more perpendicular to the surface at low coverage and more parallel to the surface at high coverage. gThis band involves both the stretching of the C-O bond and the deformation of the C-O-H angle (Hadzi, D.and Pintar, M. Spectrochim. Acta, 1958, 12, 162.) ^hSilverstein, R. M.; Clayton Bassler, G.; Morrill, T. C. Spectrometric Identification of Organic Compuonds 5th ed.; John Wiley & Sons, Inc.: New York, 1991. The direction of this $\delta(\alpha-CH_2)$ is more perpendicular to the surface at low coverage and more parallel to the surface at high coverage. ^jThe peak frequency of this band shifts from ~2855 cm⁻¹ to 2850 cm⁻¹ as increasing immersion time.

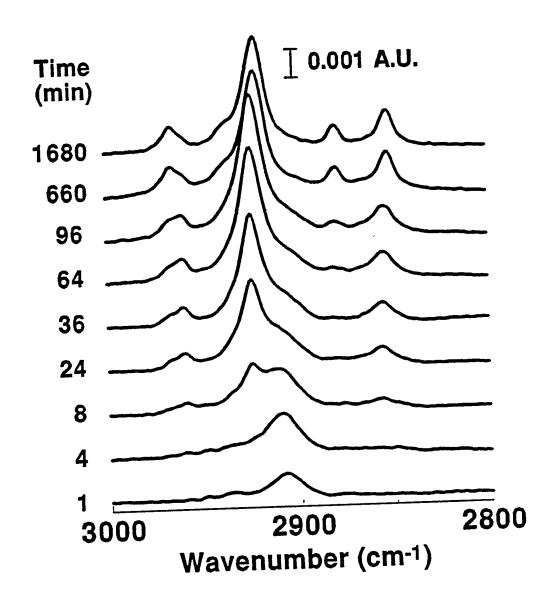


Figure 3. The IRRAS spectra in the high energy region of ODT on Au as a function of immersion time.

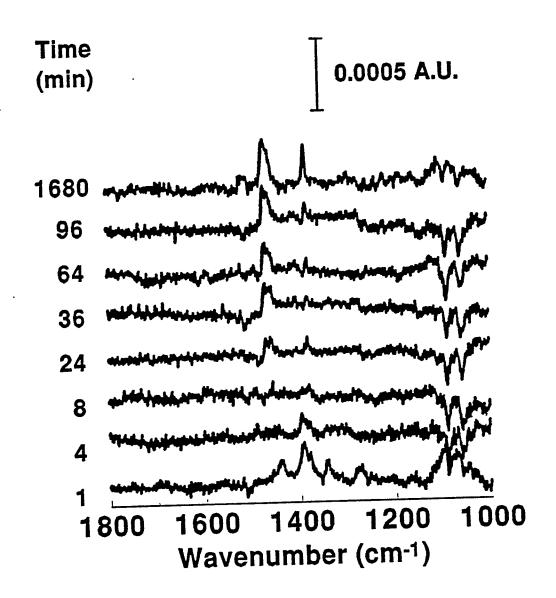


Figure 4. The IRRAS spectra in the low energy region of ODT on Au as a function of immersion time.

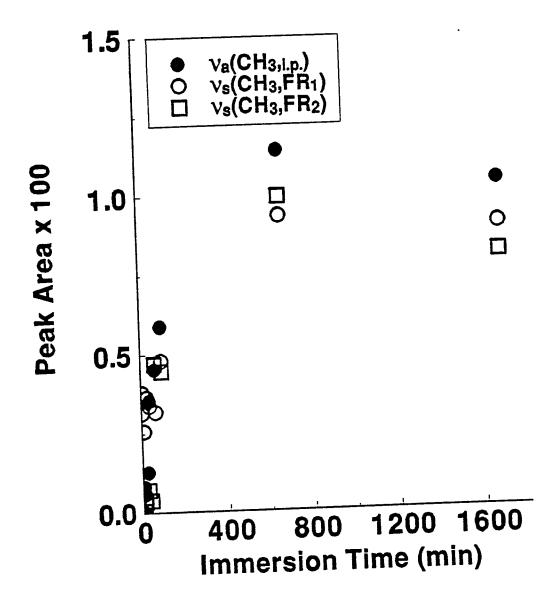


Figure 5. The peak area of $v_a(CH_{3,ip})$, $v_s(CH_3,FR_1)$, and $v_s(CH_3,FR_2)$ of ODT on Au as a function of immersion time.

The peak intensities of these three peaks increases along with the increase in that of the $\nu_a(\text{CH}_2)$ and $\nu_s(\text{CH}_2)$ modes at 2920 cm⁻¹ and 2850 cm⁻¹, respectively, while the peak intensity of the agostic $\nu_a(\text{CH}_2)$ at 2903 cm⁻¹ decreases as shown in **Figure 5**. This strongly confirms that the polymethylene chain axis lies parallel to the surface at low coverage and stands up more and more as the coverage increases.

The absence of the $\nu(\text{C-O})$ peak at 1436 cm⁻¹ indicates there is no C-O group in this monolayer. The peak at 1439 cm⁻¹ is the $\delta_a(\text{CH}_3)$ mode.

Agostic bonds

We propose that the $\nu_a(\text{CH}_2)$ band at 2903 cm⁻¹ arises from the presence of an agostic bond (metal-alkane bond) of C-H···Au. Further evidence will be provided below.

The absence of the $\nu_s(CH_2)$ and the presence of the $\nu_a(CH_2)$ strongly suggests that all the polymethylene chain must lie on the Au surface in order to have the $\nu_s(CH_2)$ dipole parallel to the Au surface and the $\nu_a(CH_2)$ perpendicular to the Au surface at the same time. The decrease in the peak frequency by 20 cm⁻¹ further supports this surface C-H...Au interaction.

ODT on Au covered by a sulfur monolayer

Gold substrates were immersed in the CCl4 solution of 10 mM of Na₂S for 4 hours and then rinsed successively with CCl4 and ethanol. These sulfur monolayers on Au were then immersed in different concentration of ODT solution for 10 min. These Au surfaces were virtually covered by the sulfur monolayer with no bare Au atoms exposed at the surface³¹⁻³³. This prevents the polymethylene chain from interacting with the Au atoms.

Obviously, only crystalline-like and liquid-like $v_a(CH_2)$ peaks with no such agostic bond at 2903 cm⁻¹ at all are observed on the Au surface which was pretreated with a sulfur monolayer as shown in **Figure 6.** This strongly confirms that the band at 2903 cm⁻¹ is associated with the bare Au atoms. Furthermore, the presence of both the $v_a(CH_2)$ and $v_s(CH_2)$ peaks concludes that the polymethylene chain axis is away from the bare Au atoms even at low coverage so that no agostic bond can be observed because no bare Au atoms available.

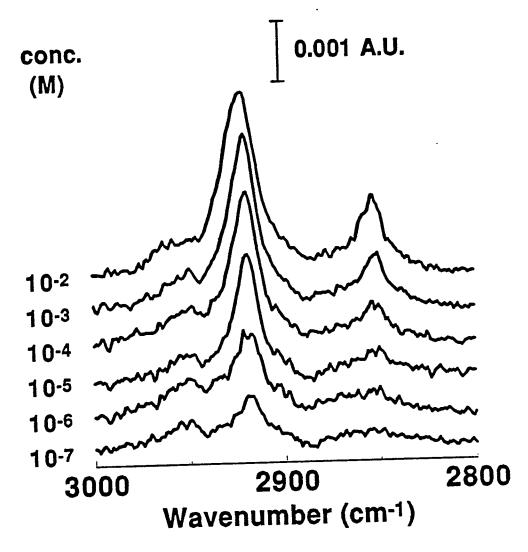


Figure 6. The IRRAS spectra in the high energy region of ODT on Au which is covered by a sulfur monolayer in different concentration of ODT for 10 min.

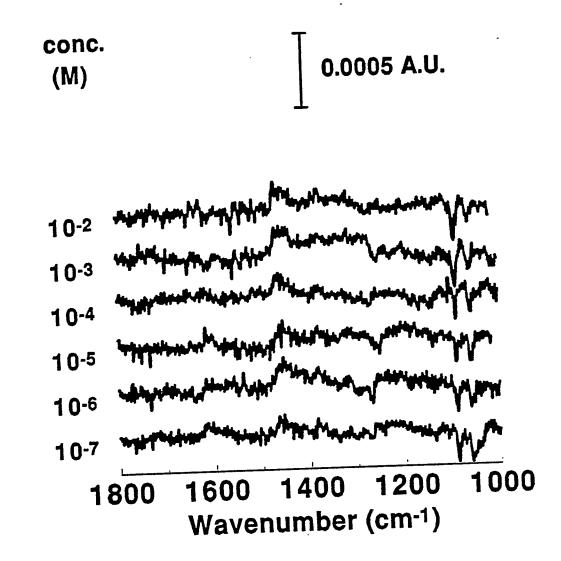


Figure 7. The IRRAS spectra in the low energy region of ODT on Au which is covered by a sulfur monolayer in different concentration of ODT for 10 min.

Unlike the MHA/Au and ODT/Au monolayers, ODT on Au which is covered by a monolayer of sulfur has only very little change in low energy region of IRRAS spectra with immersion time as shown in Figure 7. This is simply because of the lack of bare Au atoms which is already occupied by a sulfur monolayer. Clearly, when there is no bare Au atoms, there is no peak at 2903 cm⁻¹ neither. This indirectly supports the formation of the agostic bond between either MHA or ODT and bare Au surface.

Evidence for agostic bonds

The C-H···M hydrogen bonding has been verified by using neutron diffraction³⁴, crystal structure, NMR, and IR³⁵⁻⁴⁹ as well. The C-H···M hydrogen bond has an energy of the order of a few kcal/mol which is between the extremes of a chemical bond and van der Waals interactions^{50,51}. The vibration peak of this kind of hydrogen bond has lower energy by tens to several hundred wavenumbers and is greatly broadened compared to the corresponding bands of non hydrogen-bonded group⁵²⁻⁵⁹. The extent of this hydrogen bond is almost proportional to the propensity for dehydrogenation:

$$Ru(001) > Pt(111) > Pd(111) > Ni(111) > Cu(111)$$

A CH₂group contributes 5-6.5 kJ/mol to the heat on Pt(111) and $\text{Cu}(100)^{60}$. The monolayer alcohol adsorption heats increased sequentially from methanol to n-butanol (11.5-15 kcal/mol) on Pt(111)⁶¹. The heats of adsorption increase incrementally with addition of methylene groups to the hydrocarbon chain following the formula $\Delta H_{ad} = 8.6 + 1.1$ n kcal/mol for $\text{CH}_3(\text{CH}_2)_{n-1}\text{OH}$ on $\text{Ag}(110)^{62}$. In the case of adsorption of alkylthiol on Au surfaces, a lower limit of 0.8 kcal/mol is estimated for the interaction between CH₂ groups¹¹. The enthalpies of the intermolecular agostic bond between heptane and Cr, Mo, W are calculated to be 10, 9, 13 kcal/mol, respectively⁶³.

The agostic bonds between alcohols and Ag were found by using surface-enhanced Raman scattering spectroscopy ^{64,65} by Pemberton and et al. The $\nu_s(\text{CH}_2)$ agostic bond bands were shifted from 2850 cm⁻¹ down to ~2830 cm⁻¹. SERS data suggest that all four alchols studied are oriented with the α -carbon confined near the surface at all potentials studied. they proposed that

the driving force for this is the formation of CH_2 ...Ag agostic bonds. In fact, those bands at 2900 cm⁻¹ that they founded are the agostic bond of $\nu_a(CH_2)$ which is shifted down from 2928 cm⁻¹ as well although they did not point this out⁶⁵.

The evolution of the structure of MHA/Au and ODT/Au The polymethylene chain

The presence of the $\nu_s(\text{CH}_2)$ is at the expanse of the amount of $\nu_a(\text{CH}_2)$ as shown in Figures 1 and 3. This is because the polymethylene chain must turn the chain axis in order to stand up to accommodate more coverage so that the $\nu_s(\text{CH}_2)$ dipole will have more and more vertical component to the surface and the $\nu_a(\text{CH}_2)$ dipole will have less and less vertical component to the surface simultaneously. This can be confirmed by the substitution of the surface agostic $\nu_a(\text{CH}_2)$ at 2903 cm⁻¹ by the liquid-like $\nu_a(\text{CH}_2)$ at 2928 cm⁻¹ when the coverage increases.

At immersion time of ~720 min, the agostic $\nu_a(\text{CH}_2)$ peak of MHA/Au disappears and the peak frequency of the $\nu_a(\text{CH}_2)$ shifts from 2925 cm⁻¹ to 2920 cm⁻¹ as shown in Figure 1. This substantial decrease in peak frequency indicates the polymethylene chain changes from a liquid-like structure to a crystalline-like environment only when the agostic species totally disappears⁶⁶. This phenomenon implies that the polymethylene chain cannot form the most densely-pack structure as long as there are any agostic molecules because agostic molecules occupy the surface and prevent the polymethylene chain from gathering together to maximize the CH₂ interactions unless the coverage is great enough so that the total CH₂ interactions is greater than the agostic bond. The same trend is observed for ODT on Au as shown in Figure 3.

For both monolayers with agostic bonds at very low coverage, peaks found at 1396 cm⁻¹, 1436 cm⁻¹ are $\delta(\alpha\text{-CH}_2)$ and $\nu(\text{C-O})$, respectively, as shown in Figures 2 and 8. This suggests that the polymethylene chain not only lies on the Au surface but also have the $\alpha\text{-CH}_2$ twisted so that the $\delta(\alpha\text{-CH}_2)$ has more perpendicular component to the surface with the C-O bond more perpendicular to the surface at the same time.

The decrease in the intensity of the $\delta(\alpha\text{-CH}_2)$ and $\nu(\text{C-O})$ peaks with increasing the coverage suggests that the polymethylene chain must turn around to decrease the peak intensities. This is consistent with the conclusion form the $\nu(\text{CH}_2)$ analysis.

The increases in the peak intensities of $\omega(CH_2)$ peaks at 1344 cm⁻¹ and 1380 cm⁻¹ and $\delta(CH_2)$ at 1460 cm⁻¹ with dipole in the polymethylene chain axis suggests that the polymethylene chain stands up with increasing the coverage.

The carbonyl stretching mode

Only MHA/Au has carbonyl at 1717 cm⁻¹ does not appear at very low coverage as a consequence of the twist of the α -C and the turning of CO₂H with ν (C-O) dipole almost perpendicular to the Au surface.

The carboxylic acid group is partially deprotonated in ethanol solution since in addition to the carbonyl band at 1717 cm⁻¹, two carboxylate bands at ~1580 cm⁻¹ and ~1450 cm⁻¹ exist at high coverage as shown in **Figure 2**.

CONCLUSIONS

At very low coverage, all the polymethylene groups lie on the Au surface and form the C-H···Au agostic bonds which have the $\nu_a(\text{CH}_2)$ band at 2903 cm⁻¹ with the $\nu_s(\text{CH}_2)$ dipole oriented parallel to the Au surface and the $\nu_a(\text{CH}_2)$ dipole oriented perpendicular to the Au surface at the same time and the $\nu_s(\text{C-O})$, and the $\nu_s(\text{C-O})$ dipoles of MHA/Pt oriented almost perpendicular, and parallel to the Au surface, respectively. The α -CH₂ is twisted so that the $\delta(\alpha$ -CH₂) has more perpendicular component to the surface with the C-O bond pointing more perpendicular to the surface at the same time.

With increases in coverage, submonolayers must have the polymethylene chain turn the chain axis more and more in order to stand up higher and higher away from the surface in order to allow more and more molecules to be adsorbed on the surface. Eventually, all the agostic bonds will disappear.

REFERENCES

- (1) Bain, C. D.; Whitesides; G. M. Angew. Chem., Int. Ed. Engl. 1989, 28, 506, and references therein.
- (2) Whitesides, G. M.; Laibinis, P. E. Langmuir 1990, 6, 87, and references therein.
- (3) Ulman, A. An Introduction to Ultra-Thin Organic Films From Langmuir-Blodgett to Self-Assembly; Academic Press: San Diego, 1991.
- (4) Porter, M. D.; Bright, T. B.; Allara, D. L.; Chidsey, C. E. D. J. Am. Chem. Soc. 1987, 109, 3559.
- (5) Troughton, E. B.; Bain, C. D.; Whitesides, G. M.; Nuzzo, R. G.; Allara, D. L.; Porter, M. D. *Langmuir* 1988, 4, 365.
- (6) Walczak, M. M.; Chung, C.; Stole, S. M.; Widrig, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2370.
- (7) Widrig, C. A.; Alves, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2805.
- (8) Alves, C. A.; Smith, E. L.; Porter, M. D. J. Am. Chem. Soc. 1992, 114, 1222.
- (9) Widrig, C. A.; Chung, C.; Porter, M. D. J. Electroanal. Chem. 1991, 310, 335.
- (10) Dubois, L. H.; Zegarski, B. R.; Nuzzo, R. G. J. Chem. Phys. 1993, 98, 678.
- (11) Nuzzo, R. G.; Dubois, L. H.; Allara, D. L. J. Am. Chem. Soc. 1990, 112, 558.
- (12) Strong, L.; Whitesides, G. M. Langmuir 1988, 4, 546.
- (13) Bain, C. D.; Whitesides, G. M. J. Am. Chem. Soc. 1988, 110, 5897.
- (14) Bryant, M. A.; Pemberton, J. E. J. Am. Chem. Soc. 1991, 113, 8284.
- (15) Buck, M.; Eisert, F.; Fischer, J.; Grunze, M.; Träger, F. Appl. Phys. 1991, A53, 552.
- (16) Hähner, G.; Wöll, Ch.; Buck, M.; Grunze, M. Langmuir 1993, 9, 1955.
- (17) Hähner, G.; Kinzler, M.; Thümmler, C.; Wöll, Ch.; Grunze, M. J. Va. Sci. Technol. 1992, A10, 2758.
- (18) Shimazu, K.; Yagi, I.; Uosaki, K. Langmuir 1992, 8, 1385.

- (19) Cardini, G.; Bareman, J. P.; Klein, M. L. Chem. Phys. Lett. 1988, 145, 493.
- (20) Bareman, J. P.; Cardini, G.; Klein, M. L. Phys. Rev. Lett. 1988, 60, 2152.
- (21) Hautman, J.; Klein, M. L. J. Chem. Phys. 1989, 91, 4994.
- (22) Ulman, A.; Eilers, J. E.; Tillman, N. Langmuir, 1989, 5, 1147.
- (23) Chidsey, C. E. D.; Liu, G.-Y.; Rowntree, P.; Scoles, G. J. Chem. Phys. 1989, 91, 4421
- (24) Bain, C. D.; Troughton, E. B.; Tao, Y-T.; Evall, J.; Wihitesides, G. M.; Nuzzo, R. G. J. Am. Chem. Soc. 1989, 111, 321.
- (25) Buck, M.; Eisert, F.; Fischer, J. Grunze, M.; Träger, F. J. Vac. Sci. Technol. 1992, A10, 926.
- (26) Thomas, R. C.; Sun, L.; Crooks, R. M.; Ricco, A. J. Langmuir 1991, 7, 620.
- (27) Nuzzo, R. G.; Dubois, L. H.; Allara, D. L. J. Am. Chem. Soc. 1990, 112, 558.
- (28) Azzam, G. A.; Bashara, N. M. *Ellipsometry and Polarized Light*; North-Holland Publishing: Amsterdam, **1977**.
- (29) Ulman, A. J. Mater, Educ. 1989, 11, 205.
- (30) Stole, S. M.; Porter, M. D. Appl. Spectrosc. 1990, 44, 1418.
- (31) Gao, X.; Zhang, Y.; Weaver, M. J. Langmuir 1992, 8, 668.
- (32) Briceno, A.; Chander, S. J. Appl. Electrochemistry 1990, 20, 506.
- (33) Briceno, A.; Chander, S. J. Appl. Electrochemistry 1990, 20, 512.
- (34) Brown, R. K.; Williams, J. M.; Schultz, A. J.; Stucky, G. D.; Illel, S. D.; Harlow, R. L. J. Am. Chem. Soc. 1980, 102, 981.
- (35) Roe, D. M.; Bailey, P. M.; Moseley, K.; Maitlis, P. M. J. Chem. Soc. Chem. Comm. 1972, 1273.
- (36) Cotton, F. A.; Jeremic, M.; Shaver, A. Inorg. Chim. Acta 1972, 6, 543.
- (37) Cotton, F. A.; LaCour, T.; Stanislowski, A. G.; J. Am. Chem. Soc. 1974, 96, 754.
- (38) Cotton, F. A.; Day, V. W.; J. Chem. Soc. Chem. Comm. 1974, 415.
- (39) Ittel, S. D.; Van-Catledge, F. A.; Jesson, J. P. J. Am. Chem. Soc. 1979, 101, 6905.
- (40) Tachikawa, M.; Muetterties, E. L. J. Am. Chem. Soc. 1980, 102, 4542.

- (41) Dawkins, G. M.; Green, M.; Orpen, A. G.; Stone, F. G. A.; J. Chem. Soc. Chem. Comm. 1982, 412.
- (42) Dawoodi, Z.; Green, M. L. H.; Mtetwa, V. S. B.; Prout, J. Chem. Soc. Chem. Comm. 1982, 802.
- (43) Dawoodi, Z.; Green, M. L. H.; Mtetwa, V. S. B.; Prout, J. Chem. Soc. Chem. Comm. 1982, 1410.
- (44) Schultz, A. J.; Williams, J. M.; Schrock, G. Rupprecht, G.; Fellmann, J. D. J. Am. Chem. Soc. 1979, 101, 1593.
- (45) Schultz, A. J.; Brown, R. K.; Williams, J. M.; Schrock, J. Am. Chem. Soc. 1981, 103, 169.
- (46) Schrock, R. R.; Fellmann, J. D. J. Am. Chem. Soc. 1978, 100, 3519.
- (47) Wengovius, J. H.; Schrock, R. R.; Churchill, M. R.; Wasserman, H. J.; J. Am. Chem. Soc. 1982, 104, 1739.
- (48) Itoh, K.; Oshima, N.; Jameson, G. B.; Lewis, H. C.; Ibers, J. A. J. Am. Chem. Soc. 1981, 103, 3014.
- (49) Turner, H. W.; Schrock, R. R.; J. Am. Chem. Soc. 1982, 104, 2331.
- (50) Pimental, G. C.; McClellan, A. L. *The Hydorgen Bond*; Freeman: San Fransisco, 1960.
- (51) Schuster, P.; Zundel, G.; Snadorfy, C. *The Hydrogen Bond*, Vols I, II, and III, 1976.
- (52) Demuth, J. E.; Ibach, H.; Lehwald, S. *Phys. Rev. Letters* 1978, 40, 1044.
- (53) Chesters, M. A.; Parker, S. F.; Raval, R. J. Electron Spectrosc. Related Phenomena 1986, 39, 155.
- (54) Raval, R.; Pemble, M. E.; Chesters, M. A. Surface Sci. 1989, 210, 187.
- (55) Lehwald, S.; Ibach, H. Surface Sci. 1979, 89, 425.
- (56) Hoffmann, F. M.; Felter, T. E.; Thiel, P. A.; Weinberg, W. H. Surface Sci. 1983, 130, 173.
- (57) Waddill, G. D.; Kesmodel, L. L. Chem. Phys. Letters 1986, 128, 208.
- (58) Felter, T. E.; Hoffermann, F. M.; Thiel, P. A.; Weinberger, W. H. Surface Sci. 1983, 130, 163.
- (59) Raval, R.; Chesters, M. A. Surface Sci. 1989, 219, L505.
- (60) Shustorovich, E.; Baetzold, R. C.; Muetterties, J. Phys. Chem. 1983, 87, 1100.

- (61) Sexton, B. A.; Rendulic, K. D.; Hughes, A. E. Surface Sci. 1982, 121, 181.
- (62) Zhang, R.; Gellman, A. J. J. Phys. Chem. 1991, 95, 7433.
- (63) Mores, J. M.; Parker, G. H.; Burkey, T. J. Organometallies 1989, 8, 2471.
- (64) Sobocinski, R. L.; Pemberton, J. E. Langmuir 1992, 8, 2049.
- (65) Joa, S. L.; Pemberton, J. E. Langmuir 1992, 8, 2301.
- (66) Snyder, R. G.; Strauss, H. L.; Elliger, C. A. J. Phys. Chem. 1982, 86, 5145.

PAPER III

COVERAGE DEPENDENCE OF CARBONYL STRUCTURE OF 16-MERCAPTOHEXANDECANOIC ACID MONOLAYERS AT GOLD

INTRODUCTION

The most attractive beauty of these self-assembly monolayers is that these thiolate monolayers are not only very densely-packed, but also very stable¹⁻³. Furthermore, it is not only possible to change the tail group but also very easy to modify the end group for application purpose such as non-linear and active optical devices, chemical, biochemical and physical sensors, protective layers, patternable materials both for resists and for mass information storage, surface modification (e.g. wetting and electrochemical electrode properties)⁴. All these ideal properties of self-assembled monolayers have attracted worldwide attention. As a consequence, the study of the properties and construction of the self-assembled thiol monolayers on gold has grown enormously in recent years⁵⁻⁷. Although there are many different methods of preparing surface monolayers available including Langmuir-Blodgett technique⁸⁻¹⁰. The self-assembled monoalyers on Au have been well characterized.

Porter and co-workers used infrared reflection absorption spectroscopy (IRRAS), ellipsometry and contact angle measurements to demonstrate that the tilt angle is 30° with twist angle of 50° and there is an odd-even chain-length-dependent orientation of the methyl group which provides the best evidence of a conserved Au-S-C bond angle¹¹⁻¹³. Monolayers have a well ordered pattern of $\sqrt{3} \times \sqrt{3} \times \sqrt{3$

Whitesides and co-worker have shown that the distance between the nearest neighbor is 4.97 ± 0.05 Å by transmission electron diffraction spectroscopy¹⁷.

Nuzzo et al. have used low-energy electron diffraction (LEED) and concluded that these monolayers have a $\sqrt{3}$ x $\sqrt{3}$ R30° with 4.99 Å structure¹⁸. These molecular architecture is adopted independent of the chain

length or chain-terminating group and the tilt angle is ~30° with twist angle of 52° which implies that the Au surface-sulfur-carbon bond angle is near 120°. 19

The 16-mercaptohexadecanoic acid monolayer on Au has been characterized by Nuzzo et al. ¹⁹ They concluded the structure of MHA/Au monolayer is an almost perfect zigzag (all trans) conformation. From the IRRAS CH₂ stretching modes, they calculated the tilt angle of 32° and twist angle of 55° with thickness of 22 Å. The axis of C=O is tilted form the surface normal by 66°.

The usage of the monolayers is determined by their surface properties which are a combined result of the coverage, the orientation, and the composition of the end groups. A good understanding of the relationship between the surface properties of the monolayers and the coverage, orientation of a given end group of monolayers is very essential to fabricate monolayers with desired properties.

In this study, a carboxyl acid-terminated thiolate monolayer on Au $(HO_2C(CH_2)_{15}S/Au)$ was studied using IRRAS as a function of immersion time to examine the change in the relative amount of the non-hydrogen-bonded C=O, the hydrogen-bonded dimeric C=O, and hydrogen-bonded polymeric C=O as a function of coverage. The dramatical changes in wettability and ellipsometric thickness of the monolayer using contact angles and ellipsometric thickness measurements as a function of immersion time are reported.

EXPERIMENTAL SECTION

Substrate preparation

Electrodes were prepared by the resistive evaporation of 15-20 nm of chromium, followed by 200-300 nm of gold, onto 3 inch by 1 inch glass substrates. The temperature during the evaporation, measured at the plate supporting the substrates, increased to ~50° C as a result of radiative heating by the evaporation source. The evaporation rates were 0.2 and 2.0 nm/s for chromium and gold, respectively. The pressure in a cryopumped E360A Edwards Coating System during evaporation was <9x10⁻⁵ Pa. After ~45 min cooling time, the evaporator was back-filled with purified N₂. The substrates were then removed, analyzed by optical ellipsometry for the determination of the optical functions of the uncoated substrates, and immersed in the thiol-containing solution.

The surfaces of the uncoated gold electrodes have been characterized previously at both a macroscopic and microscopic level using scanning tunneling microscopy²⁰ and electrochemical measurements of surface texture and surface area¹⁶. Macroscopically, the STM images show that the electrode surface is composed of atomically-terraced crystallites ~25 nm in diameter. The electrodes have a roughness factor of ~1.3, which is given as the surface area determined from the oxidative desorption of iodine¹⁶ divided by the geometric electrode area.

Monolayer preparation

The carboxylic acid-,and methyl-terminated alkanethiolate monolayers were prepared by their spontaneous adsorption onto the evaporated gold from 0.100 mM solutions in absolute ethanol. For hydrogen-bonded C=O monolayers, the substrates were placed in solution for 4.00 min, emersed, rinsed successively with absolute ethanol and hexane, and dried under a stream of prepurified N₂. To form free C=O, longer time is needed.

Emersion measurements

The Emerson experiments were performed by the slow withdrawal of the monolayer-coated substrates from aqueous solutions of varied pH. Typically, samples were left in solution for ~1 min, with immersions of up to ~1 h yielding essentially equivalent results. Upon removal from solution, the samples were placed on a spin-coater and spun briefly at 2000 rpm to remove excess solution. The samples were then placed in the N₂-purged sample chamber of the IR spectrometer for characterization. Spectra were usually acquired within a few minutes of loading in the chamber, with the rotational vibrational bands of vapor phase water subtracted by using a reference vapor spectrum. All solutions were prepared at an ionic strength of 0.100 M by using KCl. Comparable results were obtained in both buffered and unbuffered solution.

Ellipsometric measurements of film thickness

The thicknesses of the monolayers were determined by optical ellipsometry at 632.8 nm in two steps with a computer-interfaced Gaertner Model L-116B ellipsometer. The angle of incidence was 70° from the surface normal. Upon removal from the evaporation chamber, the analyzer and polarizer angles for a reflected light beam from each uncoated substrate were measured on at least three different spots. The average complex refractive index for each substrate was then calculated with a two phase parallel layer model from classical electromagnetic theory²¹. After monolayer formation, each sample was again analyzed and the film thickness calculated from a three-phase parallel layer model, using the average complex refractive index of the individual sample and a real refractive index of 1.45 for the film. A value of 1.45, which is representative of the adsorbate precursors^{22,23}, facilitates comparison with thickness data that has been reported for a variety of monolayers. The influence of several refinements on this treatment of ellipsometric data have been recently discussed^{11,24}.

Contact angle measurements

Advancing contact angles (θ_a) were measured in air with pH = 1.0 HCl aqueous solution as probe liquids by using a Rame-Hart Model 100-00 115

goniometer. For these measurements, a 2.00 mL droplet was formed on the substrate with the needle of the syringe in the droplet. The value of q_a was determined as the volume of the droplet was slowly increased.

Infrared spectroscopy

Infrared spectra were acquired with either a Nicolet 740 or a 750 FT-IR spectrometer. Spectra of the adsorbate precursors were obtained by their dispersion in KBr. Spectra of the monolayers were obtained in a single reflection mode by using p-polarized light incident at 82° from the surface normal and a liquid nitrogen cooled HgCdTe detector. A home-built sample holder was used to position reproducibly the substrates in the sample chamber of the spectrometer²⁵. The spectrometer was purged with boil-off from liquid N₂. Spectra were obtained by referencing 1024 sample scans to 1024 background scans at 2 cm⁻¹ resolution (zero filled) with Happ-Genzel apodization. All spectra are reported as -log(R/R_o), where R is the reflectance of the coated substrate and Ro is that of a reference Au substrate. The reference substrates were prepared by immersion of an uncoated gold substrate into ~1 mM ethanolic solution of octadecanethiol-d₃₇. These films provide a substrate free from detectable IR bands in the regions relevant to these studies as determined when referenced to an uncoated Au substrate that was cleaned in a X 1:3 H₂O₂(30%):H₂SO₄(con.) solution. Further details of the preparation of the reference substrates are given elsewhere 13. Caution: The H_2O_2/H_2SO_4 solution reacts violently with organic compounds and should be handled with extreme care.

The orientation analysis, which compares the observed spectrum to that calculated for an isotropic array of oscillators of comparable packing density, was accomplished by using the frequency-dependent complex optical constants of the adsorbate precursor or a suitable substitute. The optical constants for 16-mercaptohexadecanoic acid were determined from a transmission spectrum after its dispersion as a crystalline material in KBr. From the absorbance values, a Kramers-Kronig transformation²⁶ yielded the frequency-dependent complex optical constant. The optical constants of octadecanethiol from a comparable data workup were used for tetradecanethiol, which is a liquid at room temperature.

Chemicals and reagents

Mercaptohexadecanoic acid (MHA) (HO₂C(CH₂)₁₅SH) was synthersized in this groupand characterized with IR and NMR. The n-pentyl-undecanoic acid disulfide (PUAD) (CH₃(CH₂)₄)SS((CH₂)₁₀CO₂H) was a gift from Dr. Fred Sun (Oregon Graduate Institute of Science & Technology). The rest of chemicals was obtained from Aldrich. The alkanethiols were purified prior to use by passage through an activity one neutral alumina column. All other reagents were used as received. Aqueous solutions were freshly prepared from water purified by passage through a Milli-Q purification system (Millipore, Bedford, MA).

RESULTS AND DISCUSSION

General observations of monolayer formation

Ellipsometric thickness as a function of immersion time

The thickness of the MHA monolayer on gold (MHA/Au) was measured as a function of immersion time using optical ellipsometry. The results are shown in *Figure 1*. In 0.1 mM MHA solution, the thickness increases rapidly at short immersion times, reaching a value of ~9 Å within 0.1 min. The rate of the process slows shortly thereafter with the thickness doubling within ~30 min and approaching a limiting value of 22.0 Å.

This formation process is not a simple one step Langmuir adsorption surface process because it cannot be fitted by using only one single Langmuir isotherm. Actually, this result is consistent with the two step formation process proposed by Whitesides et al.²⁷ and confirmed by Grunze et al.²⁸⁻²⁹, Shimazu et al.³⁰, and Thomas et al.³¹ The first step can be viewed as a Langmuir adsorption process, and the second step is believed to be a result of the combination of replacement of the incorporated solvent and adsorbed impurities and lateral diffusion on the surface to reduce defects and enhance packing²⁷. This second process has a much slower rate than that of the first process.

Wetting as a function of immersion time

The wetting property of the surface of MHA/Au as a function of immersion time was characterized with contact angle measurements. The contact angle of water (θ_a) and the value of $\cos\theta_a$ as a function of immersion time as shown in *Figure 2*. The contact angle decreases dramatically from 57.0 to 18.0 in less than 10 min and decreases only slightly thereafter.

A plot of the value of $\cos\theta_a$ vs. ellipsometric thickness is shown in *Figure 3*. Clearly, when the thickness is less than 15 Å, the value of $\cos\theta_a$ increases dramatically and almost linearly with increasing the ellipsometric thickness. When the thickness is greater than 15 Å, $\cos\theta_a$ levels off and approaches a limiting value of 0.96 (θ_a = 16.0).

These observations clearly demonstrate that the surface properties of a monolayer are determined by its structure even though there is no change in the

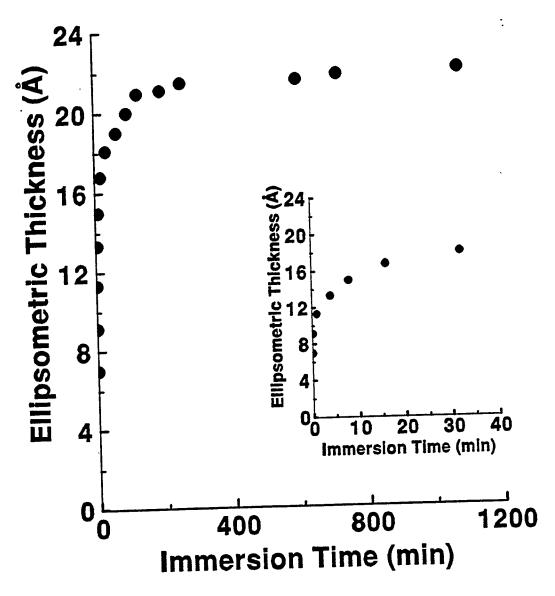


Figure 1. The ellipsometric thickness of a MHA/Au as a function of immersion time.

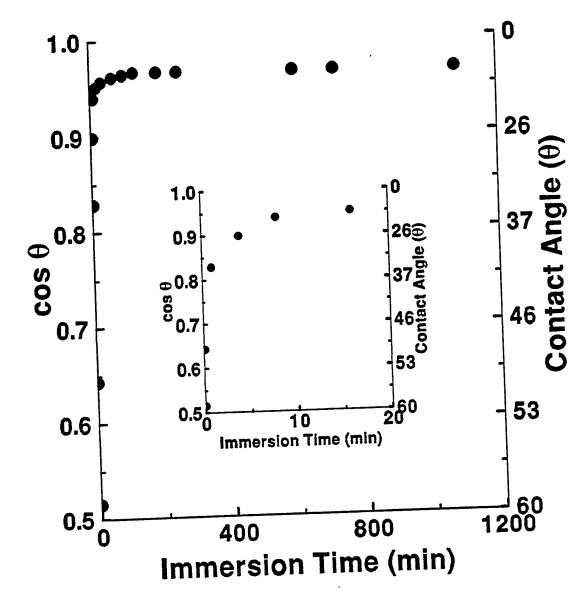


Figure 2. $\cos \theta_a$ of water at a MHA/Au surface at pH 1.0 as a function of immersion time.

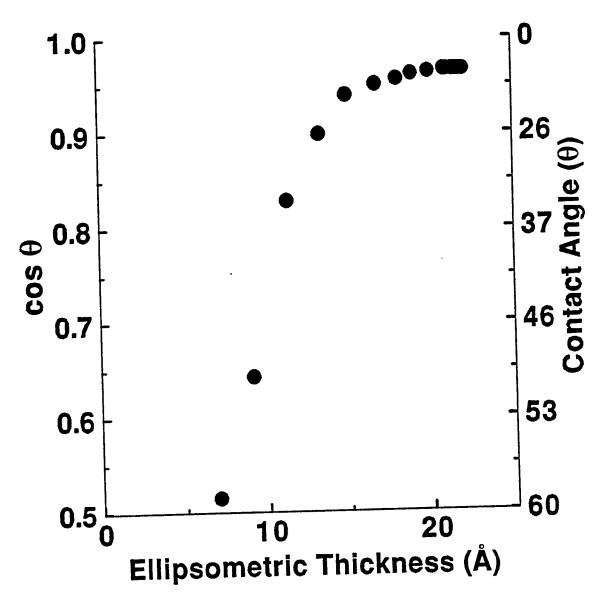


Figure 3. $\cos \theta_a$ of water at a MHA/Au surface at pH 1.0 as a function of the ellipsometric thickness.

composition of this monolayer.

One of the most important issues in this field of study at present time is to be able to control the architecture of a monolayer (i.e. the structure of a monolayer) in order to fabricate materials for research and/or application purposes. To achieve this goal, it is essential to have a detailed understanding how the monolayer structure correlate to surface properties, as discussed above. Gaining an understanding the relatioship between the monolayer structure and the surface properties is the focus of this study.

The value of $\cos \theta_a$ at 0 Å thickness, from the extrapolation in Figure 19, is ~0.25. Thus, the contact angle at 0 Å thickness is ~75° which is about the same value of water on a polymethylene alcohol surface³². It is therefore reasonable to ascribe the nearly linear increase in the values of $\cos \theta_a$ from 0.25 up to 0.95 is as a result of the increases in the ellipsometric thickness of the film going from 0 Å to 15 Å. This behavior indicates that when the coverage is low, the surface appears polymethylene. At low coverage, the monolayer structure would therefore consist of molecules laying down against one another with a very large tilt angle. This proposed structure will be supported shortly by the tilt angle analysis. With increasing coverage, the surface properties are dominated by the carboxylic acid groups on the surface.

IRRAS spectra as a function of immersion time

Figure 4 is a series of IRRAS spectra of MHA/Au as a function of immersion time. There are two main peaks and a shoulder around 2905 cm⁻¹. The peak at 2920 cm⁻¹ is assigned to the asymmetric $\nu(CH_2)$ mode, and the other peak at 2850 cm⁻¹ is the symmetric $\nu(CH_2)$ mode³¹. The shoulder at 2903 cm⁻¹ is an IR peak which has not yet been discovered and assigned in the literature (see below).

Figures 4-6 show a marked change of $\nu(CH_2)$ and $\nu(C=O)$ as a function of immersion time. In the carbonyl stretching region (1600-1750 cm), the spectrum transfers from a single band at 1717 cm⁻¹ to a doublet. The low energy band at 1717 cm⁻¹, assigned to a hydrogen-bonded dimeric C=O mode of a carboxylic acid (see below), decreases with increasing immersion time. The band at 1745 cm⁻¹, assigned to a non-hydrogen-bonded C=O mode of a carboxylic acid, i.e. a free acid, increases with immersion time.

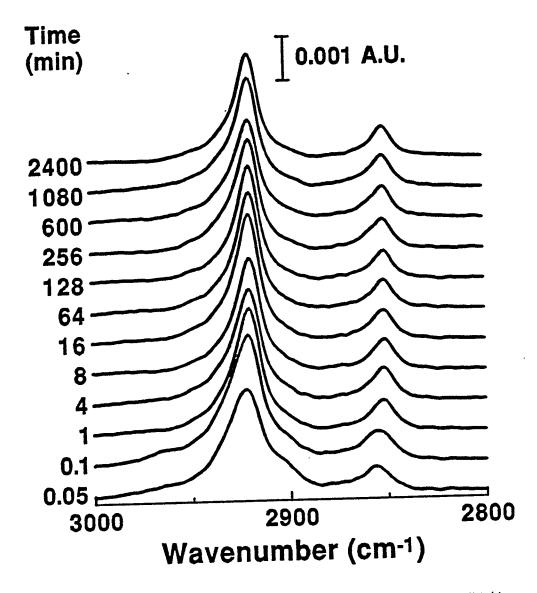


Figure 4. The IRRAS spectra in the high energy region of a MHA/Au monolayer as a function of immersion time.

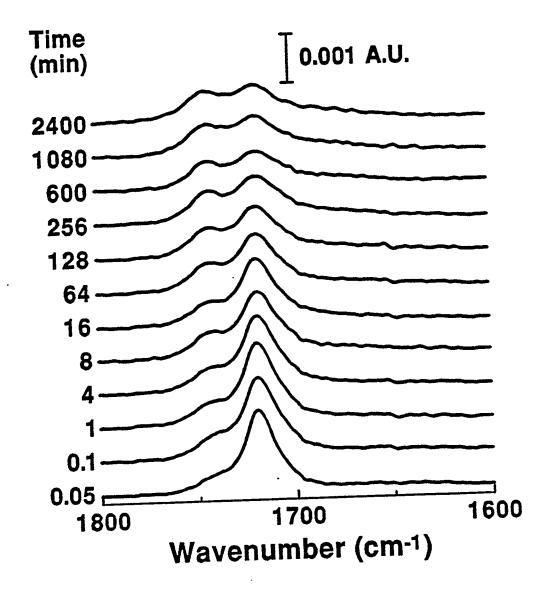


Figure 5. The IRRAS spectra in the ν (C=O) region of a MHA/Au monolayer as a function of immersion time.

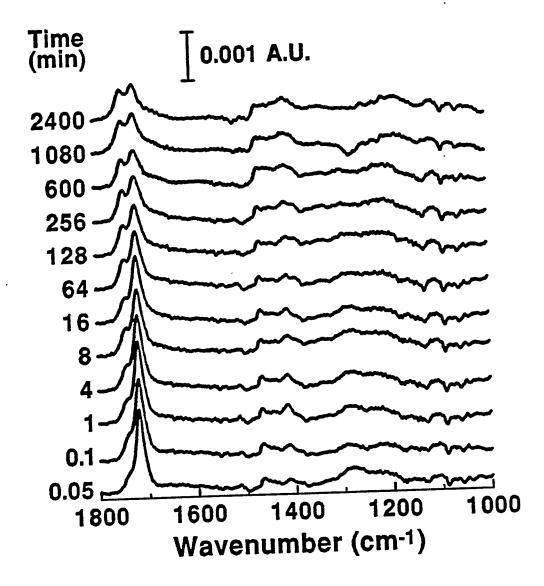


Figure 6. The IRRAS spectra in the low energy region of a MHA/Au monolayer as a function of immersion time.

IRRAS characterization

Trends

It is possible to calculate the coverage, the packing density, and the tilt angle from IRRAS spectra assuming each molecule *like* its structure in KBr and is fully extended. The IRRAS spectra of a MHA/Au obtained as a function of immersion time in the high energy region and low energy region are shown in *Figures 4 and 6*, respectively. The vibrational mode assignments are summarized in Table 1.

The average tilt angle of the axis of a monolayer can be calculated from the following equation:

$$\cos \theta_{mz} = (A_{obs}/3A_{calc}) \tag{1}$$

where A_{obs} and A_{calc} are the observed and calculated peak area, θ_{mz} is the average tilt angle between the transition dipole moments (m) and the surface normal $(z)^{25}$.

The calculated average tilt angles of this monolayer as a function of immersion time based on peak area are shown in *Figure 7*. At the early stage of the adsorption process, the large tilt angle implies that the packing density of the monolayer is low. This is consistent with the results from the thickness measurements. The tilt angle decreases to about 38° in the first 16 min and there is little subsequent decrease. After 2400 min of immersion time, the tilt angle is ~30° which is in agreement with the 32° determined by Nuzzo et al. 19

At a tilt angle of 30° , this monolayer should have a packing density of 8.0×10^{-10} mol/cm² (i.e. the distance between the nearest neighbor is 5.0 Å which is the same as that for n-alkane thiolate monolayers on Au^{14} .

The twist angle decreases from 57° to 53° in the first 16 min and thereafter decreases very slowly to a limiting value of 51°. This result indicates that the Au-S-C angle is ~120° regardless of the packing density or the tilt angle of the film¹⁹.

Table 1. Peak positions, mode assignments, and transition dipole moment directions of MHA/Au and a methyl-terminated thiolate monolayer on Au.

HO ₂ C(CH ₂) ₁₅ S/Au CH ₃ (CH ₂) ₁₃ S/Au					
Mode assignment	obs.	calc.	obs.	calc.	direction of transition dipole ^a
$\nu_{\rm a}({ m CH_{3,ip}})$			2964		[⊥] to C-CH ₃ bond in
$v_a(CH_{3,op})$			ъ	2954	C-C-C chain plane to C-CH ₃ bond C-C-C chain plane
$\nu_{\rm s}({\rm CH_3,FR_1})^{\rm c}$			2936		∥ to C-CH ₃ bond
$\nu_{\rm a}({ m CH_2})$	2918	2920	2918	2920	to C-C-C chain plane
$\nu_{\rm S}({\rm CH_3,FR_2})^{\rm C}$			2878		∥ to C-CH ₃ bond
$\nu_{\rm S}({ m CH_2})$	2850	2850	2850	2851	∥ to C-C-C plane bisceting H-C-H
ν(C=O)	1743 1717				to C=O bond
δ(CH ₂)	1680 1470	1699 1472	1465	1467	in HCH plane, bisecting HCH
ν(C-O)+δ(C-O-H) ^d		1431			.
$\delta(\alpha\text{-CH}_2)$		1410			
δ(CH ₃)			1382	1372	to C-CH ₃ bond
CH ₂ wags and twists	1350- 1170				to axis of chain

Table 1. (continued)

aOrientation of transition dipoles for C-H modes assuming all-trans conformation (R. G. Snyder, *J. Chem. Phys.* 1965, 42, 1744.). ^bThe $v_a(\text{CH}_{3,\text{op}})$ mode was not observed in the monolayer spectrum. ^cThe fermi-resonance couplet, $v_s(\text{CH}_3,\text{FR}_1)$ and $v_s(\text{CH}_3,\text{FR}_2)$, is designated by the subscripts 1 and 2, which refer to the higher and lower energy components, respectively. ^d This band involves both the stretching of the C-O bond and the deformation of the C-O-H angle (D. Hadzi and M. Pintar, *Spectrochim. Acta*, 1958, 12, 162.)

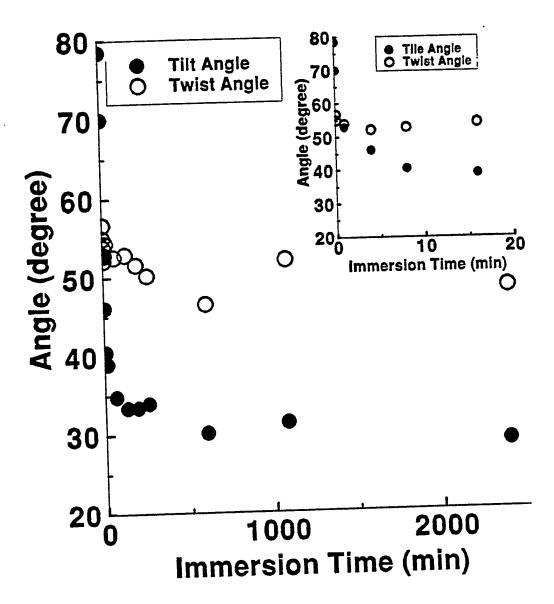


Figure 7. The claculated average tilt angle and twist angle of a MHA/Au as a function of immersion time.

Spectral deconvolution

Methylene stretching region

Examination of the IRRAS spectra in more detail shows that there are at least five different CH_2 features instead of the two features clearly evident in *Figure 4*. Five peaks yielded the best fit. The curve-fitted spectra and the experimental spectra for 1 min and 2400 min immersion time are shown in *Figures 8 and 9*. The main two peaks are the $\nu_a(CH_2)$ at 2918 cm⁻¹, and the $\nu_s(CH_2)$ at 2850 cm⁻¹. These two peaks are indicative of crystalline-like packing of the methylene groups like that in solid crystal³³.

In addition to these two peaks, there is the other set of peaks at 2928 cm⁻¹ and 2858 cm⁻¹ which are at higher energy than that of crystalline modes by ~10 cm⁻¹. These two peaks are the liquid-like $\nu_a(\text{CH}_2)$ and $\nu_s(\text{CH}_2)$ modes, respectively³³.

Finally, there is one more peak at ~2903 cm⁻¹ which has not been classified in the literature yet. We believe that this peak is the CH₂ stretching mode representing interactions of methylene groups with the gold surface, i.e. the C-H···Au agostic bond. (Details has been discussed in Paper II.) This peak is weak and finally disappears after ~16 min of immersion. At very low coverage, almost all the molecules are laying down on the surface, as shown by the agostic CH₂ mode at 2903 cm⁻¹. When there is sufficient number of molecules to cover the surface area, some of the surface agostic molecules start to stand up to increase the packing density. As the coverage increases, the average tilt angle of the molecules decreases to make more room for more molecules to be absorbed until it reaches a minimum as shown in *Figure* 7.

The main four CH_2 peaks as a function of immersion time are shown in *Figure 10*. Although the peak positions of both liquid-like $\nu(CH_2)$ modes change little, the peak positions of both crystalline-like $\nu(CH_2)$ modes decrease with immersion time, particularly in the first ~30 min, by more than two wavenumbers as shown in *Figures 11 and 12*, respectively. These data suggests that the packing of the polymethylene group becomes better and better with increasing the coverage of the monolayer.

In Figure 10, the peak areas of both crystalline-like $\nu(CH_2)$ remain roughly constant while the areas of both liquid-like $\nu(CH_2)$ decrease with

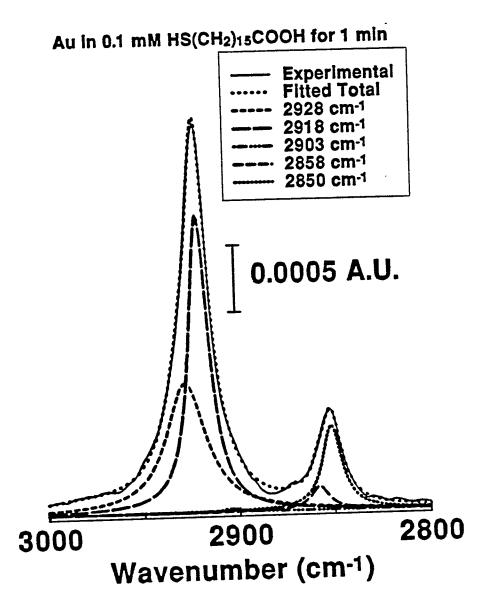


Figure 8. The curve-fitted spectra and the experimental spectrum in the $\nu(CH_2)$ region for a MHA/Au in 0.1 mM MHA solution for 1 min.

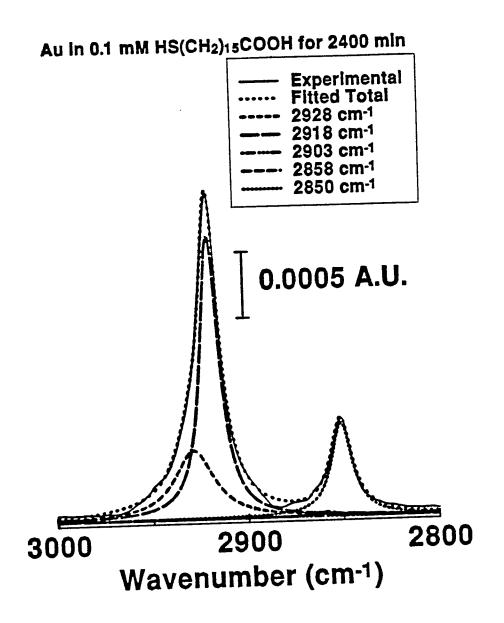


Figure 9. The curve-fitted spectra and the experimental spectrum in the $\nu(CH_2)$ region for a MHA/Au in 0.1 mM MHA solution for 2400 min.

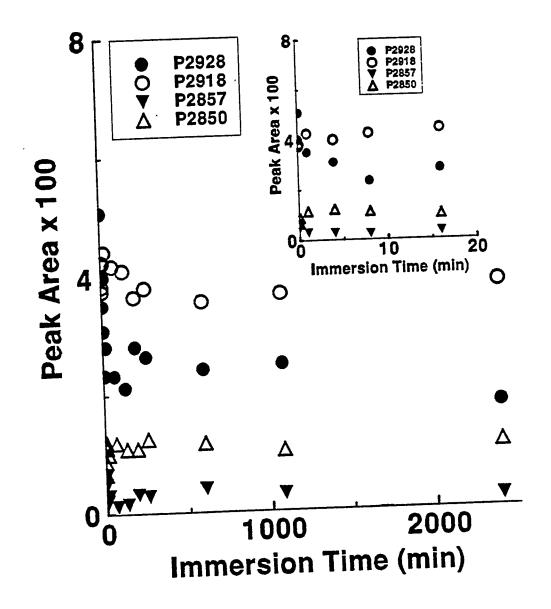


Figure 10. The peak area of deconvoluted $\nu(CH_2)$ peaks of a MHA/Au as a function of immersion time.

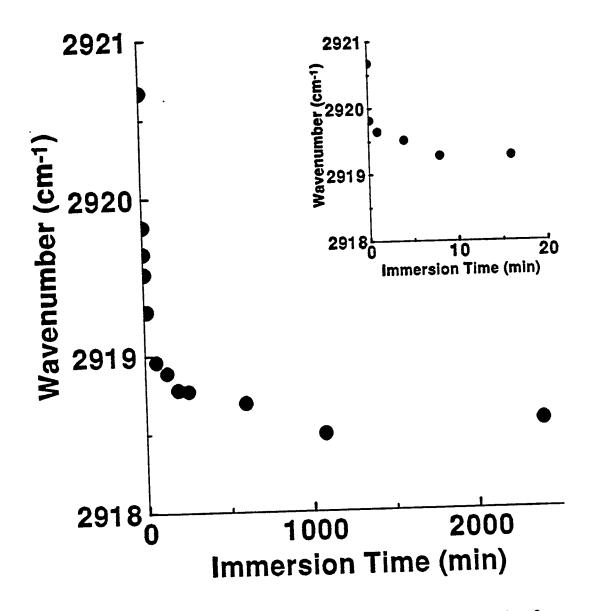


Figure 11. The peak position of the crystalline-like $\nu_a(CH_2)$ mode of a MHA/Au as a function of immersion time.

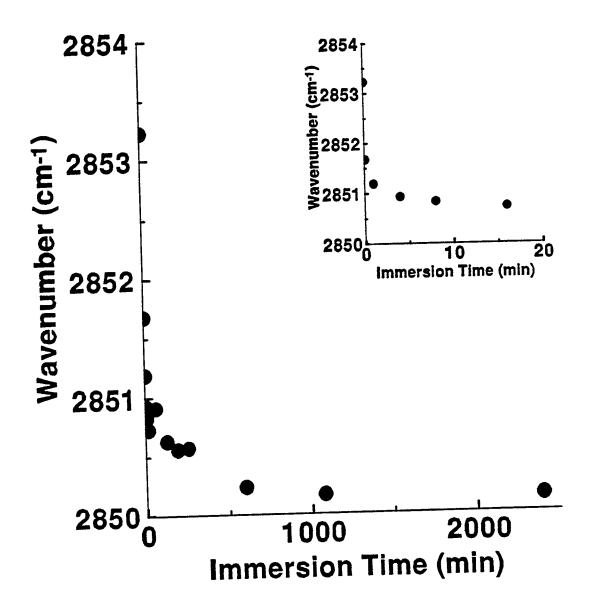


Figure 12. The peak position of the crystalline-like $\nu_s(CH_2)$ mode of a MHA/Au as a function of immersion time.

immersion time, especially during the first 30 min. These data reveal that the amount of both the liquid-like $\nu(CH_2)$ groups decrease substantially with increasing coverage while the amount of crystalline-like $\nu(CH_2)$ groups only change slightly. These results further confirm that the MHA/Au monolayer structure is changing to maximize chain-chain interactions in order to stabilize the system with increasing coverage.

The percentage of this liquid-like $\nu_a(\text{CH}_2)$ decreases from 50 % to 35 % in the first 30 min and reaches 30 % after 2400 min. Although this monolayer packs very well after 30 min, it has both trans and gauche CH₂ segments at the end of the monolayer just like that in 1,32-dotriacontanedioic acid, $\text{HO}_2\text{C}(\text{CH}_2)_{30}\text{CO}_2\text{H}$, adsorbed on Ag^{34} . It is interesting that the full width at half maximum of the crystalline-like $\nu_a(\text{CH}_2)$ decreases from ~16 cm⁻¹ to ~12 cm⁻¹ with increases in the coverage from ~30 % to 100 %. This result also suggests that the MHA/Au monolayer forms a more ordered film with increasing the coverage.

Carbonyl stretching region

The average tilt angle of carbonyl dipole

In Figure 5, the peaks between 1150 cm⁻¹ and 1300 cm⁻¹ are the progression peaks resulting from the presence of all-trans conformation sequences in the polymethylene chains of the monolayer¹⁹. Peaks between 1400 cm⁻¹ and 1500 cm⁻¹ are due to the CH₂ bending mode. The peaks between 1600 cm⁻¹ and 1750 cm⁻¹ are the ν (C=O) modes of carboxylic acid.

The experimental spectrum has all the peaks corresponding to the calculated spectrum. Interestingly, the experimental spectrum has one extra peak at 1743 cm⁻¹, which is the non-hydrogen-bonded ν (C=O) mode.

The ν (C=O) peak is shown as a function of immersion time as shown in **Figure 6**. As one can see, the peak iarea of the hydrogen-bonded dimeric peak decreases with increasing immersion time, while that of the non-hydrogen-bonded increases with time.

The average tilt angle for the C=O dipole was calculated using equation 1 with a calculated IRRAS spectrum. The average tilt angles of ν (C=O) are plotted against immersion time as shown in *Figure 13*. The average tilt angle

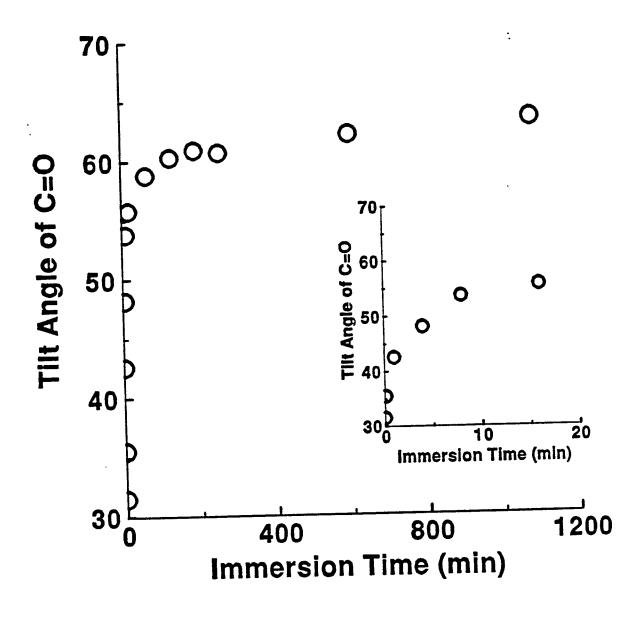


Figure 13. The average tilt angle of the ν (C=O) diploe of a MHA/Au as a function of immersion time.

starts at 32° at 0.10 min and increase sharply up to 62° in 32 min and it remains virtually the same thereafter.

Three carbonyl modes

It is impossible to obtain a good fit for the $\nu(C=O)$ envelope with only two peaks. The lower energy end cannot be fitted unless a third C=O peak is introduced. These fitted curves and the experimental curves for monolayers with 1 min and 2400 min immersion time are plotted in *Figures 14 and 15*, respectively. In both cases, the fitted curves match the experimental curves well using three $\nu(C=O)$ modes. There is a $\nu(C=O)$ mode at 1680 cm⁻¹ in addition to the non-hydrogen-bonded and the hydrogen-bonded dimeric $\nu(C=O)$ modes. It is well known that the hydrogen bonding strengths the $\nu(C=O)$ bond of the carboxylic acids and shifts the peak frequency of the $\nu(C=O)$ mode to lower energy by 20 cm⁻¹ to 80 cm⁻¹ depending upon the strength of the hydrogen bonding, i.e. the stronger the bonding the larger the shift³⁵⁻⁴⁰.

In this study, the non-hydrogen-bonded $\nu(C=O)$ is at 1743 cm⁻¹, and the dimeric hydrogen-bonded $\nu(C=O)$ is at 1717 cm⁻¹. This hydrogen-bonded dimeric C=O is believed to be the side-to-side hydrogen bonding instead of a head-to-head hydrogen bonding¹⁹.

The hydrogen-bonded polymeric C=O mode

A comparison of the peak areas of all $\nu(C=0)$ peaks as a function of immersion time is shown in *Figure 16*. There is almost no non-hydrogenbonded $\nu(C=0)$ peak in the beginning of the adsorption process when the coverage is very low, and the peak area of this peak increases with increasing immersion time and approaches a limiting value. On the other hand, that of the hydrogen-bonded dimeric $\nu(C=0)$ decreases and approaches a limiting value with increasing immersion time as shown in *Figure 16*. The peak area of the low energy $\nu(C=0)$ peak increases and approaches a limiting value with immersion time. It is very clear that most of the changes in peak area of these three peaks occurs during the first 60 min of immersion.

The decrease in the peak area is the combination result of the

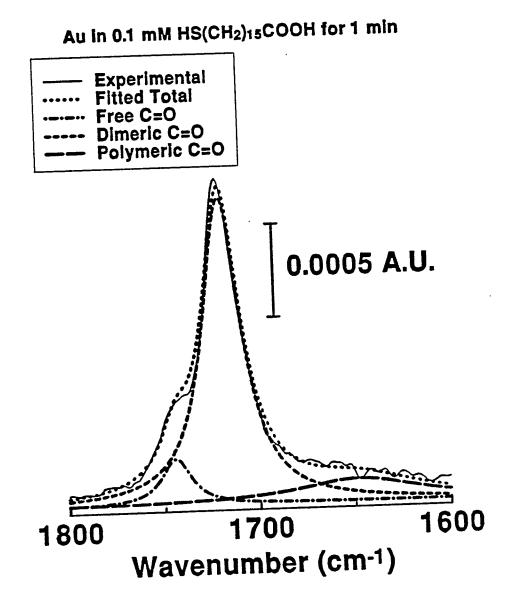
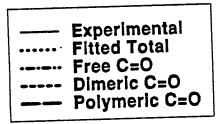


Figure 14. The curve-fitted spectra and the experimental spectrum in the ν (C=O) region for a MHA/Au in 0.1 mM MHA solution for 1 min.

Au in 0.1 mM HS(CH₂)₁₅COOH for 2400 min



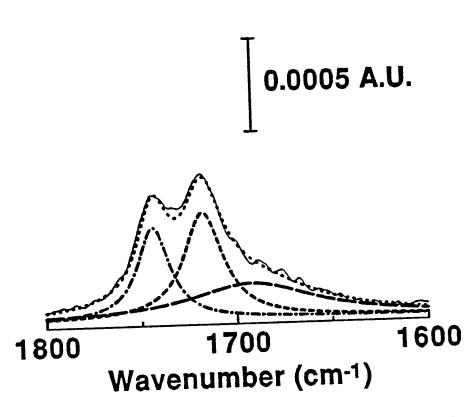


Figure 15. The curve-fitted spectra and the experimental spectrum in the ν (C=O) region for a MHA/Au in 0.1 mM MHA solution for 2400 min.

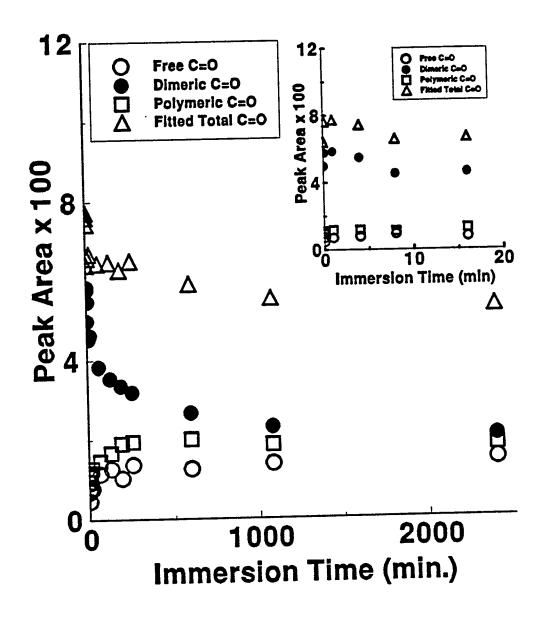


Figure 16. The peak area of deconvoluted ν (C=O) peaks of a MHA/Au as a function of immersion time.

increase of the coverage and the increase in the average tilt angle of $\nu(C=O)$ dipole. The decrease in the peak area due to the increase in the average tilt angle is greater than the increase in the peak area due to the increase in the coverage of the monolayer. As a result, the total peak area of all three v(C=O) peaks decrease with increasing the coverage. If this is taken into account, the decrease in peak area of the hydrogen-bonded dimeric $\nu(C=O)$ is almost equal to the total increase in peak area of the other two peaks particullary after 8 min of immersion. Furthermore, the peak area of the nonhydrogen-bonded $\nu(C=O)$ peak is almost the same as that of the low energy v(C=O) peak along the way. Assuming the conservation of orientation, this result strongly suggests that there is a direct link between the hydrogenbonded dimeric $\nu(C=O)$ and the other two $\nu(C=O)$ peaks i.e. the hydrogenbonded dimeric ν (C=O) mode is converted to both the non-hydrogen-bonded ν (C=O) and low energy ν (C=O) modes simultaneously. Therefore, the relative amounts of these three $\nu(C=O)$ components is a function of coverage. More precisely speaking, the composition of these v(C=O) peaks are controlled by the coverage of the monolayer.

In order to form a stable hydrogen bond for carboxylic acid, the distance between O···O must be within 2.5 to 2.8 Å range⁴¹. As the distance between the nearest COOH neighbor decreases from 7.0 Å to 6.0 Å, the COOH groups start to form stronger and stronger hydrogen bonds. Carboxylic acid groups still can form dimeric hydrogen bonds even though the distance is greater than 7.0 Å by bending the polymethylene chain slightly. Based on these data, when the coverage is greater than 50% of the full monolayer, the distance between nearest COOH neighbors is less than ~7.0 Å. (see below) In order for the COOH groups to form the most stable hydrogen bonds which is the polymeric hydrogen bonds the distance between the nearest COOH groups must be ~6.0 Å. In order for some COOH to maintain this optimal distance to form polymeric hydrogen bonds in the confined space, some of the COOH groups must be squeezed at the same time so that the distance is too short to form any hydrogen bond at all. This structural change explains why the IRRAS spectra in Figure 16 show an increase in the amount of the hydrogen-bonded polymeric $\nu(C=O)$ mode and also an increase in the

amount of the non-hydrogen-bonded $\nu(C=O)$ with increases in coverage simultaneously.

As coverage increases, some hydrogen-bonded dimeric C=O groups form polymeric hydrogen bonds and force some to have no hydrogen bond at all.

The PUAD/Au monolayer

Finally, a monolayer of (CH₃(CH₂)₄)SS((CH₂)₁₀CO₂H)/Au (PUAD/Au) with a short alkyl chain and long carboxylic acid chain was examined. The combination of the short alkyl chain and long carboxylic acid chain prevents the carboxylic acid groups from forming hydrogen-bonded polymeric even at full coverage because one pair of carboxylic acid groups is separated from another by alkyl groups.

The spectra of the $\nu(C=O)$ and the $\nu(CH_2)$ modes of a PUAD/Au as a function of immersion time are shown in *Figures 17 and 18*. There are no noticeable changes observed in the spectra and, in particularly, there is virtually no non-hydrogen-bonded C=O or hydrogen-bonded polymeric C=O observed.

The reason why the hydrogen-bonded dimeric C=O mode dominates is because the COOH groups are structurally impaired in forming any hydrogen-bonded polymeric C=O group or non-hydrogen-bonded C=O group. The monolayer from the disulfide forms so that there is a n-alkane segment in between two adjacent C=O groups. Perhaps the reason why there is no change in CH₂ stretching mode is that there is no room for CH₂ to move around because the disulfide almost already forms a densely packed structure in less than 0.1 min. Since the disulfide consists of a long carboxylic acid chain and a short alkyl chain, it is very unlikely that there will be a situation favorable for extensive Methylene chain-chain interaction between neighbors. The lack of abundant chain-chain interactions between the long carboxylic acid chains discourages these acid groups from forming any hydrogen-bonded polymeric C=O bonds. As a consequence, the hydrogen-bonded dimeric is the dominant peak at all times regardless of either the immersion time or the coverage.

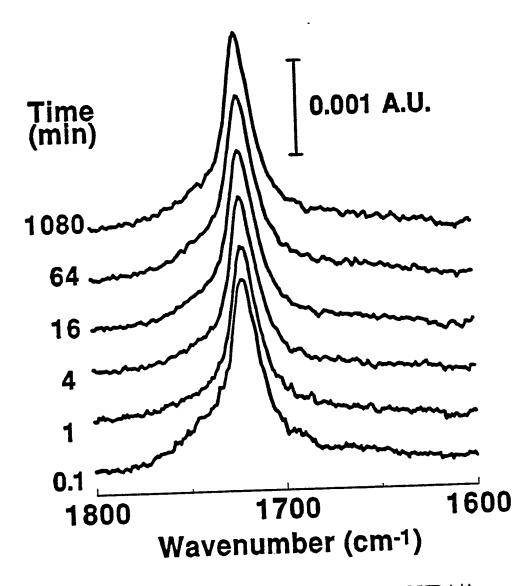


Figure 17. The IRRAS spectra in low energy region for a PUDA/Au as a function of immersion time.

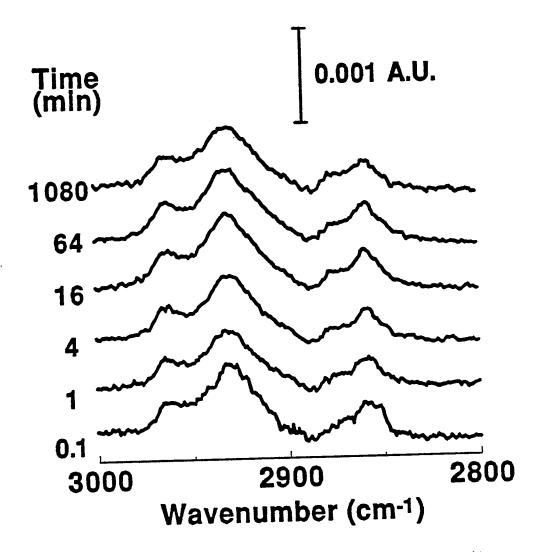


Figure 18. The IRRAS spectra in high energy region for a PUDA/Au as a function of immersion time.

This data show that when the distance between the most adjacent carboxylic acid groups is greater than 7.0 A, only hydrogen-bonded dimeric C=O group exists without any hydrogen-bonded polymeric C=O group or non-hydrogen-bonded C=O group observed. Obviously, when there is no hydrogen-bonded polymeric C=O, there is no non-hydrogen-bonded C=O either. This strongly suggests that the hydrogen-bonded polymeric C=O and non-hydrogen-bonded C=O are associated with each other. This finding is consistent with the result that shows the same increase in the amount of the peak area of both C=O peak with increasing immersion time.

All these evidences strongly confirm that the coverage-induced non-hydrogen-bonded C=O and hydrogen-bonded polymeric C=O modes evolve from the hydrogen-bonded dimeric C=O.

Coverage dependence of C=O groups

To make sure that the change in C=O structure is solely due to the change in the coverage but not because of immersion time, the solvent, or air, several more experiments were executed as following: after immersion in 0.10 mM solution for 1.0 min, MHA/Au monolayers were transferred and emersed in 0.1 M HCl ethanol solution, absolute ethanol, or air media for different time periods and then were rinsed successively with absolute ethanol and hexane, and dried under a stream of prepurified N_2 . The IRRAS spectra of the ν (C=O) modes of these samples are shown in Figures 19, 20, and 21, respectively.

Virtually no change in the $\nu(C=O)$ peaks with time in these three cases strongly suggests that the increase in the amount of both non-hydrogen-bonded $\nu(C=O)$ and hydrogen-bonded polymeric $\nu(C=O)$ modes are not due to immersion time, the solvent, or the replacement of the incorporated solvent or lateral diffusion on the surface to reduce defects and enhance packing as proposed by Bain and Whitesides²⁶.

Since the distance between the nearest COOH neighbor, d, is reciprocally proportional to the square root of the packing density of a monolayer, it can be calculated as following:

$$d = 5.0 \text{ Å/} (\Gamma/\Gamma_{\infty})^{1/2}$$
 (2)

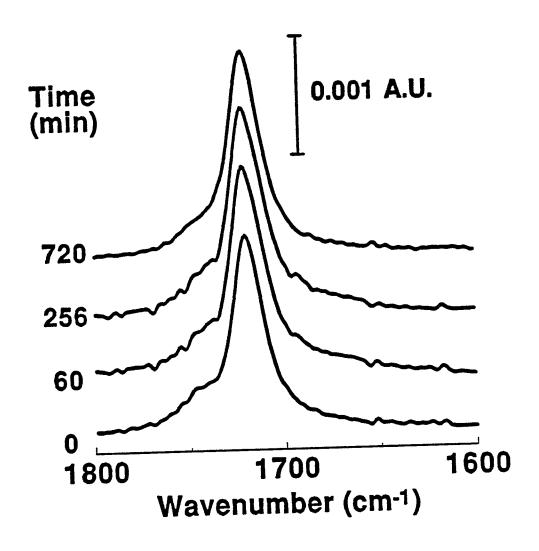


Figure 19. The IRRAS spectra in the low energy region for a MHA monolayer prepared in 0.1 mM solution for 1 min and then transferred to 0.1M HCl solution as a function of immersion time.

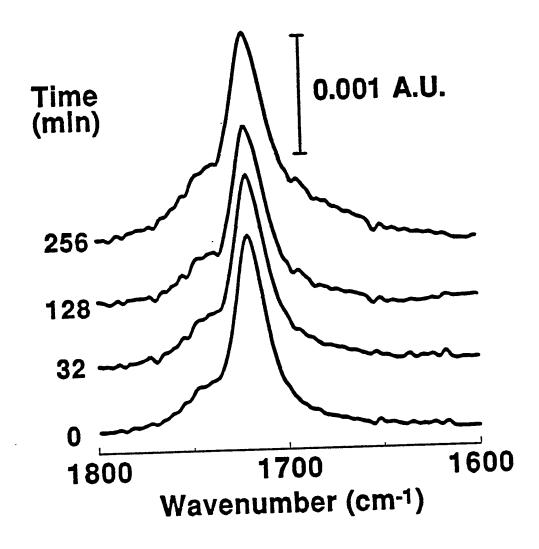


Figure 20. The IRRAS spectra in the low energy region for a MHA monolayer prepared in 0.1 mM solution for 1 min and then transferred to pure ethanol solution as a function of immersion time.

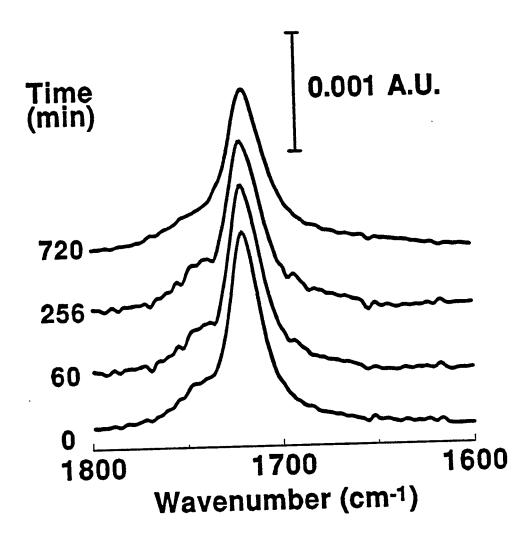


Figure 21. The IRRAS spectra in the low energy region for a MHA monolayer prepared in 0.1 mM solution for 1 min and then exposed to air as a function of immersion time.

where the Γ and Γ are the coverage of interest and the coverage of full coverage, respectively.

Since the coverage is proportional to the ellipsometric thickness equation 2 can be described as following:

$$d = 5.0 \text{ Å} / (A/A_{\infty})^{1/2}$$
 (3)

where A and A_{∞} are the thickness of interest and the thickness at full coverage, respectively. In this study, A_{∞} is 22.0 at a full coverage of 8.0 x 10^{-10} mol/cm².

The peak areas of these three ν (C=O) modes as a function of the distance, d, between the most adjacent carboxylic acid groups is shown in *Figure 22*. When the value of d is greater than 7.0 Å, there is a little increase in the area of hydrogen-bonded dimeric C=O peak resulting from the substantial increases in coverage during the early stage of adsorption. It is clear that there is a remarkable decrease in the peak area of the hydrogen-bonded dimeric C=O peak when the value of d is smaller than 7.0 Å. This decrease in peak area is accompanied by the increases in the peak areas of both free C=O and hydrogen-bonded polymeric C=O peaks especially when d approaches 5.0 Å. Therefore, we believe that the value of d plays a major role in determine the relative amount of these three distinct C=O modes, i.e., when d>7.0 Å, 7.0 Å> d>5.0 Å, and d<5.0 Å, the majority carboxyl forms are the hydrogen-bonded dimeric C=O, the hydrogen-bonded polymeric C=O, and the non-hydrogen-bonded C=O, respectively.

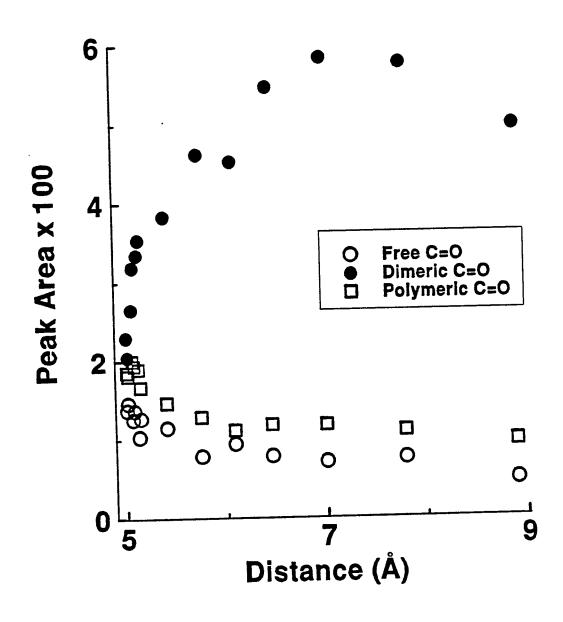


Figure 22. The peak areas of free $\nu(C=O)$, hydrogen-bonded dimeric $\nu(C=O)$, and hydrogen-bonded polymeric $\nu(C=O)$ as a function of the distance, d, between the most adjacent carbonylic acid groups.

CONCLUSIONS

The adsorption process of the MHA on Au is a two-step process with a very fast reaction rate for the first step followed by a very slow reaction rate for the second step. It was determined that this slower reaction had little to do with the replacement of the incorporated solvent or lateral diffusion on the surface to reduce defects and enhance packing.

In addition to the crystalline-like asymmetric and $\nu_s(\text{CH}_2)$ peaks, there is one pair of the liquid-like $\nu_a(\text{CH}_2)$ and $\nu_s(\text{CH}_2)$ peaks which decreases with increasing immersion time. The FWHM of the crystalline-like $\nu_a(\text{CH}_2)$ peak becomes smaller with increases in the coverage. The tilt angle of this monolayer decreases from ~78° to 32° in ~32 min in 0.1 mM solution and subsequently shows little change. On other hand, the twist angle remains almost the same with a value of 51° regardless of the coverage.

The value of $\cos\theta_a$ for pH = 1.0 HCl aqueous solution on a MHA/Au increases rapidly and almost linearly from ~0.25 to ~0.95 with increasing coverage from 0 % to ~75 % of the full coverage. When the coverage is greater than 75 %, this $\cos\theta_a$ value approaches a limiting value of 0.96. It is believed that the surface of a MHA/Au is already completely covered by carboxylic acid groups when the coverage is greater than 75 % of the full coverage.

There are three $\nu(C=O)$ modes instead of only two. In addition to the non-hydrogen-bonded and hydrogen-bonded dimeric $\nu(C=O)$ peaks, there is the hydrogen-bonded polymeric $\nu(C=O)$ bond at 1680 cm⁻¹. At low coverage, the hydrogen-bonded dimeric $\nu(C=O)$ mode is the only one observed. As the coverage increases, about 50 % of this hydrogen-bonded dimeric C=O mode splits into both non-hydrogen-bonded and hydrogen-bonded polymeric C=O modes at the same time.

The average tilt angle of the C=O dipole increases from 32° to 62° in order to form more stable side-to-side polymeric hydrogen bonds.

We believe that the three structures are a result of the distance, d. When d > 7.0 Å > 7.0 Å > d > 5.0 Å, and d < 5.0 Å, the majority carboxyl forms are the

hydrogen-bonded dimeric C=O, the hydrogen-bonded polymeric C=O, and the non-hydrogen-bonded C=O, respectively.

REFERENCES

- (1) Nuzzo, R. G.; Korenic, E. M.; Dubois, L. H. J. Chem. Phys. 1990, 93, 767.
- (2) Dubois, L. H.; Zegarski, B. R. J. Electron Spectrosc. Rel. Phenom. 1990, 54/55, 1143.
- (3) Finklea, H. O.; Ravenscroft, M. S.; Snider, D. A. *Langmuir* 1993, 9, 223.
- (4) Swalen, J. D.; Allara, D. L.; Andrade, J. D.; Chandross, E. A.; Garoff, S.; Israelachvili, J.; McCarthy, T. J.; Murray, R.; Pease, R. F.; Rabolt, J. F.; Wynne, K. J.; Yu, H. *Langmuir* 1987, 3, 932, and references therein.
- (5) Bain, C. D.; Whitesides; G. M. Angew. Chem., Int. Ed. Engl. 1989, 28, 506, and references therein.
- (6) Whitesides, G. M.; Laibinis, P. E. Langmuir 1990, 6, 87, and references therein.
- (7) Ulman, A. An Introduction to Ultra-Thin Organic Films From
 Langmuir-Blodgett to Self-Assembly; Academic Press: San Diego, 1991.
- (8) Seu, M.; Eisenberger, P.; McConnell, H. M. *Proc. Natl. Acad. Sci. U. S. A.* **1983**, *80*, 5795.
- (9) Garoff, S.; Deckman, H. W.; Dunsmiur, J. H.; Alvarez, M. S.; Bloch, J. M. J. Phys. (Pairs) 1986, 47, 701.
- (10) Meyer, E. Nature 1991, 349, 398.
- (11) Porter, M. D.; Bright, T. B.; Allara, D. L.; Chidsey, C. E. D. J. Am. Chem. Soc. 1987, 109, 3559.
- (12) Troughton, E. B.; Bain, C. D.; Whitesides, G. M.; Nuzzo, R. G.; Allara, D. L.; Porter, M. D. *Langmuir* 1988, 4, 365.
- (13) Walczak, M. M.; Chung, C.; Stole, S. M.; Widrig, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2370.
- (14) Widrig, C. A.; Alves, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2805.
- (15) Alves, C. A.; Smith, E. L.; Porter, M. D. J. Am. Chem. Soc. 1992, 114, 1222.

- (16) Widrig, C. A.; Chung, C.; Porter, M. D. J. Electroanal. Chem. 1991, 310, 335.
- (17) Strong, L.; Whitesides, G. M. Langmuir 1988, 4, 546.
- (18) Dubois, L. H.; Zegarski, B. R.; Nuzzo, R. G. J. Phys. Chem. 1993, 97(8), 1665.
- (19) Nuzzo, R. G.; Dubois, L. H.; Allara, D. L. J. Am. Chem. Soc. 1990, 112, 558.
- . (20) Widrig, C. A.; Alves, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2805.
- (21) Azzam, G. A.; Bashara, N. M. Ellipsometry and Polarized Light; Amsterdam: North-Holland Publishing, 1977.
- (22) Bain, C. D.; Whitesides, G. M.; J. Am. Chem. Soc. 1989, 111, 7164.
- (23) Handbook of Chemistry and Physics; Wsast, R. C., Ed.; Chemical Rubber Co.: Boca Raton, FL, 1981.
- (24) Ulman, A. J. Mater, Educ. 1989, 11, 205.
- (25) Stole, S. M.; Porter, M. D. Apply. Spectrosc. 1990, 102, 981.
- (26) Wilson, E. B., Jr.; Decius, J. C.; Cross, P. C. Molecular Vibrations; McGraw-Hill: New York, 1955; pp 285.
- (27) Bain, C. D.; Troughton, E. B.; Tao, Y-T.; Evall, J.; Wihitesides, G. M.; Nuzzo, R. G. J. Am. Chem. Soc. 1989, 111, 321.
- (28) Buck, M.; Eisert, F.; Fischer, J. Grunze, M.; Träger, F. Appl. Phys. 1991, A53, 552.
- (25) Buck, M.; Eisert, F.; Fischer, J. Grunze, M.; Träger, F. J. Vac. Sci. Technol. 1992, A10, 926.
- (30) Shimazu, K.; Yagi, I.; Uosaki, K. Langmuir 1992, 8, 1385.
- (31) Thomas, R. C.; Sun, L.; Crooks, R. M.; Ricco, A. J. Langmuir 1991, 7, 620.
- (32) Holmes-Farley, S. R.; Reamey, R. H.; McCarthy, T. J.; Deutch, J.; Whitesides, G. M. Langmuir 1985, 1, 725.
- (33) Snyder, R. G.; Strauss, H. L.; Elliger, C. A. J. Phys. Chem. 1982, 86, 5145.
- (34) Allara, D. L.; Atre, S. V.; Elliger, C. A.; Snyder, R. G. J. Am. Chem. Soc. 1991, 113, 1852.
- (35) Gramstad, T.; Fuglevik, J. Spectrochim. Acta 1965, 21, 343.

- (36) Pullin, J. A.; Werner, J. Spectrochim. Acta 1965, 21, 1257.
- (37) Figeys, H. P.; Nasielski, J. Spectrochim. Acta 1967, 23A, 465.
- (38) Bellamy, L. J.; Rogash, P. E. Spectrochim. Acta 1960, 16, 30.
- (39) Collins, A. J.; Morgan, K. J. J. Chem. Soc. 1963, 3437.
- (40) Brooks, C. J. W. J. Chem. Soc. 1960, 661.
- (41) Jones, R. E.; Templeton, D. H. Acta Cryst. 1958, 11, 484.

PAPER IV

INFRARED REFLECTION ABSORPTION SPECTROSCOPIC STUDIES OF THE ACID-BASE REACTIVITY OF ORGANIC MONOLAYER FILMS: EMERSED CARBOXYLIC ACID-TERMINATED MONOLAYERS

INTRODUCTION

Monolayer films prepared by the adsorption of thiol-containing compounds ($X(CH_2)_nSH$) at gold provide unique opportunities for the construction of organic interfaces with a well-defined composition and thickness¹⁻³. These films form as gold(I) thiolates⁴⁻⁸ to yield densely-packed structures for end groups that are of a size comparable with the polymethylene chain. In such cases, the spatial arrangement of the layer reflects the bonding of the thiolate head group with the underlying gold lattice. For alkanethiolates, both diffraction^{9,10} and direct atomic-scale imaging^{11,12} techniques have established that the layers adopt a ($\sqrt{3} \times \sqrt{3}$) R30° overlayer structure at Au(111) (the predominant crystal face of the surface of evaporated gold films¹³⁻¹⁸). Because of these characteristics, thiolates at gold have proven extremely attractive in studies aimed at delineating the structural basis of processes occurring at solid-aqueous phase interfaces, e.g. electron-transfer¹⁹⁻²³, wetting^{14,24-25}, and bioadhesion^{26,27}.

One of the most important issues of these monolayers is their surface properties which are mainly affected by the end group of these monolayers²⁸. Therefore, it is very important to understand and study the end group chemistry. The chemistry of the carboxylic acid end group of these carboxylic acid-terminated alkanethiolates is the target of this study.

Central to the above efforts is the need for detailed descriptions of the interactions between the adsorbed layer and the contacting aqueous phase. It is therefore of fundamental importance to develop techniques for the in situ characterization of such interfaces at a molecular level. For electrochemically-driven molecular transformations^{29,30}, infrared reflection-absorption spectroscopy (IRRAS), coupled with electrochemical³¹⁻³³ or optical ^{34,35} modulation techniques, has proven particularly useful. However, as recently discussed, the properties of most thiolate monolayers limit the applicability of the noted modulation techniques³⁶. Electrochemical modulation schemes are ineffective because the structure of these densely-packed monolayers is insensitive to applied voltages up to the onset of oxidative or reductive desorption³⁷.

Optical modulation techniques suffer from overlap of diagnostic spectral features (e.g. C-H stretching modes) of these monolayer and the O-H stretching mode of aqueous solutions, leading to band shape and intensity distortions that complicate a structural interpretation³⁶.

We recently described an approach that reduced the complications of obtaining IRRAS spectra in the C-H stretching region of monolayer films under a thin layer of a contacting aqueous solution 36,38 . The approach is based on a cell design that facilitates the interchange of sample and reference substrates while maintaining a solution layer of effectively constant thickness. However, during studies of monolayers composed of various end-group functionalities 36 , we found characterizations of the reactivity of carboxylic acid-terminated thiolate monolayers to be particularly problematic using our approach. The principal complication arises from the proximity of the O-H bending mode of aqueous solutions and the $\nu(C=O)$ of these layers, which reveals that we will need to gain further control in the repositioning of sample and reference substrates for such a measurement scheme to function effectively. A flow cell may work and we are pursuing such a possibility.

Interestingly, recent reports suggest another possibility for examining the reactivity of these layers under a thin aqueous solution layer³⁸. This approach uses substrates that have been carefully emeresed from the contacting liquid. Early work with this approach has focused on examining the ionic structure at the electrode surface³⁹⁻⁴², the contention being that such a structure remains intact upon removal of the electrode from solution. Though questions concerning the maintenance of the molecular structure of the interface upon introduction into an ultra high vacuum environment are not yet fully resolved^{43,44}, recent Raman spectroscopic studies of adsorbed organic films have established that the molecular structure in solution is retained at electrodes immersed into environments at atmospheric pressure⁴⁵.

EXPERIMENTAL SECTION

Substrate preparation

Electrodes were prepared by the resistive evaporation of 15-20 nm of chromium, followed by 200-300 nm of gold, onto 3 inch by 1 inch glass substrates. The temperature during the evaporation, measured at the plate supporting the substrates, increased to ~50° C as a result of radiative heating by the evaporation source. The evaporation rates were 0.2 and 2.0 nm/s for chromium and gold, respectively. The pressure in a cryopumped E360A Edwards Coating System during evaporation was <9x10⁻⁵ Pa. After ~45 min cooling time, the evaporator was back-filled with purified N₂. The substrates were then removed, analyzed by optical ellipsometry for the determination of the optical functions of the uncoated substrates, and immersed in the thiol-containing solution.

The surfaces of the uncoated gold electrodes have been characterized previously at both a macroscopic and microscopic level using scanning tunneling microscopy⁴⁶ and electrochemical measurements of surface texture and surface area⁴. Macroscopically, the STM images show that the electrode surface is composed of atomically-terraced crystallites ~25 nm in diameter. The electrodes have a roughness factor of ~1.3, which is given as the surface area determined from the oxidative desorption of iodine⁴ divided by the geometric electrode area.

Monolayer preparation

The carboxylic acid-terminated, and methyl-terminated alkanethiolate monolayers were prepared by their spontaneous adsorption onto the evaporated gold from 0.1 mM solutions in absolute ethanol. For the hydrogen-bonded dimeric C=O monolayers, the substrates were placed in solution for 4 min., immersed, rinsed successively with absolute ethanol and hexane, and dried under a stream of prepurified N_2 . More than 30 min is needed to form free C=O form monolayers.

Emersion measurements

Prior to the emersion measurements, all the samples were immersed in pH 1.0 HCl aqueous solution for 30 min. in order to obtain more stable structures. The emersion experiments were performed by the slow withdrawal of the monolayer-coated substrates from aqueous solutions of varied pH. Typically, samples were left in solution for ~1 min, with immersions of up to ~1 h yielding essentially equivalent results. Upon removal from solution, the samples were placed on a spin-coater and spun briefly at 2000 rpm to remove excess solution. The samples were then placed in the N2-purged sample chamber of the IR spectrometer for characterization. Spectra were usually acquired within a few minutes of loading in the chamber, with the rotational vibrational bands of vapor phase water subtracted by using a reference vapor spectrum. All solutions were prepared at an ionic strength of 0.1 M by using KCl. Comparable results were obtained in both buffered and unbuffered solution.

Infrared spectroscopy

Infrared spectra were acquired with either a Nicolet 740 or a 750 FT-IR spectrometer. Spectra of the adsorbate precursors were obtained by their dispersion in KBr. Spectra of the monolayers were obtained in a single reflection mode by using p-polarized light incident at 82° from the surface normal and a liquid nitrogen cooled HgCdTe detector. A home-built sample holder was used to position reproducibly the substrates in the sample chamber of the spectrometer⁴⁷. The spectrometer was purged with boil-off from liquid N₂. Spectra were obtained by referencing 1024 sample scans to 1024 background scans at 2 cm⁻¹ resolution (zero filled) with Happ-Genzel apodization. All spectra are reported as -log(R/R_o), where R is the reflectance of the coated substrate and Ro is that of a reference Au substrate. The reference substrates were prepared by immersion of an uncoated gold substrate into ~1 mM ethanolic solution of octadecanethiol-d₃₇ for 24 hours. These films provide a substrate free from detectable IR bands in the regions relevant to these studies as determined when referenced to an uncoated Au substrate that was cleaned in a X 1:3 H₂O₂(30%):H₂SO₄(con.) solution. Further details of the preparation of the reference substrates are given elsewhere⁴⁸. Caution:

The H_2O_2/H_2SO_4 solution reacts violently with organic compounds and should be handled with extreme care.

The orientation analysis, which compares the observed spectrum to that calculated for an isotropic array of oscillators of comparable packing density, was accomplished by using the frequency-dependent complex optical constants of the adsorbate precursor or a suitable substitute. The optical constants for 16-mercaptohexadecanoic acid was determined from a transmission spectrum after its dispersion as a crystalline material in KBr. From the absorbance values, a Kramers-Kronig transformation⁴⁹ yielded the frequency-dependent complex optical constant. The optical constants of octadecanethiol from a comparable data workup were used for tetradecanethiol, which is a liquid at room temperature.

Chemicals and reagents

Chemicals of CH₃(CH₂)₁₃SH (TDT), HO₂C(CH₂)₂SH (MPA), HCl and KOH were obtained from Aldrich. The carboxylic acid-terminated thiols inculding HO₂C(CH₂)₁₀SH (MUA), and HO₂C(CH₂)₁₅SH (MHA) were synthesized in this group and characterized with IR and NMR. The disulfide (CH₃(CH₂)₄)SS((CH₂)₁₀CO₂H) (PUAD) was a gift from Dr. Fred Sun (Oregon Graduate Institute of Science & Technology). The alkanethiols were purified prior to use by passage through an activity one neutral alumina column. All other reagents were used as received. Aqueous solutions were freshly prepared from water purified by passage through a Milli-Q purification water system (Millipore Corp. Bedford, MA).

RESULTS AND DISCUSSION

We have demonstrated that the coverage and the orientation of a MHA monolayer on gold are a function of immersion time⁵⁰. After in 0.1 mM solution for only 4 min, the fractional coverage of the MHA/Au monolayer increases from 0 % to 65 % and the average tilt angle of the polymethylene chain of the monolayer decreases from 78° to 46°. On the other hand, the twist angle remains virtually the same with a value of ~52° 16 regardless the coverage. This indicates that although there is a substantial decrease in the average tilt angle, the Au-S-C angle is always almost 120°. In addition to the non-hydrogen-bonded and the hydrogen-bonded dimeric $\nu(C=O)$ peaks, there is a hydrogen-bonded polymeric $\nu(C=O)$ bond at 1680 cm⁻¹. When the coverage is low, the hydrogen-bonded dimeric $\nu(C=O)$ mode is the only one observed. As the coverage increases, the hydrogen-bonded dimeric $\nu(C=O)$ peak gradually splits into both the non-hydrogen-bonded v(C=O) and the hydrogen-bonded polymeric $\nu(C=0)$ peaks at the same time. The average tilt angle of the $\nu(C=O)$ dipole increases from 32° to 64° in order to form more stable side-to-side network type hydrogen bond. The result of this is a significant increase in the wetting properties of the film⁵⁰.

Peak positions, mode assignments, and transition dipole moment directions (for an all-trans chain) of the MHA/Au monolayer are listed in Table $1^{19,51}$.

The spectra in Figure 1 contain several important compositional and structural details. From a compositional viewpoint, the absence of the $\nu_a(\text{CH}_{3,\text{ip}})$ at 2964 cm⁻¹, the $\nu_s(\text{CH}_3,\text{FR}_1)$ at 2936 cm⁻¹, the $\nu_s(\text{CH}_3,\text{FR}_2)$ at 2878 cm⁻¹, and the $\delta(\text{CH}_3)$ at 1382 cm⁻¹ in Figure 1 indirectly confirms the formation of the carboxylic acid-terminated layer, and the presence of the $\nu_a(\text{CH}_2)$ at 2918 cm⁻¹ with $\nu_s(\text{CH}_2)$ at 2850 cm⁻¹ indicates the existence of the methylene chain in this monolayer.

The $\nu_a(\text{CH}_2)$ at 2918 cm⁻¹ and the $\nu_s(\text{CH}_2)$ at 2850 cm⁻¹ modes remain almost independent of the pH of immersion solution as shown in

Table 1. Peak positions, mode assignments, and transition dipole moment directions of carboxylic acid- and methyl-terminated alkanethiolate monolayers at gold exposed prior to emersion experiments.

	HO ₂ C(CH ₂) ₁₅ S/Au		CH ₃ (CH ₂) ₁₃ S/A		u —	
Mode assignment dipole ^a	obs.	calc.	obs.	calc.	direction of transition	
$\nu_{\rm a}({ m CH}_{3,{ m ip}})$			2964		[⊥] to C-CH ₃ bond in C-C-C chain plane	
$v_a(CH_{3,op})$			ь	2954	[⊥] to C-CH ₃ bond [⊥] C-C-C chain plane	
$v_{\rm s}({\rm CH_3,FR_1})^{\rm c}$		•	2936		∥ to C-CH ₃ bond	
$v_a(CH_2)$	2918	2920	2918	2920	to C-C-C chain plane	
$\nu_{\rm S}({ m CH_3,FR_2})^{\rm c}$			2878		to C-CH ₃ bond	
$\nu_{\rm s}({ m CH_2})$	2850	2850	2850	2851	∥ to C-C-C plane bisceting H-C-H	
ν(C=O)	1743 1717,1680	1699)			to C=O bond	
δ (CH ₂)	1460	1472	1468	1467	in HCH plane, bisecting HCH	
ν(C-O)+δ(C-O-H) ^d 14	431			to C-O bond	
$\delta_{\rm s}({ m CH_3})$	1382	1382	1380		to C-CH ₃ bond	
$\omega(CH_2)$ and twists	1382 1170	1382	1380 1344	1380 1170	to axis of chain	

Table 1. (continued)

aOrientation of transition dipoles for CH modes assuming all-trans conformation (Snyder, R. G. J. Chem. Phys. 1965, 42, 1744). bThe $\nu_a(\text{CH}_{3,\text{op}})$ mode was not observed in the monolayer spectrum. cThe fermi-resonance couplet, $\nu_s(\text{CH}_3,\text{FR}_1)$ and $\nu_s(\text{CH}_3,\text{FR}_2)$, is designated by the subscripts 1 and 2, which refer to the higher and lower energy components, respectively. d This band involves both the stretching of the C-O bond and the deformation of the C-O-H angle (D. Hadzi and M. Pintar, Spectrochim. Acta, 1958, 12, 162). Silverstein, R. M.; Clayton Bassler, G.; Morrill, T. C. Spectrometric Identification of Organic Compuonds 5th ed.; John Wiley & Sons, Inc.: New York, 1991.

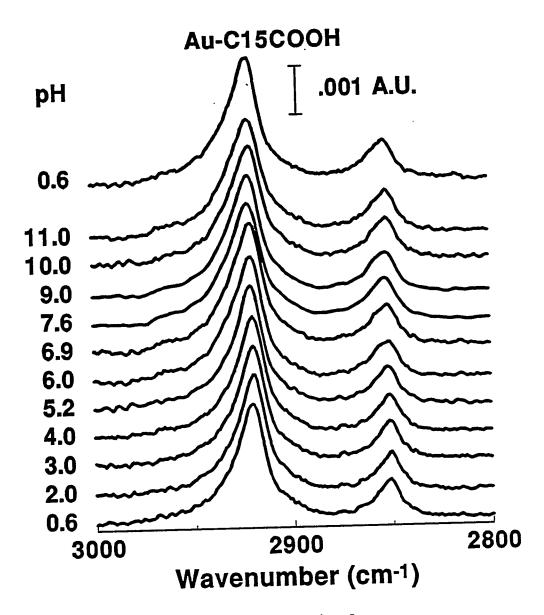


Figure 1. IRRAS spectra in the high energy region for a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 as a function of solution pH.

Figure 1. This implies that the orientation of the polymethylene chain is always the same regardless of the solution pH. That is, there is no solvent or electrolyte incorporated in the film due to the deprotonation of the carboxylic group. This further confirms that the monolayer has a densely-packed structure. This conservation of the orientation of the polymethylene chain is the fundamentally important corner stone for the IRRAS studies of the C=O modes since the absorption is a function of the orientation as well.

In Figure 2, the presence of the $\nu(C=0)$ envelope between 1650 and 1760 cm⁻¹, coupled with the absence of the aforementioned methyl bands, verify the construction of the acid-terminated layer.

Figure 3 shows the expanded IRRAS spectra in the ν (C=O) region as a function of solution pH.

Methyl-terminated monolayers

Spectra for the methyl-terminated layer are given in Figures 4 and 5 for the sequential emersion from pH 1.0, 6.0, and 12.7 solutions. Spectra are also given for the emersion from the pH 1.0 solution after emersion from the pH 12.7 solution. The spectra in all instances are essentially identical to that measured initially at pH 1.0. This is further supported by the different spectra given in the insert of Figures 4 and 5, which were calculated by dividing the spectrum at a given pH by that of the initial pH 1.0 emersion. These results indicate that the methyl-terminated layer is unaffected by immersion into aqueous solutions of varied pH. Immersion times up to ~1 hour did not alter the observations and agree with extensive stability studies reported earlier^{1,14}.

MHA monolayers

MHA monolayers with a fractional coverage of 0.65

The IRRAS spectra for the emersion into solutions of varied pH of the MHA/Au monolayer which has the hydrogen-bonded dimeric C=O form as the majority are shown in the low and high energy regions in

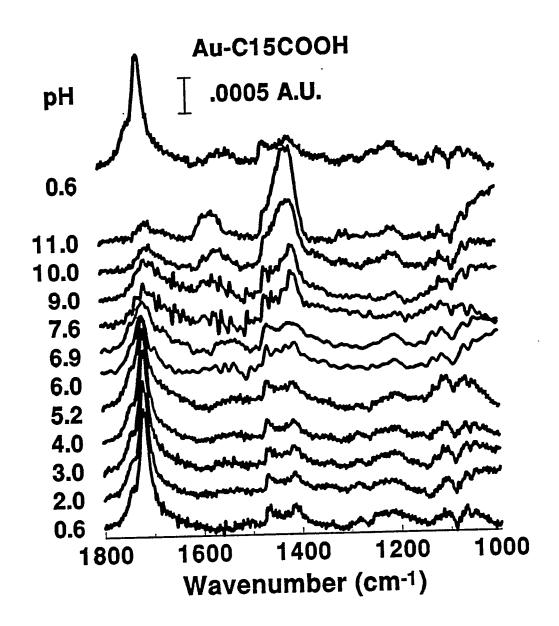


Figure 2. IRRAS spectra in the low energy region for a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 as a function of solution pH.

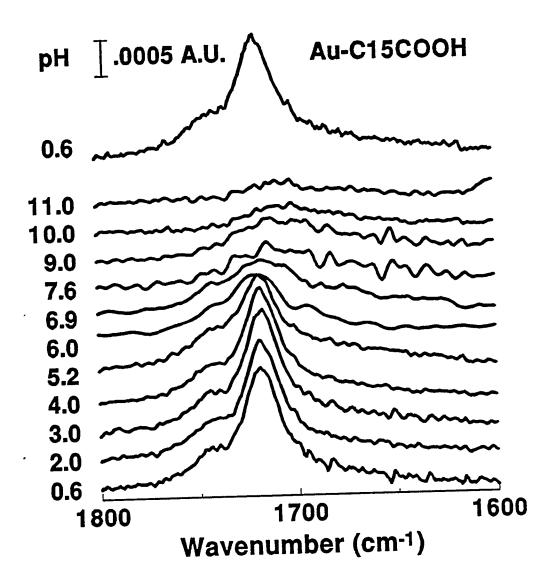


Figure 3. IRRAS spectra in ν (C=O) region for a HO₂C(CH₂)₁₅S/Au monolayer with a fraction coverage of 0.65 as a function of solution pH.

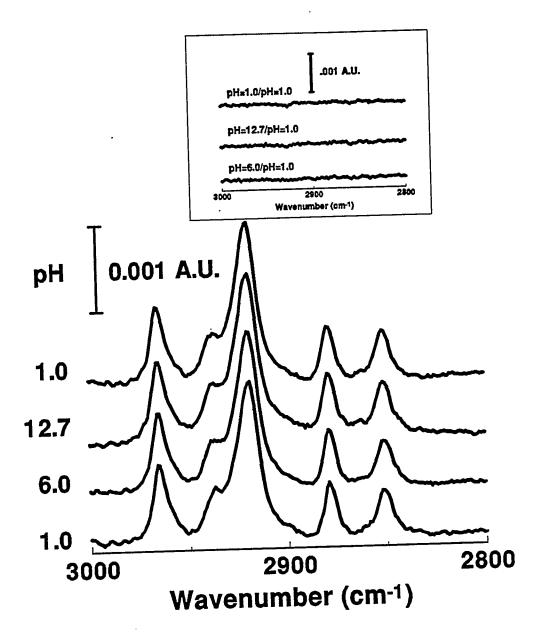


Figure 4. IRRAS spectra in the high energy region for a monolayer of $CH_3(CH_2)_{13}S$ at gold as a function of solution pH. Inset: difference spectra using the spectrum at pH 1.0 as the reference spectrum.

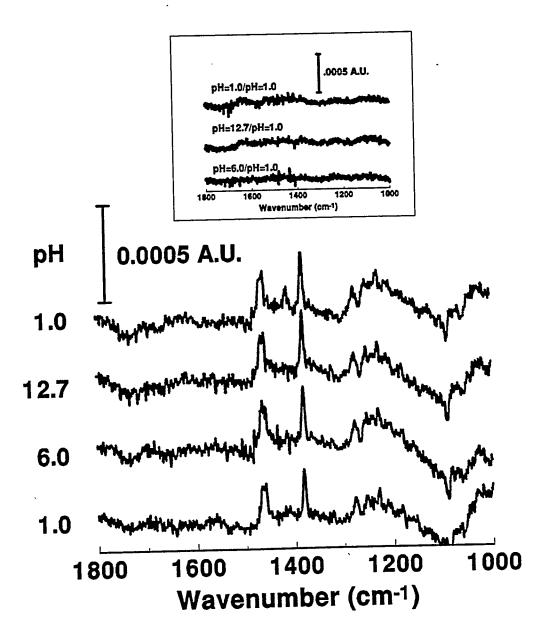


Figure 5. IRRAS spectra in the low energy region for a monolayer of $CH_3(CH_2)_{13}S$ at gold as a function of solution pH. Inset: difference spectra using the spectrum at pH 1.0 as the reference spectrum.

Figures 2 and 1, respectively. The pH of the immersion solutions was increased from an initial value of 0.6 to a final value of 11.0. A spectrum is also given for the re-immersion of the sample into the pH 0.6 solution after cycling through the noted sequence. Spectra above a pH of 11.0 for these layers were masked by intense, poorly-defined features, which we attributed to the relatively large amount of residual electrolyte as shown in Figure 6. Changes in the spectra are most evident in the carbonyl stretching region at a pH of 6.0 and greater. Little change is observable in the spectra below this value. Following the earlier band assignments, the changes in the spectra are consistent with the progressive ionization of the acidic end groups as the pH of the immersion solution increases.

The changes in the carbonyl stretching region are, however, not accompanied at an observable level by the expected changes in the spectral regions attributed to the symmetric $v_a(COO-)$ or asymmetric v_s (COO-) carboxylate stretching modes until above a pH of 9.0. The former is generally observed between 1600 cm⁻¹ and 1550 cm⁻¹, and the latter between 1450 cm⁻¹ and 1400 cm⁻¹. 52-55 Though the overlap with δ (CH₂), δ (a-CH₂), and ι (C-O) + δ (O-H) modes complicates an interpretation of the $v_s(COO-)$ region, no observable change is apparent in the $v_a(COO-)$ region. Based on the assumed orientation of the end group, this observation is unexpected, suggesting the possible structural rearrangement of the chain terminus upon ionization. Such changes would be driven by charge repulsion and/or counter-ion association effects. Experiments with diluted layers and with immersion solutions of varied ionic strength and composition are planned to examine this issue further. The MHA monolayer with not only the dimeric and the polymeric forms but also a free form will be discussed. Although the peak intensity of the dimeric C=O form is smaller than that of the monolayer without free form, it behaves similarly to the latter.

To gain further insight into the reactivity of these layers, the envelope in the $\nu(C=0)$ region was deconvoluted into its three major

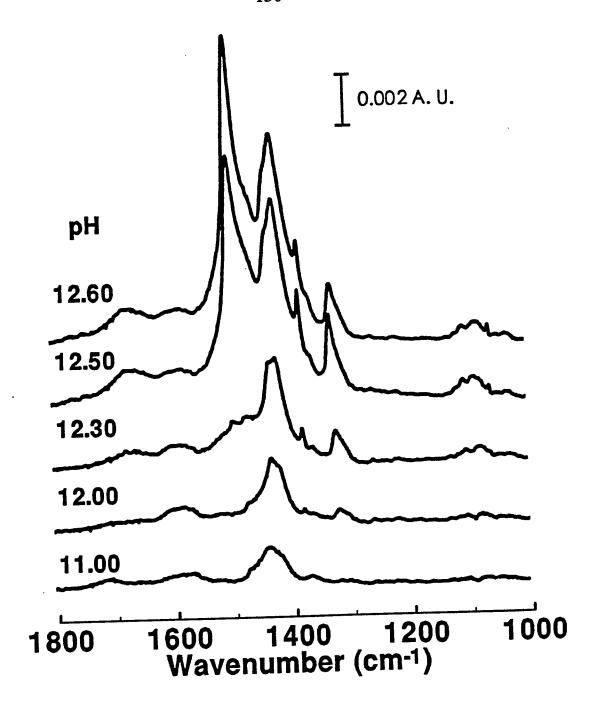


Figure 6. IRRAS spectra in the low energy region for multilayers of KOH on top of a HO₂C(CH₂)₁₅S/Au monolayer as a function of solution pH.

component peaks: the free acid, the hydrogen-bonded dimeric acid, and the hydrogen-bonded polymeric acid. The results of the deconvolution for the initial pH 0.6 emersion sample is shown in Figure 7. Fits to the data assuming three peaks could be varied to a small degree, but the trends in the pH dependence were consistent. Assuming that the oscillator strengths and orientation of both forms of the acids are comparable yields a layer that is ~10% of the free acid, ~10% of the hydrogen-bonded polymeric acid and ~80% of the hydrogen-bonded dimeric acid. For a monolayer in the same solution for 24 hours, there are ~20% of free, ~20% of hydrogen-bonded polymeric acid and ~60% of the hydrogenbonded dimeric acid. We also note the large width of the hydrogenbonded polymeric acid suggests the possibility of a distribution of chainlengths of the linear network bonding. The results of the deconvolution are summarized as a function of pH in Figures 8-10. The peak frequency of each peak remains the same with the peak area of each peak decreases quite differently with increasing the solution pH. Figure 11 provides a comparison of the peak area of the free acid, the hydrogen-bonded dimeric acid and the hydrogen-bonded polymeric acid as a function of the solution pH. Assuming the orientation of the each C=O dipole does not change, the pKa of each acid can be calculated and is equal to the pH where the decrease in the peak area of each peak is 50 %. The calculated pKa values for the free acid, the dimeric acid, and the polymeric acid are 6.6, 7.0 and 11.0, respectively, as listed in Table 2. The transition range of the free acid, the dimeric acid, and the polymeric acid are 3, 7 and 3 pH units, respectively. Interestingly, the peak area of the free acid is virtually lost by pH 8.0 while that of polymeric acid remains almost intact up to pH 9.0.

Superimposed is an experimental fit to the data that assumes the reactivity of a simple monobasic acid in a solution of high ionic strength. This data suggest two interesting points. First, the acid has a pK_a value of 7.1 which is far greater than that, 5.0, in solution. Second, the transition occurs over an interval of \sim 6.0 pH units. Together, these results point to

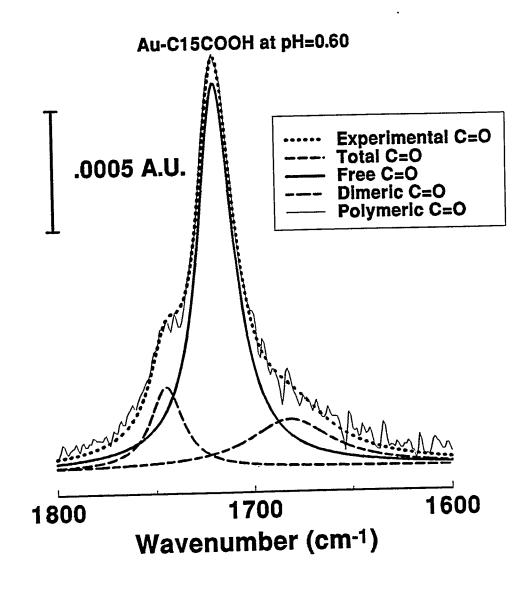


Figure 7. Deconvoluted IRRAS spectra in the ν (C=O) region of a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 emersed in pH 0.60 HCl aqueous solution.

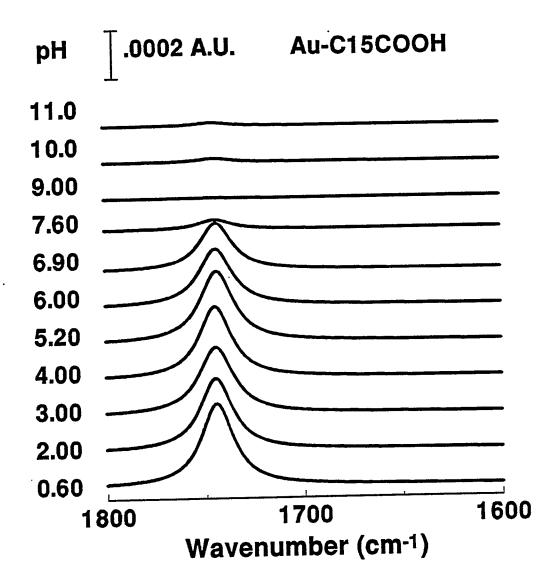


Figure 8. Deconvoluted IRRAS spectra of the free ν (C=O) of a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 as a function of solution pH.

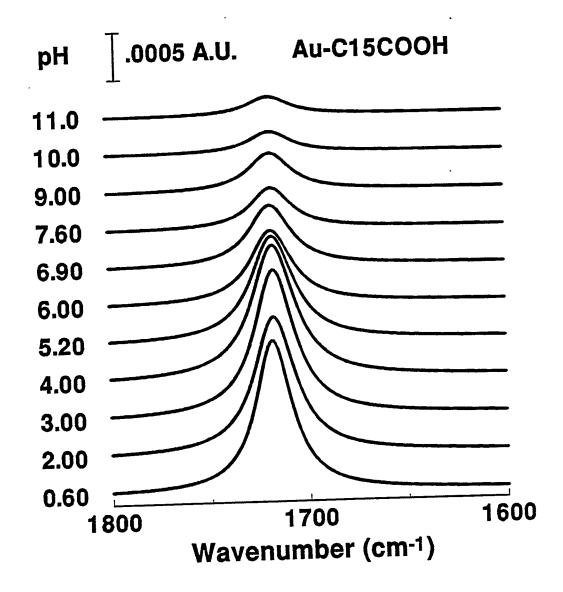


Figure 9. Deconvoluted IRRAS spectra of the hydrogen-bonded dimeric $\nu(C=O)$ of a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 as a function of solution pH.

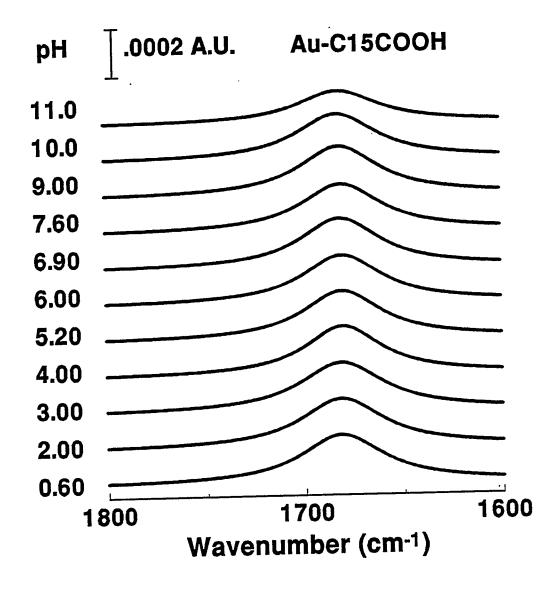


Figure 10. Deconvoluted IRRAS spectra of the hydrogen-bonded polymeric $\nu(C=0)$ of a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 as a function of solution pH.

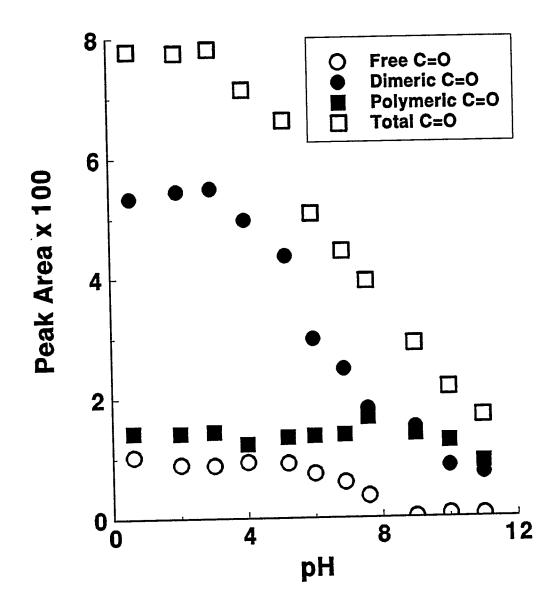


Figure 11. The IRRAS peak area of the ν (C=O) peaks of a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 as a function of solution pH.

the free acid acting as a simple monobasic acid which has a strength roughly 10-2 times less than carboxylic acids in solution (consistent with noninteracting site considerations).

The data for the hydrogen-bonded acid is more interesting. It exhibits a continual decrease up to the limiting pH. This behavior is similar to dimers in solution like malonic acid. Actually, there are two different kinds of hydrogen-bonded acids: a dimeric acid and a polymeric acid. The dimeric acid has higher pH value than that in solution, so it is a weaker acid when it is hydrogen bonded. To the other extremely, polymeric acid has the highest pK_a of all, because the hydrogen bonding is not just between two molecules but a network of many acid molecules. This strong hydrogen bond makes any acid very difficult to be ionized to give away H⁺.

The pK_a and transition rang of the whole monolayer are 7.1 and 9 pH units. In other words, the acid-terminated monolayers have not only weaker acidity but broader transition pH units than this acid in aqueous solution.

Effective pKa

To confirm these points, we calculate the degree of ionization, α_i , of acid and plotted it as a function of the solution pH as shown in Figure 12. There is only little ionization when solution pH is less than 4 for all carboxylic acids. The extent of ionization for the free acid starts to rise when the solution pH is greater than 4 and reaches a value of 1 when the solution pH is greater than 8. This trend is very similar to that in aqueous solution. That for the dimeric acid starts to increase after pH 4 and never reaches a value of 1 even at pH 12. That is, there is a transition of at least 8 pH units. This is very possible since these densely-packed molecules are so close together, the electrostatic repulsion among the deprotonated molecules becomes stronger and stronger. As deprotonation continues, there is a greater charge build up in this very small confined space. On the other hand, the value of α_i of polymeric acid is virtually 0 until pH 9.

Table 2. The pK_a calculated from IRRAS data for the MHA/Au with a fraction coverage of 0.65 (MHA/Au/0.65), MHA/Au, MUA/Au, MPA/Au, and PUAD/Au.

pKa	free acid	dimeric acid	polymeric acid	overall pK _a
MHA/Au/0.65	6.6	7.0	11.0	7.1
MHA/Au	6.5	7.0	11.0	7.2
MUA/Au	6.1	7.8	10.5	8.0
MPA/Au	5.3	5.7	9.5	6.0
PUAD/Au	8	6.5 ^a	a	a

^a Not available because of the change in the orientation of the monolayer.

because of the strong hydrogen-bonding.

A plot of the effective pK_a value for each acid against the degree of ionization, α_i , of acid will be able to give us a better idea on how theses three different acids behave. The effective pK_a is calculated using the following equation:

$$pK_{a}^{eff} = pH - \log(\alpha_{i}/(1-\alpha_{i}))$$
 (1)

where α_i is the extent of ionization. This equation relates the extent of ionization at a particular value of pH to the "effective pK," of monolayers. This "effective pK," is the one that would be attributable to the extent of ionization for a simple monobasic acid. In fact, the pK, is independent of α_i for a monobasic acid. For carboxylic acids, the value of pK, is reported from 3 to 11⁵⁶⁻⁶⁶. The effective pK₂ of these C=O as a function of the extent of ionization are shown in Figure 13. Since both the free acid and the hydrogen-bonded dimeric acid have the effective pK_a of ~5, comparable to that in aqueous solution, at the very early step of ionization, it is clear that the low dielectric constant of these monolayer does not affect this ionization very much⁵¹. As the extent of ionization increases from 0.4 to 1.0, the effective pK, of the hydrogen-bonded dimeric acid increases from 5 to 10. This increase in the effective pK, is comparable to that for polyethylene carboxylic acid studied by Whitesides and co-workers⁵¹. They believed that the increase in the effective pK. might reflect either a change in the ease of ionization of a homogeneous population of carboxylic acid groups as the electrically neutral carboxylic acid groups are converted to anionic carboxylate groups or heterogeneity in the population of carboxylic acid groups being examined. We believe that it is the electrostatic repulsion but not the dielectric constant that plays the most important role for the acid-base chemistry of these acidterminated alkanethiol monolayers.

In these analysis, the error of the values of the effective pKa are too

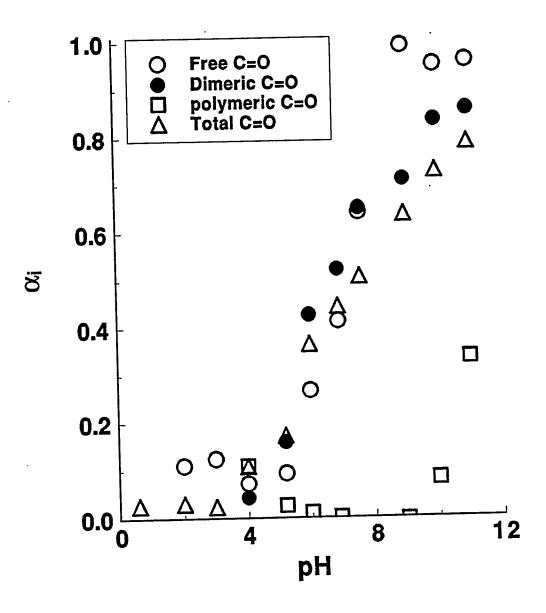


Figure 12. The degree of ionization α_i of a HO₂C(CH₂)₁₅S/Au monolayer with a fraction coverage of 0.65 as a function of solution pH.

large to be meaningful when α_i is either smaller than 0.1 or greater than 0.9. The values of effective pK, for both the free acid and the dimeric acid are about the same and have a constant value of 5.0 when the extent of ionization is less than 0.4. It is very clear that the effective pK, of hydrogen-bonded polymeric acid is around 11. Interestingly, the effective pK, of the total acid is almost the same as that of the hydrogen-bonded dimeric acid. We will see the same trend for different monolayers as well. The effective pK, values at 50 % of ionization for the free acid, the hydrogen-bonded dimeric acid, the hydrogen-bonded polymeric acid, and the overall are 6.9, 7.0, 11, and 7.2, respectively.

MHA monolayers with a full coverage

No noticeable change in the IRRAS spectra of $\nu(CH_2)$ modes in Figures 14 for a MHA/Au prepared in 0.10 mM solution for 24 hours as a function of solution pH indicates that the orientation of the polymethylene chain remains the same through out this pH windows studied. The IRRAS spectra of v(C=O) modes of the same monolayer as a function of solution pH are shown in Figure 15. As you can see, the ν (C=O) peaks of this monolayer are quite different from the MHA/Au monolayer with a fractional coverage of 0.65. These ν (C=O) peaks have ~20 % of the free ν (C=O), ~20% of the hydrogen-bonded polymeric ν (C=O), and only 60 % of the hydrogen-bonded dimeric $\nu(C=O)$. All the amount of these acids decrease with increasing the solution pH and disappear when the solution pH is greater than 11. The recovery of these acids after re-immersion of these monolayers back into pH 1 solution suggests that not only there is virtually no change in the orientation of the film but also there is almost no loss of these acid monolayers during the ionization process. There is a slight increase in the amount of the hydrogen-bonded dimeric acid after it is re-immersed.

This monolayer has three different acid groups behavior similar to that of the monolayer with a fractional coverge of 0.65 although there is a difference in the peak area as shown in **Figure 16**. The free acid starts to

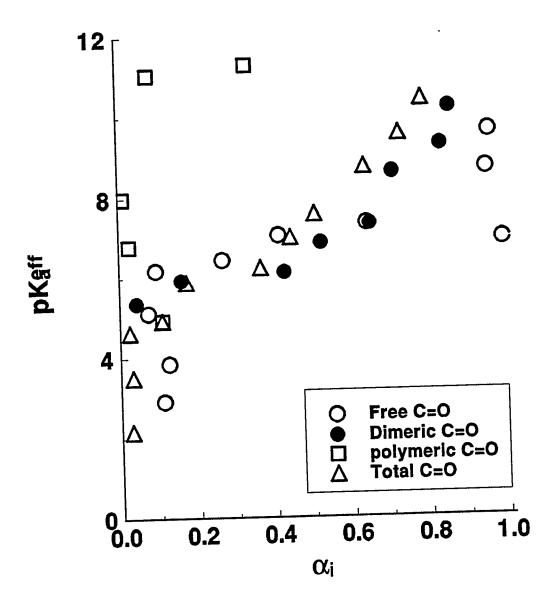


Figure 13. The effective pK_a of a $HO_2C(CH_2)_{15}S/Au$ monolayer with a fraction coverage of 0.65 as a function of the extent of ionization α_i .

ionize at pH 4 and almost completes the ionization process at pH 8. The hydrogen-bonded dimeric acid starts to ionize at pH 4 and has finished by pH 11. The hydrogen-bonded polymeric acid remains intact until pH 10. The pK_a for the non-hydrogen-bonded acid, the hydrogen-bonded dimeric acid, the hydrogen-bonded polymeric acid, and the overall pK_a are 6.5, 7.0, 11.0, and 7.2, respectively.

The calculated values of the extent of ionization for all these acid groups are shown in Figure 17. It is clear that the free acid has the highest value at all pH and virtually reaches a value of 1 when the solution pH is greater than 8. The extent of ionization of the dimeric acid has a value higher than that of polymeric acid but smaller than that of free acid. The extent of ionization of the hydrogen-bonded polymeric acid not only has the smallest value and very little till pH 8. This clearly demonstrates that these three different acid groups have remarkably different properties as far as the extent of ionization is concerned. That is, in terms of acidity: the free acid > the hydrogen-bonded dimeric acid > the hydrogen-bonded polymeric acid.

Obviously, the effective pK, of these three different acid groups behave very differently as shown in Figure 18. The effective pK, of the free acid has a value of 5 at the beginning of ionization and increases to ~7 when the free acid is almost fully ionized. The effective pK, of the hydrogen-bonded dimeric acid has a initial value of ~6 and increases to ~8. The effective pK, of hydrogen-bonded polymeric acid is ~10. Surprisingly, the effective pK, of the overall acid group has almost the same value as that of the hydrogen-bonded dimeric acid again. At 50 % of ionization, the effective pK, values of the free acid, the hydrogen-bonded dimeric acid, the hydrogen-bonded polymeric acid, and the overall value are 6.5, 7.8, 11.0 and 7.8, respectively. So, it is clear that these three acids have significant differences in their acid properties. Interestingly, the overall properties of these three acids are virtually the same as that of the hydrogen-bonded dimeric acid.

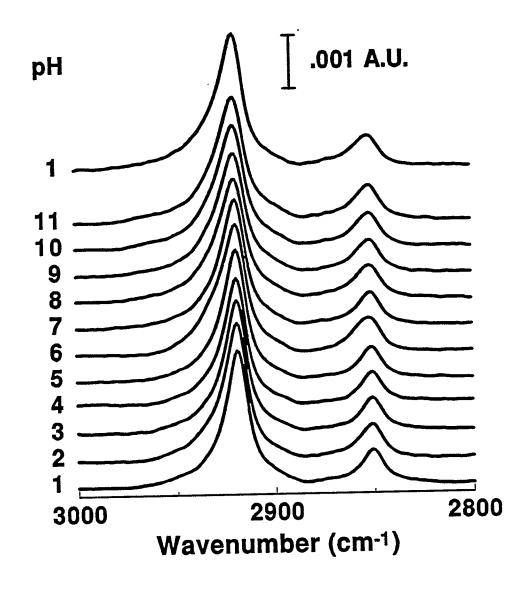


Figure 14. IRRAS spectra in high energy region for a HO₂C(CH₂)₁₅S/Au monolayer as a function of solution pH.

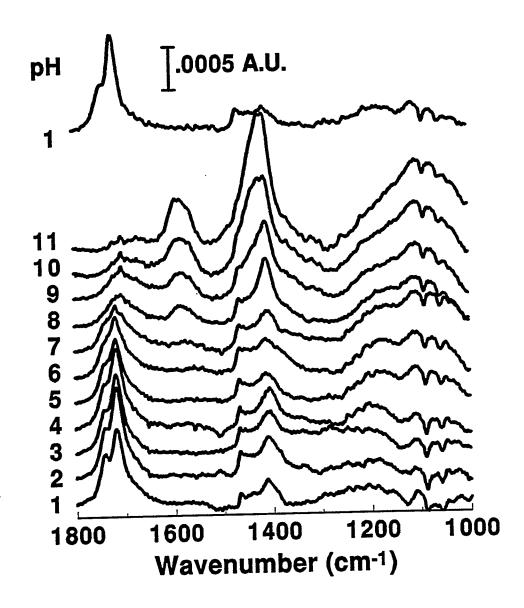


Figure 15. IRRAS spectra in low energy region for a HO₂C(CH₂)₁₅S/Au monolayer hours as a function of solution pH.

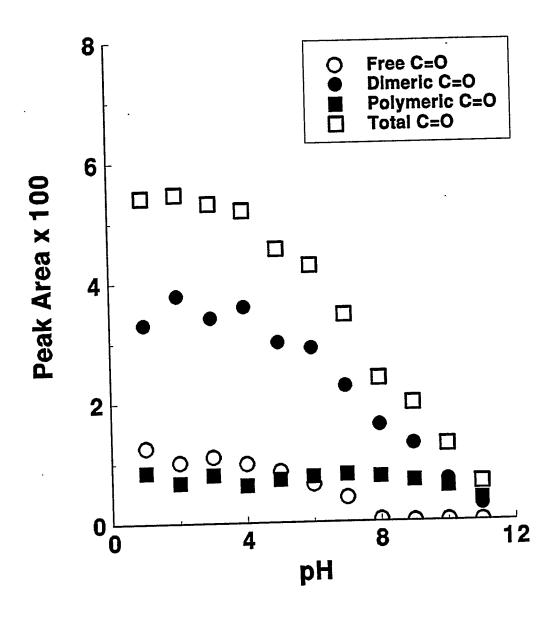


Figure 16. The IRRAS peak area of the C=O peaks of a $HO_2C(CH_2)_{15}S/Au$ monolayer as a function of solution pH.

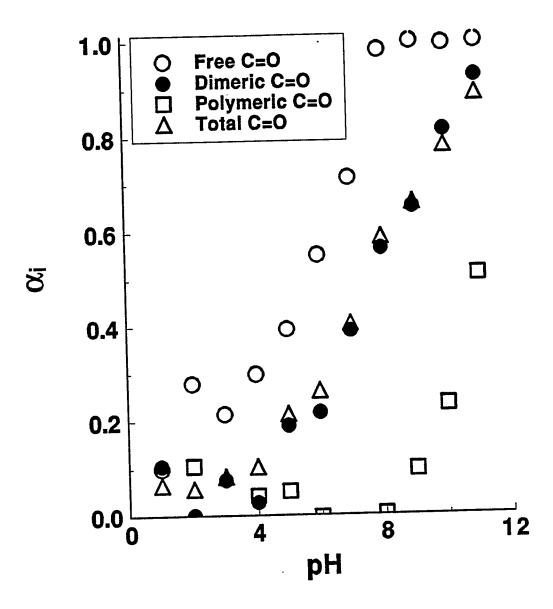


Figure 17. The degree of ionization α_i of a HO₂C(CH₂)₁₅S/Au monolayer as a function of solution pH.

The comparison between the effective pK_s for the MHA/Au monolayer with a fractional coverge of 0.65 and 0.98 shows not much difference between these two monolayers although there is a slightly higher value for free acid of the monolayer with a fraction coverage of 0.65. The slight difference most likely is due to the detection limit of the very small amount of the free acid in the monolayer.

MUA monolayers

No change in the $\nu(CH_2)$ modes in Figure 19 for a 24-hour-immersed MUA/Au monolayer indicates the conservation of the orientation of the polymethylene chain of this monolayer. The recovery of all these three $\nu(C=0)$ modes in Figure 20 after re-immersion back in pH 1 solution confirms this point. It is apparent that there is a slight increase in the amount of the hydrogen-bonded dimeric acid after it is re-immersed.

In this film, the relative amount of the free acid, the hydrogenbonded polymeric acid, and the hydrogen-bonded dimeric acid are 25%, 25 %, and 50 %, respectively, as shown in Figure 21. This indicates that in this monolayer, more hydrogen-bonded dimeric acids are converted to both the free and hydrogen-bonded polymeric acids than a MHA/Au monolayer after 24 hours immersion time in 0.1 mM solution. The peak area of the free v(C=0) mode starts decreasing at pH 5 and disappears at pH 8. The peak area of the hydrogen-bonded dimeric ν (C=O) mode starts decreasing at pH 4 and keeps decreasing until the solution pH is greater than 11. Clearly, the peak area of the hydrogen-bonded polymeric ν (C=O) mode is constant until the solution pH is greater 9. The calculated pKa values for the free acid, the dimeric acid, the polymeric acid, and the overal pKa are 6.1, 7.8, 10.5 and 8.0, respectively. The transition range of the free acid, the dimeric acid, and the polymeric acid are 3, 7 and 3 pH units, respectively. By pH 8, the peak area of the free acid disappears completely while that of polymeric acid remains almost intact.

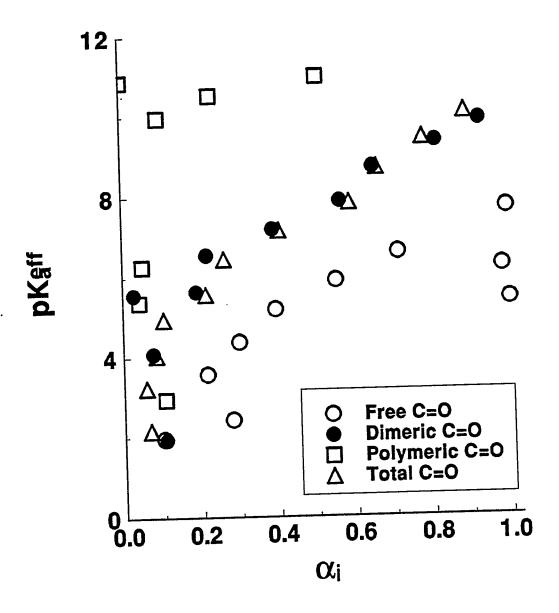


Figure 18. The effective pK_a of a $HO_2C(CH_2)_{15}S/Au$ monolayer as a function of the extent of ionization α_i .

The extent of ionization as a function of solution pH is shown in Figure 22 which shows almost the same trend as that for the MHA/Au monolayer in Figures 12 and 17. Once again, the free acid has the highest values at almost any pH and the hydrogen-bonded polymeric acid has the lowest values with the hydrogen-bonded dimeric acid in between. The free acid starts to ionize at pH 5 and almost completes at pH 8. The hydrogen-bonded dimeric acid starts to ionize at pH 4 and barely finishes at pH 11. The hydrogen-bonded polymeric acid remains almost intact until pH 9. The overall performance is basically the same as that of the hydrogen-bonded dimeric acid.

The values of the effective pK, for all these acid groups as a function of the extent of ionization are shown in Figure 23. The free acid has an initial value of ~5 and increases up to ~8. The hydrogen-bonded dimeric acid with a initial value of ~5 increases with increasing the extent of ionization and levels off with a finial value of ~10. The value for the hydrogen-bonded polymeric acid is ~10 though there is only one data point. Surprisingly, the overall values are very close to that of the hydrogen-bonded dimeric acid again. The effective pK, values at 50 % of ionization for free acid, the hydrogen-bonded dimeric acid, the hydrogen-bonded polymeric acid, and the overall are 6.3, 8.3, 11.0, and 8.3, respectively.

The comparison between the effective pK, values of MUA/Au with that of MHA/Au monolayer shows a slightly higher values for MUA/Au monolayer by ~10 % in the overall effective pK, value although there is not much difference in the beginning of ionization. Both monolayers have almost the same effective pK, values for both the free acid and the polymeric acid, but the MUA/Au monolayer has a higher value for the dimeric acid particularly at high extent of ionization. These higher effective pK, values in MUA/Au monolayers most likely is because the MUA/Au monolayer has higher packing density than MHA/Au monolayer. If this is true, then this further confirms that the electrostatic repulsion is the dominant factor in this ionization process.

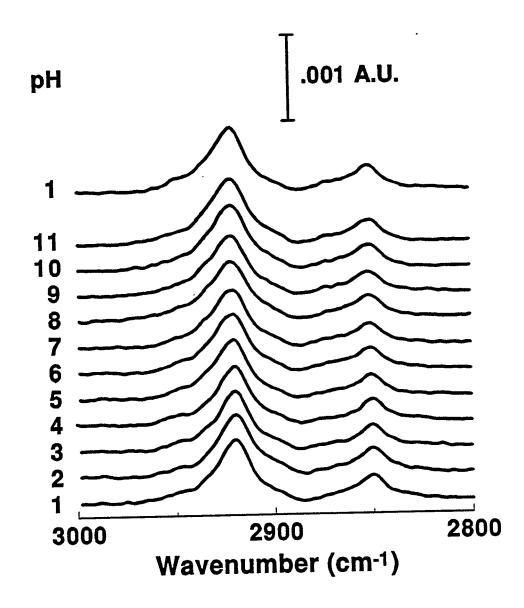


Figure 19. IRRAS spectra in high energy region of a HO₂C(CH₂)₁₀S/Au monolayer as a function of solution pH.

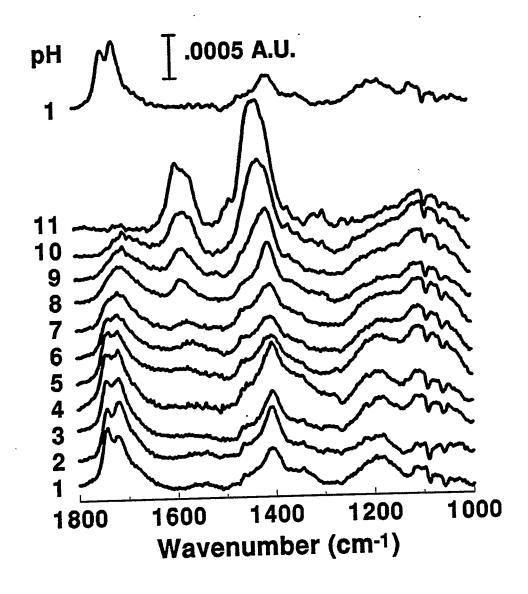


Figure 20. IRRAS spectra in low energy region for a $HO_2C(CH_2)_{10}S/Au$ monolayer as a function of solution pH.

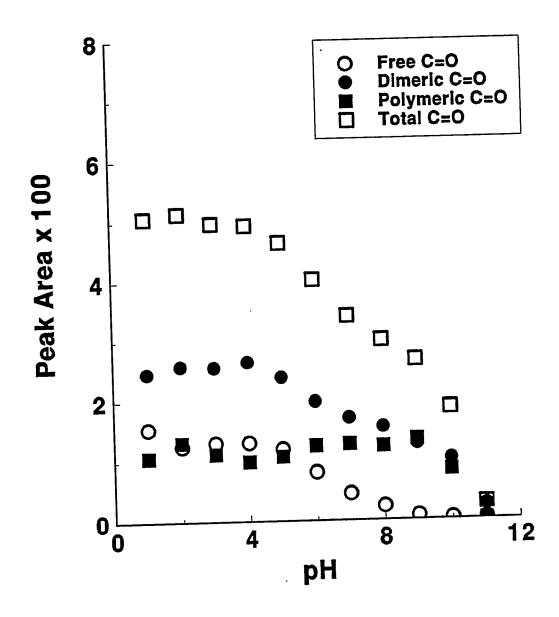


Figure 21. The IRRAS peak area of the C=O peaks of a $HO_2C(CH_2)_{10}S/Au$ monolayer as a function of solution pH.

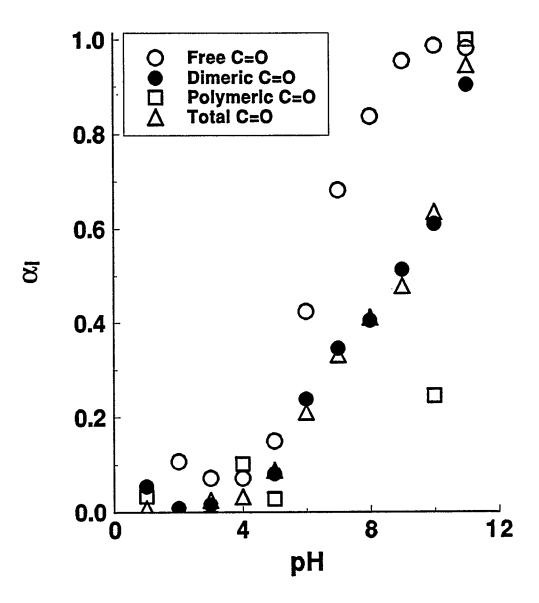


Figure 22. The degree of ionization α_i of a HO₂C(CH₂)₁₀S/Au monolayer as a function of solution pH.

MPA monolayers

There is virtually no $\nu(CH_2)$ peaks for the MPA/Au monolayer as shown in Figure 24. This indicate that axis of the two CH_2 groups are almost parallel to the surface normal so that the plane of each CH_2 is parallel to the surface. No change in this IRRAS region during the ionization process implies this chain axis remains the same all the time. This is further supported by the full recovery of the all the IRRAS peaks by re-immersion back to pH 0.50 solution as shown in Figure 25.

Both the $v_a(COO^-)$ and $v_s(COO^-)$ peaks do not appear until pH ~6 where the v(C=O) peaks almost disappear as shown in Figures 25 and 26.

The peak area of the three $\nu(C=O)$ peaks as a function of solution pH are shown in Figure 27. Apparently, the free $\nu(C=O)$ starts to be ionized after pH 3.5 and finishes the ionization process by pH 7.5. The hydrogen-bonded dimeric $\nu(C=O)$ starts ionization around pH 2 all the way up to pH 11. This transition of 9 pH units is very broad. The hydrogen-bonded polymeric $\nu(C=O)$ dose not have any significant ionization until pH 7.

With an initial value of about 0.05 at pH 3, the extent of ionization for both the free C=O and the hydrogen-bonded dimeric C=O starts to increase all the way to a finial value of 1 by pH 8 and pH 11, respectively, as shown in Figure 28. Meanwhile, the value for the hydrogen-bonded polymeric C=O is virtually 0 until around pH 7. The values of the effective pK_a for all three acid groups as a function of the extent of ionization are shown in Figure 29. The effective pK_a of the free acid always has the lowest value, although it is not very clear due to the too small amount of this free acid present in this monolayer. The effective pK_a of the free acid has an initial value of ~4.5 and remains almost the same value till 50 % of ionization and starts to increase to a finial value of ~8. The effective pK_a of the hydrogen-bonded dimeric acid has an initial value of 4.5 and increases up to ~10. It is clear that the has a initial value of about 8 and increase up to 11, though the ionization is only 66 %

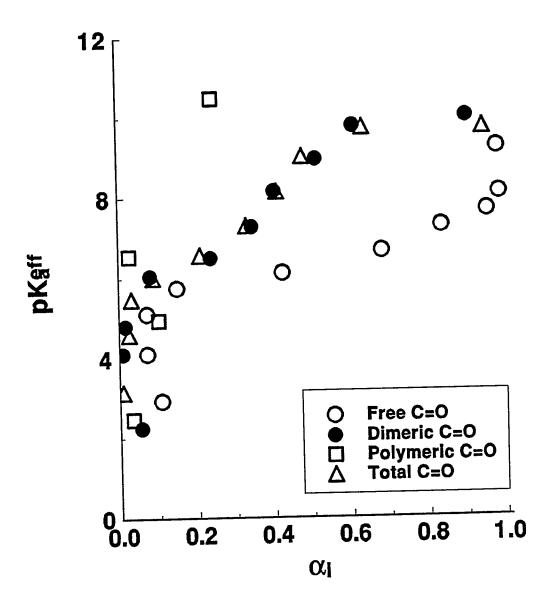


Figure 23. The effective pK_a of $HO_2C(CH_2)_{10}S/Au$ monolayer as a function of the extent of ionization α_i .

remain the same or decrease but not to increase as increasing solution pH as seen for all the monolayers mentioned previously. In this disulfide monolayer, there is a free space between two neighbor C=O groups since the alkane chain is shorter by 6 CH₂ in length. This provides the room for those carboxylate group which has one negative charge to eliminate the strong electrostatics repulsion. This suggests that the carboxylate group like to turn around to minimize the electrostatic repulsion if possible. Therefore, the two peaks cannot be used for the calculation of the extent complete. The large deviation for hydrogen-bonded polymeric acid is due to the very weak signal for it. The overall effective pK, is almost has the same values as that for the hydrogen-bonded dimeric acid. The effective pK, values at 50 % of ionization for the free acid, the hydrogen-bonded dimeric acid, the hydrogen-bonded polymeric acid, and the overall are 5.3, 5.7, 9.5, and 6.0, respectively.

PUAD monolayers

Unlike carboxylic acid-terminated thiolate monolayers, the PUAD/Au monolayer has a change in the $\nu(C-H)$ peaks intensity of the IRRAS spectra during the ionization process as shown in Figure 30. The peak intensity at 2926 cm⁻¹ almost has a same value of ~0.0010 A.U. when pH is less than 4 and starts increasing and levels off with a limiting value of 0.0018 A.U. by pH 6 as shown in Figure 31. Upon ionization, the peak intensity of the $\nu(C-H)$ mode at 2926 cm⁻¹ jumps from 0.0010 to 0.0018 A.U. as shown in Figure 32. Apparently, the monolayer structure has been disturbed by the electrolyte and the buffer after it is ionized. This is further supported by the change in the $\nu(C=0)$ peaks after reimmersion back to pH 1 solution and the presence of the extra phosphate (as a buffer) peaks around 1110 cm⁻¹, 1265 cm⁻¹, and 1410 cm⁻¹ as shown in Figure 33. The incorporation of the electrolyte and the buffer into this monolayer after ionization can be confirmed further from the increase in the peak area of the free v(C=O) mode and the hydrogenbonded polymeric v(C=0) mode with increasing the extent of ionization as shown in Figure 34. These two peak areas are supposed to either

Au-S(CH2)2COOH at 0.60 pH

.0005 A.U.

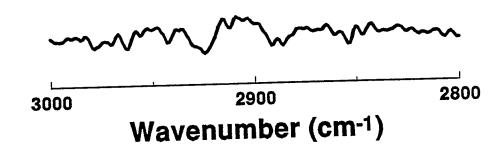


Figure 24. IRRAS spectra in the high energy region of a $HO_2C(CH_2)_2S/Au$ monolayer as a function of solution pH.

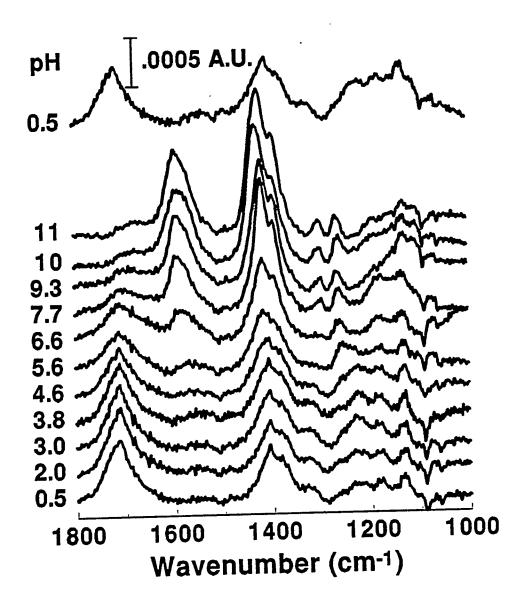


Figure 25. IRRAS spectra in the low energy region for a $HO_2C(CH_2)_2S/Au$ monolayer as a function of solution pH.

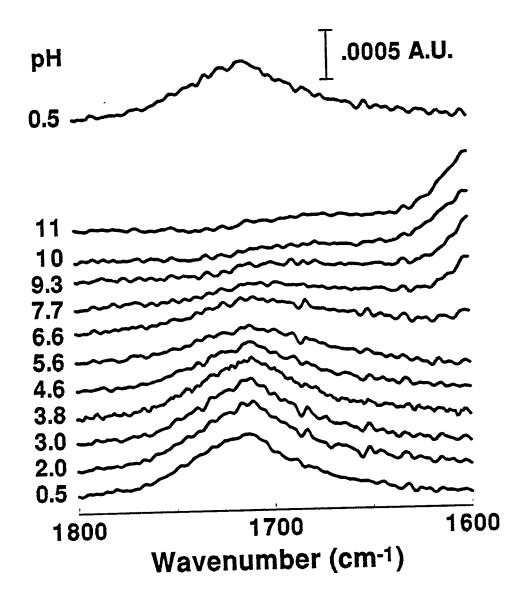


Figure 26. IRRAS spectra in the C=O stretching region for a $HO_2C(CH_2)_2S/Au$ as a function of solution pH.

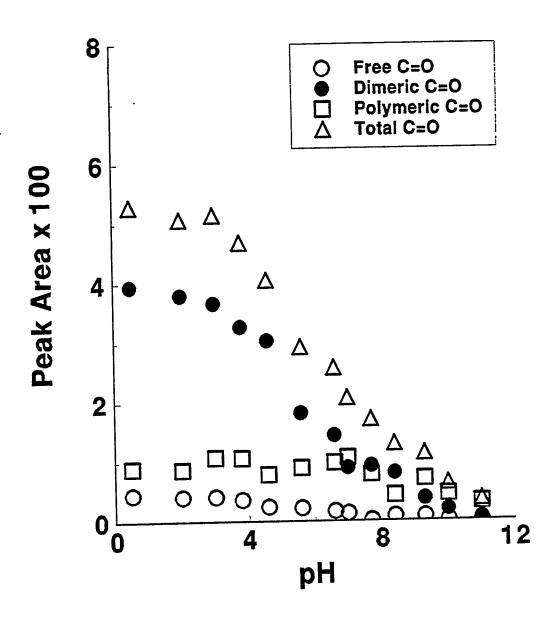


Figure 27. The IRRAS peak area of the C=O peaks of the IRRAS spectra for a HO₂C(CH₂)₂S/Au monolayer as a function of solution pH.

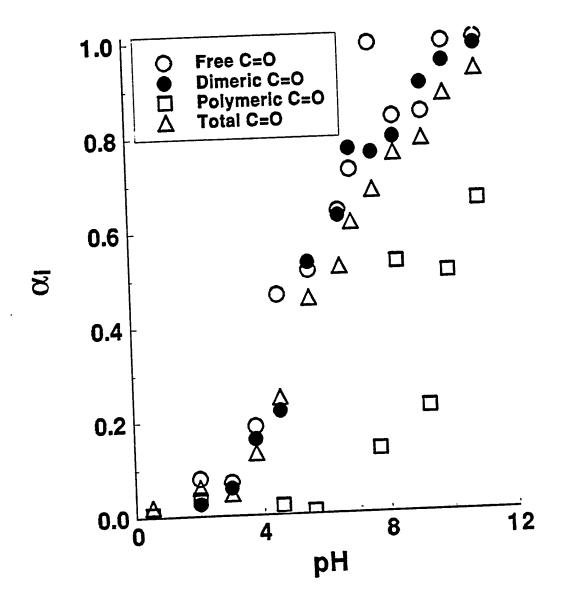


Figure 28. The degree of ionization α_i of a HO₂C(CH₂)₂S/Au monolayer as a function of solution pH.

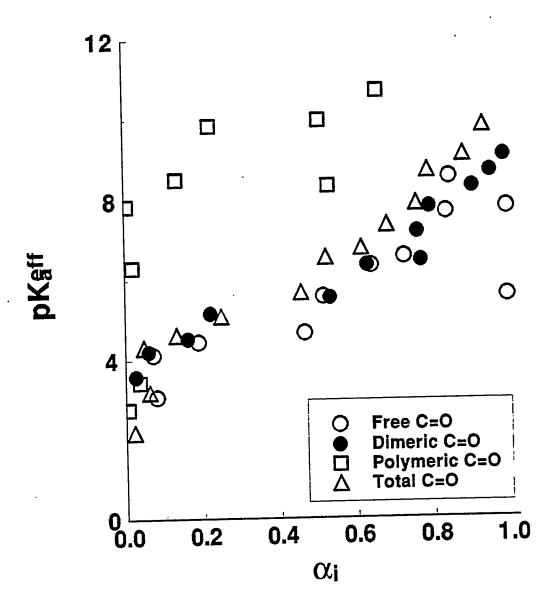


Figure 29. The effective pK_a of a $HO_2C(CH_2)_2S/Au$ monolayer as a function of the extent of ionization α_i .

of ionization or the effective pK_a. Fortunately, the free $\nu(C=O)$ and the hydrogen-bonded polymeric $\nu(C=O)$ peaks are relatively small, the hydrogen-bonded dimeric $\nu(C=O)$ peak is still can be used to estimate the overall acid property assuming the value of the overall effective pK_a is virtually the same as that of the dimeric acid. The extent of ionization is very small till pH 5 and starts to increase rapidly with a transition of at least 8 pH units as shown in **Figure 35**. The pK_a from IRRAS measurement for the dimeric acid is 6.5.

The value of the effective pK_a is ~7 in the beginning of ionization and starts to increase after 40 % of ionization to a value of 11 at 80 % of ionization as shown in Figure 36. The effective pK_a value at 50 % of ionization is 6.6. Since the orientation of this monolayer has changed upon ionization, there is no guarantee that the $\nu(C=O)$ dipole remains the same. The above analysis will not be valuable unless the $\nu(C=O)$ dipole remains the same orientation during the titration process. There is no $\nu(COO)$ peak observed for the PUAD/Au monolayer. This does not mean there is no $\nu(COO)$ groups at all, but instead the $\nu(COO)$ groups is in the monolayer with both the $\nu(COO)$ and $\nu(COO)$ dipole parallel to the surface so that there is no IRRAS signal can be seen.

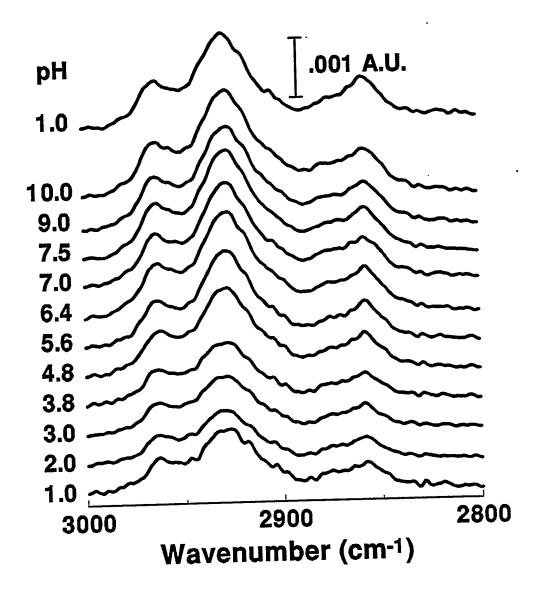


Figure 30. IRRAS spectra in the high energy region for a monolayer of $HO_2C(CH_2)_{10}SS(CH_2)_4CH_3$ at gold as a function of solution pH.

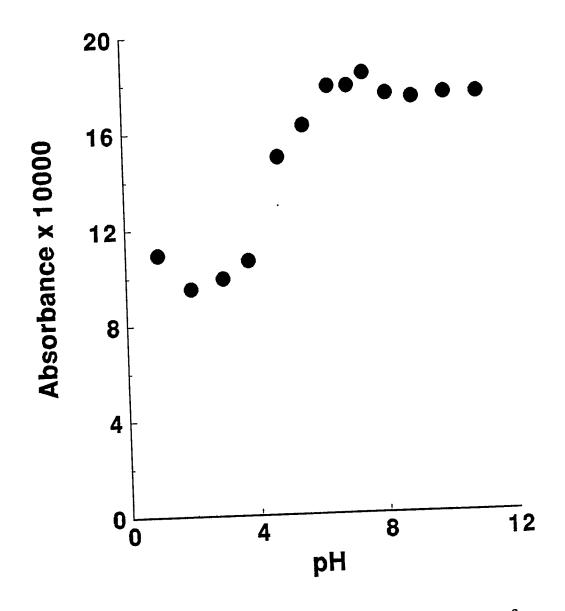


Figure 31. The IRRAS peak intensity at 2926 cm⁻¹ for a monolayer of HO₂C(CH₂)₁₀SS(CH₂)₄CH₃ at gold as a function of solution pH.

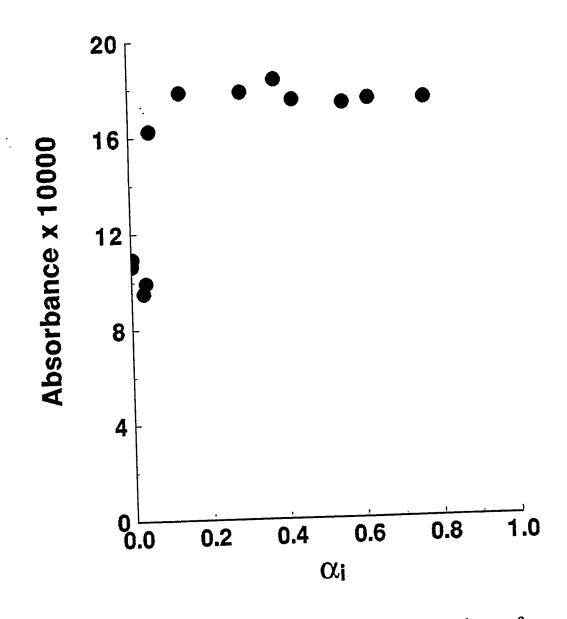


Figure 32. The IRRAS peak intensity at 2926 cm⁻¹ for a monolayer of $HO_2C(CH_2)_{10}SS(CH_2)_4CH_3$ at gold as a function of the extent of ionization α_i .

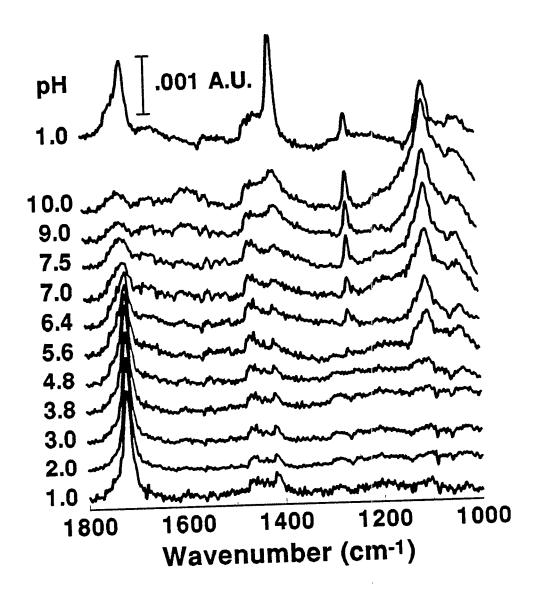


Figure 33. IRRAS spectra in the low energy region for a monolayer of $HO_2C(CH_2)_{10}SS(CH_2)_4CH_3$ at gold as a function of solution pH.

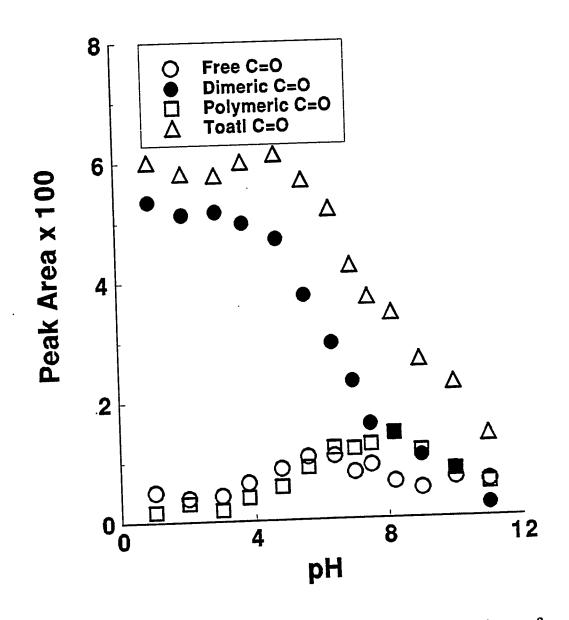


Figure 34. The IRRAS peak area of the ν (C=O) peaks of a monolayer of $HO_2C(CH_2)_{10}SS(CH_2)_4CH_3$ at gold as a function of solution pH.

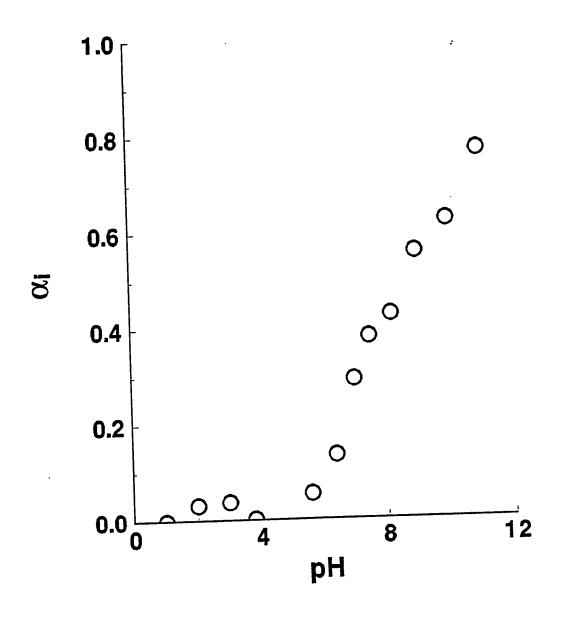


Figure 35. The degree of ionization α_i of a monolayer of $HO_2C(CH_2)_{10}SS(CH_2)_4CH_3$ at gold as a function of solution pH.

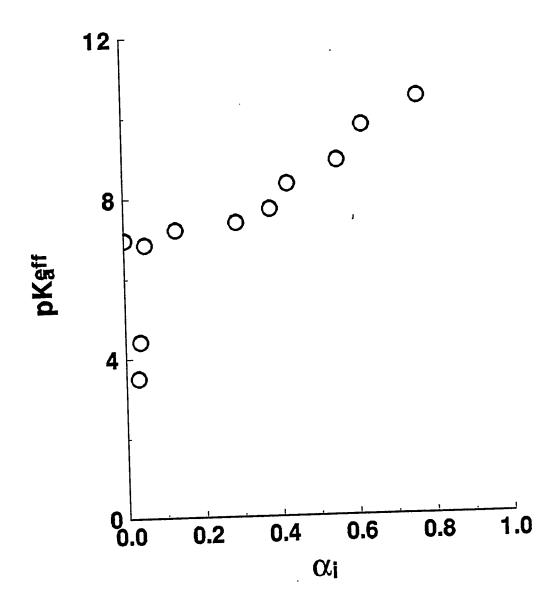


Figure 36. The effective pK_a of a monolayer of $HO_2C(CH_2)_{10}SS(CH_2)_4CH_3$ at gold as a function of the extent of onization α_i .

CONCLUSIONS

The spatial structure of densely-packed carboxylic acid-terminated monolayers remains intact, but that of a loosely-packed monolayer is disturbed by the incorporation of the electrolyte and buffer upon ionization. The carboxylate plane of the latter is oriented parallel to surface so that there is no IRRAS signal observed, though monolayers have been ionized.

For all acid studied, the effective pK, increases as the extent of ionization α_i , and the effective pK, values at 50 % of ionization for the free acid, the hydrogen-bonded dimeric acid, and the hydrogen-bonded polymeric acid are around 6, 8, and 10, respectively. The difference in pK_a values is mainly due to the difference in the strength of the hydorgen bond.

All three acid groups are weaker acids than the corresponding adsorbate precursor in aqueous solution. The values of the effective pK, at 50 % of ionization for MHA, MUA, PUAD, and MPA are 7.2, 8.0, 6.5 and 6.0, respectively.

REFERENCES

- (1) Bain, C. D.; Whitesides; G. M. Angew. Chem., Int. Ed. Engl. 1989, 28, 506, and references therein.
- (2) Whitesides, G. M.; Laibinis, P. E. Langmuir 1990, 6, 87, and references therein.
- (3) Ulman, A. An Introduction to Ultra-Thin Organic Films From Langmuir-Blodgett to Self-Assembly; Academic Press: San Diego, 1991.
- (4) Widrig, C. A.; Chung, C.; Porter, M. D. J. Electroanal. Chem. 1991, 310, 335.
- (5) Bryant, M. A.; Pemberton, J. E. J. Am. Chem. Soc. 1991, 113, 8284.
- (6) Bain, C. D.; Biebuyck, H. A.; Whitesides, G. M. Langmuir 1989, 5, 723.
- (7) Whitesides, G. M.; Laibinis, F. E. Langmuir, 1990, 6, 87.
- (8) Nuzzo, R. G.; Zegarski, B. R.; Dubois, L. H. J. Am. Chem. Soc. 1987, 109, 733.
- (9) Fenter, P.; Eisenberger, P.; Li, J.; Camillone, N. III; Bernasek, S.; Scoles, G.; Ramanarayanan, T. A.; Liang, K. S. *Langmuir* 1991, 7, 2013.
- (10) Chidsey, C. E. D.; Liu, G.-Y.; Rowntree, P.; Scoles, G. J. Chem. Phys. 1989, 91, 4421.
- (11) Widrig, C. A.; Alves, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2805.
- (12) Alves, C. A.; Smith, E. L.; Porter, M. D. J. Am. Chem. Soc. 1992, 114, 1222.
- (13) Widrig, C. A.; Chung, C.; Porter, M. D. J. Electroanal. Chem. 1991, 310, 335.
- (14) Bain, C. D.; Troughton, E. B.; Tao, Y-T.; Evall, J.; Wihitesides, G. M.; Nuzzo, R. G. J. Am. Chem. Soc. 1989, 111, 321.
- (15) Nuzzo, R. G.; Zegarski, B. R.; Dubois, L. H. J. Am. Chem. Soc. 1987, 109, 733.
- (16) Dubios, L. H.; Zegarski, B. R.; Nuzzo, R. G. J. Chem. Phys. 1993, 98(1), 678.

- (17) Hallmark, V. M.; Chiang, S.; Rabolt, J. F.; Swalen, J. D.; Wilson. R. J. Phys. Rev. Lett. 1987, 59, 2879.
- (18) Alves, C. A.; Porter, M. D. Langmuir 1993, 9, 3507.
- (19) Porter, M. D.; Bright, T. B.; Allara, D. L.; Chidsey, C. E. D. J. Am. Chem. Soc. 1987, 109, 3559.
- (20) Tarlvo, M. J.; Bowden, E. F. J. Am. Chem. Soc. 1991, 113, 1847.
- (21) Li, T. T.-T.; Weaver, M. J. J. Am. Chem. Soc. 1984,106, 6107.
- (22) De Long, H. C.; Donohue, J. J.; Buttry, D. A. Langmuir 1991, 7, 2196.
- (23) De Long, H. C.; Buttry, D. A. Langmuir 1990, 6, 1319.
- (24) Nuzzo, R. G.; Allara, D. L. J. Am. Chem. Soc. 1983, 105, 4481.
- (25) Nuzzo, R. G.; Dubois, L. H.; Allara, D. L. J. Am. Chem. Soc. 1990, 112, 558.
- (26) Czanderna, A. W.; King, D. E.; Spaulding, D. J. Vac. Sci. Technol. 1991, A9, 2607.
- (27) Pale-Grosdemange, C.; Simmon, E. S.; Prime, K. L.; Whitesides, G. M. J. Am. Chem. Soc. 1991, 113, 12.
- (28) Bain, C. D.; Whitesides, G. M. J. Am. Chem. Soc. 1988, 110, 5897.
- (29) Popenoe, D. D.; Deinhammer, R. S.; Porter, M. D. Langmuir, 1992, 8, 2521.
- (30) Finklea, H. O.; Avery, S.; Lynch, M.; Furtsh, T. Langmuir 1987, 3, 409.
- (31) Foley, J. K.; Pons, B. S. Anal. Chem. 1985, 57, 945A.
- (32) Pons, S.; Foley, J. K.; Russell, J.; Seversion, M. Mordern Aspects of Electrochemistry Eds. Bochris, J, O'M.; Conway, B. E.; White, R. E. 1986, 17, 223.
- (33) Bewick, A.; Kunimatsu, K.; Pons, B. S.; Russell, J. W. J. Electroanal. Chem. 1984, 160, 47.
- (34) Goldern, W. G. Fourier Transform Infrared Spectroscopy, Ed. Ferraro, J. R.; Basile, L. J. Vol. 4; New York: Academic Press, 1985.
- (35) Kemp, J. C. Polarized Light and Its Interaction with Modulating Devices, a Methology Review; Hillsboro, OR: HINDS International, Inc. 1987.
- (36) Popenoe, D. D.; Stole, S. M.; Porter, M. D. Appl. Spectroscopy 1992, 46, 79.

- (37) Wu, C.-C.; Porter, M. D. unpublished data.
- (38) Stole, S. M.; Porter, M. D. Langmuir 1990, 6, 1199.
- (39) Hansen, W. N.; Wang, C. L.; Humpherys, T. W. J. Electroanal. Chem. 1978, 90, 137.
- (40) Hansen, W. N.; Wang, C. L.; Humpherys, T. W. J. Electroanal. Chem. 1978, 93, 87.
- (41) Lolb, D. M.; Hansen, W. N. Surface Sci. 1979, 79, 205.
- (42) Hansen, W. N. Surface Sci. 1980, 101, 109.
- (43) Fujihira, M.; Kuwana, T. *Electrochimica Acta* 1975, 20, 565, and references therein.
- (44) D'Agostino, A. T.; Hansen, W. N. Surface Sci. 1986, 165, 268.
- (45) Sobocinski, R. L.; Bryant, M. A.; Pemberton, J. E. J. Am. Chem. Soc. 1990, 112, 6177.
- (46) Widrig, C. A.; Alves, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2805.
- (47) Stole, S. M.; Porter, M. D. Appl. Spectrosc. 1990, 44, 1418.
- (48) Walczak, M. M.; Chung, C.; Stole, S. M.; Widrig, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2370.
- (49) Wilson, E. B., Jr.; Decius, J. C.; Cross, P. C. Molecular Vibrations; McGraw-Hill: New York, 1955; pp 285-6.
- (50) Paper III of this dissertation.
- (51) Laibinis, P.; Whitesides, G. M.; Allara, D. L.; Tao, Y.-T.; Parik, A. N.; Nuzzon, R. G. J. Chem. Phys. 1991, 113, 7152.
- (52) Allara, D. L.; Nuzzo, R. G. Langmuir 1985, 1, 45.
- (53) Allara, D. L.; Nuzzo, R. G. Langmuir 1985, 1, 52.
- (54) Scowen, R. V.; Leja, J. Can. J. Chem. 1967, 45, 2829.
- (55) Nakamoto, K. Infrared and Raman Spectra of Inorganic and Coordination Compounds; Third ed.; Wiley: New YorK, NY, 1978, 269.
- (56) Peter, R. A. Proc. R. Soc. London, Ser. A 1931, 133, 140.
- (57) Danielli, J. F. Proc. R. Soc. London, Ser. B. 1937, 122, 155.
- (58) Shulman, J. H.; Hughes, A. H. Proc. R. Soc. London, Ser. A. 1932, 138, 430.
- (59) Glazer, J.; Dogan, M. Z. Trans. Fraday Soc. 1953, 49, 448.

- (60) Sander, J. V.; Spink, J. A. Nature (London) 1955, 175, 644.
- (61) Betts, J. J.; Pethica, B. A. Trans. Fraday Soc. 1956, 52, 1581.
- (62) Mille, M. J. Colloid Interface Sci. 1981, 81, 168.
- (63) Laskowaski, M.; Scheraga, H. A. J. Am. Chem. Soc. 1954, 76, 6305.
- (64) Mille, M.; Van der Kooi, G. J. Colloid Interface Sci. 1977, 61, 475.
- (65) Lukac, S. J. Phys. Chem. 1983, 87, 5045.
- (66) Holmes-Farley, S. R.; Reamey, R. H.; McCarthy, T. J.; Deutch, J.; Whitesides, G. M. Langmuir 1985, 1, 725.

PAPER V

STUDY OF FORMATION AND ESTERIFICATION OF 11-MERCAPTOUNDECANOIC ACID MONOLAYERS AT GOLD USING IRRAS AND CONTACT ANGLE MEASUREMENT

INTRODUCTION

The self-assembly thiol monolayers on gold has attracted enormous world-wide attention for the last ten years due to the good stability, well-characterized systems, easy preparation, and excellent modification properties for applications of these monolayers¹⁻³.

The 16-mercaptohexadecanoic acid (MHA) monolayer has been characterized by Nuzzo et al.⁴ They concluded that the structure of a MHA/Au monolayer is almost a perfect zigzag (all trans) conformation. These bands are assigned to the progression of CH_2 twisting and wagging modes⁵. From the IRRAS $\nu(CH_2)$ modes, the tilt angle and twist angle were calculated to be 32° and 55°, respectively, with the axis of C=O tilted from the surface normal by 66°.⁴ The thickness of the MHA/Au monolayer was measured with ellisometry method to be 16 Å⁴.

For application purposes, it is very likely to modify a monolayer to construct a new one with the right properties. Therefore, it is essential to understand the surface reaction fully in order to facilitate these modification processes. Although esterification at a polymer surface has been studied by Whitesides et al, the complicated polymer system can not serve as a good model for surface reaction study because of the limited understanding regarding the coverage and the orientation of carboxylic acids at these polymer surface⁶.

In this study, two 11-mercaptoundecanoic acid monolayers on Au with different fractional coverages of 0.50 and 1.00 were used as models for a surface reaction study that probed the relationship between the coverage and extent of reaction using IRRAS and contact angle measurements.

EXPERIMENTAL SECTION

Substrate preparation

Electrodes were prepared by the resistive evaporation of 15-20 nm of chromium, followed by 200-300 nm of gold, onto 3 inch by 1 inch glass substrates. The temperature during the evaporation, measured at the plate supporting the substrates, increased to ~50° C as a result of radiative heating by the evaporation source. The evaporation rates were 0.1 and 0.5 nm/s for chromium and gold, respectively. The pressure in a cryopumped E360A Edwards Coating System during evaporation was <9x10⁻⁵ Pa. After ~45 min cooling time, the evaporator was back-filled with purified N₂. The substrates were then removed, analyzed by optical ellipsometry for the determination of the optical functions of the uncoated substrates, and immersed in the thiol-containing solution.

The surfaces of the uncoated gold electrodes have been characterized previously at both a macroscopic and microscopic level using scanning tunneling microscopy⁷ and electrochemical measurements of surface texture and surface area⁸. Macroscopically, the STM images show that the electrode surface is composed of atomically-terraced crystallites ~25 nm in diameter. The electrodes have a roughness factor of ~1.3, which is given as the surface area determined from the oxidative desorption of iodine⁸ divided by the geometric electrode area.

Monolayer preparation

The MUA monolayers were prepared by their spontaneous adsorption onto the evaporated gold from 0.100 mM solutions in absolute ethanol for 10 sec and 24 hours in order to obtain two film thickness of 8.0 Å and 16.0 Å, emersed, rinsed successively with absolute ethanol and hexane, and dried under a stream of prepurified N₂.

Esterification

Samples were left in solution for different lengths of time in 20 % (v/v) of conc. HCl ethanol solution. Upon removal from solution, the samples were rinsed with pH 1.0 HCl aqueous solution and then placed on a spin-coater and

spun briefly at 2000 rpm to remove excess solution. The samples were then placed in the N_2 -purged sample chamber of the IR spectrometer for characterization. Spectra were usually acquired within a few minutes of loading in the chamber, with the rotational vibrational bands of vapor phase water subtracted by using a reference vapor spectrum.

Ellipsometric measurements of film thickness

The thickness of the monolayers were determined by optical ellipsometry at 632.8 nm in two steps with a computer-interfaced Gaertner Model L-116B ellipsometer. The angle of incidence was 70° from the surface normal. Upon removal from the evaporation chamber, the analyzer and polarizer angles for a reflected light beam from each uncoated substrate were measured on at least three different spots. The average complex refractive index for each substrate was then calculated with a two phase parallel layer model from classical electromagnetic theory. After monolayer formation, each sample was again analyzed and the film thickness calculated from a three-phase parallel layer model, using the average complex refractive index of the individual sample and a real refractive index of 1.46 for the film. A value of 1.46, which is representative of the adsorbate precursors facilitates comparison with thickness data that has been reported for a variety of monolayers 2,11-15. The influence of several refinements on this treatment of ellipsometric data have been recently discussed 11,12,16.

Contact angle measurements

Advancing contact angles (θ_a) were measured in air with pH = 1.0 HCl aqueous solution as probe liquids by using a Rame-Hart Model 100-00 115 goniometer. For these measurements, a 2.0 mL droplet was formed on the substrate with the needle of the syringe in the droplet. The value of θ_a was determined as the volume of the droplet was slowly increased.

Infrared spectroscopy

Infrared spectra were acquired with either a Nicolet 740 or a 750 FT-IR spectrometer. Spectra of the adsorbate precursors were obtained by their dispersion in KBr. Spectra of the monolayers were obtained in a single

reflection mode by using p-polarized light incident at 82° from the surface normal and a liquid nitrogen cooled HgCdTe detector. A home-built sample holder was used to position reproducibly the substrates in the sample chamber of the spectrometer¹⁷. The spectrometer was purged with boil-off from liquid N₂. Spectra were obtained by referencing 1024 sample scans to 1024 background scans at 2 cm⁻¹ resolution (zero filled) with Happ-Genzel apodization. All spectra are reported as -log(R/R_o), where R is the reflectance of the coated substrate and Ro is that of a reference Au substrate. The reference substrates were prepared by immersion of an uncoated gold substrate into ~1 mM ethanolic solution of octadecanethiol-d₃₇. These films provide a substrate free from detectable IR bands in the regions relevant to these studies as determined when referenced to an uncoated Au substrate that was cleaned in a X 1:3 H₂O₂(30%):H₂SO₄(con.) solution. Further details of the preparation of the reference substrates are given elsewhere 18. Caution: The H₂O₂/H₂SO₄ solution reacts violently with organic compounds and should be handled with extreme care.

The orientation analysis, which compares the observed spectrum to that calculated for an isotropic array of oscillators of comparable packing density, was accomplished by using the frequency-dependent complex optical constants of the adsorbate precursor or a suitable substitute. The optical constants for 11-mercaptoundecanoic acid was determined from a transmission spectrum after its dispersion as a crystalline material in KBr. From the absorbance values, a Kramers-Kronig transformation yielded the frequency-dependent complex optical constant.

Chemical and reagents

The 11-mercaptoundecanoic acid (HO₂C(CH₂)₁₀SH) (MUA) was synthesized in this group and characterized with IR and NMR. Ethyl alcohol was from Quantum Chemical Corp. (IL). The rest of chemicals was obtained from Aldrich. All other reagents were used as received. Aqueous solutions were freshly prepared from water purified by passage through a Milli-Q purification system (Millipore Corp., Bedford, MA).

RESULTS AND DISCUSSION

Monolayer formation

Ellipsometric thickness as a function of immersion time

The thickness of the MUA monolayer on gold (MUA/Au) was measured as a function of immersion time using optical ellipsometry. The results are shown in *Figure 1*. In 0.1 mM MUA solution, the thickness of MUA/Au increases rapidly at short immersion times, reaching a value of ~6 Å within 0.1 min. The rate of the process slow thereafter with the thickness doubling within ~4 min and approaching a limiting value of 16.0 Å.

This formation process is not a simple one step Langmuir isotherm surface process. Actually, this result is consistent with a two-step formation process proposed by Whitesides et al.²⁰ and confirmed by Shimazu et al.²¹ and Grunze et al.²²⁻²⁴ The first step is a Langmuir adsorption process, and the second step is believed to be a result of the combination of replacement of the incorporated solvent and adsorbed impurities and lateral diffusion on the surface to reduce defects and enhance packing²⁰. This second process has a much slower rate than that of the first process.

Wetting as a function of immersion time

The wetting property of the surface as a function of immersion time was characterized with contact angle measurements. The probe liquid of a pH 1.0 HCl aqueous solution was used to keep the acid monolayers in acidic form and the ionic strength the same. The contact angle of water (θ_a) and $\cos\theta_a$ as a function of immersion time are shown in *Figure 2*. The contact angle decreases very dramatically from 47.0° to 10.0° in less than 4 min and decreases only slightly thereafter.

The observation clearly demonstrates that surface properties of the film are determined by the structure of the monolayer. Therefore, it is possible to manipulate the coverage and the structure of a monolayer to control the surface properties of this film by varying immersion time. In this study, two MUA/Au monolayers with different ellipsometric thicknesses of 8.0 Å and 16.0 Å were prepared in 0.1 mM solution for 10 sec and 24 hours, respectively. Since the

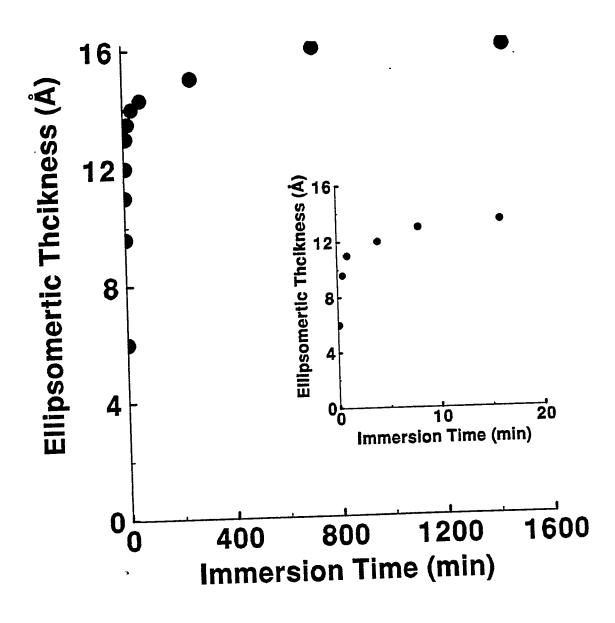


Figure 1. The ellipsometric thickness of a MUA/AU in 0.1mM MUA ethanol solution as a function of immersion time.

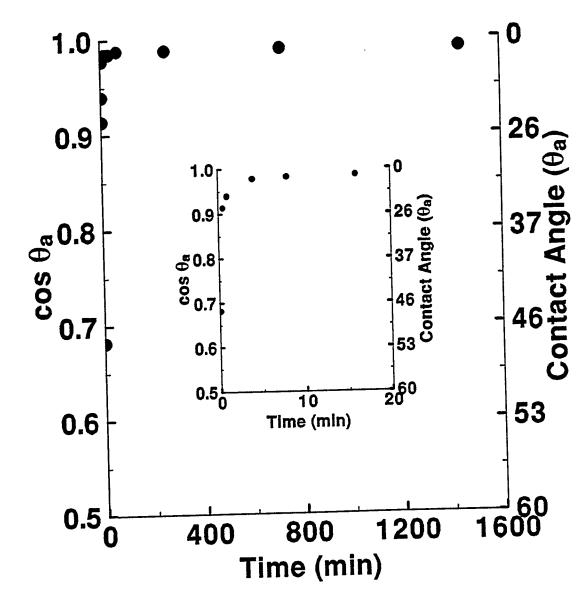


Figure 2. $\cos \theta_a$ of pH 1.0 HCl probe solution at a MUA/Au prepared in 0.1mM MUA ethanol solution as a function of immersion time.

limiting thickness of the MUA/Au monolayer is 16.0 Å, the fractional coverage of the MUA/Au monolayers with the ellipsometric thicknesses of 8.0 Å and 16.0 Å are 0.50 and 1.0, respectively. The contact angle of water at the half-coverage MUA/Au monolayer and the full-coverage MUA/Au monolayer are 25.0 ° and 8.0°, respectively.

IRRAS spectra as a function of immersion time

Figure 3 is a series of IRRAS spectra in the high energy region of a MUA/Au monolayer as a function of immersion time. The peak at 2920 cm⁻¹ is assigned to the $v_a(CH_2)$ mode, and the other peak at 2850 cm⁻¹ is the $v_s(CH_2)$ mode²⁵. The shift to low energy of the peak frequencies of both modes with increasing immersion time indicates that the MUA/Au monolayer becomes more crystalline-like²⁵ with increasing the coverage. The decrease in the peak area of both modes with increasing immersion time suggests that the average tilt angle of the polymethylene chain decreases with increasing the coverage.

IRRAS spectra in the low region as a function of immersion time is shown in **Figure 4**. The weak peaks between 1150 cm⁻¹ and 1300 cm⁻¹ are the progression peaks resulting from an all-trans conformation⁴. Peaks between 1400 cm⁻¹ and 1500 cm⁻¹ are due to the CH₂ bending mode. In the carbonyl stretching region (1600 cm⁻¹ -1750 cm⁻¹), the spectrum transfers clearly from a single band at 1717 cm⁻¹ to a doublet. The low energy band at 1717 cm⁻¹, assigned to a hydrogen-bonded dimeric ν (C=O) mode of a carboxylic acid (see below), decreases with increasing immersion time. The band at 1745 cm⁻¹, assigned to a non-hydrogen-bonded ν (C=O) mode of a carboxylic acid, i.e. a free acid, increases with immersion time. In fact, there is another peak at 1680 cm⁻¹ which will be discussed later.

IRRAS characterization

The average tilt angle of the polymethylene chain

The coverage, the packing density, and the tilt angle can be calculated from the IRRAS data assuming the experimental spectrum has all the peaks corresponding to that in the KBr spectrum. The experimental and the

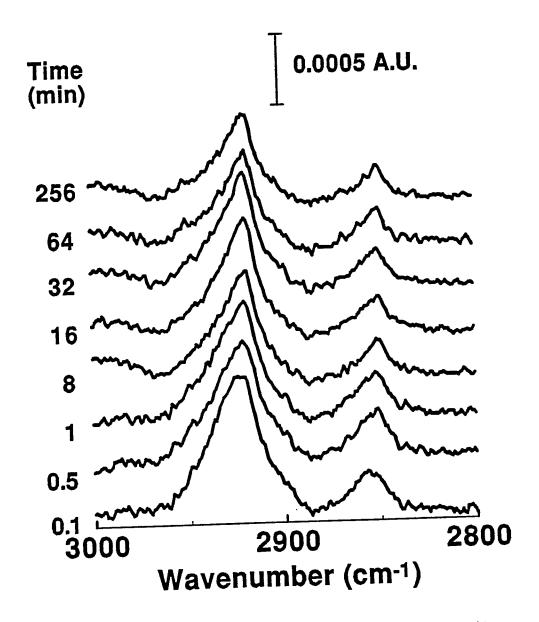


Figure 3. The IRRAS spectra in the high energy region of a MUA/Au prepared in 0.1mM MUA ethanol solution as a function of immersion time.

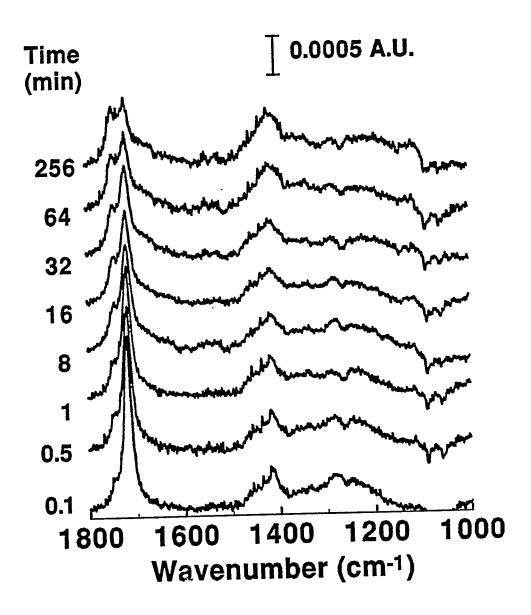


Figure 4. The IRRAS spectra in the low energy region of a MUA/Au prepared in 0.1mM MUA ethanol solution as a function of immersion time.

calculated IRRAS spectra of MUA/Au obtained as a function of immersion time in the high energy region and the low energy region are shown in *Figures* 5 and 6, respectively. The vibrational mode assignments are summarized in **Table 1**. The ν (C=O) peak at 1713 cm⁻¹ in KBr spectrum is a hydrogen-bonded dimeric ν (C=O) mode which is transformed to three different ν (C=O) peaks when MUA is on the Au surface as shown in **Figures 6 and 7**.

The average tilt angle of the axis of this monolayer can be calculated from the following equation

$$\cos \theta_{mz} = (A_{obs}/3A_{calc}) \tag{1}$$

where A_{obs} and A_{calc} are the observed and the calculated peak area, θ_{mz} is the average tilt angle between the transition dipole moments (m) and the surface normal $(z)^{13}$.

The calculated average tilt angle of the polymethylene chain of the MUA/Au monolayer as a function of immersion time is shown in **Figure 8**. At the early stage of the adsorption process, the tilt angle of the polymethylene chain is large, suggesting a low packing density. This is consistent with the results from the ellipsometric thickness measurement data. The tilt angle decreases from 57 ° to 25° in the first 4 min and there is not much decrease thereafter. After 1440 min of immersion time, this tilt angle of the polymethylene chain of the MUA/Au is 24° which is less than the value of 32° determined by Nuzzo et al. for the MHA/Au monolayer⁴. The less tilt angle for short chain monolayers is a result of the less chain-chain interaction for these monolayers than long chain monolayers.

At this tilt angle, this monolayer should have a packing density of $8.0~\rm x$ $10^{-10}~\rm mol/cm^2$, i.e. the distance between the nearest neighbor is $5.0~\rm \mathring{A}$ which is the same as that for n-alkane thiolate monolayers on $\rm Au^{7,8}$.

The average tilt angle of carbonyl dipole

The experimental IRRAS spectrum has all the peaks corresponding to the calculated spectrum. Interestingly, the experimental spectrum has two extra peaks: one at 1743 cm⁻¹ which is the non-hydrogen-bonded $\nu(C=O)$

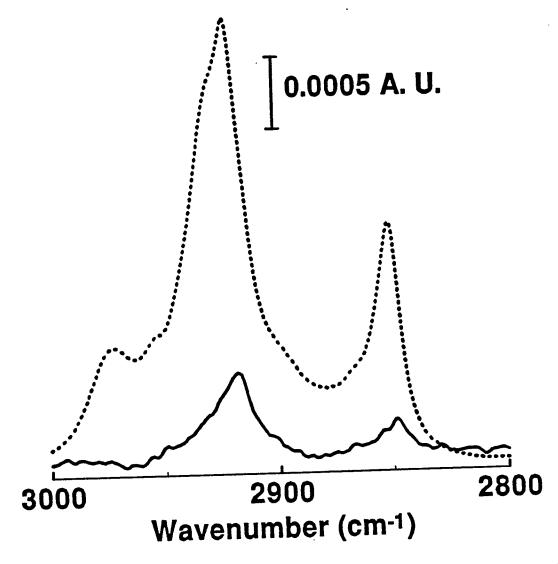


Figure 5. The experimental (solid line) and calculated (dashed line) IRRAS spectra in the high energy region for a full-coverage MUA/Au monolayer.

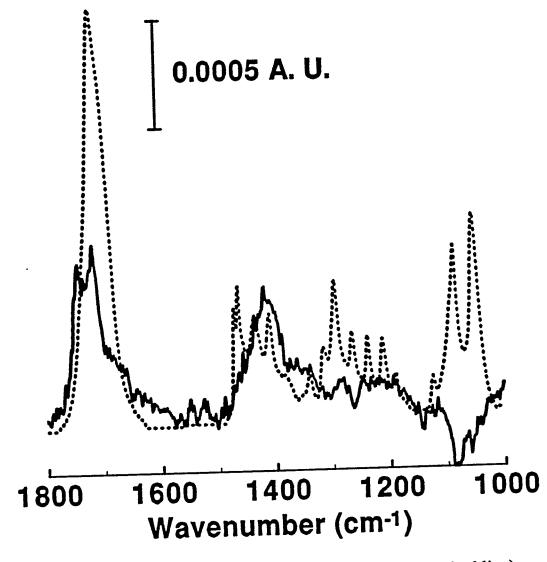


Figure 6. The experimental (solid line) and calculated (dashed line) IRRAS spectra in the low energy region for a full-coverage MUA/Au monolayer.

Table 1. Peak positions, mode assignments, and transition dipole moment directions of carboxylic acid- and ester-terminated alkanethiolate monolayers at gold.

]	HO ₂ C(CH ₂) ₁₀ S/Au		C ₂ H ₅ O ₂ C(CH ₂) ₁₀ S/Au		Au
Mode Assignment	obs. c	alc.	obs.	calc.	direction of transition dipole ^a
$v_a(CH_{3,ip})$			2964 ^w		to C-CH₃ bond in C-C-C chain plane
$v_a(CH_{3,op})$			b	2954	to C-CH ₃ bond C-C-C chain plane
$\nu_{\rm s}({ m CH_3,FR_1}$)c		2936		to C-CH3 bond
$v_a(CH_2)$	2918	2921	2918	2920	to C-C-C chain
$\nu_{\rm s}$ (CH ₃ ,FR ₂	r)c		2878 ^w		∥ to C-CH ₃ bond
$\nu_{\rm S}({ m CH_2})$	2850	2851	2850	2851	to C-C-C plane bisceting H-C-H
ν(C=O)	1743		1743	1740	to C=O bond
(0 0)	1717	1713			
	1680				
δ (CH ₂)	w	1471	1465	1467	in HCH plane, bisecting HCH
ν(C-O)+		1437			
δ(C-O-H)d		1441		1438	
δ (a-CH ₂)		1410			a arr 1 1
δ (CH ₃)			1382	1372	to C-CH ₃ bond
CH ₂ wags and twists	1200-13	1200-1350		50	to axis of chain
ν(C-O)	е	1086	1208	1212	
ν(C-O)	е	1051	1179	1176	

Table 1. (continued)

aOrientation of transition dipoles for CH modes assuming all-trans conformation (R. G. Snyder, J. Chem. Phys. 1965, 42, 1744). bThe $\nu_a(\text{CH}_{3,\text{op}})$ mode was not observed in the monolayer spectrum. cThe fermiresonance couplet, $\nu_s(\text{CH}_3,\text{FR}_1)$ and $\nu_s(\text{CH}_3,\text{FR}_2)$, is designated by the subscripts 1 and 2, which refer to the higher and lower energy components, respectively. d This band involves both the stretching of the C-O bond and the deformation of the C-O-H angle (D. Hadzi and M. Pintar, Spectrochim. Acta, 1958, 12, 162). The peak is canceled by the reference peaks. Peak intensity is weak.

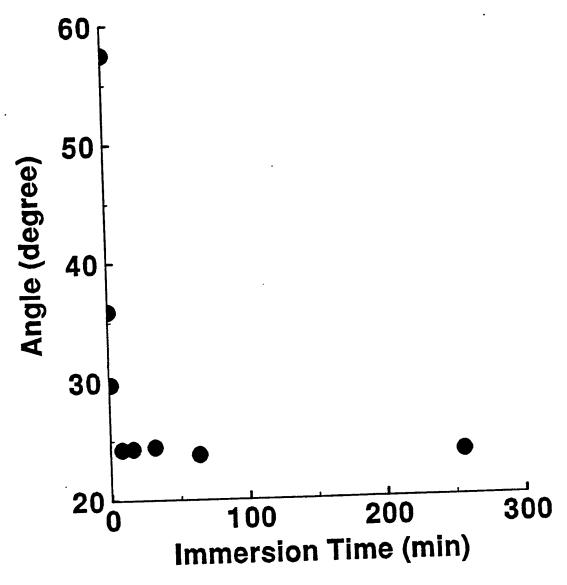
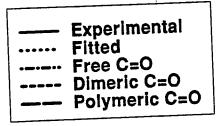


Figure 7. The experimental ν (C=O) mode and deconvoluted ν (C=O) modes for a half-coverage MUA/Au in 20 % (ν / ν) HCl ethanol solution for 48 min.



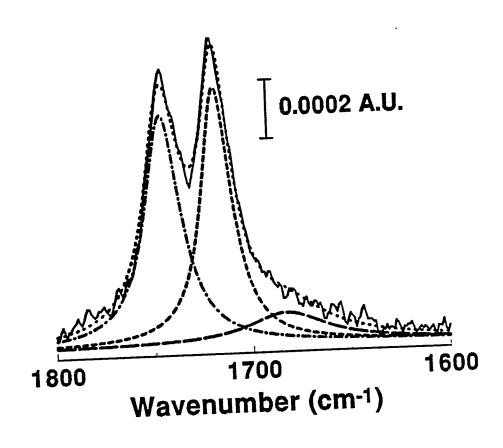


Figure 8. The calculated average tilt angle of a MUA/Au monolayer as a function of immersion time in 20 % (v/v) HCl ethanol solution.

mode and the other one at 1680 cm⁻¹.

The $\nu(C=O)$ peaks as a function of immersion time are shown in *Figure* 4. As one can see, the peak intensity of the hydrogen-bonded dimeric $\nu(C=O)$ peak decreases with increasing immersion time, while that of the non-hydrogen-bonded $\nu(C=O)$ increases with time. The average tilt angle for the $\nu(C=O)$ dipole was calculated using equation 1 with a calculated IRRAS spectrum.

The average tilt angle of the $\nu(C=0)$ dipole starts at 52° at 0.10 min. and increase rapidly up to 60° in 4 min. There is only little increase in the tilt angle thereafter.

Three carbonyl modes

The $\nu(C=O)$ curve cannot be fitted unless a third C=O peak is introduced to match the low energy region of the $\nu(C=O)$ envelope. The fitted curves match the experimental curves very well using three $\nu(C=O)$ modes as shown in **Figure 7**. It is well known that the hydrogen bonding strengths the $\nu(C=O)$ bond of the carboxylic acids and shifts the peak frequency of the $\nu(C=O)$ mode to lower energy by 20 cm⁻¹ to 80 cm⁻¹ depending upon the strength of the hydrogen bonding, i.e. the stronger the bonding the larger the shift²⁶⁻³¹.

There is a ν (C=O) mode at 1680 cm⁻¹ in addition to the non-hydrogen-bond ν (C=O) mode and the hydrogen-bond dimeric ν (C=O) mode. The ν (C=O) mode at 1680 cm⁻¹ is assigned to the hydrogen-bonded polymeric ν (C=O) mode which has been discussed in detail elsewhere³². It is believe that the three structures are a function of the distance between the two most adjacent carboxylic acid neighbors, d. When d>7.0 Å, 7.0 Å> d>5.0 Å, and d<5.0 Å, the majority carboxyl groups are the hydrogen-bonded dimeric C=O, the hydrogen-bonded polymeric C=O, and the non-hydrogen-bonded C=O, respectively.

For a MUA/Au, the peak frequencies of the non-hydrogen-bonded $\nu(C=O)$ mode, the hydrogen-bonded dimeric $\nu(C=O)$ mode and the hydrogen-bonded polymeric $\nu(C=O)$ mode are at 1743 cm⁻¹, 1717 cm⁻¹, 1680 cm⁻¹, respectively. The hydrogen-bonded dimeric C=O is believed to have the side-to-side hydrogen bonding instead of a head-to-head hydrogen bonding⁴.

Esterification of MUA/Au monolayers IRRAS

Figures 9 and 10 show the IRRAS spectra in the high energy and the low energy region for a MUA/Au with fractional coverage of 0.50 prior to and after esterification, respectively. In Figure 9, there are very weak CH₃ modes at 2965 cm⁻¹ and 2875 cm⁻¹ after 330 min of reaction time. This weak intensity is mainly a result of the small cross section found for related ester-terminated monolayers and their adsorbate precursors of MUA⁴. After 330 min of reaction time, the peak energies for asymmetric ν (CH₂) mode and symmetric ν (CH₂) mode shift to lower energy by 8 cm⁻¹ and 6 cm⁻¹, respectively. This indicates that the esterification process strengthen the polymethylene chainchain interaction from a liquid-like phase to a crystalline-like phase²⁵. At the same time, the decrease in the peak area of these two modes after esterification suggests that this film has better ordering than the one before esterification.

Before esterification, MUA/Au has only one strong $\nu(C=O)$ peak at 1717 cm⁻¹ which is the hydrogen-bonded dimeric $\nu(C=O)$ mode of the acid-terminated thiolate monolayer as shown in **Figure 10**. After esterification, MUA/Au has several strong peaks between 1000 cm⁻¹ and 1400 cm⁻¹ which are the fingerprint of the ester⁴ in addition to a different $\nu(C=O)$ mode. The peak at 1180 cm⁻¹ and 1208 cm⁻¹ are associated with the two newly formed $\nu(C=O)$ modes⁴. The hydrogen-bonded $\nu(C=O)$ peak at 1717 cm⁻¹ is converted to the non-hydrogen-bonded $\nu(C=O)$ peak at 1743 cm⁻¹ after esterification.

The conversion of the $\nu(C=O)$ from a hydrogen-bonded mode to a non-hydrogen-bonded mode allows the extent of esterification to be monitored and calculated as shown in Figure 11.(see below) In fact, there is a relatively small hydrogen-bonded polymeric $\nu(C=O)$ peak at 1680 cm⁻¹ in addition to these two strong $\nu(C=O)$ peaks as shown in Figure 7. No obvious change in the hydrogen-bonded polymeric $\nu(C=O)$ peak as a function of reaction time, as shown in Figure 12, indicates that the hydrogen-bonded polymeric $\nu(C=O)$ group behaves totally differently from the hydrogen-bonded dimeric $\nu(C=O)$ group due to the strong network-type hydrogen bond in terms of its reactivity.

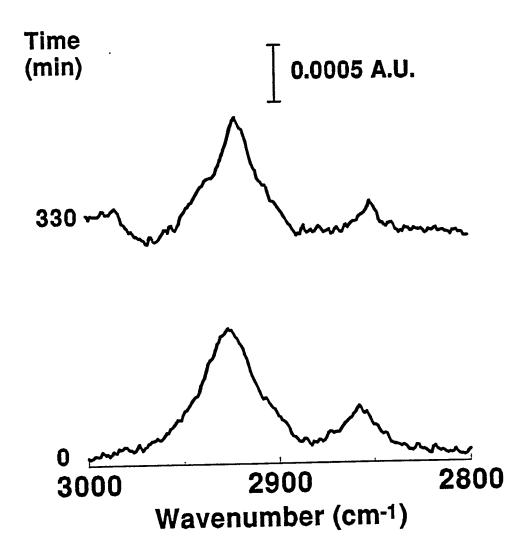


Figure 9. The IRRAS spectra in the high energy region of a half-coverage MUA/Au monolayer in 20 % (v/v) HCl ethanol solution for 0 and 330 min.

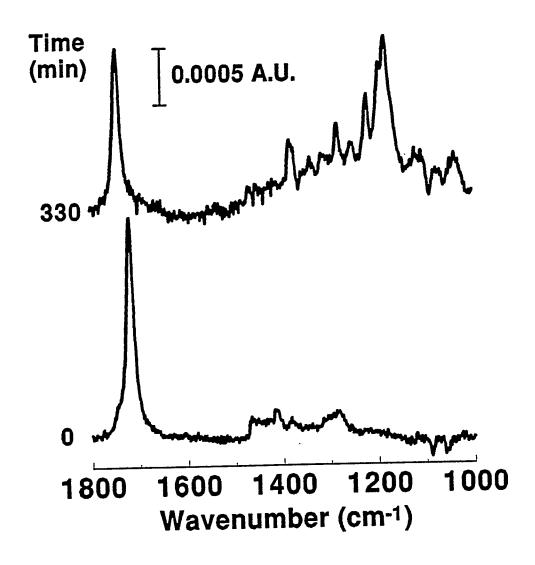


Figure 10. The IRRAS spectra in the low energy region of a half-coverage MUA/Au in 20 % (v/v) HCl ethanol solution for 0 and 330 min.

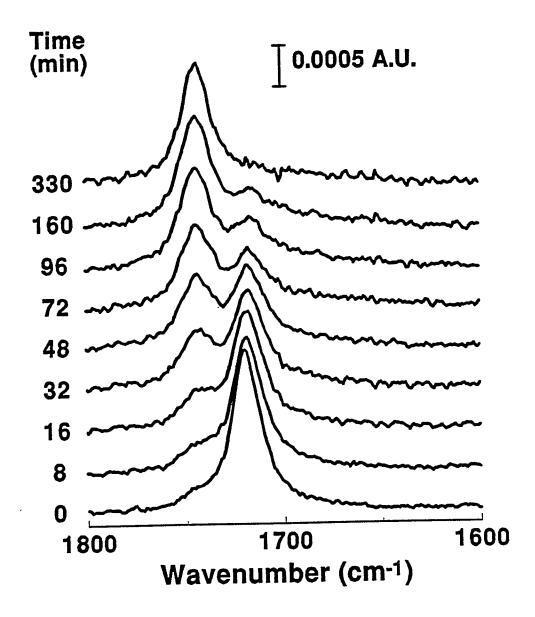


Figure 11. The IRRAS spectra in the $\nu(C=0)$ region of a half-coverage MUA/Au monolayer as a function of immersion time in 20 % (ν/ν) HCl ethanol solution.

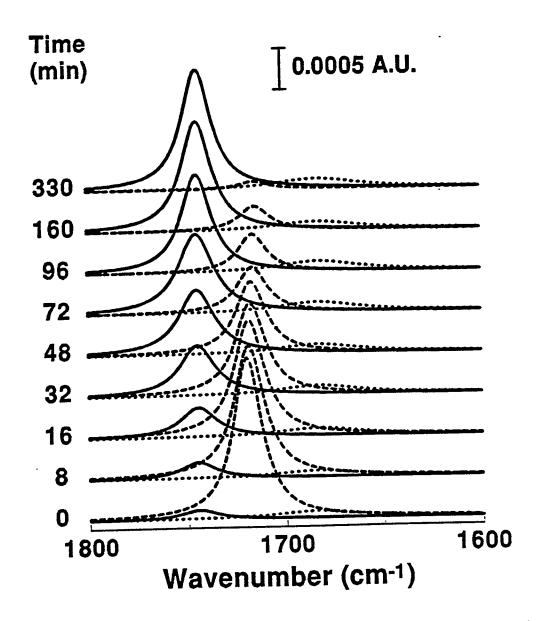


Figure 12. The deconvoluted ν (C=O) modes of a half-coverage MUA/Au in 20 % (ν / ν) HCl ethanol solution as a function of immersion time.

This result is consistent with the finding from the surface titration experiment discussed in **Paper III** of this dissertation. The kinetic data can be obtained from the deconvoluted peaks for the half-coverage monolayer as a function of reaction time, as shown in **Figures 11-13**, but not for the full-coverage monolayer because the IRRAS peak intensities of both the hydrogen-bonded dimeric $\nu(C=O)$ at 1717 cm⁻¹ and the non-hydrogen-bonded $\nu(C=O)$ at 1743 cm⁻¹ are weak for the full-coverage MUA/Au monolayer.

Contact angle measurements

As expect, the conversion of the hydrogen-bonded dimeric $\nu(C=0)$ to the non-hydrogen-bonded $\nu(C=0)$ due to esterification dramatically changes the wetting properties of the surface. This is shown in Figure 14. The contact angle, θ_a , increases from 25.0° to 66.0° and from 8.0° to 66.0° at the half-coverage and full-coverage acid monolayers due to esterification, respectively. This strongly confirms that the higher surface energy acid monolayers are indeed transferred to a lower surface energy ester monolayer⁶ due to esterification. Like the IRRAS data, these data also provide an opportunity to evaluate the rate of esterification as a function of reaction time and the kinetic information. The next section examines the kinetics.

Kinetic analysis

IRRAS

The extent of esterification, θ , of the half-coverage MUA/Au monolayer can be calculated from the IRRAS spectra with the following equations:

$$\theta = (A_{acid} - A_{ta})/A_{acid}$$
 (2)

where A_{ta} is the peak area of the hydrogen-bonded dimeric $\nu(C=0)$ at 1717 cm⁻¹ at a given time in the course of the reaction and A_{acid} is the peak area before the onset of esterification, or

$$\theta = A_{te}/A_{ester} \tag{3}$$

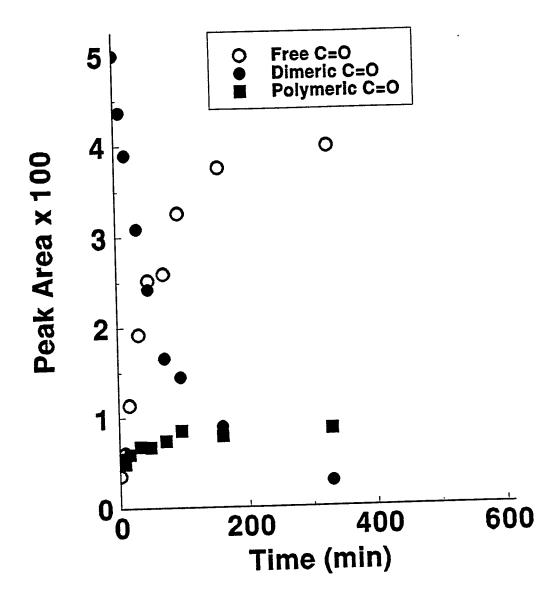


Figure 13. The peak area of the deconvoluted v(C=O) modes of a half-coverage MUA/Au Å as a function of immersion time in 20 % (v/v) HCl ethanol solution.

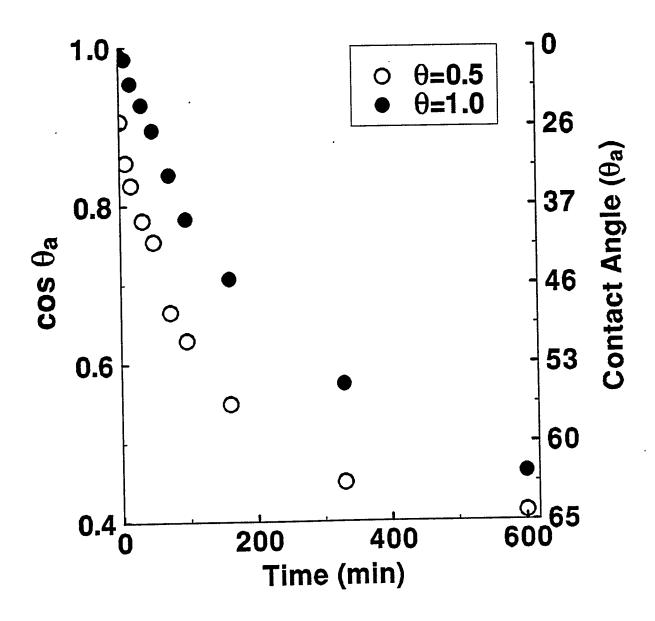


Figure 14. $\cos\theta_a$ of pH 1.0 HCl aqueous solution at a half-coverage MUA/Au monolayer and a full-coverage MUA/Au monolayer as a function of immersion time in 20 % (v/v) HCl ethanol. solution.

where A_{te} is the peak area of the non-hydrogen-bonded $\nu(C=O)$ at 1743 cm⁻¹ at a given time in the course of the reaction and A_{ester} is the peak area after esterification is complete.

Equation 2 follows the process via reactant loss, and Equation 3 via product formation. Consistent results from both equations indicates that the conservation of the orientation of the monolayer is valid.

The linear relationship between the value of $\ln (1-\theta)$ and reaction time suggests that the surface esterification reaction is a first-order reaction as shown in Figure 15.

Unfortunately, these two equations were not applied for full-coverage MUA/Au monolayers because the IRRAS peak intensities of both the hydrogen-bonded dimeric $\nu(C=O)$ peak at 1717 cm⁻¹ and the non-hydrogen-bonded $\nu(C=O)$ peak at 1743 cm⁻¹ are weak for full-coverage MUA/Au monolayers.

Contact angle measurements

The extent of esterification, θ , at MUA/Au monolayers were calculated from contact angle measurement data by using the Cassie equation as follow:

$$\cos \theta_{\rm a} = \theta \cos \theta_{\rm a,ester} + (1 - \theta) \cos \theta_{\rm a,acid} \tag{4}$$

where $\theta_{a,ester}$ and $\theta_{a,acid}$ are the contact angles at the 100 % ester monolayer and the 100 % acid monolayer, respectively.

The linear relationship between the $\ln{(1-\theta)}$ and reaction time from both IRRAS and contact angle measurement data concludes this surface esterification is a first-order reaction as shown in Figure 15. The contact angle measurement data have better reproducibility than the IRRAS data simply because the contact angle measurement data are the average of four repeat measurements while the IRRAS data are from a single measurement. The values of the error bar for the contact angle measurements and IRRAS are 1° and 5×10^{-5} A.U., respective. Since both techniques give the same result for the half-coverage MUA/Au, both can be used to measured the rate for the full-coverage MUA/Au monolayer as well. Only the contact angle

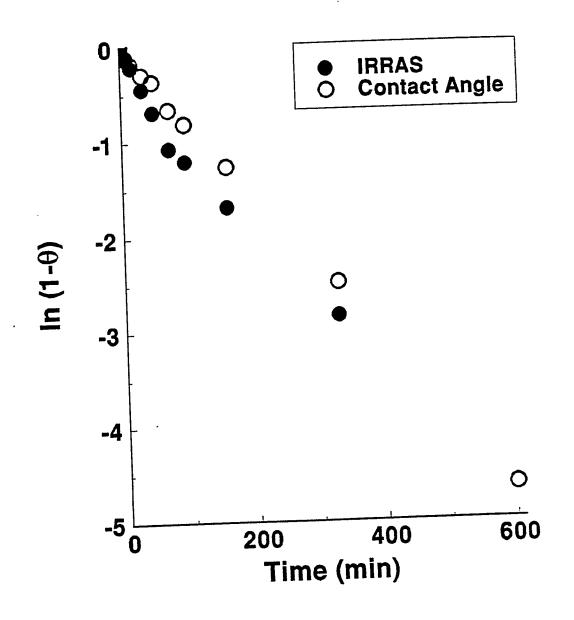


Figure 15. ln (1-θ) from IRRAS and contact angle measurement data for a half-coverage MUA/Au monolayers in 20 % (v/v) HCl ethanol solution as a function of immersion time.

measurements technique was used rather the IRRAS because the IRRAS peak intensities of both the hydrogen-bonded dimeric $\nu(C=O)$ peak and the non-hydrogen-bonded $\nu(C=O)$ peak are weak for full-coverage MUA/Au monolayers.

Since ethanol was used as the solvent, this first-order reaction most likely is a pseudo first-order reaction.

For a first-order reaction, the rate constants are equal to the slope of the $\ln (1-\theta)$ vs. reaction time as shown in Figure 16 and were calculated to be 7.62 x 10^{-3} sec⁻¹ and 3.87 x 10^{-3} sec⁻¹ for a packing density of 4.0 x 10^{-10} mol/cm² and 8.0 x 10^{-10} mol/cm² MUA/Au monolayers, respectively. This indicates that the rate constant is a reciprocal function of the packing density of the thiolate monolayer.

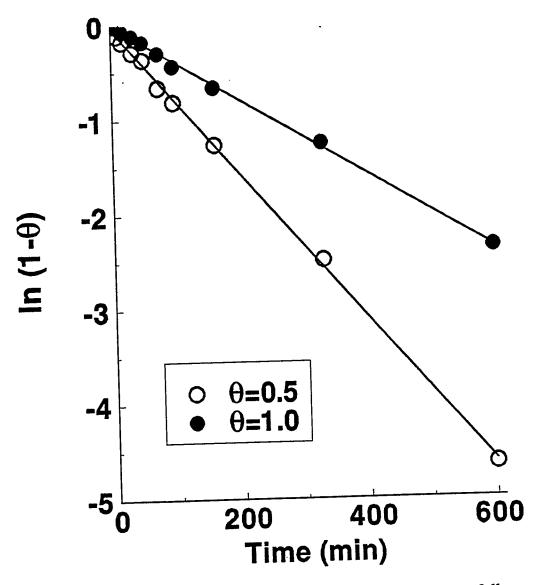


Figure 16. ln (1-θ) of a half-coverage MUA/Au monolayer and a full-coverage MUA/Au monolayer as a function of immersion time in 20 % (v/v) HCl ethanol solution.

CONCLUSIONS

The adsorption process of the MUA on Au is at least a two-step process with a very fast reaction rate for the first step followed by a very slow reaction rate for the second step.

Esterification transforms the hydrogen-bonded dimeric $\nu(C=O)$ peak at 1717 cm⁻¹ completely to the non-hydrogen-bonded $\nu(C=O)$ peak at 1743 cm⁻¹ without change the hydrogen-bonded polymeric $\nu(C=O)$ group due to the strong network-type hydrogen bond of the hydrogen-bonded polymeric $\nu(C=O)$ group. Esterification process stabilizes the polymethylene chain from liquid-like phase to crystalline-like phase.

Surface reaction can be monitored with the IRRAS or the contact angle measurement. This surface esterification is a pseudo-first-order reaction with a rate constant of $7.62 \times 10^{-3} \, \mathrm{sec^{-1}}$ and $3.87 \times 10^{-3} \, \mathrm{sec^{-1}}$ for a packing density of $4.0 \times 10^{-10} \, \mathrm{mol/cm^2}$ and $8.0 \times 10^{-10} \, \mathrm{mol/cm^2}$ MUA/Au monolayers, respectively. The rate constant of surface esterification is a reciprocal function of the packing density of the thiolate monolayer.

REFERENCES

- (1) Bain, C. D.; Whitesides; G. M. Angew. Chem., Int. Ed. Engl. 1989, 28, 506 and references therein.
- (2) Whitesides, G. M.; Laibinis, P. E. Langmuir 1990, 6, 87 and references therein.
- (3) Ulman, A. An Introduction to Ultra-Thin Organic Films From Langmuir-Blodgett to Self-Assembly; Academic Press: San Diego, 1991.
- (4) Nuzzo, R. G.; Dubois, L. H.; Allara, D. L. J. Am. Chem. Soc. 1990, 112, 558.
- (5) Snyder, R. G.; Schachtschneider, J. H. Spectrochim. Acta 1963, 19, 85.
- (6) Holmes-Farley, S. R.; Whitesides, G. M. Langmuir 1987, 3, 60.
- (7) Widrig, C. A.; Alves, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2805.
- (8) Widrig, C. A.; Chung, C.; Porter, M. D. J. Electroanal. Chem. 1991, 310, 335.
- (9) Azzam, G. A.; Bashara, N. M. Ellipsometry and Polarized Light; North-Holland Publishing: Amsterdam, 1977.
- (10) Handbook of Chemistry and Physics; Weast, R. C., Ed.; Chemical Rubber Cor.: Boca Raton, FL, 1981.
- (11) Porter, M. D.; Bright, T. B.; Allara, D. L.; Chidsey, C. E. D. J. Am. Chem. Soc. 1987, 109, 3559.
- (12) Finklea, H. O.; Avery, S.; Lynch, M.; Furtsch, T. Langmuir 1987, 3, 409.
- (13) Troughton, E. B.; Bain, C. D.; Whitesides, G. M.; Nuzzo, R. G.; Allara, D. L.; Porter, M. D. *Langmuir* 1988, 4, 365.
- (14) Tao, Y.-T.; Lee, M.-T.; Chang, S.-C. J. Am. Chem. Soc. 1993, 115, 9547.
- (15) Bain, C. D.; Whitesides, G. M. J. Am. Chem. Soc. 1989, 111, 7164.
- (16) Ulman, A. J. Mater, Educ. 1989, 11, 205.
- (17) Stole, S. M.; Porter, M. D. Appl. Spectrosc. 1990, 44, 1418.
- (18) Walczak, M. M.; Chung, C.; Stole, S. M.; Widrig, C. A.; Porter, M. D. J. Am. Chem. Soc. 1991, 113, 2370.
- (19) Wilson, E. B., Jr.; Decius, J. C.; Cross, P. C. Molecular Vibrations; McGraw-Hill: New York, 1955, pp 285.

- (20) Bain, C. D.; Troughton, E. B.; Tao, Y-T.; Evall, J.; Wihitesides, G. M.; Nuzzo, R. G. J. Am. Chem. Soc. 1989, 111, 321.
- (21) Shimazu, K.; Yagi, I.; Uosaki, K. Langmuir 1992, 8, 1385.
- (22) Buck, M.; Eisert, F.; Fischer, J. Grunze, M.; Träger, F. Appl. Phys. 1991, A53, 552.
- (23) Buck, M.; Eisert, F.; Fischer, J. Grunze, M.; Träger, F. J. Vac. Sci. Technol. 1992, A10, 926.
- (24) Thomas, R. C.; Sun, L.; Crooks, R. M.; Ricco, A. J. Langmuir 1991, 7, 620.
- (25) Snyder, R. G.; Strauss, H. L.; Elliger, C. A. J. Phys. Chem. 1982, 86, 5145.
- (26) Gramstad, T.; Fuglevik, J. Spectrochim. Acta 1965, 21, 343.
- (27) Pullin, J. A.; Werner, J. Spectrochim. Acta 1965, 21, 1257.
- (28) Figeys, H. P.; Nasielski, J. Spectrochim. Acta 1967, 23A, 465.
- (29) Bellamy, L. J.; Rogash, P. E. Spectrochim. Acta 1960, 16, 30.
- (30) Collins, A. J.; Morgan, K. J. J. Chem. Soc. 1963, 3437.
- (31) Brooks, C. J. W. J. Chem. Soc. 1960, 661.
- (32) Paper III of this dissertation.

GENERAL SUMMARY AND PROSPECTS

Deprotonation causes the heterogeneous electron transfer rate of ferricyanide at phenylcarboxylic acids and pyridylcarboxylic acids coated Pt electrodes to decrease by 0 % to 40 % and 85%, respectively, and decreases the potential at reaction plane of the phenylcarboxylic acid and the pyridylcarboxylic acid monolayers on platinum by 0 to 5 mV and ~20 mV, respectively. The observed $\Delta \phi_r$ values due to deprotonation for phenylpropanoic acid/Pt and phenylbutanoic acid/Pt are only 0.7 % of the theoretical values and that for isonicotinic acid/Pt and nicotinic acid/Pt are only 1.2 %. No change in ϕ_r was observed for benzoic acid/Pt and phenylacetic acid/Pt electrodes. Pyridylcarboxylic acids becomes weaker acids upon immobilization onto a Pt surface primarily due to hydrogen bonding.

At very low coverage, all the polymethylene chains of thiolate monolayers lay on gold and form the C-H···Au agostic bond at 2903 cm⁻¹ with $\nu_a(\text{CH}_2)$ and $\nu_s(\text{CH}_2)$ dipole perpendicular and parallel to the Au surface, respectively. At increasing coverage, the polymethylene chain must turn the chain axis in order to stand up to accommodate more thiol molecules to be adsorbed on Au surface so that the amount of the agostic $\nu_a(\text{CH}_2)$ decreases and disappears at high coverage.

The hydrogen-bonded dimeric C=O is the dominated C=O peak of carboxylic acid-terminate thiloate monolayers on Au at low coverage. With increasing coverage, the amount of the hydrogen-bonded dimeric C=O decreases while the amount of both the non-hydrogen-bonded and the hydrogen-bonded polymeric C=O groups increase, meanwhile the surface wetting property increases remarkably with increasing coverage.

The pK_a and the titration transition range of the non-hydrogen-bonded C=O, the hydrogen-bonded dimeric C=O, and the hydrogen-

bonded polymeric C=O groups of carboxylic acid-terminate thiloate monolayers on Au are determined from IRRAS data to be \sim 6, \sim 8, and \sim 10, and 3, 7, and 3 pH units, respectively. The effective pK_a of the non-hydrogen-bonded C=O and the hydrogen-bonded dimeric C=O groups increases from 5 up to \sim 10 with increasing the extent of deprotonation.

Esterification of the 11-mercaptoundecanoic acid (MUA) monolayer on Au converts the $\nu(C=O)$ at 1717 cm⁻¹ of the carboxylic acid to the $\nu(C=O)$ at 1743 cm⁻¹ of ester without change the hydrogen-bonded polymeric $\nu(C=O)$ at 1680 cm⁻¹ at all, meanwhile the contact angle at this monolayer increases from ~10° to 62°. Both IRRAS and contact angle measurements can be used to measure the reaction rate of this esterification process. The rate constant is proportional reciprocally to the packing density of the monolayer. For example the rate constants are $7.62 \times 10^{-3} \, \text{sec}^{-1}$ and $3.87 \times 10^{-3} \, \text{sec}^{-1}$ for a packing density of 4.0 x10 $^{-10} \, \text{mol/cm}^2$ and $8.0 \times 10^{-10} \, \text{mol/cm}^2$ MUA/Au monolayers, respectively.

The use of ion-pair electrolytes might be able to increase the change in the potential at the reaction plane due to less screening effect than conventional electrolytes. The use of different metal surfaces should form agostic bonds also with methylene groups. The reaction rate of different surface reactions can be measured in the same fashion using IRRAS and/or contact angle measurements.

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