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Author(s): Robinson, Aaron Jon

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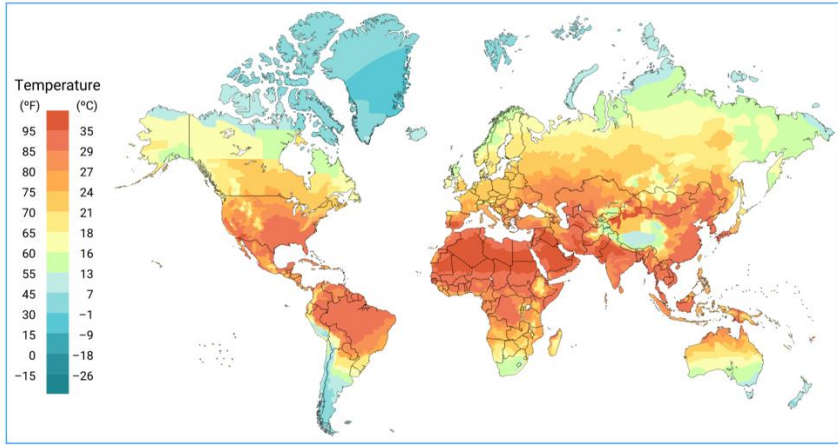
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Using fungi to evaluate threats associated with climate change

Aaron Robinson
Scientist II B-GEN

September 18, 2024

Fungi and Climate Change



The future of fungi: threats and opportunities

Nicola T. Case ¹, Judith Berman ², David S. Blehert ³, Robert A. Cramer ⁴, Christina Cuomo ⁵, Cameron R. Currie ⁶, Iuliana V. Ene ⁷, Matthew C. Fisher ⁸, Lillian K. Fritz-Laylin ⁹, Aleeza C. Gerstein ¹⁰, N. Louise Glass ¹¹, Neil A. R. Gow ¹², Sarah J. Gurr ¹², Chris Todd Hittinger ¹³, Tobias M. Hohl ¹⁴, Iliyan D. Iliev ¹⁵, Timothy Y. James ¹⁶, Hailing Jin ¹⁷, Bruce S. Klein ^{18,19,20}, James W. Kronstad ²¹, Jeffrey M. Lorch ³, Victoria McGovern ²², Aaron P. Mitchell ²³, Julia A. Segre ²⁴, Rebecca S. Shapiro ²⁵, Donald C. Sheppard ²⁶, Anita Sil ²⁷, Jason E. Stajich ¹⁷, Eva E. Stukenbrock ^{28,29}, John W. Taylor ³⁰, Dawn Thompson ³¹, Gerard D. Wright ³², Joseph Heitman ³³, Leah E. Cowen ^{1,*}

Global Health

- 1.5 million deaths annually
- ~12 billion annual economic burden
- No vaccines, poor diagnostics, resistance

Global Security

- Agricultural losses -10 to 20% (\$100 to \$200 billion)
- Expanding geographic range and emerging pathogens
- Antifungal resistance

Ecological Stability

- Species extinction and collapse (land and sea)
- Pathogens in the environment

Motivations for studying thermophilic fungi

- Rare unifying trait among fungal pathogens
 - Survive mammalian body temperatures
 - Therapeutic design
 - Agnostic detection of emerging pathogens
- Modeling impacts of climate change
 - Evolutionary trade-offs
 - Effects on metabolism
 - Susceptible populations
- Biotechnology
 - Biomass transformation
 - Protein engineering

Thermophilic: optimal growth above 45°C

Thermotolerant: can grow above 35°C, but not necessarily optimal

Mesophilic: optimal growth at 20 – 34°C

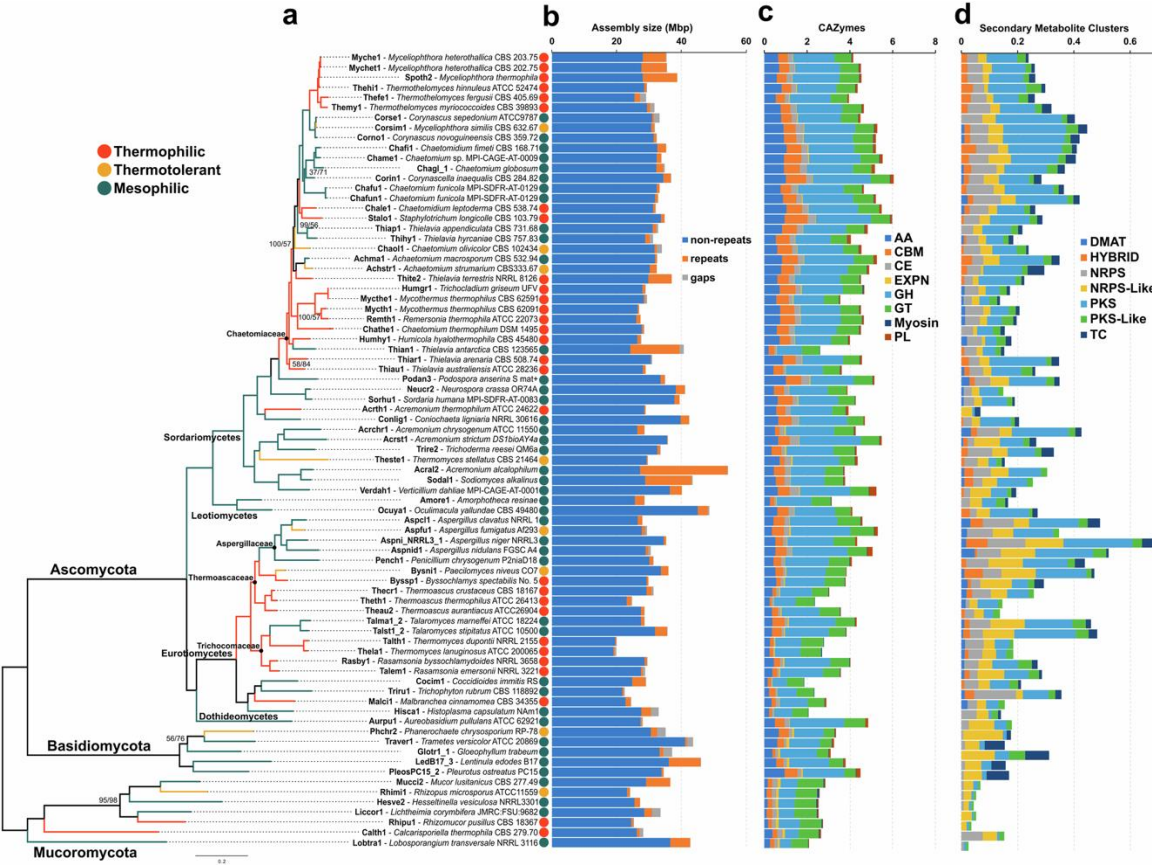
Comparative genomic analysis of thermophilic fungi reveals convergent evolutionary adaptations and gene losses

[Andrei S. Steindorff](#), [Maria Victoria Aguilar-Pontes](#), [Aaron J. Robinson](#), [Bill Andreopoulos](#), [Kurt LaButti](#), [Alan Kuo](#), [Stephen Mondo](#), [Robert Riley](#), [Robert Otilar](#), [Sajeet Haridas](#), [Anna Lipzen](#), [Jane Grimwood](#), [Jeremy Schmutz](#), [Alicia Clum](#), [Ian D. Reid](#), [Marie-Claude Moisan](#), [Gregory Butler](#), [Thi Truc Minh Nguyen](#), [Ken Dewar](#), [Gavin Conant](#), [Elodie Drula](#), [Bernard Henrissat](#), [Colleen Hansel](#), [Steven Singer](#), ... [Igor V. Grigoriev](#) 

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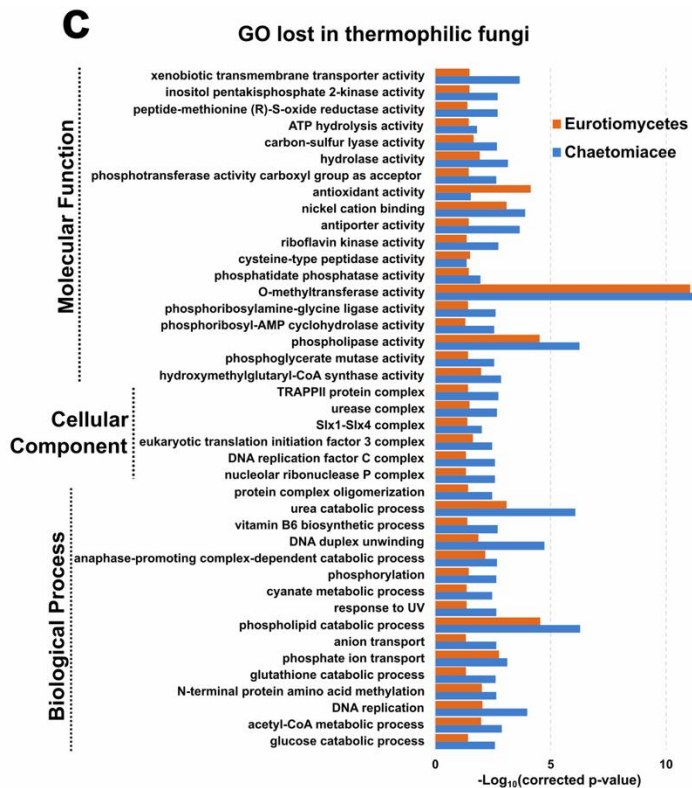
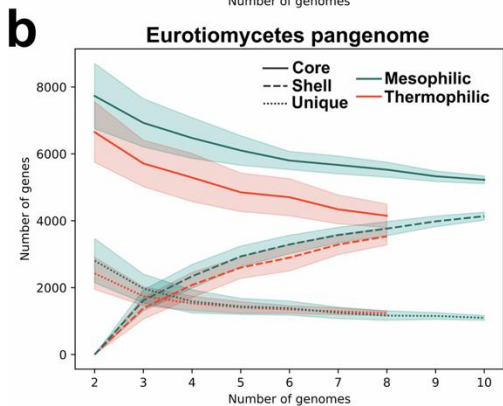
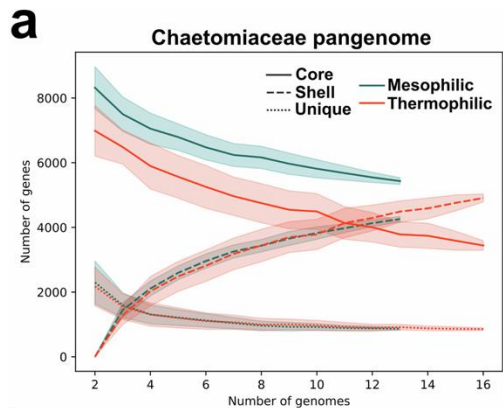
Comparative Genomics



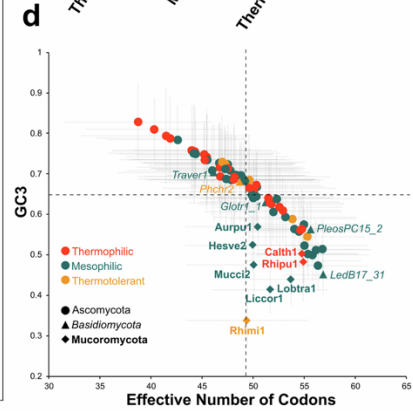
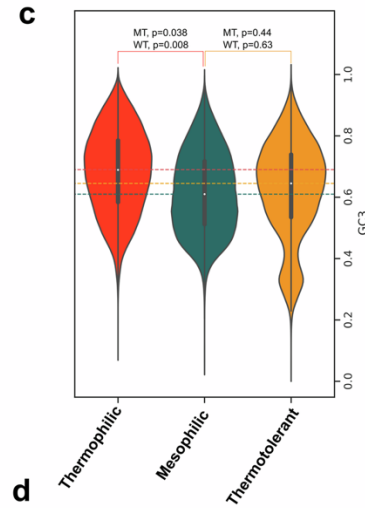
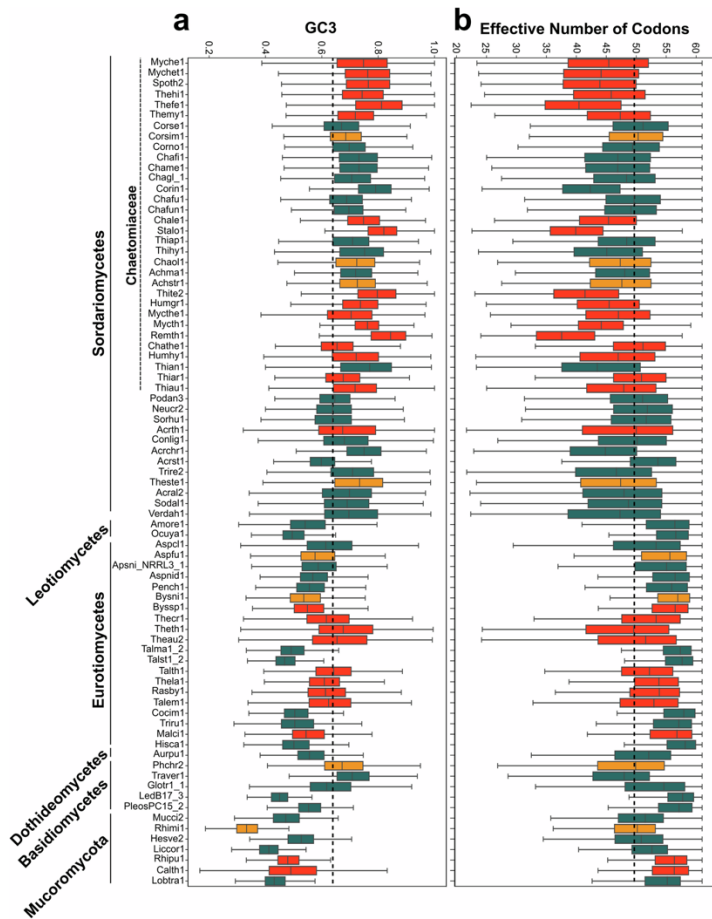
29 thermophiles
8 thermotolerant
42 mesophiles

Ancestral states thermophilic
(likelihood reconstruction model)

Thermophilic genomes generally
have smaller size (38%)

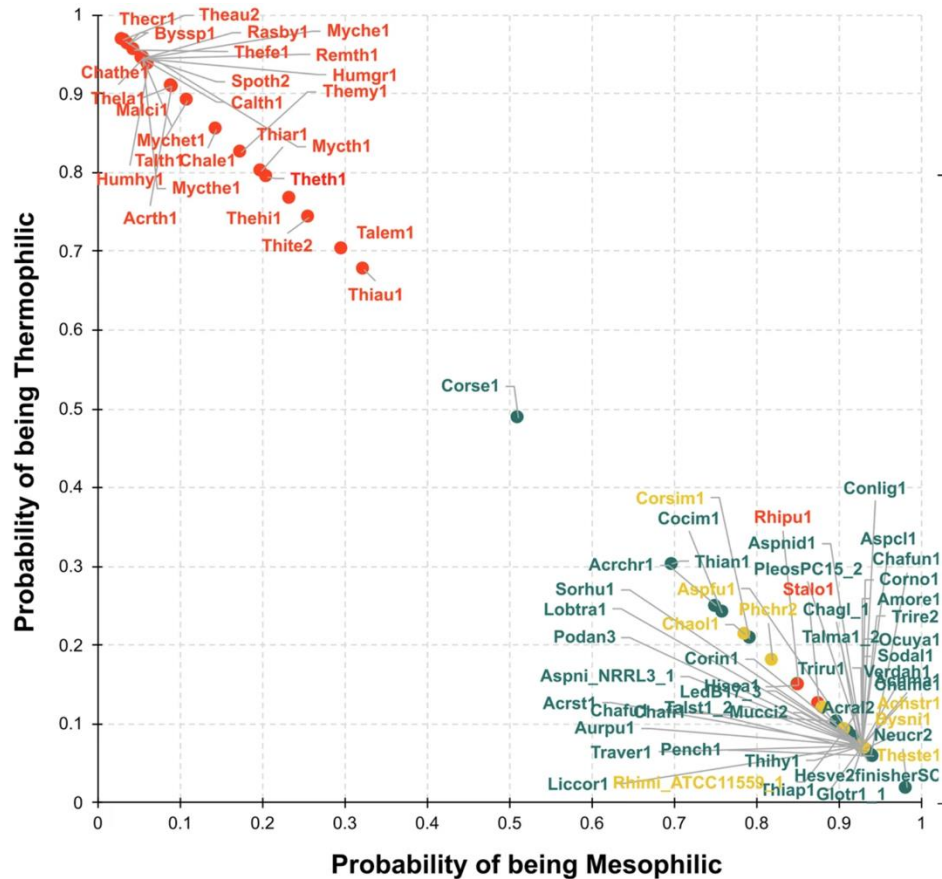


Too many differences...need to constrain the comparisons



Limited examinations showed potential signatures based on GC content of third base of codon (GC3) and ENC....

Again, trends driven by phylogenetic relationships



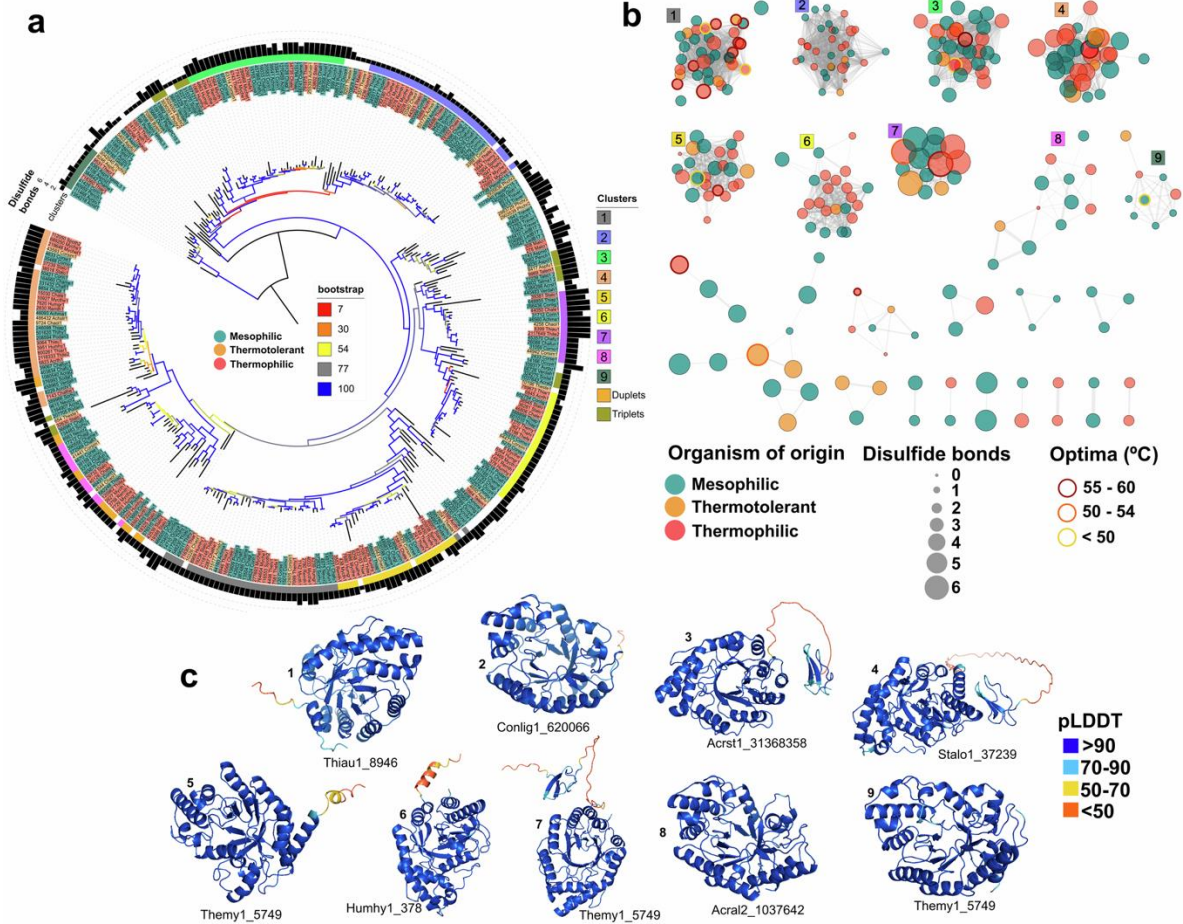
Support Vector Machine (SVM) to identify correlations between phenotype and gene content

20 thermophilic and 30 mesophilic genomes used for training

100 Orthogroups produced an accuracy of 70%

Identified 8 clusters with 94% accuracy

Thermotolerant consistently classified as mesophilic



GH10 endoxylanase examined with Foldseek (structural similarity) and Alphafold2

Intermixing of sequences from thermophiles and mesophiles – some long branches (hi evo rate)

Optimal temperature data (heterologous expression) mapped on top

High optima in same cluster

Summary

- Identified some genomic and evolutionary traits of thermophilic fungi
- Unfortunately – identification of things missing from the genome, rather than contributing to thermotolerance
- More detailed examinations and finer scales would be helpful...

Genetics of thermotolerance in a model fungus

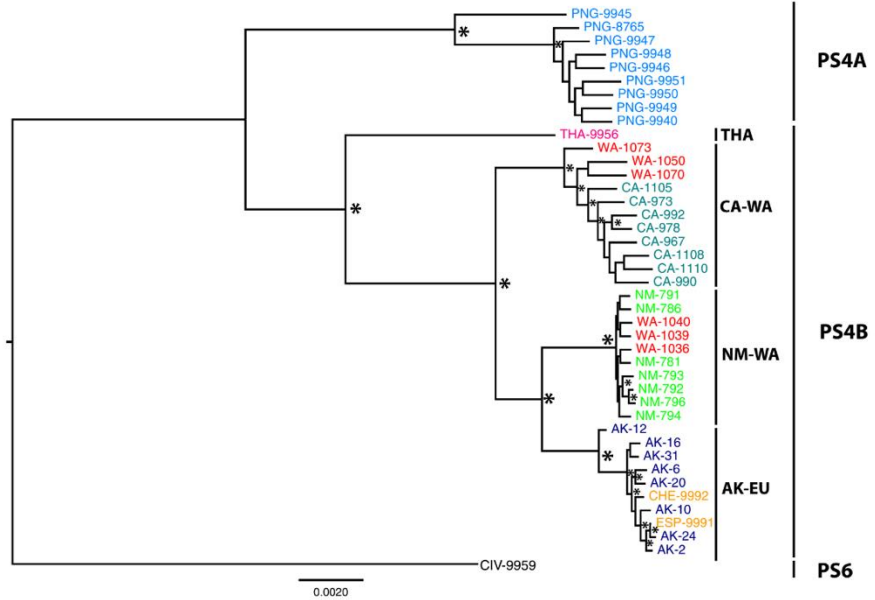
Neurospora discreta

- Found following forest fires
- Phylogenomic studies show evidence of endemic populations
- Lots of genetic resources

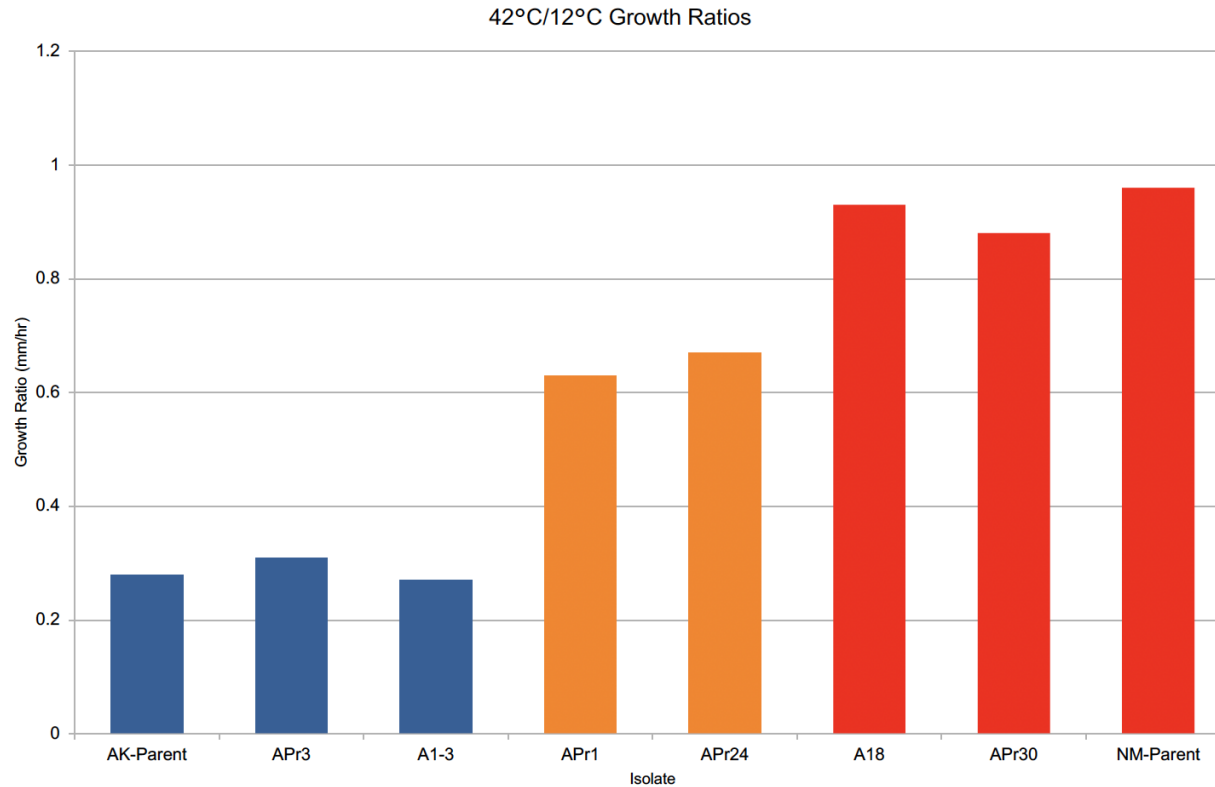


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(A)

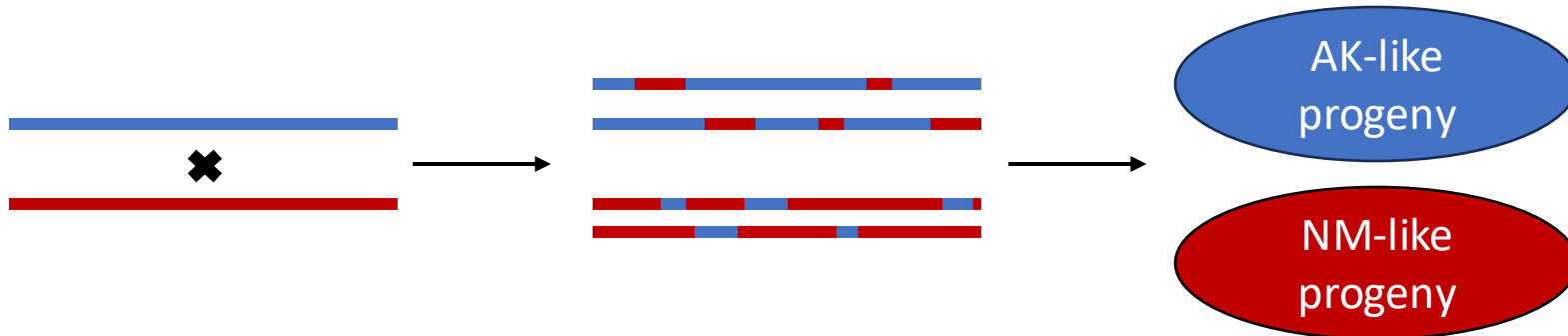


Thermotolerance is heritable and controlled by multiple loci



Identifying genomic regions contributing to thermotolerance

Isolate	Mating genotype
NM - 799	MAT A
NM - 798	MAT a
AK - 852	MAT A
AK - 853	MAT a
AK1	Unknown
AK10	Unknown



Parental Genome Comparison

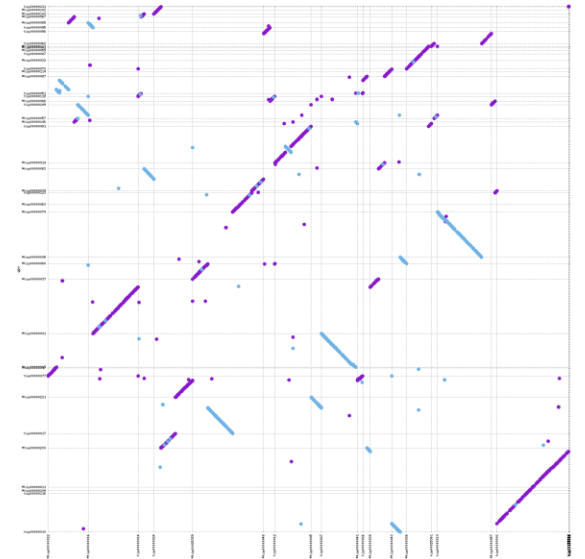
Isolate	Genome Size	BUSCO
NM – FGSC 10342	37.5 Mb	C:94.9%[S:94.8%,D:0.1%],F:1.4%,M:3.7%,n:3817
AK – FGSC 10343	36.7 Mb	C:96.4%[S:96.2%,D:0.2%],F:1.3%,M:2.3%,n:3817



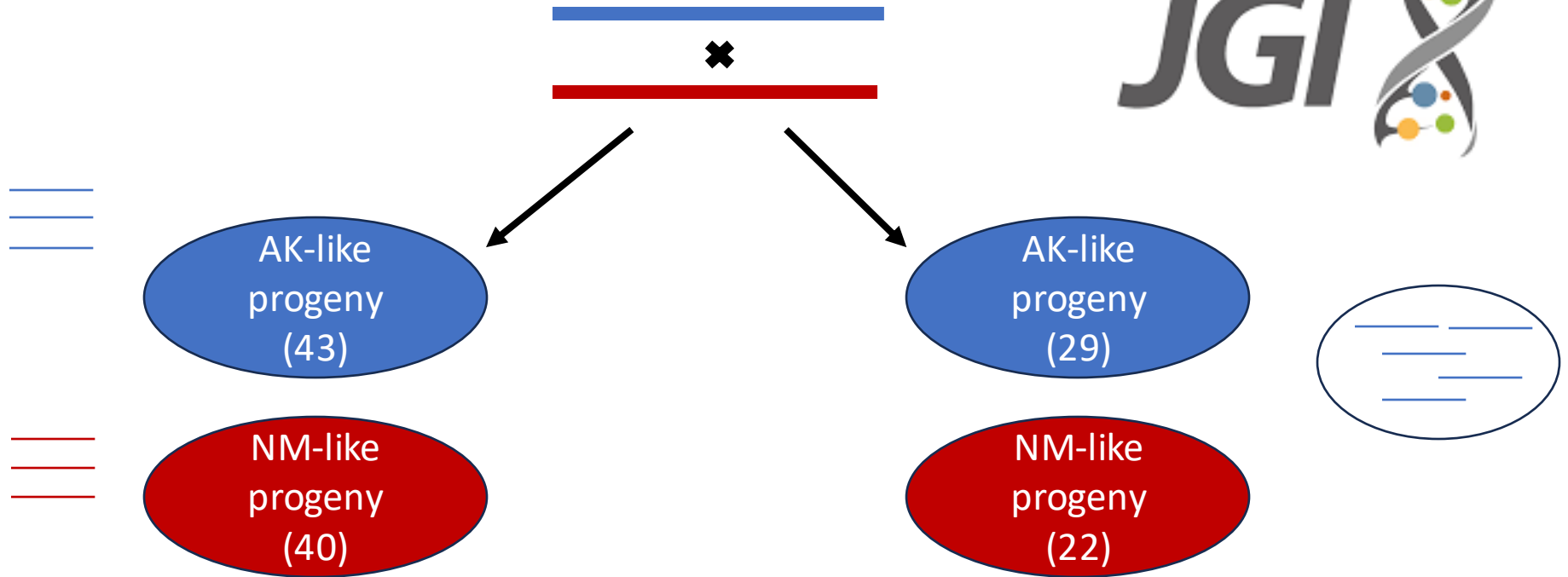
NM Predicted Gene # - 8,397

AK Predicted Gene # - 8,520

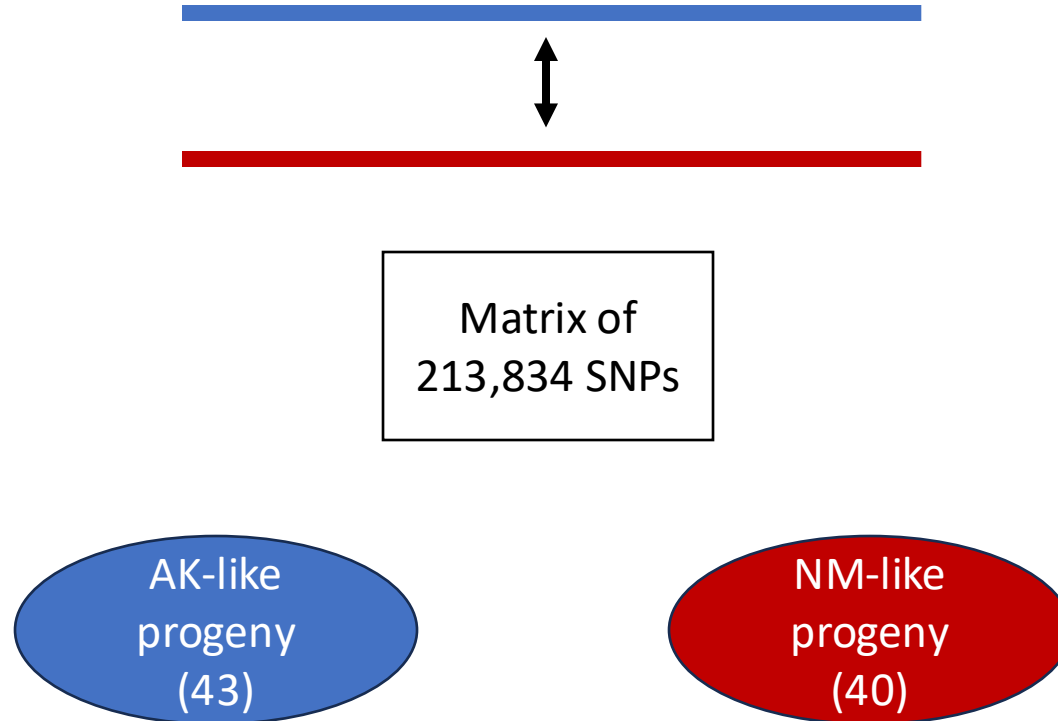
Number of genes	Number of genes in orthogroups	Number of unassigned genes
16,917	16,522 (97.7%)	395 (2.3%)



Genome-based approaches

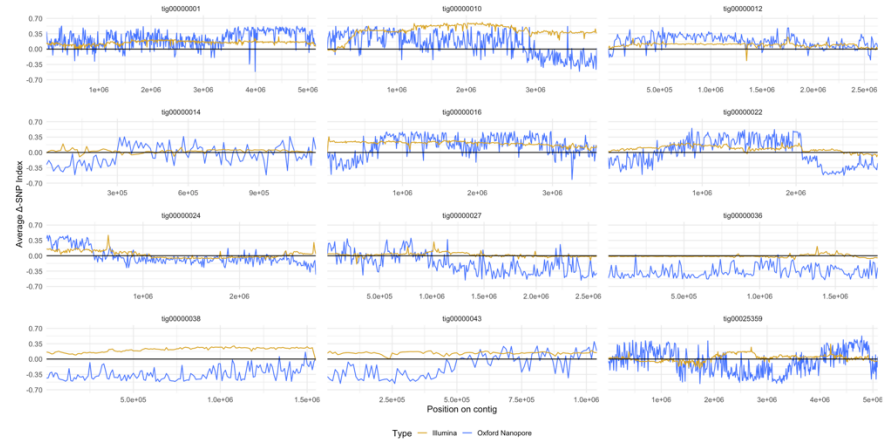
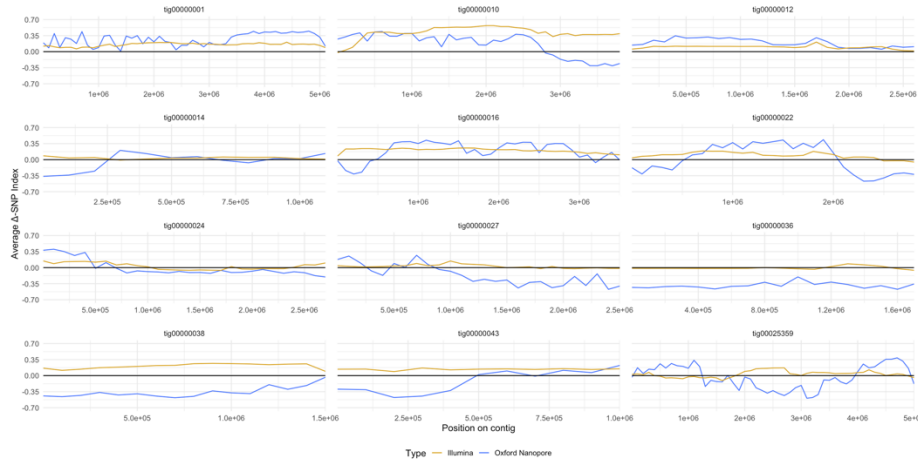


Characterizing Genotypes



Largest difference
between phenotypes

Relationships between genotype and phenotype



Identification of genes associated with thermotolerance

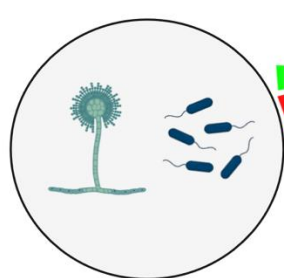
- 1 Mb region with significant correlations between genotype and phenotype
- 83 putative genes – mix of known and unknown function
- Closest blastp matches to other thermophiles
- Highest significance in NM groups – phenotyping needs improvement in AK groups
- 91 missense mutations, 64 synonymous, 11 other (intron, intergenic, splice)

Lots of intrinsically disordered proteins – next challenge for AlphaFold team!

2283881 C G 0 0 0.41379310344827586 0.5862068965517241

Remote Sensing of Fungi in Complex Environments

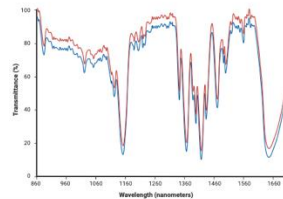
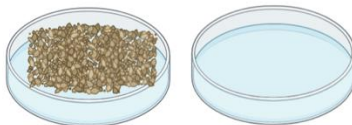
Laboratory



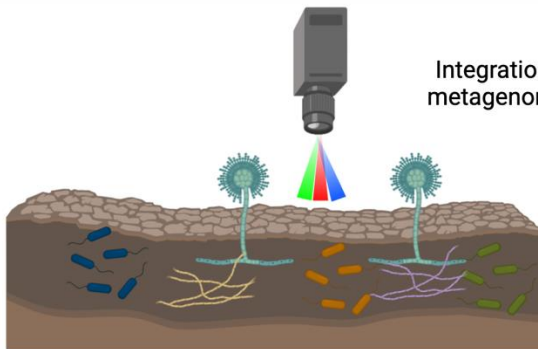
Spectral profiling of bacterial and fungal spores and cells

Identify spectral signatures

Low complexity backgrounds

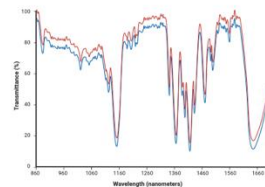


Field



High complexity background

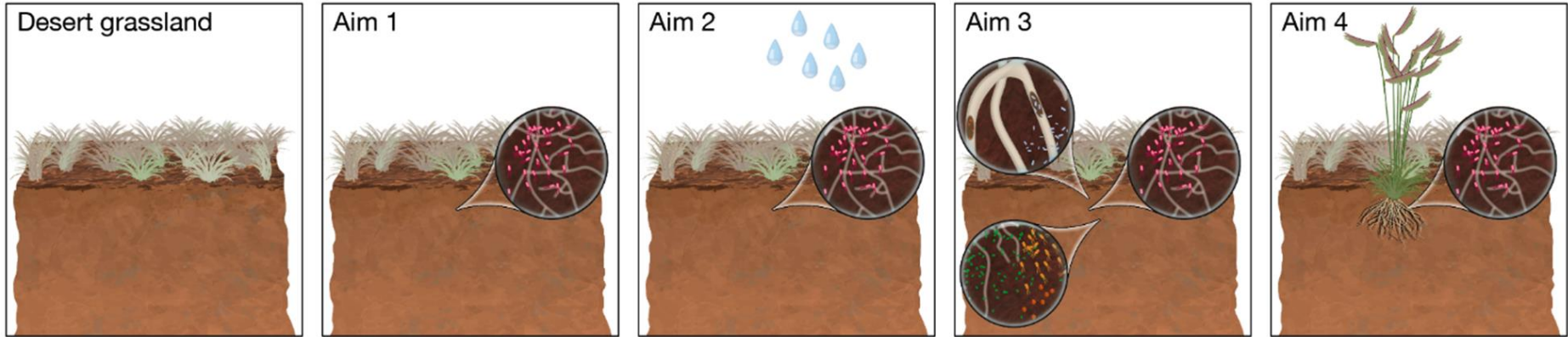
Integration of metagenomics



In final stages of approval for BSL-2 lab (BLOOM) for working with pathogenic fungi and culturing from soils and wastewater

Integrating other forms of characterization

Fungal Interactions



Bioinformatic Resources for BFI

- Had any interactions been previously described between bacteria and fungi from our system?
- Lack of centralized resources and knowledgebases for BFI research...so we built our own!




<https://sfa-bfi.edgebioinformatics.org/>



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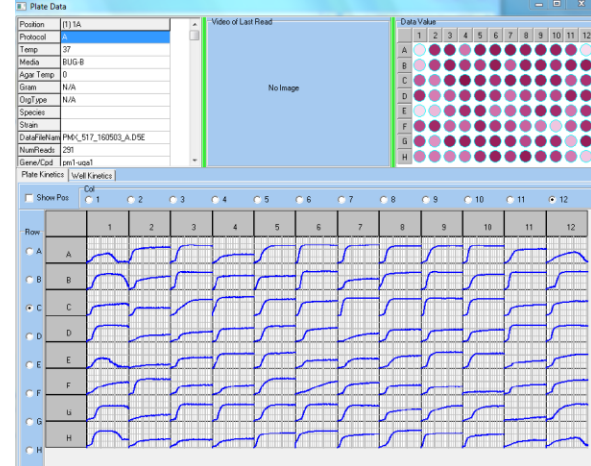
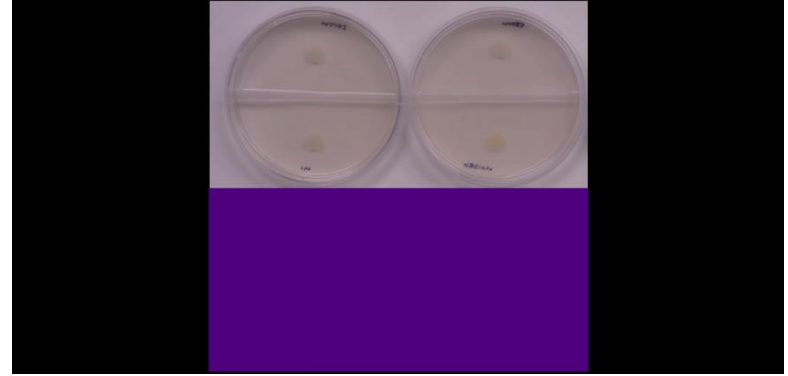


A centralized resource for bacterial–
fungal interactions research

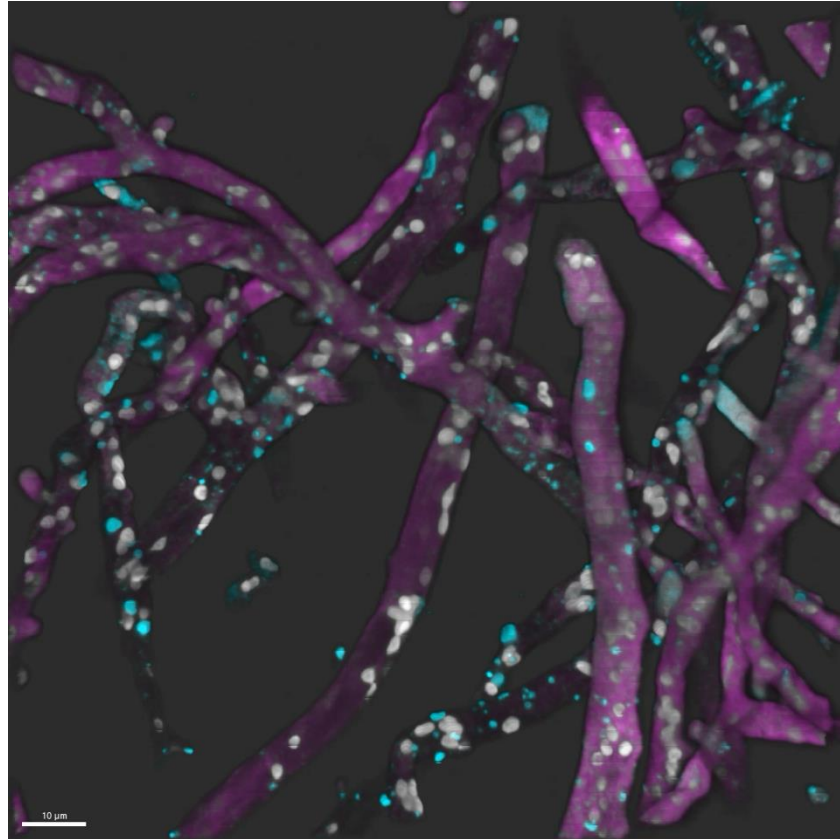
[Aaron Robinson](#)¹  , [Michal Babinski](#)¹, [Yan Xu](#), [Julia Kelliher](#), [Reid Longley](#),
[Patrick Chain](#)  

Developing methods to efficiently identify and characterize interactions

- Mix of imaging, metabolic, and omics to characterize how fungi interact with bacteria and other organisms
- **Environmental** – understand community dynamics toward more accurate models (abiotic and biotic)
- **Security** – Signatures of pathogens and potential countermeasures
- Interactions to **enhance** or **deter**



Hyphal networks in soil and health



Acknowledgements



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Dean Morales
La Verne Gallegos-Graves
Buck Hanson
Reid Longley
Leah Johnson
Kelli Feeser
Justine Macalindong
Jim Brunner
Kaelan Prime

Remote Sensing DI

Patrick Chain
Nicholas Parra-Vasquez
Jeffrey Klein
Daniel Jacobsen

