



Experimental and Modeling Informed Data Analytics Platform to Identify Viral Features Indicative of Pandemic Potential



PRESENTED BY

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Emerging infectious disease pose an imminent threat to human health, economic and national security



“Pandemics are for the most part disease outbreaks that become widespread as a result of the spread of human-to-human infection. Beyond the debilitating, sometimes fatal, consequences for those directly affected, pandemics have a range of negative social, economic and political consequences. These tend to be greater where the pandemic is a novel pathogen, has a high mortality and/or hospitalization rate and is easily spread. According to Lee Jong-wook, former Director-General of the World Health Organization (WHO), pandemics do not respect international borders.

Therefore, they have the potential to weaken many societies, political systems and economies simultaneously.”

United Nations Chronical, 2008 (<https://www.un.org/en/chronicle/article/national-security-and-pandemics>)

1918: Influenza

2002: West Nile Virus

2003: SARS

2005: Bird flu

2009: Swine flu

2014: Ebola

2016: Zika virus

2019 - : Covid 19

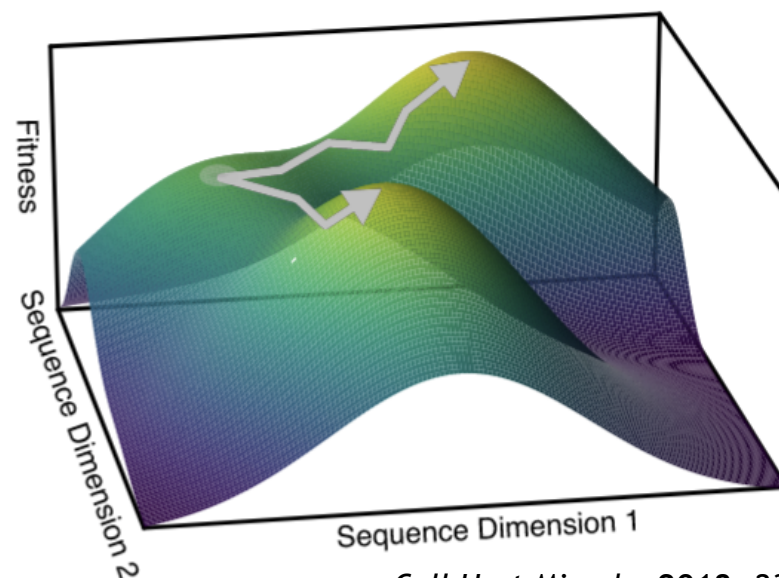
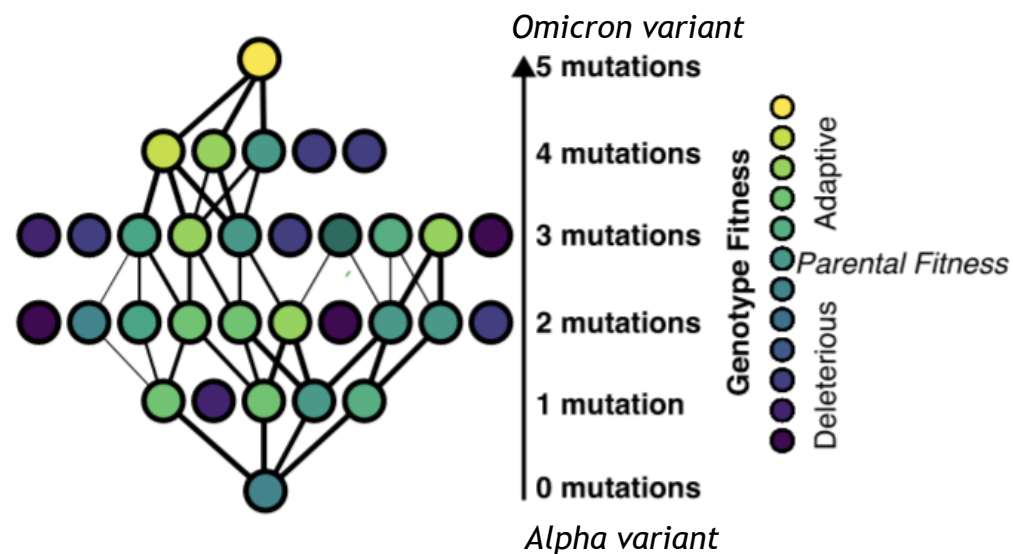
The current Covid-19 pandemic highlights the devastating potential of new and emerging infectious diseases.

And the need to develop methods to predict the pandemic potential of emerging pathogens.

We are trying to understand and predict viral evolution



Evolution occurs along a genotype/phenotype – fitness landscape



Cell Host Microbe 2018, 23 (4), 435-446.

Simple 5 mutation site, 2 possible mutations network

- 2^5 (32) genotype combinations
- Connected by single mutations
- $\text{Prob}\{\text{mutation}\} \sim \text{line width}$
- Network complexity \sim Interactions among mutations (epistasis)

Populations explore the topography of the fitness landscape

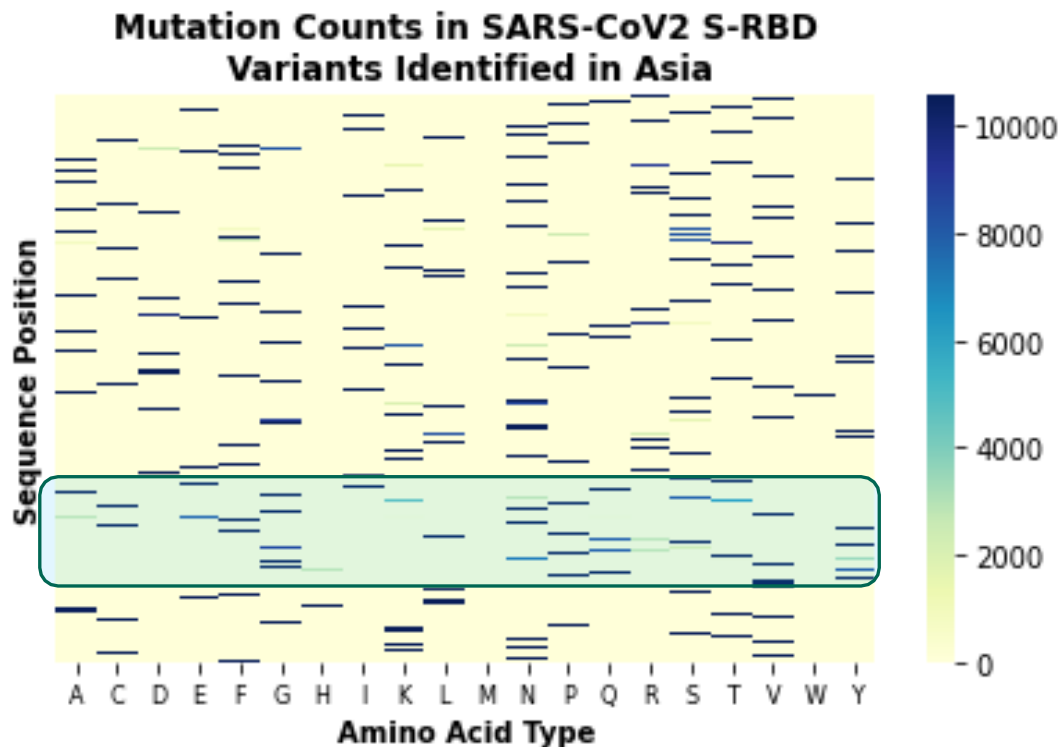
- By acquiring mutations
- Natural selection drives populations toward local maxima
- Swarms of variants simultaneously exploring the fitness landscape

We are essentially trying to identify (predict) the local maxima on the fitness landscape

Why is it so challenging?



Sequence alignment of 10,570 SARS-CoV2-S-RBD variants identified in Asia



| | | | | | |
|--|--------------------|-------------------|-------------------|--------------------|--------------------|
| Consensus Conservation Consensus RBD asia: reference sequence | 1 RVQPTESIVR | 11 FPNITNLCPF | 21 GEVFNATRFA | 31 SVYAWNRRKI | 41 SNCVADYSVL |
| Consensus Conservation Consensus RBD asia: reference sequence | 51 YNSASFSTFK | 61 CYGVSP TKLN | 71 DLCFTNVYAD | 81 SFVIRGDEV R | 91 QIAPGQTGKI |
| Consensus Conservation Consensus RBD asia: reference sequence | 101 ADYNYKLPDD | 111 FTGCVIAWNS | 121 NNLDSKVGGN | 131 YNYLYRLFRK | 141 SNLKPFERDI |
| Consensus Conservation Consensus RBD asia: reference sequence | 151 STEIYQAGS x | 161 PCNGVEGFNC | 171 YFPLQSYGFQ | 181 PT x GVGYPY | 191 RVVVL SFELL |
| Consensus Conservation Consensus RBD asia: reference sequence | 201 HAPATVCGPK | 211 KSTNLVKNKC | 221 VNF | | |

Most sequence positions are relatively invariant

Consensus sequence generated from alignment of the 10,570 variants identified in Asia

- Conservation does reduce the search space or space of possible variants.
- Space of variants is still huge $\sim 20^n$, where n is the number of possible mutation sites.
- Highly fit variants are very rare and predicting the pathway along the fitness landscape difficult.
- Fitness landscape is very high dimension with multiple objective functions.

Using Published Data



Starr, Tyler N., et al. "Deep mutational scanning of SARS-CoV-2 receptor binding domain reveals constraints on folding and ACE2 binding." *Cell* 182.5 (2020): 1295-1310.

- Receptor binding domain (RBD) expression on cell surface of yeast
 - RBD consists of spike amino acids 331-531 (201 total)
- PCR-based mutagenesis introduces mutations

Binding

- Titration curves for 16 ACE2 concentrations
- Binding Endpoint: Change in $\log_{10}(K_a)$ from wildtype (K_a is inverse dissociation constant)

Expression

- Fluorescence-activated cell sorting
- Expression Endpoint: Change in mean fluorescence intensity from wildtype

Global epistasis models predict effects on expression/binding for single mutations



Greaney, Allison J., et al. "Comprehensive mapping of mutations in the SARS-CoV-2 receptor-binding domain that affect recognition by polyclonal human plasma antibodies." *Cell host & microbe* 29.3 (2021): 463-476.

- Similar experiment to before
- 10 neutralizing antibodies
 - 9 from SARS-CoV-2 patients, 1 from SARS-CoV-1
- Also used to build global epistasis models

Endpoint: $\log_{10}(\text{Escape Fraction})$

- Escape Fraction $E_v = F * \frac{n_v^{post}/n_v^{pre}}{N^{post}/N^{pre}}$
 - F is total fraction of library that escapes antibody binding
 - n_v^{pre}, n_v^{post} are the counts for variant v before and after enriching for antibody escape plus a pseudo-count .5
 - $N^{pre} = \sum n_v^{pre}, N^{post} = \sum n_v^{post}$



Binding

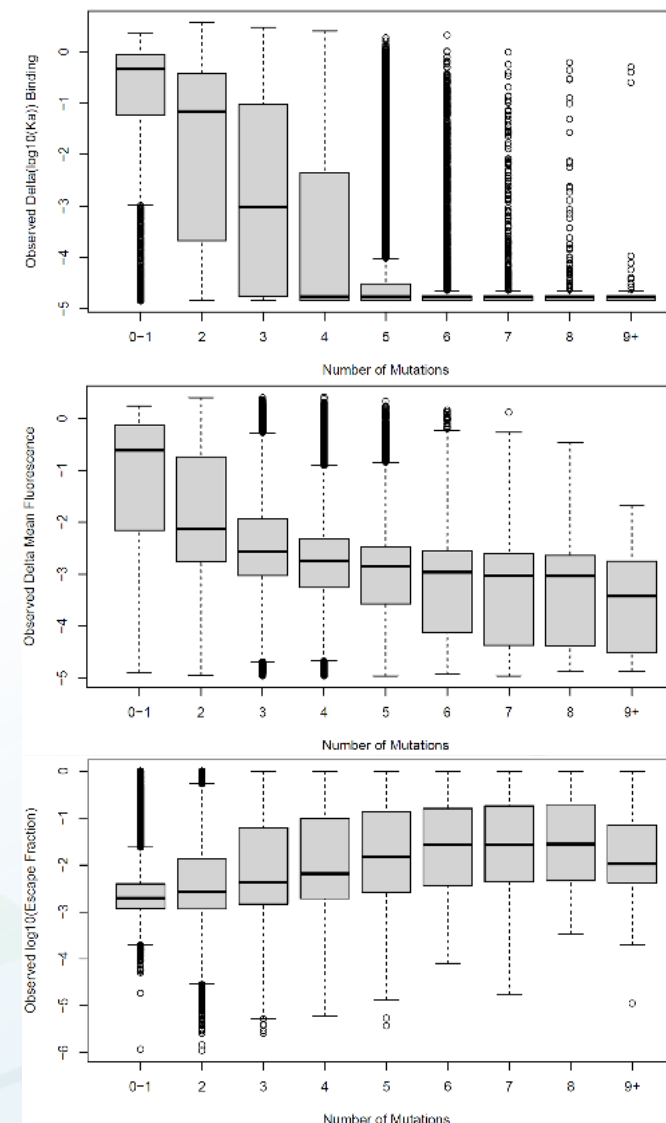
- 146,437 observations of 105,526 unique variants
- 0-10 mutations per variant with median 3
- 3,802 unique mutations represented (out of $19 \times 201 = 3,819$ possible)
 - No deletions/insertions

Expression

- 177,759 observations of 135,386 unique variants
- 0-12 mutations per variant with median 3
- 4,002 unique mutations represented (of 4,020 possible)
 - Deletions included, but no insertions

Antibody Escape

- 714,797 observations of 50,795 unique variants
- 10 antibodies with 66,403 - 79,126 observations each
- 0-10 mutations per variant with median 2
- 3,954 unique mutations represented (of 4,020 possible)
 - Deletions included, but no insertions



One Variant per row

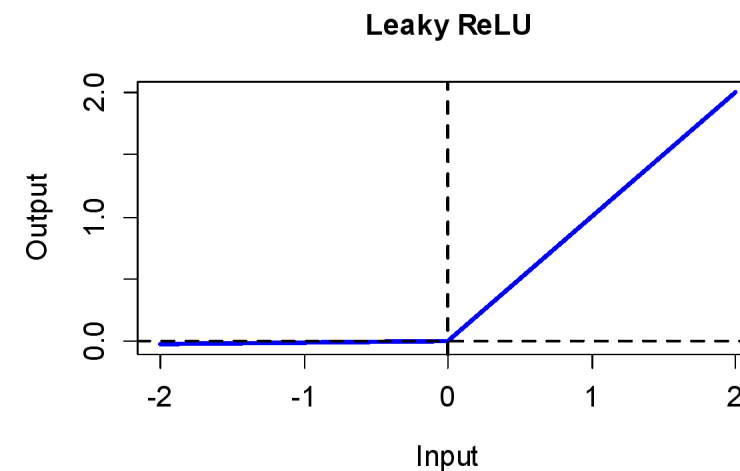
Features

- Individual mutations: e.g. N1A, N1C, N1D, ...
- Antibody type
- Currently no feature selection
- Use sparse matrices
- Also tried:
 - Location and types of mutations: e.g. 1,2,3,...,201, AC, AD, ...
 - Derived features: e.g. Moreau-Broto autocorrelation, conjoint triad descriptors, etc.

| Variant | N1* | N1A | N1C | ... | T201Y | COV2-2082_400 | ... | CR3022_400 | Endpoint |
|---------|-----|-----|-----|-----|-------|---------------|-----|------------|----------|
| 1 | 0 | 0 | 0 | | 0 | 0 | | 0 | -2.93 |
| 2 | 0 | 0 | 0 | | 0 | 1 | | 0 | -2.57 |
| 3 | 0 | 0 | 0 | | 0 | 0 | | 0 | -2.81 |
| 4 | 0 | 0 | 0 | | 0 | 0 | | 0 | -3.19 |

Use Keras neural net machine learning model

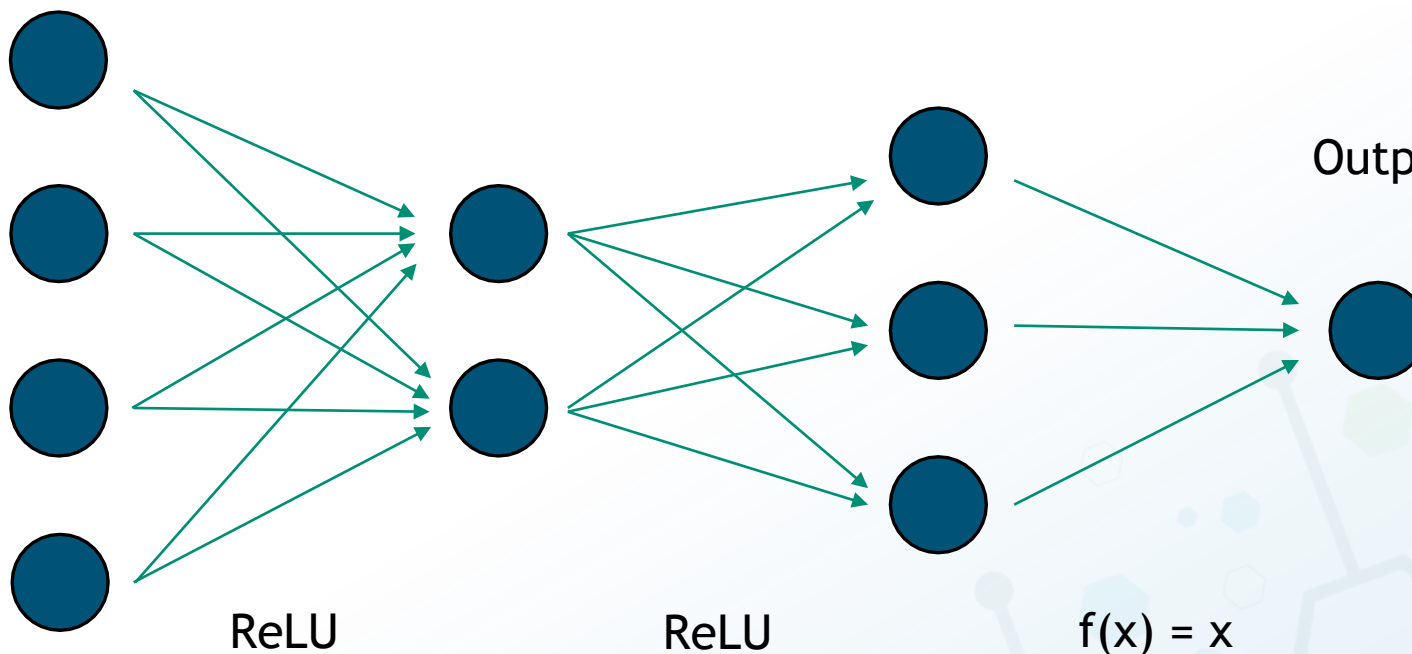
- Tensorflow backend
- Thousands of parameters
- Also tried:
Xgboost, random forest, support vector machines (too much data)



Input Layer

Hidden Layers

Output





Five-fold cross-validation

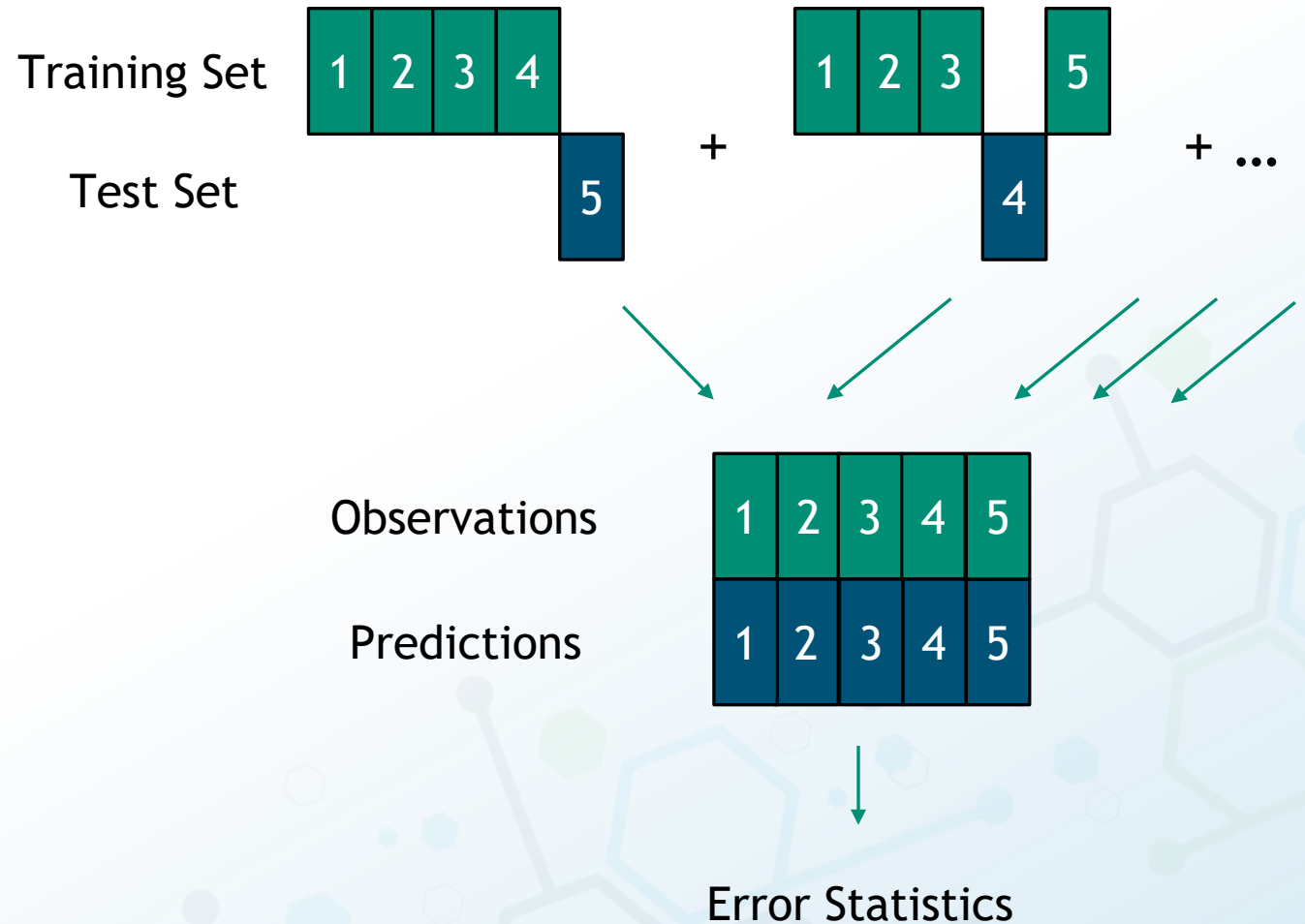
- Estimates predictive ability on data outside of training set
- Hidden assumption: future data is similar to data you have

Hyperparameters

- Layers: 2 or 3
- Sizes: $2^2, 2^3, 2^4, 2^5, 2^6, 2^7, 2^8$
- ReLU: Leaky or Regular
- 784 possible combinations

Tuning

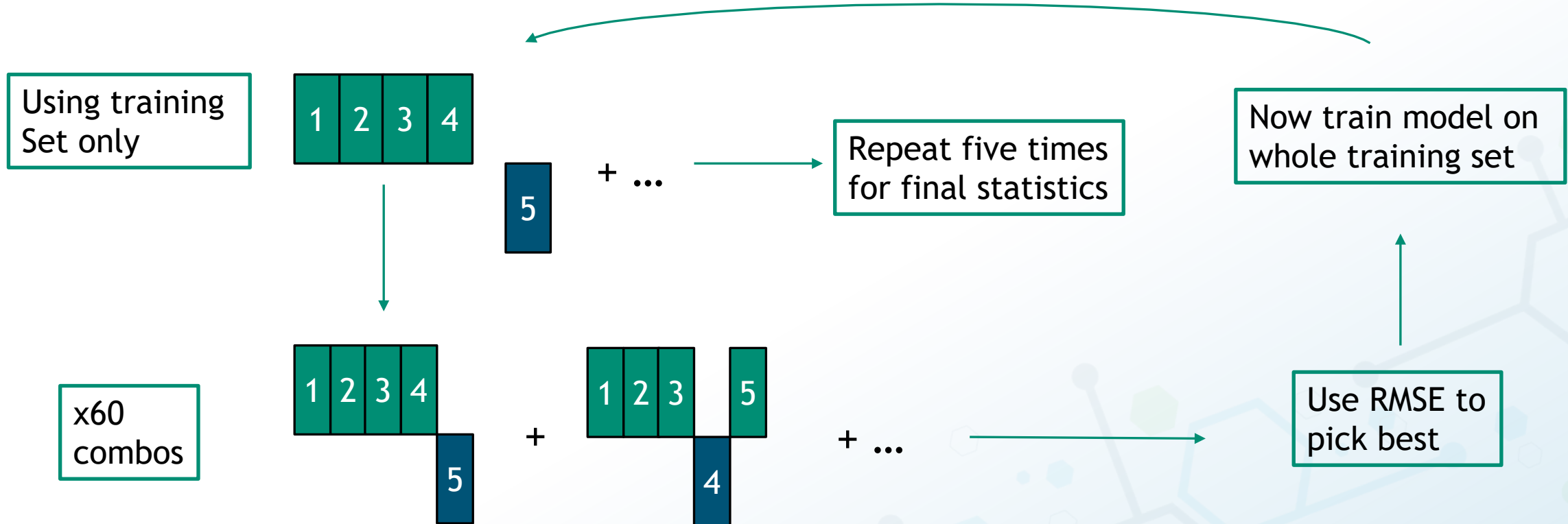
- 60 randomly selected combinations
- Choose parameters with best root-mean-square-error (RMSE)





Tuning in Loop

- Tune independently within each training set
- Avoids overfitting in results



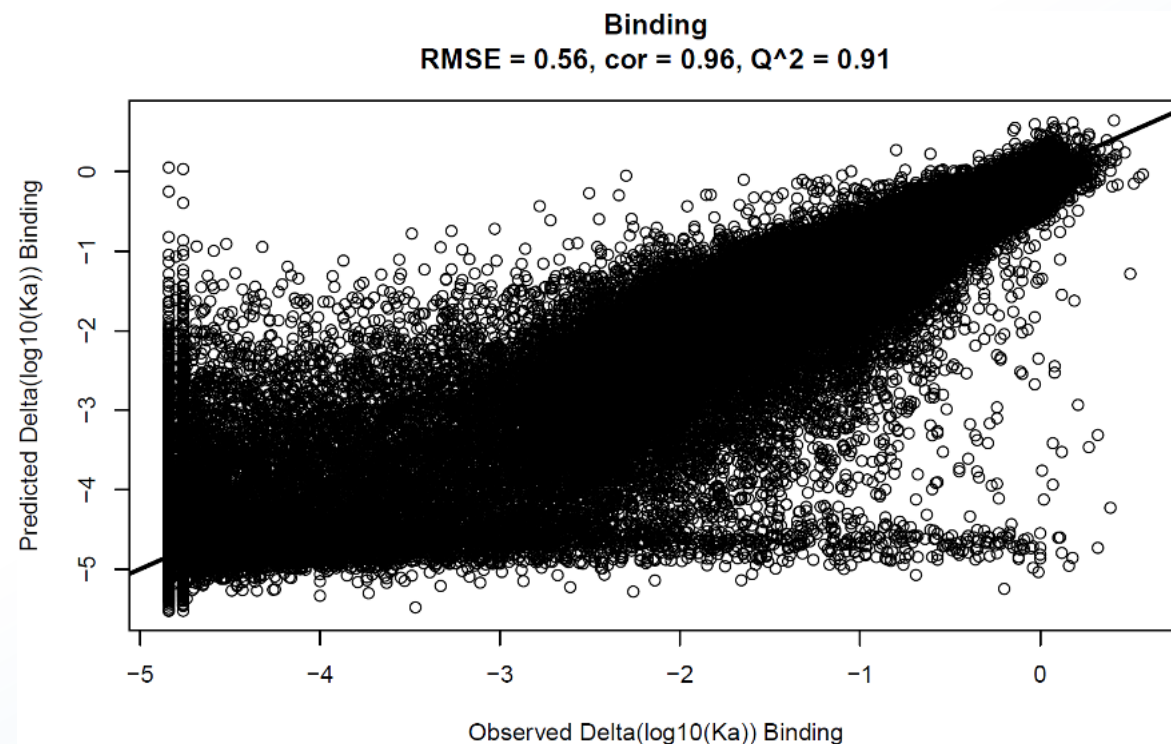


Statistics used

- Root-mean-square error (RMSE)
 - $\sqrt{\sum_i (y_i - f_i)^2 / n}$
 - Where y_i are observed endpoints and f_i are predictions based on other data points
- Pearson Correlation
- Cross-validated coefficient of determination (R^2)
 - $Q^2 = 1 - \sum_i (y_i - f_i)^2 / \sum_i (y_i - \bar{y})^2$

Binding results:

- RMSE = 0.56 $\Delta\log_{10}(K_a)$
- Pearson correlation of 0.96
- $Q^2 = 0.91$



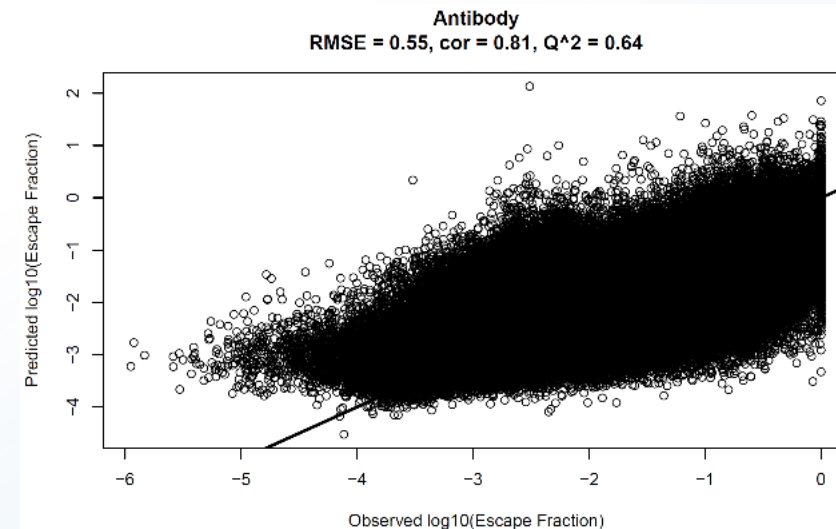
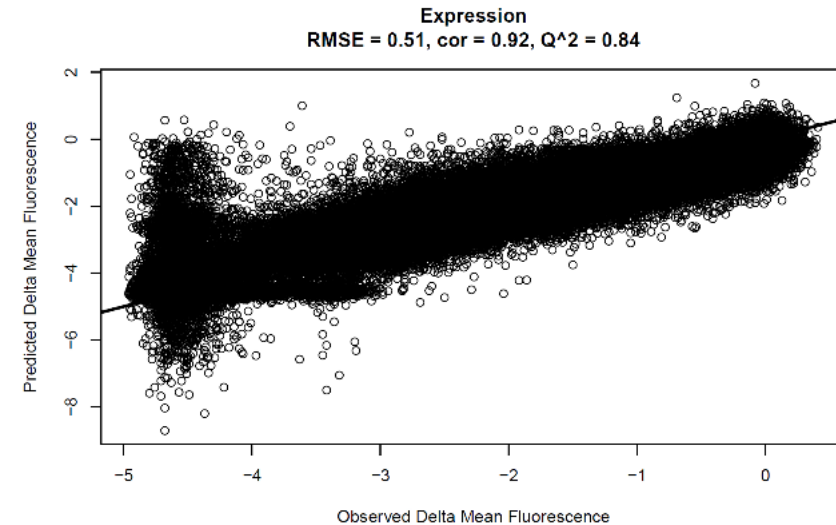


Expression results:

- RMSE = 0.51 Δ Mean Fluorescence
- Pearson correlation = 0.92
- $Q^2 = 0.84$

Antibody results:

- Untuned, using 128 x 32 hidden layers and regular ReLU
- RMSE = 0.55 $\Delta\log_{10}$ (Escape Fraction)
- Pearson correlation = 0.81
- $Q^2 = 0.64$



Antibody Model Issues



Originally made one model for each antibody

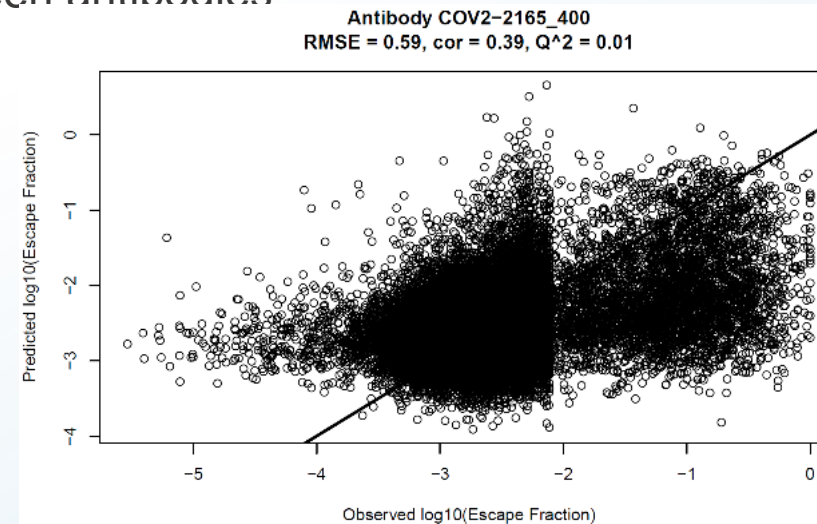
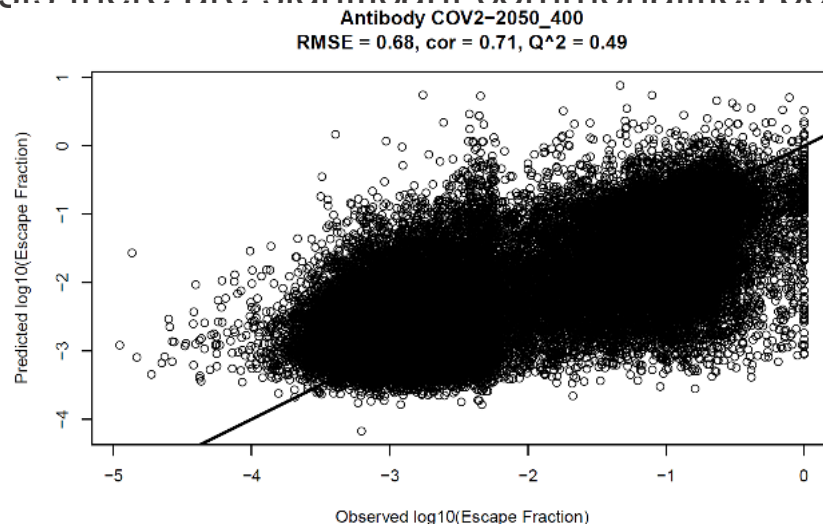
- But half of models were mediocre and other half were poor

Why?

- Antibody escape is based on counts before and after an antibody is applied
- Lowest count observations are discarded, but the bulk of observations are low-count and high uncertainty
- Removing them degrades model quality even further
- Weighting observations did not help
- The five poor antibody models are dominated by these observations

Combining model had better statistics than any single antibody model

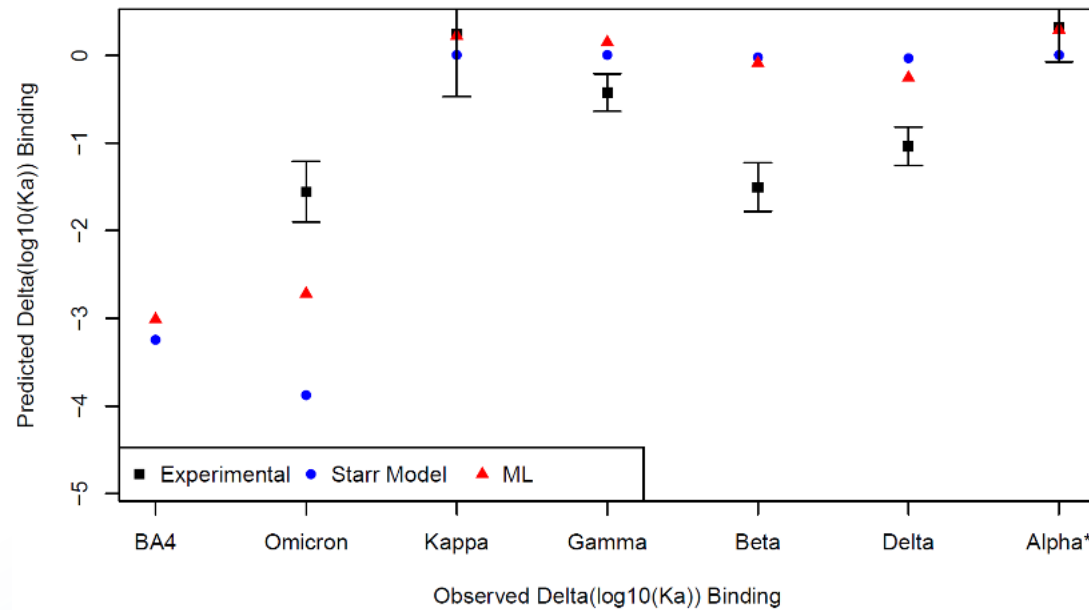
- Suggests there are significant commonalities between antibodies





Our group performed binding assays on wild variants

- Only six can be compared so far
 - Can't predict insertions
 - Alpha is only variant present in training data
- Omicron BA.1.1 has 16 RBD mutations
- BA4/5 has 17, and only 11 in common with BA.1.1

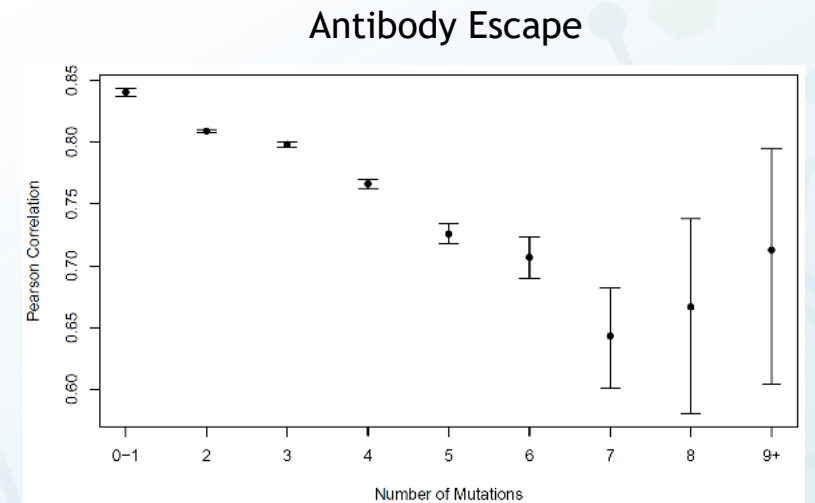
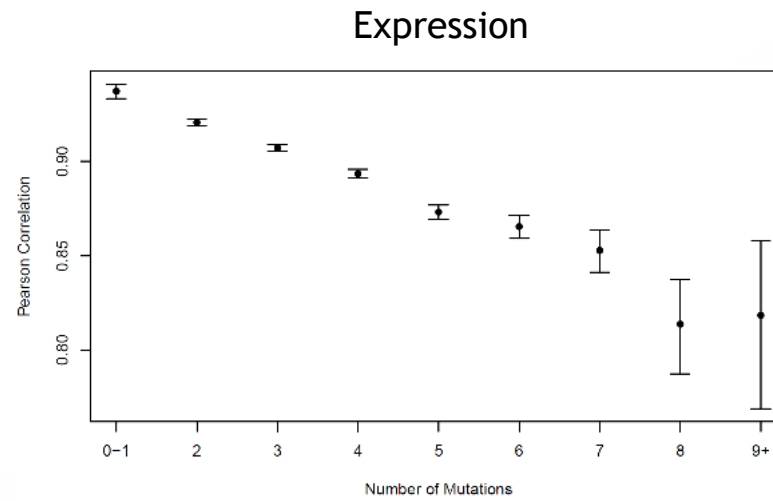
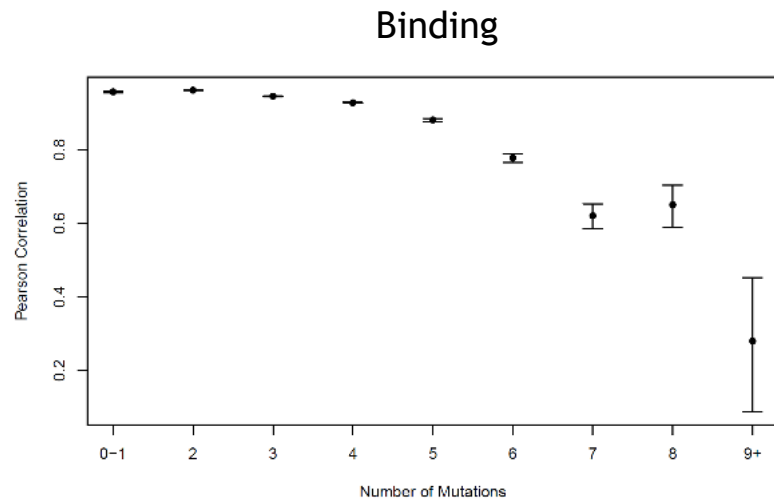




Model quality tends to drop with increasing numbers of mutations for all models

Single mutations and small combinations are well-represented in the data

More complicated mutation combinations are not present, and hard to predict the effect of

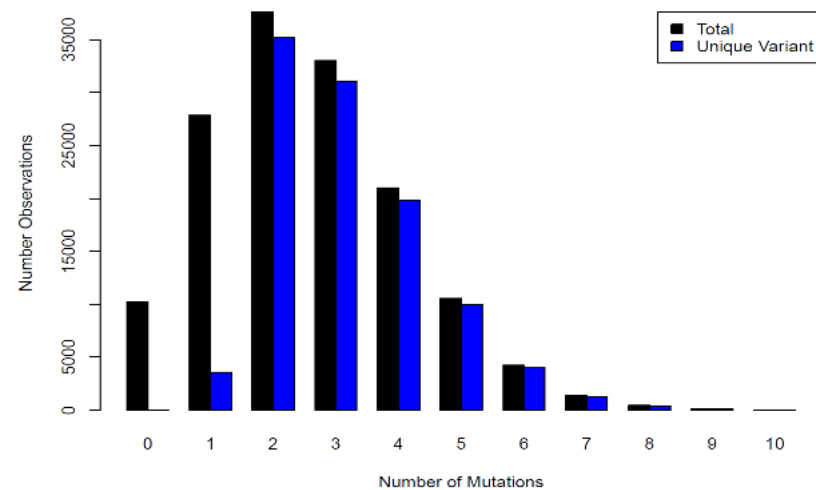




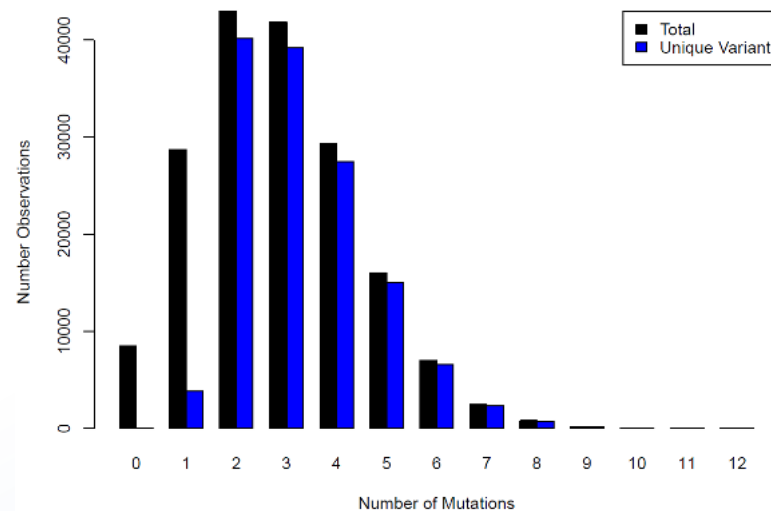
Original experiment was designed to find single mutation effects

- Space of variants near Wuhan is well-covered
- Coverage drops quickly with more mutations
 - Especially relative to all possible combinations
- Space near omicron variants is unexplored

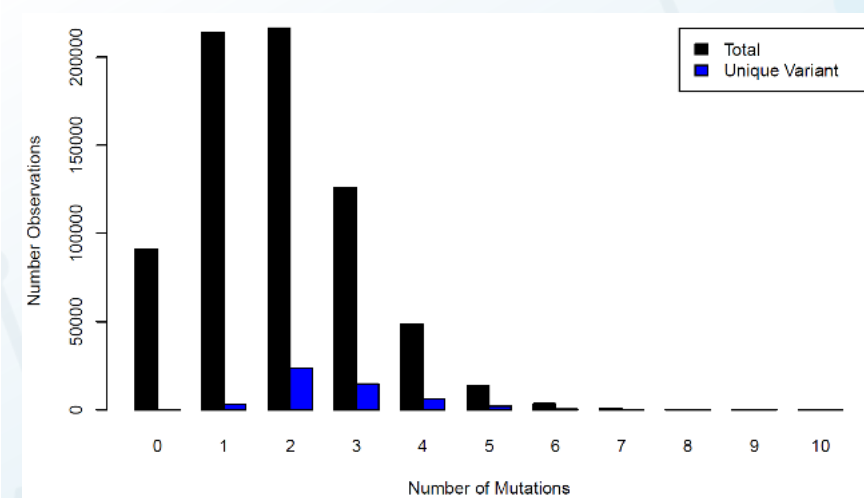
Binding



Expression



Antibody



Site Region Selection For Future Experiments



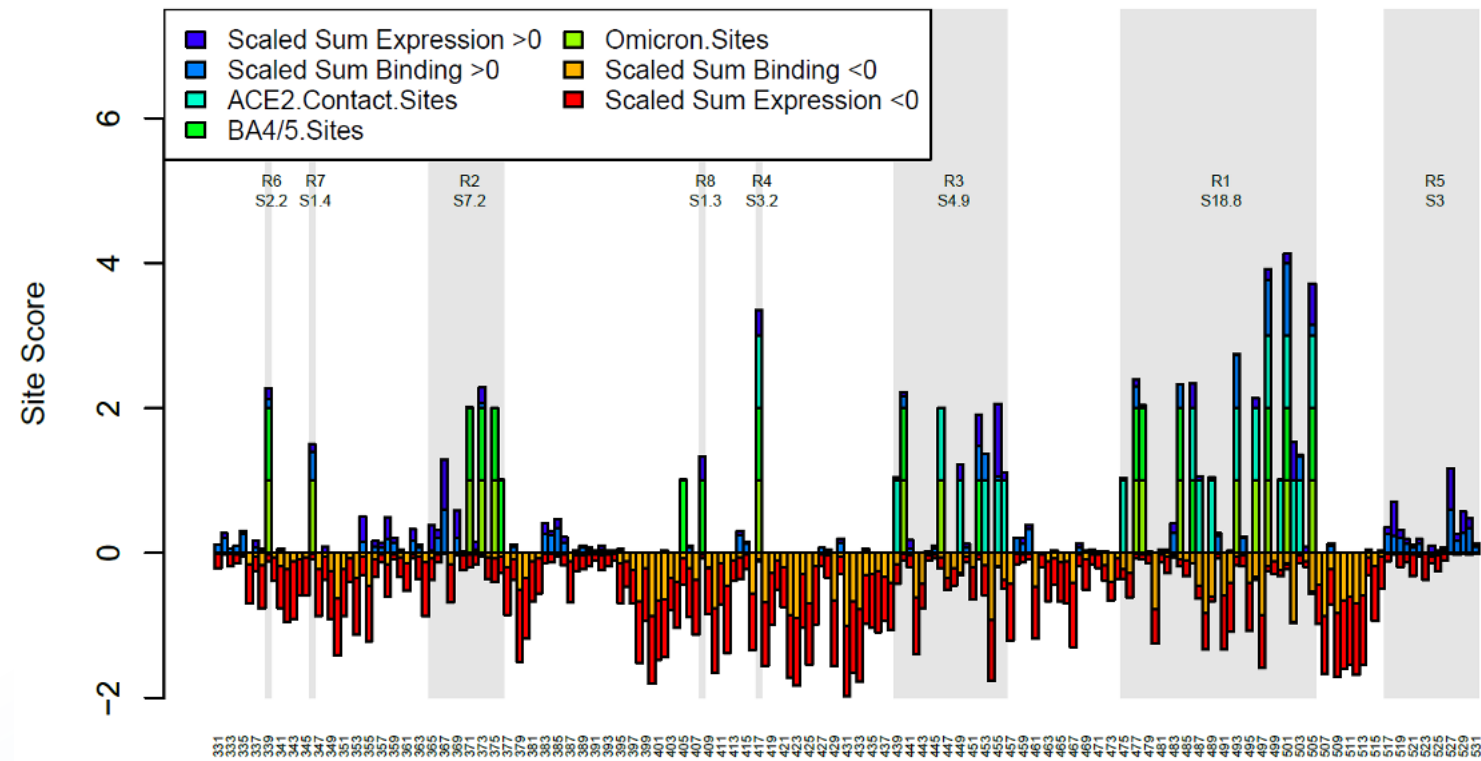
Use Starr model single mutation binding/expression effects

Sum positive and negative binding/expression effects for each site and scale to a maximum of 1 over all sites

Increase score by one for site presence in Omicron BA1.1, BA4/5, or ACE2 contact site

Find regions with maximum total scores iteratively

- Top four regions: 475-505, 365-376, 439-356, 417
- Top four scores: 18.8, 7.2, 4.9, 3.2





Further binding experiments centered on BA4/5

- Train wider mutation set
- Closer to current state of virus
- Further model validation

Antibody tuning

Antibody binding based on antibody sequence/characteristics



Acknowledgements



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