



# Synthetic Biology Enabled Modification of Bone Cells and the Impact on Bone Mineral Formation

Dulce Hayes, Kimberly Butler, Kalista Blake Pattison, Jerilyn A. Timlin

## Purpose of Study

Modification of inorganic biomaterial by engineering bone producing osteoblasts to increase levels of the signaling molecule, osteoprotegerin (OPG).

**Impact:** If successful, this is a first step toward tuning bone cells to produce bone material to desired properties *in-vitro*.

## Introduction

- Bone is a hybrid inorganic material composed of the mineral hydroxyapatite and collagen.
- Bone tissue is composed of osteoblasts (form bone), osteocytes (sense mechanical stress), osteoclasts (remove bone)
- *In vivo*, bone regenerates in response to mechanical stress and transduces to biochemical response, specifically modifies the ratio of OPG and Receptor Activator of Nuclear factor Kappa-B Ligand (RANKL) which makes it attractive for advanced materials applications
- K7M2 bone cells are osteoblasts which secrete OPG and a high ratio of RANKL/OPG enhances the differentiation of pre-osteoclasts to mature osteoclasts while a low ratio of RANKL/OPG inhibits this differentiation of pre-osteoclasts such as RAW cells (monocytes).

Engineered K7M2 cells to overexpress OPG signaling molecule in the RANKL/RANK/OPG pathway and then assessed the effect on osteoclast formation and activity with a resorption pit assay using calcium phosphate as a synthetic biomimetic material. K7M2 is an osteosarcoma cell line that promotes osteoclastogenesis. In this study, we questioned whether engineering OPG is sufficient to alter the osteoclastogenic capacity of K7M2 cells.

## Findings

- OCs in UCM had more than 8 nuclei and OCs in CM generally had less than 3 nuclei
- There is a reduction in number of pits formed in CM from K7M2 cells

## Conclusions

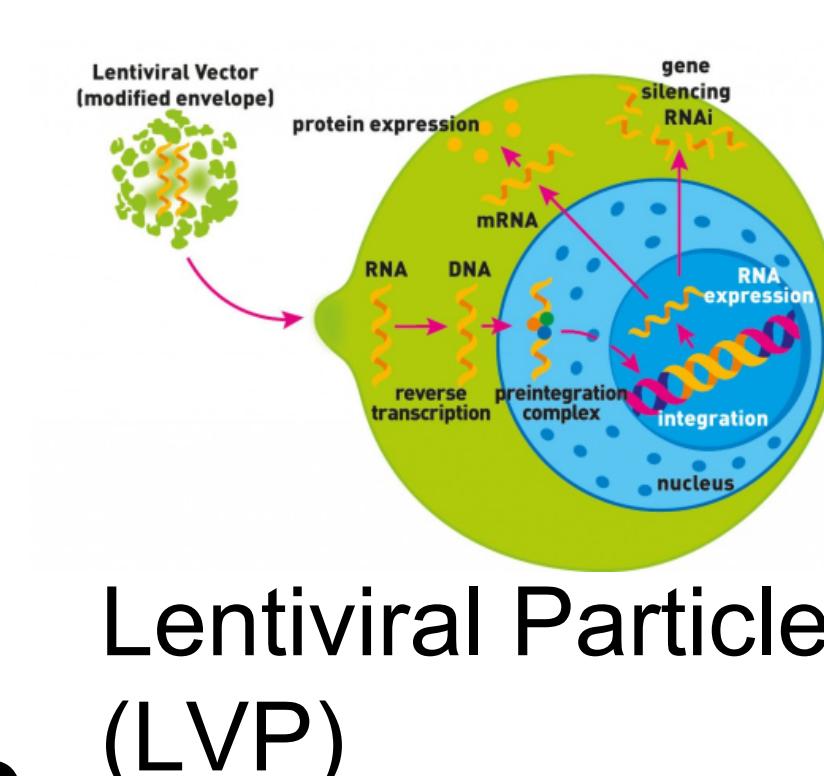
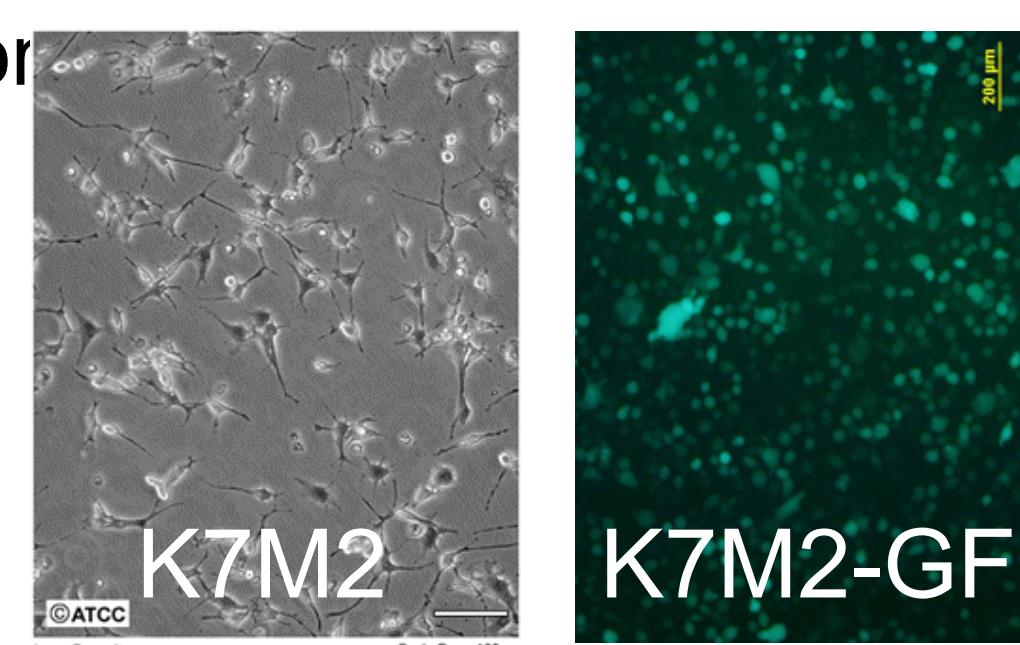
- Successfully engineered and characterized K7M2 cell line.
- The OCs in CM did not generally differentiate into mature OCs compared to OCs induced in UCM.
- Induction in K7M2 CM needs to be optimized.

This work was supported by the Laboratory Directed Research and Development program at Sandia National Laboratories, a multimission laboratory managed and operated by National Technology and Engineering Solutions of Sandia, LLC, a wholly owned subsidiary of Honeywell International Inc., for the U.S. Department of Energy's National Nuclear Security Administration under contract DE-NA0003525.

## Experimental Details & Results

### Genetic Engineering of K7M2

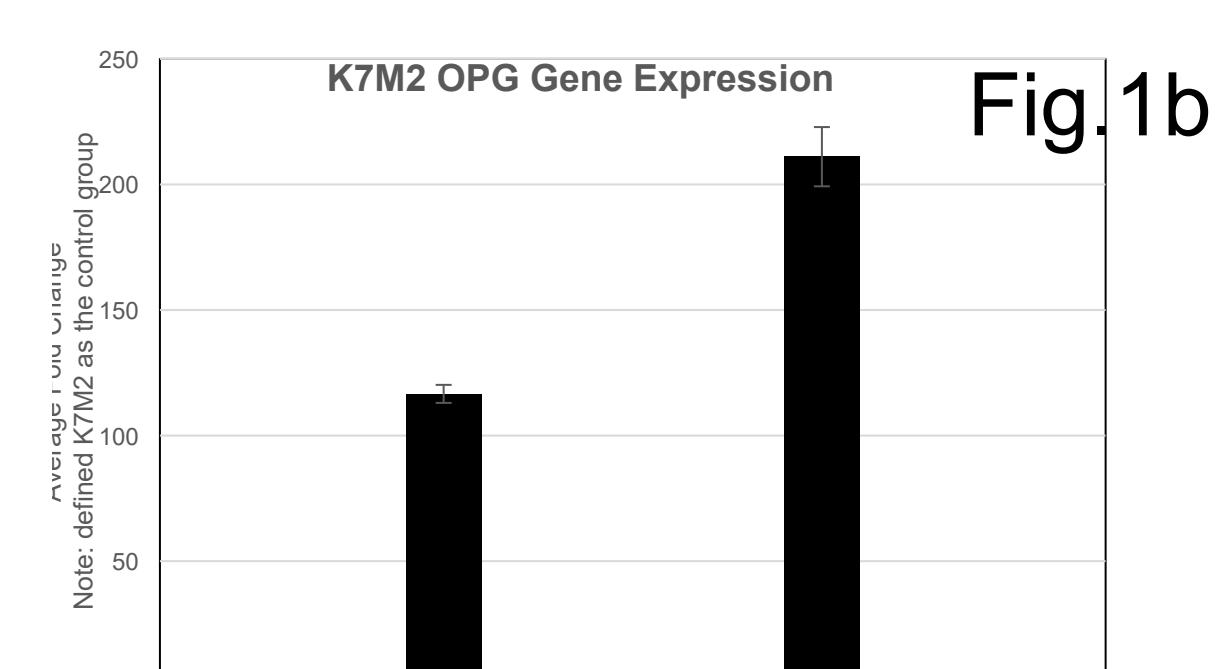
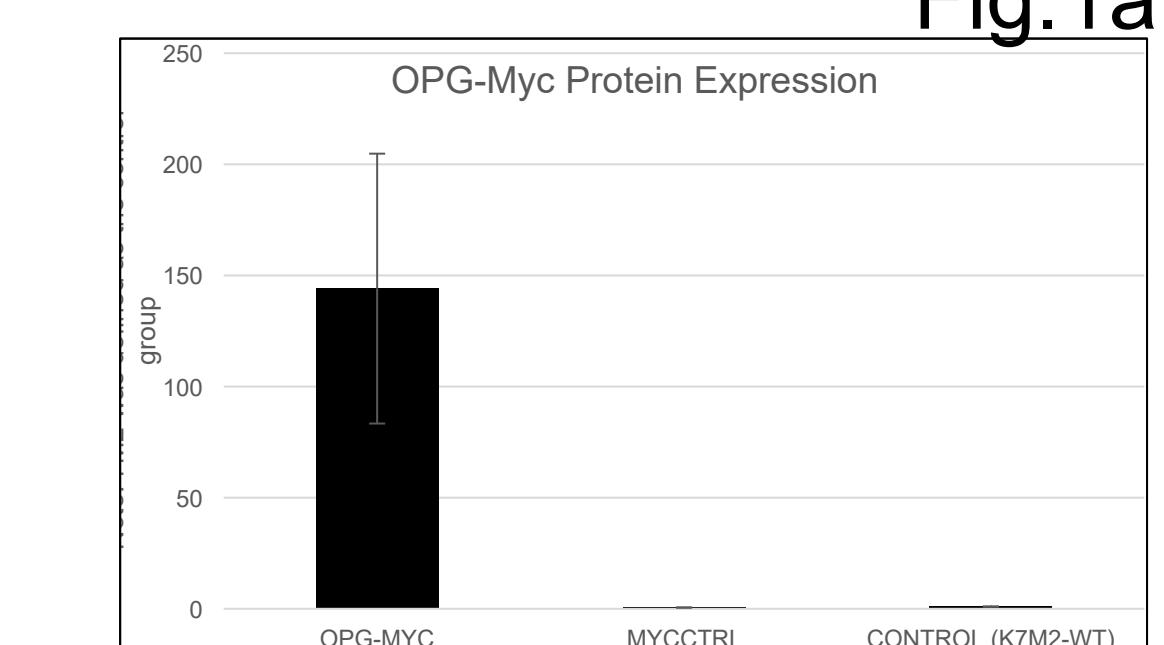
K7M2 cells were incubated with LVP's from Origene which introduced OPG gene into K7M2 chromosomes.



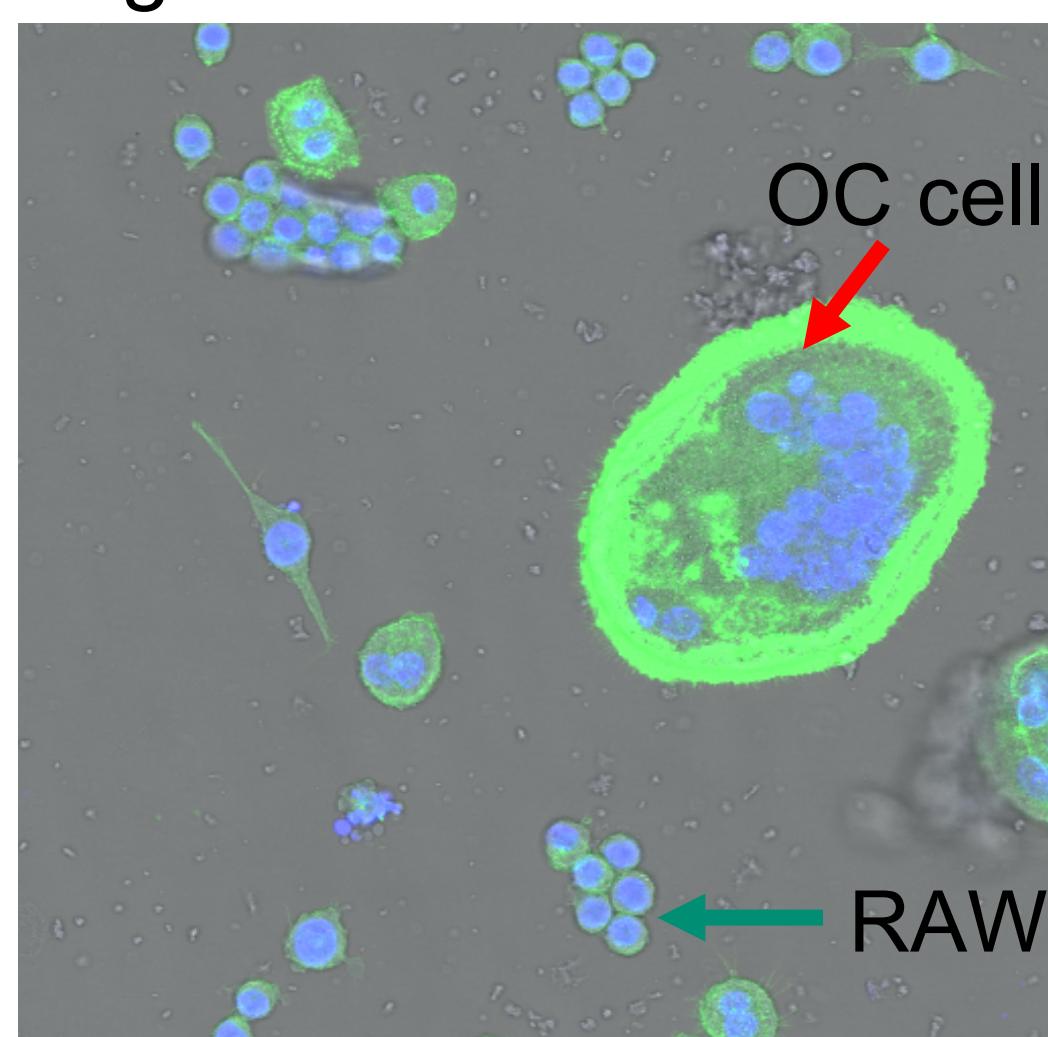
### Verification of Increased OPG Expression

K7M2-GFP modified cell line was cultured in DMEM medium for 48 hours.

- Medium was assayed using Enzyme-linked immunosorbent assay (ELISA) to measure OPG protein concentration compared to cells modified with a control LVP which should not increase OPG (K7M2-Myc Tag), and the parent cell line (K7M2) (Fig.1a)
- Polymerase Chain Reaction (PCR) was used to assess gene expression. (Fig.1b)



### Differentiation of RAW cells to Osteoclasts (OC)



### Effect of Increased OPG on Osteoclast Differentiation

RAW cells were grown overnight in DMEM medium. The following day, this conditioned medium (CM) was added to RAW cells. A resorption pit assay and quantitative confocal microscopy was used to determine the effect.

### Resorption Pit Assay

6 well plates were coated with simulating body fluid for 3 days and then 1 day with  $\text{CaPO}_4$ . RAW cells were seeded on these plates and induced with RANKL with conditioned medium from K7M2-OPG cells (Fig.3 c) or K7M2-Myc ctrl cells (Fig.3b) and as a positive control, RAW cells were seeded with unconditioned medium (UCM) (Fig. 3 a). After 6-7 days, plates were stained with 5% Silver Nitrate and treated with UV for an hour and then imaged with inverted microscope to visualize pits.

### Resorption by Osteoclasts

