

1 A Miniature Multilevel Structure for a Lossless Ion Manipulation Ion 2 Mobility Spectrometer with Wide Mobility Range Separation 3 Capabilities

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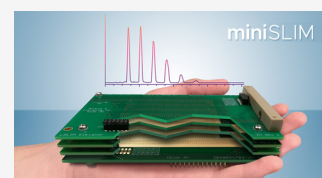


Article Recommendations



Supporting Information

6 **ABSTRACT:** Ion mobility spectrometry employing structures for lossless ion manipulations
7 (SLIM-IMS) is an attractive gas-phase separation technique due to its ability to achieve
8 unprecedented effective ion path lengths (>1 km) and IMS resolving powers in a small footprint.
9 The emergence of multilevel SLIM technology, where ions are transferred between vertically
10 stacked SLIM electrode surfaces, has subsequently allowed for ultralong single-pass path lengths
11 (>40 m) to be achieved, enabling ultrahigh resolution IMS measurements to be performed over
12 the entire mobility range in a single experiment. Here, we report on the development of a 1 m path
13 length miniature SLIM module (miniSLIM) based on multilevel SLIM technology. Ion trajectory simulations were used to optimize
14 SLIM board spacings and SLIM board thicknesses, and a new method of efficiently transferring ions between SLIM levels using
15 asymmetric traveling waves (TWs) was demonstrated. We experimentally characterized the performance of the miniSLIM IMS-MS
16 relative to a drift tube IMS-MS using Agilent tuning mixture cations and tetraalkylammonium cations. The miniSLIM achieved a
17 resolving power of up to 131 ($CCS/\Delta CCS$), which is $\sim 1.5\times$ higher than achievable with a 78 cm path length drift tube IMS.
18 Additionally, the entire ion mobility range was successfully transmitted in a single separation. We also demonstrated the miniSLIM's
19 performance as a standalone IMS system (i.e., without MS), which showed baseline separation between all AgTM cations and a clear
20 differentiation between different charge states of a standard peptide mixture. Overall, the miniSLIM provides a compact alternative
21 to high performance IMS instruments possessing similar path lengths.



22 ■ INTRODUCTION

23 Miniature ion mobility spectrometers (IMS) are highly
24 attractive for their small size, light weight, and lower power
25 consumption compared to their larger counterparts. The
26 challenge when transitioning to a miniature IMS is maintaining
27 sufficiently high separation power (i.e., resolving power and
28 resolution).¹ Typically, IMS separation power can be enhanced
29 by increasing (i) pressure, (ii) ion path length (absolute² and/
30 or effective³), (iii) electric field strength, or some combination
31 of the three.^{4,5} High separation powers can also be achieved by
32 trapping ions between a counteracting gas flow and voltage
33 gradient and slowly decreasing the voltage gradient, as in
34 trapped ion mobility spectrometry (TIMS),⁶ or by using
35 mixtures of buffer gases and/or slow changes in compensation
36 voltage, as in field asymmetric waveform IMS (FAIMS).⁷
37 While FAIMS and TIMS instruments can be quite small, they
38 incur limitations due to ion transmission efficiency or space
39 charge capacity. Voltage breakdown at or above certain
40 pressures limits the maximum usable voltage, and a lack of
41 effective confinement at elevated pressures limits the total
42 usable path length (e.g., in drift tubes). The general
43 miniaturization of IMS remains challenging, and a straightfor-
44 ward method for maintaining higher performance in
45 conjunction with miniaturization is highly desirable.

Increasing IMS separation power (at low pressure) without
46 changing instrument size is commonly performed by passing or
47 cycling ions around the same ion path multiple times. 48
49 Multipass methods provide increased effective path lengths
50 (i.e., the path length ions actually travel), allowing higher
51 resolving powers and resolutions to be achieved when many
52 passes are used (via a \sqrt{L} relationship, where L is the path
53 length for one pass or cycle).^{8–10} The main disadvantage of ion
54 multipass methods is that faster ions eventually overtake (i.e.,
55 “lap”) and pass slower ions if enough passes are allowed,
56 resulting in convoluted mobility spectra. This means that
57 multipass methods are best used to analyze a narrow ion
58 mobility range in a single experiment, keeping in mind that ion
59 mobility windows become narrower as the number of passes
60 increases. Ion lapping issues are especially pronounced for
61 cyclic miniature IMS systems, which normally employ short
62 absolute path lengths.

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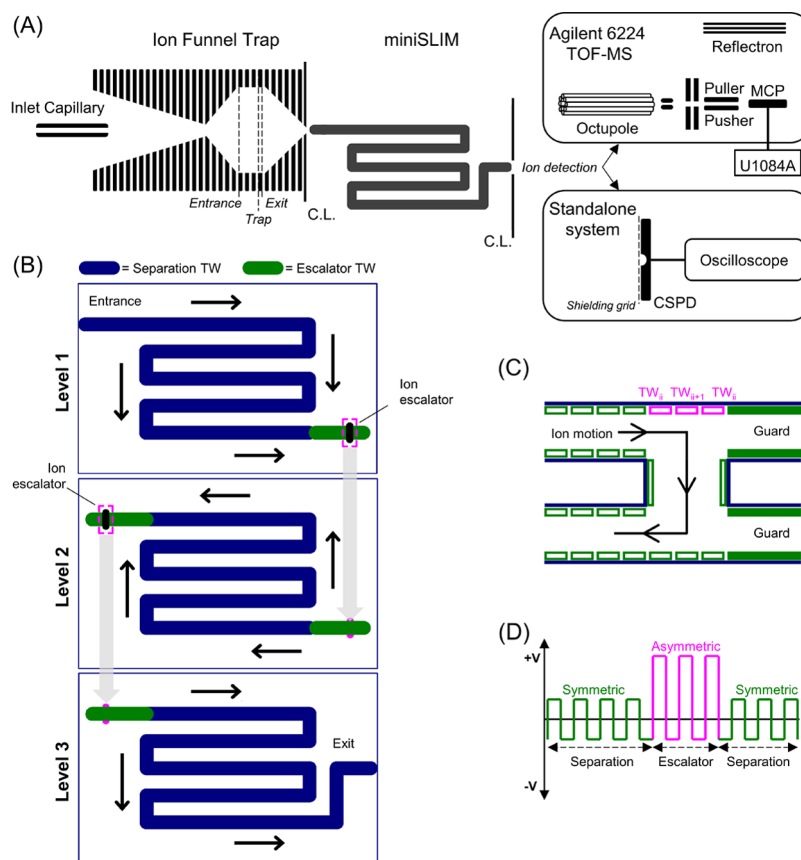


Figure 1. (A) Experimental schematic of the multilevel miniSLIM module and two ion detection methods. (B) Board layouts of all three miniSLIM levels. (C) Illustration of ion motion through an ion escalator employing asymmetric traveling waves. The leftmost and rightmost TW electrodes possess the same phase TW (ii), while the middle TW electrode possesses a 45° phase-shifted TW (ii + 1). (D) Depiction of symmetric and asymmetric traveling waves in the multilevel miniSLIM.

63 Long absolute path lengths have been established in IMS by
 64 using traveling wave (TW)-based IMS employing structures
 65 for lossless ion manipulations (SLIM-IMS).^{11,12} In SLIM-IMS,
 66 TWs propel ions between two mirror-image surfaces
 67 possessing TW, RF, and DC guard electrodes. The electrodes
 68 can be arranged on printed circuit boards (PCBs) to create
 69 long serpentine path lengths (>10 m absolute path length) in a
 70 small footprint (e.g., 45.9 cm × 32.5 cm).¹³ Additionally, ions
 71 that reach the end of the serpentine path can be sent back to
 72 the start of the SLIM path and allowed to pass through the
 73 serpentine path again to achieve truly unprecedented effective
 74 path lengths (e.g., >1 km).¹⁴ However, issues associated with
 75 ion lapping remain even with such long absolute path lengths,
 76 and multiple separations are required to cover a broad mobility
 77 range.

78 To achieve ultrahigh resolution ion mobility separations
 79 without ion multipass methods, we developed a multilevel
 80 SLIM by vertically stacking dual-surface SLIM boards (i.e.,
 81 PCBs with electrode arrays on each side).¹⁵ Ions transfer
 82 between levels using “ion escalators” located at the ends of
 83 each ion track.¹⁶ The long absolute path length (~43 m)
 84 achieved with a multilevel conventionally sized SLIM
 85 previously provided high resolution separations of phosphazene
 86 standards and complex mixtures of phosphopeptides over
 87 the entire mobility range of the sample.¹⁵ Such ion escalator
 88 technology has provided substantial increases to the absolute
 89 path lengths already achievable in SLIM-IMS without changing

the instrument footprint or using multipass methods, and a 90
 miniature version is a logical extension. 91

Here, we present the first miniature SLIM instrument 92
 (miniSLIM) using ion escalator technology to provide high 93
 resolution IMS separations over a wide mobility range without 94
 ion lapping. We used ion trajectory simulations to optimize 95
 and improve the performance of the ion escalators to ensure 96
 essentially lossless ion transmission using a new method for 97
 transferring ions between SLIM levels. The miniSLIM used 98
 three ion levels (33 cm path length per level), which 99
 established a total path length of 1 m. We characterized the 100
 performance of the optimized multilevel miniSLIM IMS when 101
 coupled to time-of-flight mass spectrometry (TOF-MS) using 102
 standard compounds and then as a fully functional standalone 103
 system. In all, the miniSLIM implementation provides a 104
 compact alternative to larger IMS instruments possessing 105
 similar path lengths. 106

EXPERIMENTAL SECTION 107

Chemicals and Electropray Ionization. Tetraalkylam- 108
 monium salts (TAA_C₂ – TAA_C₈) and solvents were 109
 purchased from Millipore-Sigma (St. Louis, MO, USA). 110
 Tetraalkylammonium salts (TAA_C₂ – TAA_C₈) were 111
 purchased as chloride, bromide, or hydrate salts and were 112
 diluted to a 1 μM equimolar mixture in acetonitrile. Nine 113
 peptide standards (angiotensin I, angiotensin II, bradykinin, 114
 fibrinopeptide A, kemptide, melittin, neurotensin, renin 115
 substrate tetradecapeptide, and substance P) were also 116

117 purchased from Millipore-Sigma and diluted to a 10 μM
118 equimolar mixture in 70:30 methanol:water. A low concen-
119 tration Agilent tuning mixture was purchased from Agilent
120 Technologies (Santa Clara, CA, USA) and used as received.
121 Nanoelectrospray ionization (nESI) emitters (2–4 μm o.d.)
122 were pulled from borosilicate glass capillaries (10 cm length,
123 1.5 mm o.d., 0.86 mm i.d.) using a P-2000 laser tip puller
124 (Sutter Instruments, Novato, CA, USA). High voltage was
125 applied to the solutions via a stainless-steel wire inserted into a
126 microelectrode holder (Warner Instruments, Hamden, CT,
127 USA).

128 **Ion Optics and SLIM Instrumentation.** The main
129 components of the miniSLIM IMS module included a single
130 inlet capillary (500 μm i.d.), an ion funnel trap (IFT),¹⁷ and
131 four vertically stacked PCBs comprising the miniSLIM (Figure
132 1A,B). When mass analysis was desired, ions were detected and
133 digitized using an Agilent 6224-TOF-MS and U1084A 8-bit
134 digitizer (Acqiris, Plan-les-Ouates, Switzerland) operating at 1
135 GS/s. When the miniSLIM was operated as a standalone
136 system, an AcroMass charge-sensing particle detector (CSPD,
137 Taipei, Taiwan) and Tektronix oscilloscope (DPO 5204B,
138 Beaverton, OR, USA) were used. The TWs shown in the blue
139 regions always possessed high speeds and low amplitudes for
140 mobility separation, while the TWs shown in the green regions
141 possessed either high or low speeds and amplitudes, depending
142 on the experiment being performed. The pink regions indicate
143 the locations of ion escalators.¹⁵ Each ion level possessed ~ 33
144 cm path length, yielding a total path length of ~ 1 m for the
145 miniSLIM. Board spacings and thicknesses were 2.80 and 1.65
146 mm, respectively. The dimensions of the assembled miniSLIM
147 were 11.1 cm \times 6.7 cm \times 1.4 cm ($L \times W \times H$). Table S1 in the
148 Supporting Information provides experimental dimensions for
149 all electrode types used in the miniSLIM. Photographs of the
150 assembled and disassembled miniSLIM are also provided in
151 the Supporting Information (Figure S1).

152 **Ion Escalator Using Asymmetric Traveling Waves.**
153 The miniSLIM utilized asymmetric TWs (i.e., TWs with higher
154 amplitudes and a DC bias than the other TWs) applied to the
155 three TW electrodes above an ion escalator to facilitate
156 efficient ion transfer between levels. A diagram depicting the
157 new ion escalator region is shown in Figure 1C, and Figure 1D
158 shows examples of symmetric and asymmetric TWs as applied
159 to the separation and escalator TW electrodes, respectively.
160 The TW electrodes in pink show the TW electrodes above the
161 ion escalator that are independent from the surrounding TW
162 electrodes in green. Note that the rightmost TW electrode
163 possessed the same phase TW (designated TW_{ii}) as the
164 leftmost TW electrode, and the middle TW electrode
165 possessed a 45 phase-shifted TW (designated TW_{ii+1}). It was
166 also possible to apply TWs with different speeds to the ion
167 escalator. Different from our previously published design,¹⁵ we
168 also removed all but one reverse TW electrode across the ion
169 escalator orifice and replaced the area with a guard electrode.

170 **Data Acquisition and Handling.** IMS-MS spectra were
171 collected using a custom data acquisition algorithm and viewed
172 using the Pacific Northwest National Laboratory (PNNL)
173 unified ion mobility format (.UIMF) viewer. Standalone IMS
174 spectra were collected using an oscilloscope (DPO 5204B,
175 Tektronix Inc., Beaverton, OR, USA). All data were exported
176 as .csv files and imported into MATLAB (Mathworks, Natick,
177 MA, USA) for data processing and plotting. Baseline
178 corrections for standalone data were performed in Origin
179 2021b (OriginLab Corporation, Northampton, MA, USA).

RESULTS AND DISCUSSION

180

Optimization of Ion Escalator Geometries Using Ion Trajectory Simulations. Ion escalators allow ultralong path lengths (>40 m) to be established in SLIM without ion cycling and have enabled separations over wide mobility ranges with ultrahigh resolution (resolving power ~ 560).¹⁵ Although ion escalators yield high ion transmission efficiency for high mobility ions, some low mobility ions can be lost even when general ion surfing conditions are used (i.e., low speed, high amplitude TWs). We hypothesize that ion losses arise from the fact that ions must traverse a long distance in one TW period to be efficiently transferred through an ion escalator, lest they roll-over a TW and encounter deleterious effects, such as space-charge from incoming ions. This distance was ~ 5.5 mm in previous multilevel SLIM modules (board spacings and thicknesses were 3.15 and 2.35 mm, respectively), which is 5.5 \times longer than a standard TW electrode. When designing the miniSLIM, ion trajectory simulations were used to explore how the traversal distance affects ion transmission efficiency through ion escalators.

The effects of SLIM board thickness and board spacing (i.e., ion escalator length) were evaluated independently. Geometry files (.GEM) were used to create eight different ion escalator workspaces: five different board spacings (2.55, 2.70, 2.85, 3.00, and 3.15 mm at 2.35 mm board thickness) and three different board thicknesses (1.65, 2.00, and 2.35 mm at 3.15 mm board spacing). The transmission efficiencies of the same negative Agilent tuning mixture ions (m/z 602, 1034, 1334, 1634, 1934, 2234, 2534, and 2834; 1000 ions per m/z) previously used to evaluate transmission efficiency were evaluated. Table S1 in the Supporting Information also provides the geometric and electronic simulation parameters for the previously developed multilevel SLIM system (as starting parameters) and the miniSLIM. Simulated ion transmission efficiency was defined as the number of ions traversing the ion escalator in one TW period divided by the total number of ions simulated.

Plots of ion transmission efficiency versus SLIM board spacing show that the transmission efficiencies of high mobility ions (e.g., m/z 602, 1034, and 1334) remain high regardless of board spacing (Figure 2A). However, the transmission efficiencies of moderate-to-low mobility ions (e.g., m/z 1634–2834) decrease when board spacings exceeded 2.55 mm, and transmission efficiencies were lowest when the largest board spacing was evaluated (i.e., 3.15 mm). This data indicates that ion transmission efficiencies through ion escalators are highest when board spacings are small. Similarly, high ion transmission efficiencies were obtained for all ions when smaller board thicknesses (e.g., 1.65 mm) were used compared to larger board thicknesses (e.g., 2.00 and 2.35 mm) (Figure 2B). The main takeaway from these simulations is that ion transmission efficiencies are best when small ion traversal distances (i.e., board spacings + board thicknesses) are used. Based on these simulations, we implemented board spacings and thicknesses of 2.80 and 1.65 mm, respectively, for the miniSLIM. This establishes a total ion traversal distance of 4.45 mm, which is 1.05 mm smaller than used in previous SLIM modules. We note that, while 2.55 mm board spacings yielded the highest simulated transmission efficiencies, we chose to use slightly larger board spacings because they provide room for more ions to traverse the SLIM than smaller board spacings. Another reason we used 2.80 mm board

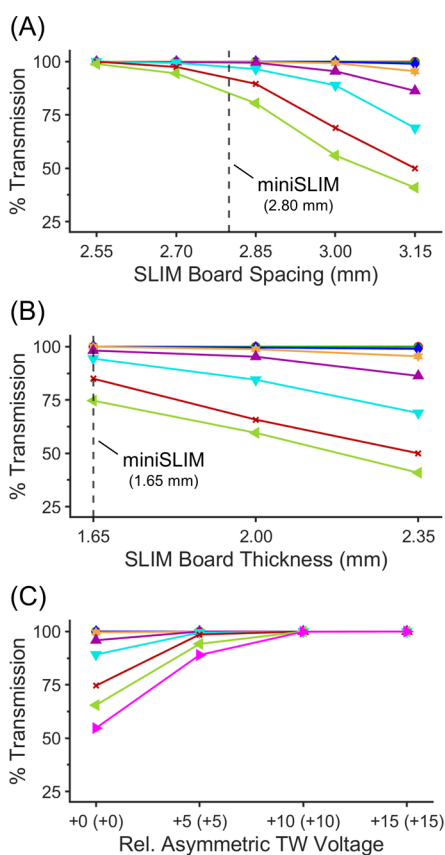


Figure 2. Simulated percent ion transmission efficiency plots of (–) Agilent tuning mixture ions traversing an ion escalator as a function of (A) SLIM board spacing and (B) SLIM board thickness using surfing traveling wave speeds. RF = 1 MHz, 300 V_{pp}, TW = 32 m/s, 15V_{0-p}, guard = –5V_{dc}, P = 3.5 Torr He. Colors in panels (A, B) represent (–) AgTM ions with *m/z* (red) 602, (dark green) 1034, (blue) 1334, (orange) 1634, (purple) 1934, (light blue) 2234, (dark red) 2534, and (light green) 2834. (C) Simulated percent ion transmission efficiency plots of (+) Agilent tuning mixture ions traversing an ion escalator as a function of asymmetric TW bias (amplitude). RF = 1.5 MHz, 300 V_{pp}, TW = 24 m/s, 15V_{0-p}, guard = +5V_{dc}, P = 2.5 Torr N₂. Colors in panel (C) represent (+) AgTM ions with *m/z* (red) 322, (dark green) 622, (blue) 922, (orange) 1222, (purple) 1522, (light blue) 1822, (dark red) 2122, (light green) 2422, and (pink) 2722.

242 spacings is to maximize the capturing of ions by the SLIM
243 given that the conductance limiting orifice of the ion funnel
244 trap preceding the miniSLIM has an i.d. of 2.50 mm. We also
245 note that we utilized 1.65 mm board thickness for practical
246 reasons (sturdiness and the multilayer construction of these
247 double-sided PCBs).

248 A few notable advantages are apparent when using
249 optimized SLIM board spacings and thicknesses. Ions
250 possessing high and low mobilities can be efficiently trans-
251 ferred through the escalator in one TW period, which
252 minimizes peak broadening and ion losses. However, what is
253 more interesting is that better transmission efficiencies enable
254 the use of nitrogen buffer gas for ion mobility separations. This
255 is important because transmission efficiencies were only high
256 when helium was used in previous multilevel SLIM systems.
257 Nitrogen is more practical to use and better suited for
258 miniature (and eventually portable) IMS systems, and the
259 remainder of the data presented here (simulations and
260 experiments) was collected using nitrogen.

Ion Transmission Efficiency Using Asymmetric TWs. 261

262 After optimizing SLIM board spacings and thicknesses, a new
263 method that uses asymmetric TWs to transfer ions between
264 SLIM levels (instead of surfing TWs) was evaluated. We note
265 that ion surfing happens when ions do not experience roll-over
266 events during their movement with the TW and instead move
267 at the same speed as the TWs. Ion surfing conditions can be
268 achieved when the TW speed is sufficiently low (lower than
269 ion velocity), or the TW amplitude is sufficiently high. The
270 new method of transferring ions through the ion escalator
271 involves applying asymmetric TWs to the three TW electrodes
272 located directly above the ion escalator while keeping all other
273 TW parameters the same (see Figure 1C,D). This idea was
274 based on decreasing the complexity of using TWs with
275 different speeds while maintaining similar or improved
276 performance. Ion transmission efficiencies using different TW
277 speeds (simulated and experimental) have been previously
278 described.¹⁵

279 Ion trajectory simulations of positive AgTM ions (*m/z* 322,
280 622, 922, 1222, 1522, 1822, 2122, 2422, and 2722) were
281 performed to explore how asymmetric TWs affect ion
282 transmission efficiency through ion escalators. The buffer gas
283 was 2.5 Torr nitrogen. Asymmetric TWs were established by
284 adding a DC bias and providing larger TW amplitudes to the
285 TWs applied to the three electrodes located directly above the
286 ion escalator. We use the nomenclature +V_{dc} (+V_{0-p}) to
287 indicate relative increases in asymmetric TW voltages, and we
288 note that these voltages are added on top of the already
289 existing TW amplitude being applied to the other TW
290 electrodes. For example, if a 15V_{0-p} TW is being applied to
291 the separation TWs, then +5V_{dc} (+5V_{0-p}) means that the
292 asymmetric TW will possess a +5V_{dc} bias and 20V_{0-p} TW
293 amplitude. The asymmetric TW voltages were increased with a
294 1:1 ratio (e.g., +5V_{dc} (+5V_{0-p}), +10V_{dc} (+10V_{0-p}), etc.), which
295 allowed us to obtain higher amplitude crests (i.e., tops of the
296 square waves) while maintaining identical amplitude troughs
297 (i.e., bottoms of the square waves). This was desired so that
298 ions could be pushed through the ion escalator with more
299 force than that of lower amplitude TWs.

300 Ion trajectory simulations using asymmetric TWs were
301 performed, and plots of ion transmission efficiency versus
302 relative asymmetric TW voltage show that asymmetric TWs
303 work well to transfer ions between SLIM levels (Figure 2C).
304 As can be seen, applying TWs with the same speed and
305 amplitude as the surrounding TW electrodes (i.e., no
306 asymmetric TW) results in ion losses, mostly associated with
307 medium and low mobility ions (i.e., *m/z* 1522–2722).
308 However, the transmission efficiencies greatly increase when
309 a +5V_{dc} (+5V_{0-p}) asymmetric TW is used, meaning that more
310 ions are traversing the ion escalator in one TW period.
311 Applying a +10V_{dc} (+10V_{0-p}) asymmetric TW further increases
312 transmission efficiencies to near 100%, and applying a +15V_{dc}
313 (+15V_{0-p}) achieves 100% transmission efficiency for all tested
314 ions. Additionally, the transmission efficiencies of nine
315 tetraalkylammonium cations were evaluated using the same
316 asymmetric TW voltages. However, all TAA ions transferred
317 through the ion escalators in one TW period under all
318 conditions, which is attributed to their higher mobilities as
319 compared to most AgTM ions.

320 **Characterization of miniSLIM-MS Performance.** The
321 results of the ion trajectory simulations showed that small PCB
322 spacings and PCB thicknesses should be used when employing
323 ion escalators and that asymmetric TWs work well to transfer

ions between levels. This knowledge was used to fabricate and assemble a multilevel miniSLIM platform that possessed three ion levels (two ion escalators) with a total path length of ~ 1 m. The main advantage of this miniSLIM is that the 1 m path length is established in a very compact format and without having to cycle ions, meaning that a wide mobility range can be analyzed. Initial experiments were performed by attaching the miniSLIM platform to TOF-MS and introducing the same positive AgTM ions examined in simulations using 3.5 Torr nitrogen buffer gas. Relative asymmetric voltages of $+20V_{dc}$ ($+20V_{0-p}$) were applied to the three ion escalator TW electrodes, and ions were injected using an IFT (488 μ s injection time). Voltages, frequencies, and pressures were optimized to produce the highest signal intensities. A full characterization of asymmetric TW performance is provided later.

A mobility spectrum of AgTM cations was produced after summing 500 individual separations (Figure 3A, green trace).

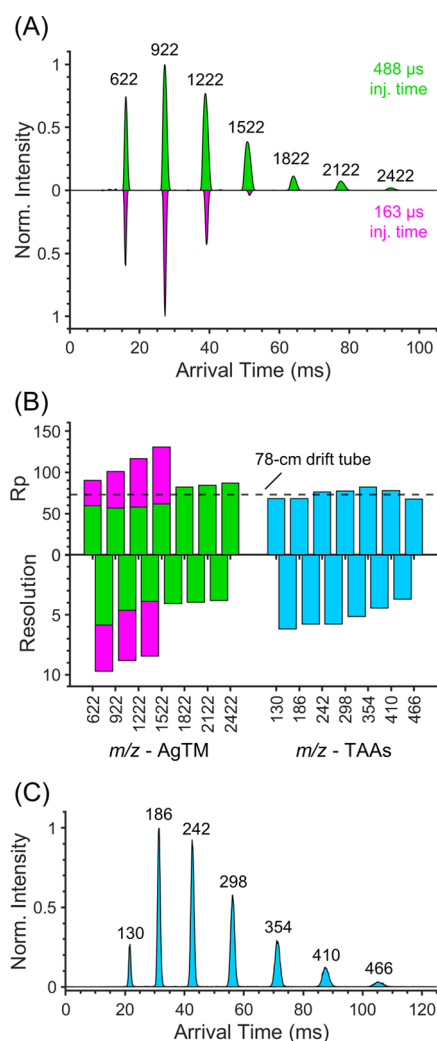


Figure 3. (A) IMS spectra of AgTM cations acquired using a multilevel miniSLIM module and injection times of (green) 488 and (pink) 163 μ s. $TW_{asym} = +20V_{dc}$ ($+20V_{0-p}$). $Pressure_{SLIM} = 3.5$ Torr nitrogen. (B) Resolving power and resolution calculations of AgTM and TAA cations. The dashed line indicates the highest resolving power obtained in ref 19 with a 78 cm drift tube. (C) IMS spectra of TAA cations. $TW_{asym} = +10V_{dc}$ ($+10V_{0-p}$). $Pressure_{SLIM} = 2.3$ Torr nitrogen.

As can be seen, AgTM ions with m/z 622–2422 were readily observed and baseline-separated, which represents a clear separation of ions possessing widely varying mobilities with a miniature SLIM instrument. The data also represents the successful transfer of high and low mobility ions through two ion escalators in a nitrogen buffer gas. Collisional cross section-based resolving powers ($R_p(CCS)$) were calculated by converting the arrival times of the AgTM ions to the CCS domain using CCS values from Stow et al.¹⁸ and applying eq 1

$$R_p(CCS) = \frac{CCS}{\Delta CCS} \quad (1)$$

where CCS is taken at the peak center and ΔCCS is the full width at half-maximum (fwhm) of the peak in the CCS domain. The resolving powers of the high mobility AgTM ions (m/z 622, 922, 1222, and 1522) were between 58 and 62, while the low mobility AgTM ions (m/z 1822, 2122, and 2422) were between 83 and 87 (Figure 3B, green bars). The resolving powers obtained for the miniSLIM using 488 μ s injection times were similar to the highest resolving power acquired by Ibrahim et al. using a 78 cm drift tube and singly charged ions, which was ~ 73 .¹⁹ These data represent the resolving powers and resolutions expected during routine analysis using a miniSLIM-MS module.

To determine the highest resolving power achievable using the miniSLIM, AgTM ions were injected using a 163 μ s injection time at 3.5 Torr nitrogen and 500 individual separations were summed (Figure 3A, pink trace). As expected, peaks were thinner when lower injection times were used, although there was a notable absence of lower mobility AgTM ions (i.e., m/z 1822–2422), which is attributed to ion gating effects. Resolving powers of the four observed AgTM ions at 163 μ s injection times ranged between 90 (m/z 622) and 131 (m/z 1522) with an average of 110 (Figure 3B, pink bars). These values demonstrate that the miniSLIM can produce resolving powers up to $\sim 1.5\times$ higher than a previously used 78 cm drift tube operating at a similar pressure but at a fraction of the total size. It may be possible to improve resolving powers by further reducing injection times, possibly by using the tristate ion shutter developed by Kirk et al., which showed that resolving powers of ~ 140 could be achieved when using 1 μ s injection times in a 30 cm drift tube.^{20,21} However, the drift tube used by Kirk et al. was operated at high electric field strength and 15 Torr, which was 4 to $5\times$ higher pressure than used in the miniSLIM.

Peak–peak resolutions were also calculated according to eq 2 using both injection times^{22,23}

$$Res_{pp} = 1.18 \times \frac{td_2 - td_1}{fwhm_2 + fwhm_1} \quad (2)$$

where td_i is the center of the ion arrival time distribution (ATD) for ion i and $fwhm$ is the full width at half-maximum of the ion ATD in the time domain. Resolutions ranged between 3.8 and 5.9 using 488 μ s injection times and 8.5 and 9.7 using 163 μ s injection times (Figure 3B, green and pink bars).

The miniSLIM was also tuned to transmit very high mobility ions, which required applying higher RF frequencies to the IFT and miniSLIM (1.5 MHz for both) and lower RF amplitude in the IFT ($\sim 50 V_{pp}$). The pressure in the SLIM chamber was set to 2.3 Torr N_2 to achieve optimal signal intensities. A mixture of seven TAAs was introduced using a 488 μ s injection time and relative asymmetric voltages of $+10V_{dc}$ ($+10V_{0-p}$) (Figure 399

3C, blue trace). As can be seen, all seven TAA cations were observed and baseline-separated. Resolving powers of the TAAs ranged between 68 and 92 with an average of 74, and resolutions ranged between 3.7 and 6.2 with an average of 5.2 (Figure 3B, blue bars). CCS values for TAAs were obtained from Campuzano et al.²⁴ These values are again similar to the resolving powers and resolutions obtainable using 78 cm path length drift tubes and showcase the miniSLIM's ability to separate high mobility ions in a nitrogen buffer gas.

Experimental Characterization of Asymmetric TWs.

The purpose of applying asymmetric TWs to the ion escalator is to ensure efficient ion transfer between SLIM levels. A systematic study of the effect of asymmetric TW on experimental ion transmission was performed by varying relative asymmetric TW voltages and monitoring the number and intensities of AgTM cations. Ions were injected with 488 μ s injection times, and separation TWs possessed speeds and amplitudes of 128 m/s and 17.5V_{0-p}, respectively.

Experiments began by applying high relative asymmetric TW voltages (+20V_{dc} (+20V_{0-p})) to the ion escalator and then decreasing by +5 (+5V_{0-p}) until separation TW voltages (+0V_{dc} (+0V_{0-p})) were reached (Figure 4A). Note that the

+15V_{dc} (+15V_{0-p}) resulted in similar signal intensities for all ions compared to +20V_{dc} (+20V_{0-p}), which means that high asymmetric TW voltages do not appear to cause a high mobility cutoff (i.e., by giving high mobility ions enough momentum to crash into the electrodes on the bottom board). However, a noticeable drop in ion signal intensities was observed when decreasing to +10V_{dc} (+10V_{0-p}), indicating that some ions do not successfully traverse the ion escalators. Further decreasing to +5V_{dc} (+5V_{0-p}) results in the total loss of the lowest mobility ion (m/z 2422), lower signal intensities overall, and peak tailing for the three highest mobility ions (m/z 622, 922, and 1222). The tailing indicates that ions did not traverse the ion escalator in one TW period but were also not lost. Last, removing the relative asymmetric TW voltages shows a total loss of the low mobility ions and significant peak tailing for all other ions.

This data demonstrates that applying even a small asymmetric TW significantly improves ion transmission through ion escalators and that high asymmetric TWs successfully transmit ions without observable deleterious effects (over the voltage range used). Since one of the goals of implementing the asymmetric TWs was to be an alternative to the previous method of transferring ions between SLIM levels (i.e., using separation and surfing TWs), a set of comparison mobility spectra was acquired by keeping the speed of the TW applied to the separation region constant (i.e., 128 m/s, 17.5V_{0-p}) and varying the speed of the TW applied to the ion escalator region (e.g., 128, 64, 32, and 16 m/s). The ion escalator region includes the TW electrodes surrounding the ion transfer region (i.e., the orifice) and the electrodes inside the orifice. Schematics depicting separation and ion escalator TW electrodes and the speed of TWs applied to each region are given in the Supporting Information (Figure S2). The mobility spectra acquired by varying the speed of the ion escalator TW along with extracted peak areas are also shown in the Supporting Information (Figure S3). As can be seen, peak areas and ATD shapes generally improved as lower TW speeds were used in the ion escalator. However, high mobility ions can be slightly lost, especially when a low TW speed is used (i.e., 128:16 m/s). This means that the speed of the TW applied to the ion escalator must be tuned depending on the mobility range being transferred to avoid losses.

An experiment to compare the highest resolving powers and resolutions achievable using the asymmetric and TW speed ratio methods was performed using +20V_{dc} (+20V_{0-p}) and 128:16 m/s, respectively. Overlaid mobility spectra collected using AgTM cations and injection times of 163 μ s at 3.5 Torr nitrogen using both methods and a plot of resolving power and resolution calculations are shown in the Supporting Information (Figure S4). The highest asymmetric TW voltage yielded approximately 10% (and up to 25%) higher resolving powers and resolutions than those obtained by applying a low TW speed to the ion escalator. We note however that both methods are viable ways to transfer ions between SLIM levels.

Experiments were also performed where combinations of asymmetric TWs and surfing TWs were used to evaluate if wider mobility ranges and more symmetric ATD shapes could be obtained. The mobility spectra of 12 combinations of asymmetric TWs (+0V_{dc} (+0V_{0-p}), +5V_{dc} (+5V_{0-p}), and +10V_{dc} (+10V_{0-p}) and TW speed ratios (128:128, 128:64, 128:32, and 128:16 m/s) are shown in the Supporting Information (Figure S5). TAA cations were used for this study. As can be seen, there is little difference between when an asymmetric TW

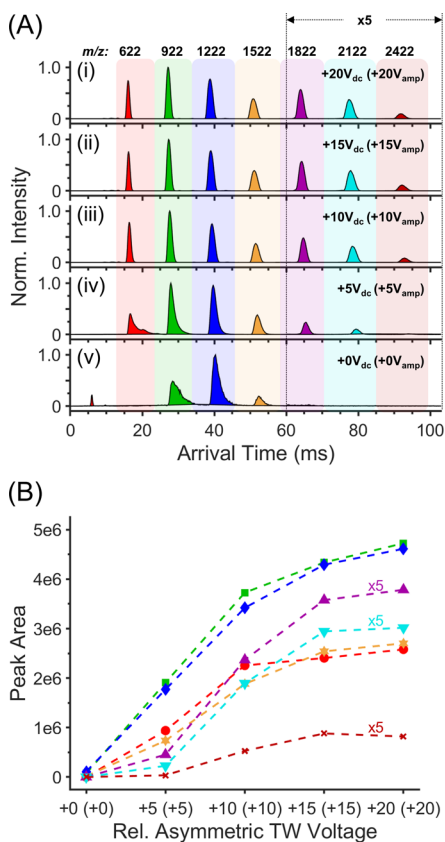


Figure 4. (A) IMS spectra and (B) extracted peak areas of AgTM cations as a function of asymmetric TW voltage (V_{dc} (V_{0-p})).

mobility regions around m/z 1822, 2122, and 2422 (i.e., >60 ms) were multiplied five times to observe ion ATDs more easily. As can be seen, applying +20V_{dc} (+20V_{0-p}) resulted in the observation of seven AgTM cations, each with a relatively symmetric ATD (Figure 4A, inset (i)). Ion peak areas were extracted and plotted as a function of relative asymmetric TW voltage (Figure 4B) to better illustrate the effect of changing voltage. Decreasing the relative asymmetric TW voltage to

493 was combined with a TW speed ratio and when one was not.
 494 We rationalize that ions are moving under surfing conditions
 495 when a slow TW is being applied, and increasing the amplitude
 496 (and DC offset) of the surfing TW will not change this. We
 497 note that our power supply could not produce asymmetric TW
 498 voltages above $+10V_{dc}$ ($+10V_{0-p}$) when combined with surfing
 499 TWs.

500 A final set of characterization experiments explored the
 501 effects of the asymmetric TW bias and amplitude independent
 502 from one another. Nine ion mobility spectra acquired using
 503 combinations of 0, 5, and $10V_{dc}$ and 0, 5, and $10V_{0-p}$ are shown
 504 in the Supporting Information (Figure S6). The results
 505 indicate that applying a DC bias without an increased TW
 506 amplitude (i.e., $+5V_{dc}$ ($+0V_{0-p}$) and $+10V_{dc}$ ($+0V_{0-p}$))
 507 improved ion ATD shapes and the transmitted mobility
 508 range. Alternatively, increasing the TW amplitude without a
 509 DC bias minimally improved spectral quality. However, the
 510 combination of DC bias and increased TW amplitude (equal
 511 values in particular) yielded mobility spectra with the most
 512 symmetric ion ATD shapes and widest mobility range, which
 513 agrees well with ion trajectory simulations.

514 **Operation of miniSLIM as a Standalone System.** A
 515 significant benefit of high performance miniature IMS systems
 516 is their ability to operate as standalone systems (i.e., IMS
 517 separations only). The performance of the miniSLIM as a
 518 standalone system was evaluated by detaching the miniSLIM
 519 from TOF-MS and using a charge sensing particle detector
 520 (CSPD) (see Figure 1A). A photograph of the standalone
 521 miniSLIM assembly is shown in Figure 5A.

522 The mobility spectra of AgTM ions were acquired using 488
 523 μs injection times, and 450 individual separations were
 524 averaged using the standalone system (Figure 5B). Relative
 525 asymmetric TW voltages were $+20V_{dc}$ ($+20V_{0-p}$). Baseline
 526 separation was again achieved, and the full mobility range of
 527 AgTM ions was observed, including the lowest mobility ion
 528 (m/z 2722). Sodiated adducts of several low mobility ions
 529 were also observed adjacent to the protonated species. We
 530 note that the IMS spectra collected using the CSPD required
 531 the application of a digital Fourier transform filter to remove
 532 interference from RF and TWs and also a baseline correction
 533 to account for CSPD response. The raw arrival time
 534 measurements and data processing workflow for AgTM cations
 535 are shown in the Supporting Information (Figure S7).

536 A preliminary demonstration of the miniSLIM's ability to
 537 separate biologically relevant compounds was performed by
 538 analyzing a mixture of nine standard peptides and averaging
 539 500 individual separations (Figure 5C, red trace). For
 540 comparison, mobility spectra were obtained using the IMS-
 541 TOF-MS configuration (Figure 5C, blue trace) and a list of
 542 detected peptides with charge states and arrival times is
 543 provided in the Supporting Information (Table S2). Note that
 544 two sections of the IMS-MS spectra were zoomed in to
 545 observe small features more easily ($72\text{--}130\text{ ms} = 10\times$ and $>$
 546 $130\text{ ms} = 1000\times$). As can be seen, there is good correlation
 547 between the standalone IMS and IMS-MS spectra. Eight of the
 548 nine peptides were successfully detected in the IMS-MS
 549 spectra as either the 1+, 2+, 3+, or 4+ charge state with
 550 protonated and/or sodiated adducts. The smallest and largest
 551 ions detected (and able to be assigned) were 386.5
 552 ($kemptide^{2+}$) and 1423 ($melittin^{2+}$), respectively. While
 553 standards would need to be analyzed to assign peaks in the
 554 standalone system, these IMS spectra establish a strong
 555 foundation for using multilevel SLIM technology to analyze

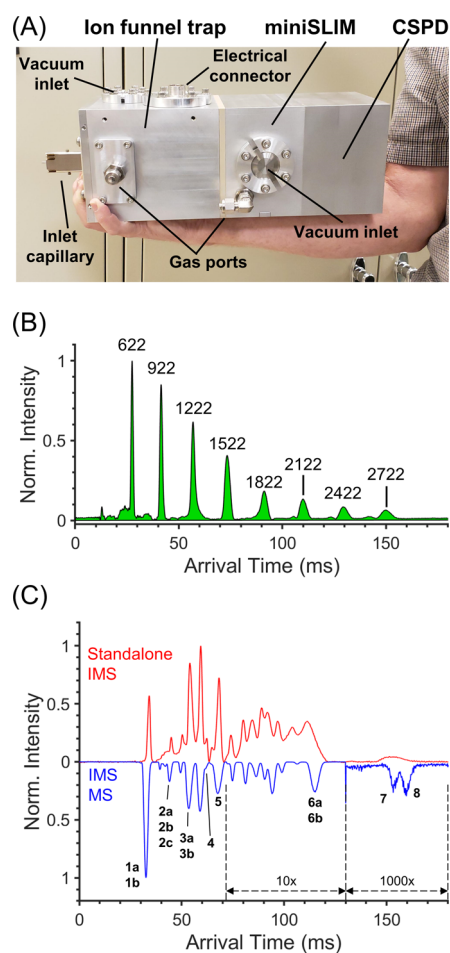


Figure 5. (A) Photograph of the standalone miniSLIM assembly. (B) IMS spectra of AgTM cations acquired using a standalone miniSLIM. $TW_{sep} = 192\text{ m/s}$, $15V_{0-p}$. (C) IMS spectra of a mixture of nine standard peptides acquired using (red) a standalone miniSLIM and (blue) miniSLIM-TOF-MS. $TW_{sep} = 192\text{ m/s}$, $15V_{0-p}$. $Pressure_{SLIM} = 3.5\text{ Torr}$ nitrogen.

biologically relevant mixtures over a wide mobility range 556 without having to cycle ions to obtain high quality separations. 557

CONCLUSIONS 558

We report the performance of a 1 m path length miniSLIM as a 559 hybrid IMS-MS instrument and as a standalone system. We 560 demonstrate a new method of transferring ions between SLIM 561 levels using asymmetric TWs. Optimizing the ion escalator 562 geometry and TW design facilitated high ion transmission 563 efficiency for a wide range of ion mobilities and allowed 564 nitrogen buffer gas to be used instead of helium. The 565 miniSLIM yielded resolving powers of up to $1.5\times$ higher 566 than obtainable with a previously reported 78 cm drift tube for 567 singly charged ions but at a fraction of the size. 568

A logical extension of this work could be to further increase 569 the path length by adding SLIM levels while maintaining a 570 similarly compact overall size. The present vacuum chamber 571 can accommodate approximately nine levels ($\sim 3\text{ m}$ ion path 572 length), which should provide a resolving power of ~ 220 . 573 Further improvements to ion transmission and ATD shapes 574 are likely obtainable using higher relative asymmetric TW 575 voltages. While the miniSLIM is quite small, it is not 576 completely portable due to the current ancillary components, 577

578 and improvements to make it so are underway, including
579 miniaturizing and simplifying the power supplies, reducing the
580 size of the ion detector housing, and implementing alternative
581 ion injection and detection methods. For instance, the ion
582 funnel trap can also be omitted and replaced by an ion trapping
583 and accumulation region integrated into one of the miniSLIM
584 levels. A Faraday detector can also help reduce the size and the
585 need for baseline correction. Pulsed ion and gas introduction,
586 as implemented by Cooks and co-workers,²⁵ as well as
587 alternative ionization and ion introduction methods can be
588 implemented to greatly reduce pumping requirements.
589 Optimizing mobility separations for air as the buffer gas will
590 also significantly enhance the attractiveness of the multilevel
591 miniSLIM IMS system. Such large additional gains serve to
592 make the miniSLIM IMS highly attractive for future standalone
593 instrumentation.

594 ■ ASSOCIATED CONTENT

595 ■ Supporting Information

596 The Supporting Information is available free of charge at
597 <https://pubs.acs.org/doi/10.1021/acs.analchem.1c04700>.

598 Geometric and electronic parameters of experimental
599 miniSLIM, simulated original multilevel SLIM, and
600 simulated miniSLIM systems; photographs of the
601 miniSLIM analyzer; schematics of the ion escalator
602 operating with TW speed ratios; ion mobility spectra
603 and extracted peak areas of Ag⁺ cations acquired
604 using four different TW speed ratios; performance
605 comparison of asymmetric TWs and TW speed ratios;
606 ion mobility spectra of TAA cations obtained using a
607 combination of asymmetric TWs and TW speed ratios;
608 effects of independently varying the DC voltage and TW
609 amplitude components of an asymmetric TW; data
610 filtering and processing workflow for the standalone
611 miniSLIM; and a list of detected peptides in a nine-
612 peptide mixture using IMS-MS (PDF)

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634 Notes

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■ REFERENCES

645

- (1) Ahrens, A.; Hitzemann, M.; Zimmermann, S. *Int. J. Ion Mobility Spectrom.* **2019**, *22*, 77–83.
- (2) Bohrer, B. C.; Merenbloom, S. I.; Koeniger, S. L.; Hilderbrand, A. E.; Clemmer, D. E. *Annu. Rev. Anal. Chem.* **2008**, *1*, 293–327.
- (3) Glaskin, R. S.; Ewing, M. A.; Clemmer, D. E. *Anal. Chem.* **2013**, *85*, 7003–7008.
- (4) Cumeras, R.; Figueras, E.; Davis, C. E.; Baumbach, J. I.; Gràcia, I. *Analyst* **2015**, *140*, 1376–1390.
- (5) Cumeras, R.; Figueras, E.; Davis, C. E.; Baumbach, J. I.; Gràcia, I. *Analyst* **2015**, *140*, 1391–1410.
- (6) Fernandez-Lima, F.; Kaplan, D. A.; Suetering, J.; Park, M. A. *Int. J. Ion Mobility Spectrom.* **2011**, *14*, 93–98.
- (7) Shvartsburg, A. A.; Clemmer, D. E.; Smith, R. D. *Anal. Chem.* **2010**, *82*, 8047–8051.
- (8) Giles, K.; Ujma, J.; Wildgoose, J.; Pringle, S.; Richardson, K.; Langridge, D.; Green, M. *Anal. Chem.* **2019**, *91*, 8564–8573.
- (9) Ben Faleh, A.; Warnke, S.; Rizzo, T. R. *Anal. Chem.* **2019**, *91*, 4876–4882.
- (10) Bansal, P.; Yatsyna, V.; AbiKhodr, A. H.; Warnke, S.; Ben Faleh, A.; Yalovenko, N.; Wysocki, V. H.; Rizzo, T. R. *Anal. Chem.* **2020**, *92*, 9079–9085.
- (11) Hamid, A. M.; Ibrahim, Y. M.; Garimella, S. V. B.; Webb, I. K.; Deng, L.; Chen, T.-C.; Anderson, G. A.; Prost, S. A.; Norheim, R. V.; Tolmachev, A. V.; Smith, R. D. *Anal. Chem.* **2015**, *87*, 11301–11308.
- (12) Ibrahim, Y. M.; Hamid, A. M.; Deng, L.; Garimella, S. V. B.; Webb, I. K.; Baker, E. S.; Smith, R. D. *Analyst* **2017**, *142*, 1010–1021.
- (13) Deng, L.; Ibrahim, Y. M.; Hamid, A. M.; Garimella, S. V. B.; Webb, I. K.; Zheng, X.; Prost, S. A.; Sandoval, J. A.; Norheim, R. V.; Anderson, G. A.; Tolmachev, A. V.; Baker, E. S.; Smith, R. D. *Anal. Chem.* **2016**, *88*, 8957–8964.
- (14) Deng, L.; Webb, I. K.; Garimella, S. V. B.; Hamid, A. M.; Zheng, X.; Norheim, R. V.; Prost, S. A.; Anderson, G. A.; Sandoval, J. A.; Baker, E. S.; Ibrahim, Y. M.; Smith, R. D. *Anal. Chem.* **2017**, *89*, 4628–4634.
- (15) Hollerbach, A. L.; Li, A.; Prabhakaran, A.; Nagy, G.; Harrilal, C. P.; Conant, C. R.; Norheim, R. V.; Schimelfenig, C. E.; Anderson, G. A.; Garimella, S. V. B.; Smith, R. D.; Ibrahim, Y. M. *Anal. Chem.* **2020**, *92*, 7972–7979.
- (16) Ibrahim, Y. M.; Hamid, A. M.; Cox, J. T.; Garimella, S. V. B.; Smith, R. D. *Anal. Chem.* **2017**, *89*, 1972–1977.
- (17) Ibrahim, Y.; Belov, M. E.; Tolmachev, A. V.; Prior, D. C.; Smith, R. D. *Anal. Chem.* **2007**, *79*, 7845–7852.
- (18) Stow, S. M.; Causon, T. J.; Zheng, X.; Kurulugama, R. T.; Mairinger, T.; May, J. C.; Rennie, E. E.; Baker, E. S.; Smith, R. D.; McLean, J. A.; Hann, S.; Fjeldsted, J. C. *Anal. Chem.* **2017**, *89*, 9048–9055.
- (19) Ibrahim, Y. M.; Baker, E. S.; Danielson, W. F., III; Norheim, R. V.; Prior, D. C.; Anderson, G. A.; Belov, M. E.; Smith, R. D. *Int. J. Mass Spectrom.* **2015**, *377*, 655–662.
- (20) Kirk, A. T.; Grube, D.; Kobelt, T.; Wendt, C.; Zimmermann, S. *Anal. Chem.* **2018**, *90*, 5603–5611.
- (21) Kirk, A. T.; Bohnhorst, A.; Raddatz, C.-R.; Allers, M.; Zimmermann, S. *Anal. Bioanal. Chem.* **2019**, *411*, 6229–6246.
- (22) Dodds, J. N.; May, J. C.; McLean, J. A. *Anal. Chem.* **2017**, *89*, 12176–12184.

- 701 (23) Jeanne Dit Fouque, K.; Ramirez, C. E.; Lewis, R. L.; Koelmel, J.
702 P.; Garrett, T. J.; Yost, R. A.; Fernandez-Lima, F. *Anal. Chem.* **2019**,
703 *91*, 5021–5027.
- 704 (24) Campuzano, I.; Bush, M. F.; Robinson, C. V.; Beaumont, C.;
705 Richardson, K.; Kim, H.; Kim, H. I. *Anal. Chem.* **2012**, *84*, 1026–
706 1033.
- 707 (25) Gao, L.; Cooks, R. G.; Ouyang, Z. *Anal. Chem.* **2008**, *80*,
708 4026–4032.