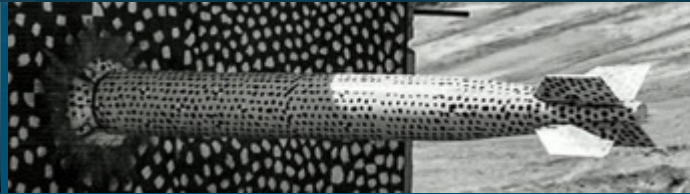
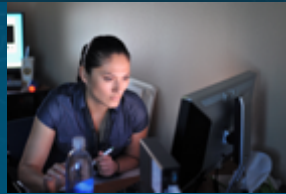


# UT Austin | Sandia Technical Exchange



PRESENTED BY



# Synthetic Biology

Anne Ruffing ([aruffin@sandia.gov](mailto:aruffin@sandia.gov) )



# Synthetic Biology for Biofuel and Biochemical Production



## CRISPRi modification of *Synechococcus* sp. PCC 7002 for enhanced biomass production

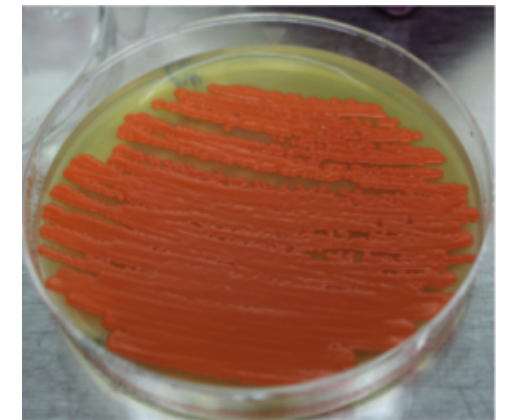
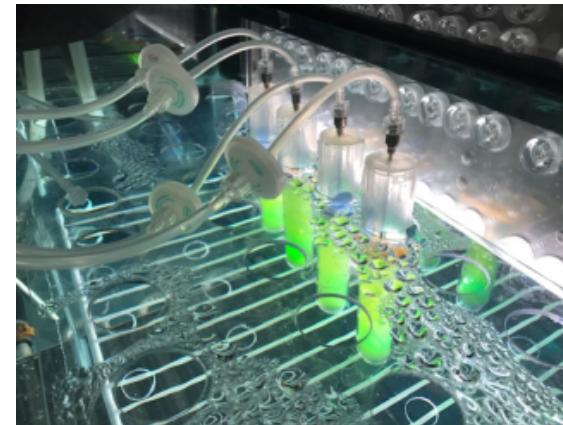
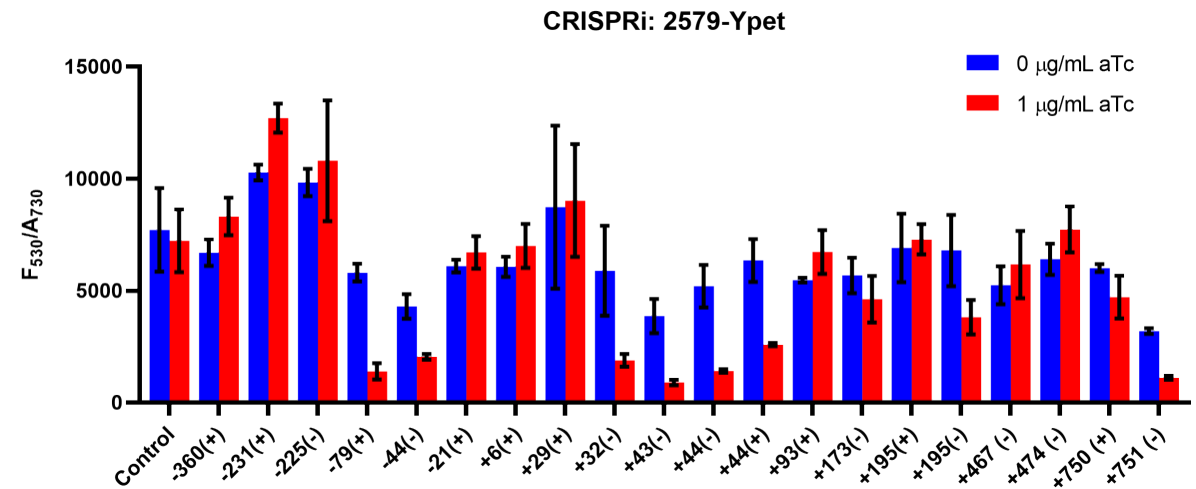
- CRISPRi design rules for cyanobacteria
- Shuttle vector for gRNA expression in 7002
- Improved operon prediction using RNA-seq data and machine learning algorithms
- Metabolic Flux Analysis (Collaborator: John Morgan, Purdue University)

## Engineering *Microchloropsis gaditana* to reduce dark loss for increased biomass production

- CRISPR/Cas9 editing for gene knockout
- CRISPRi (dCas12a) for gene knockdown

## Development of RNAi and CRISPRi tools for gene knockdown in *Rhodospiridium toruloides* (Agile Biofoundry)

- Improve terpene production



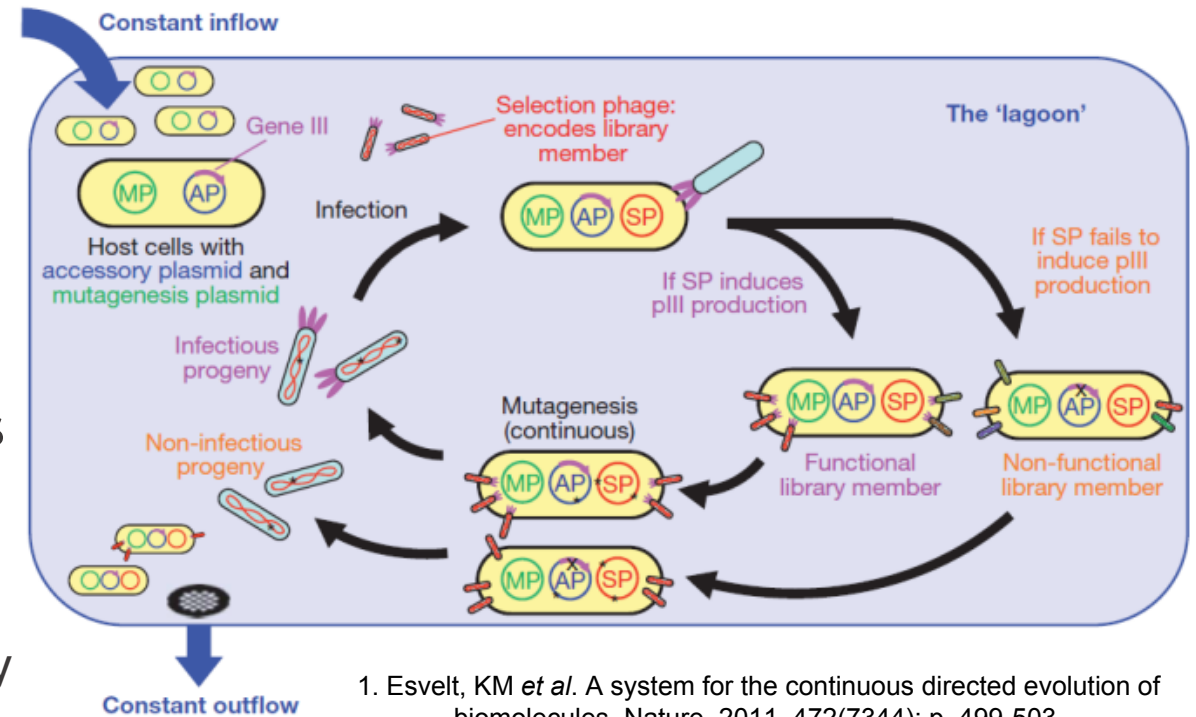


## Engineering noble gas biosensors using phage-assisted continuous evolution (PACE)

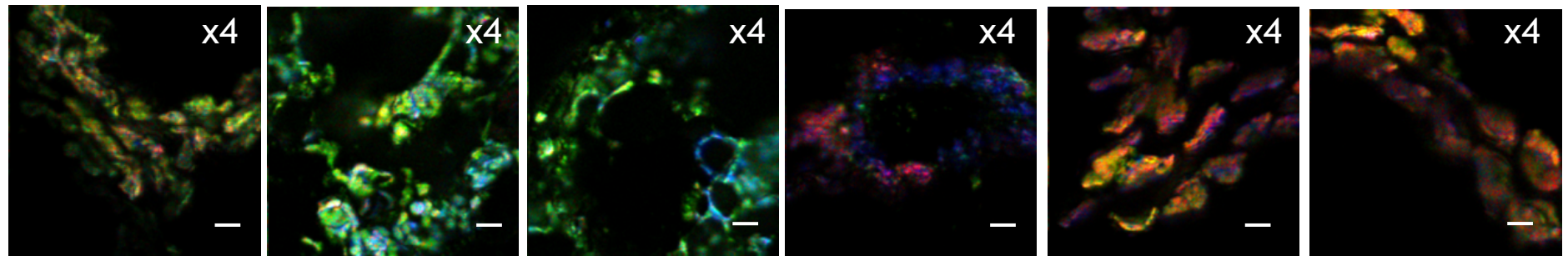
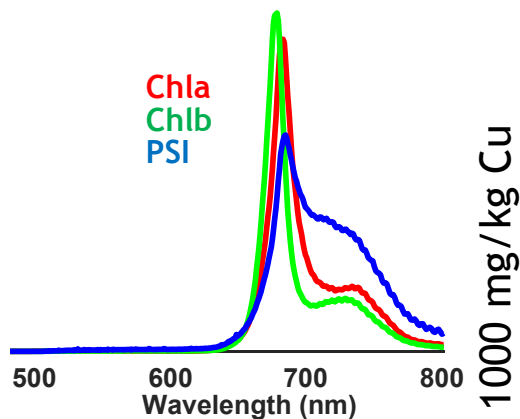
- Understanding mechanisms of noble gas interactions in *E. coli*
- PACE for rapid protein evolution (Collaborator: Ahmed Badran, Broad Institute)

## Using bioindicators to detect source emissions

- Hyperspectral imaging and multivariate curve resolution analysis to detect spectra in tall fescue
- Hyperspectral confocal fluorescence microscopy



1. Esvelt, KM *et al.* A system for the continuous directed evolution of biomolecules. *Nature*, 2011. 472(7344): p. 499-503.







# Sandia Biomanufacturing- the Agile BioFoundry

John Gladden ([jmgladd@sandia.gov](mailto:jmgladd@sandia.gov))

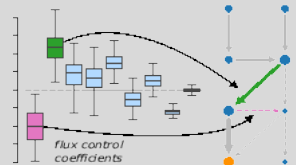
# ABF DBTL Infrastructure



Design



Sequence  
Validation



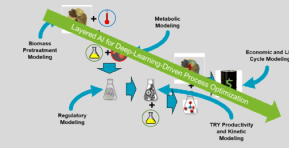
Bayesian Inference:  
Metabolic Kinetics



Kinetic Learning



Metabolic Modeling



Deep Learning

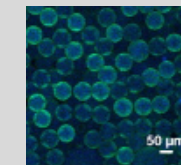
Learn



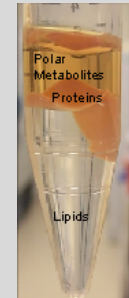
Build



EASy



Biosensors



Transcriptomics,  
Metabolomics,  
Proteomics, &  
Lipidomics



Test



EDD



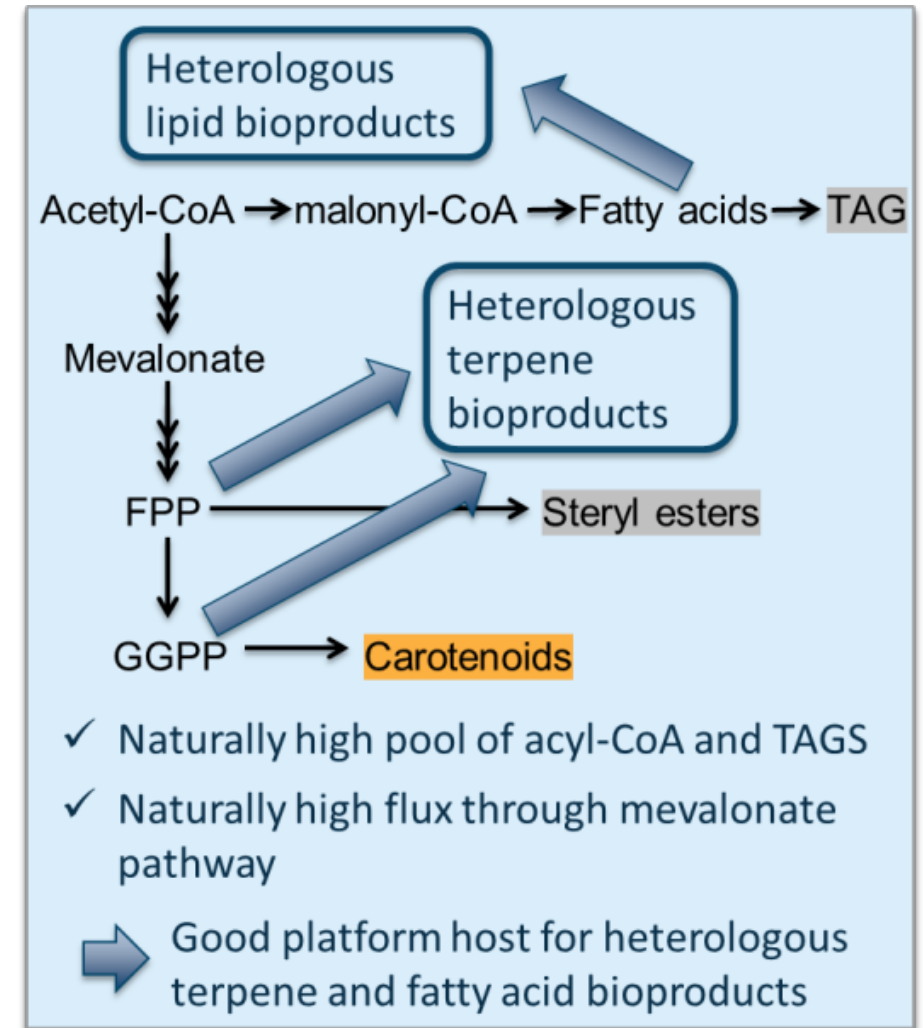
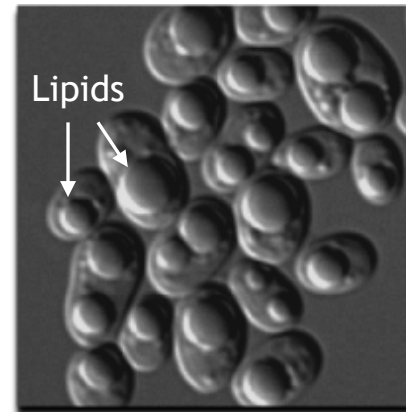
Data Quality  
Assessment

# ABF Host- *Rhodospiridium toruloides*



## Rhodospiridium toruloides

- Utilized lignocellulos
- Fast growing
- Oleaginous, carotenogenic
- Metabolically versatile
- Genetically tractable





- als





# Potential Partnerships



## **Research interests:**

- Developing non-model microbial biomanufacturing hosts
- Expertise in fungal and bacterial systems

## **Potential areas for partnership:**

- Characterization on non-model organisms (growth, physiology, genomics, metabolic models, etc.)
- Synthetic biology tool development
- Automation- development of high-throughput engineering methods
- Metabolic engineering for production of biofuels and bioproducts



# Biotechnology & Bioengineering

Ramdane Harouaka (rharoua@sandia.gov)

# Microfluidics & Microfabricated Platforms



## High-throughput screening

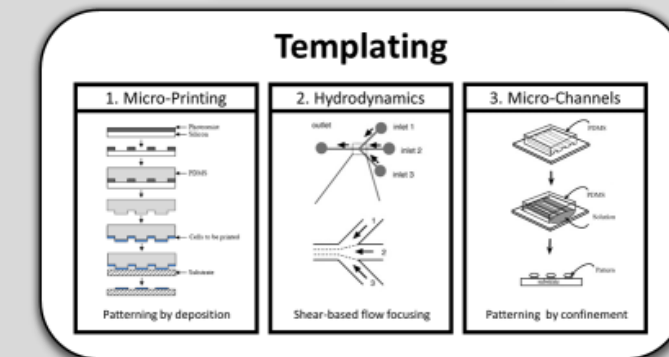
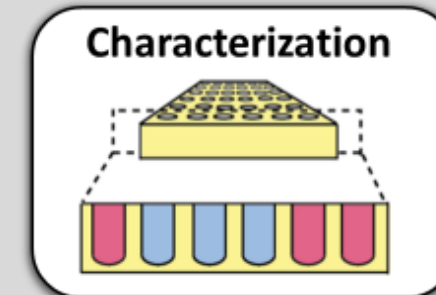
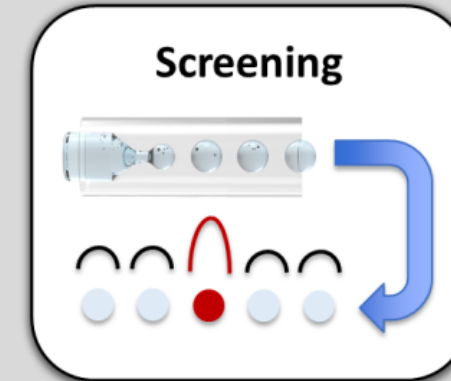
- Droplet assays for clonal identification/selection
- Optical, chemical, functional, proteomic, etc

## Single-cell Characterization

- Multi-species Combined / Dual RNA-Seq
  - Massively multiplexed probe libraries
  - Semirandom amplification
- Microwell platforms for combined Imaging + Transcriptomics

## Microfluidic Templating

- On-chip biosynthesis and packaging
- Fabrication of ordered structures







# Virus Engineering

Todd W. Lane | DMTS ([twlane@sandia.gov](mailto:twlane@sandia.gov))



## Target 1: Polinton Like Viruses

- Polintons are virus-like self-synthesizing transposons wide spread in the genomes of diverse eukaryotes
  - All encode a protein-primed DNA polymerase and retrovirus-like integrase
  - Most encode:
    - an ATPase and maturation protease homologous to enzymes of NCLDV
    - homologs of major and minor capsid proteins
- PLVs share characteristics and some core genes with both Polintons and virophages
- Tetraselmis striata* virus (originally *Tetraselmis viridis* virus)
- 31 kb genome dsDNA
- 60 nm Icosahedral virion
- 33 ORFs
- “Restore” the polymerase and integrase functions of the Polinton either by complementation in the host or engineering of the virus
- Remove either or both of the major and minor capsid proteins.
- Create a production host for viral packaging
- and a insertion competent host.

