

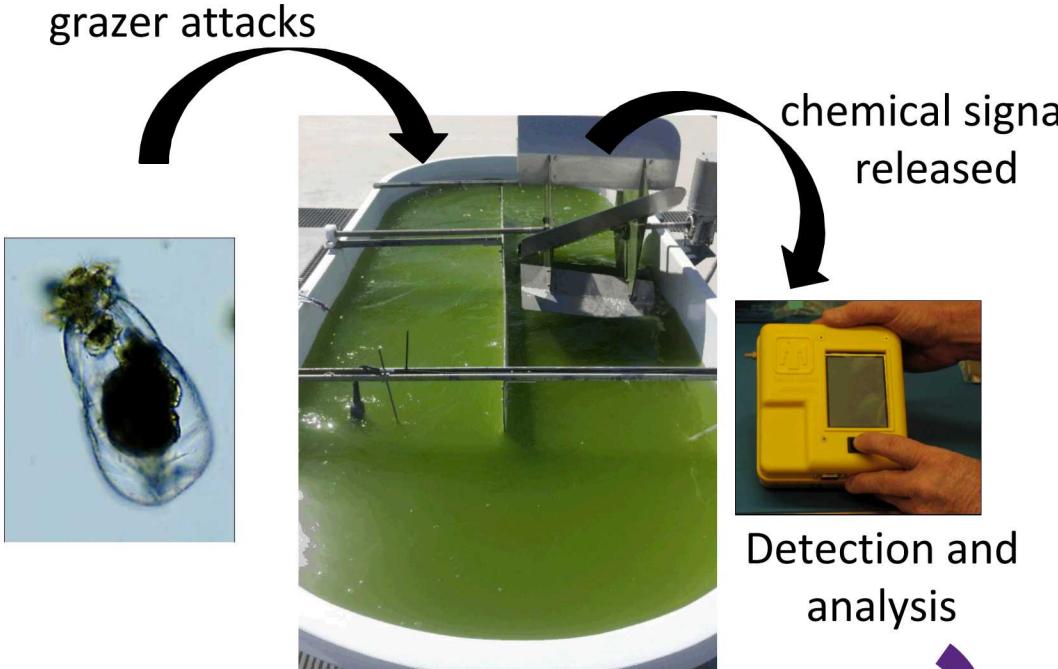
# Chemical and Genetic Detection Systems for Early Warning of Pond Crashes

**Carolyn L. Fisher, Kristen Reese, Kristina M. Mahan, Pamela D. Lane, James D. Jaryenneh, Marion Russell, Randy Maddalena, Matthias Frank, A. Daniel Jones,  
Todd W. Lane**

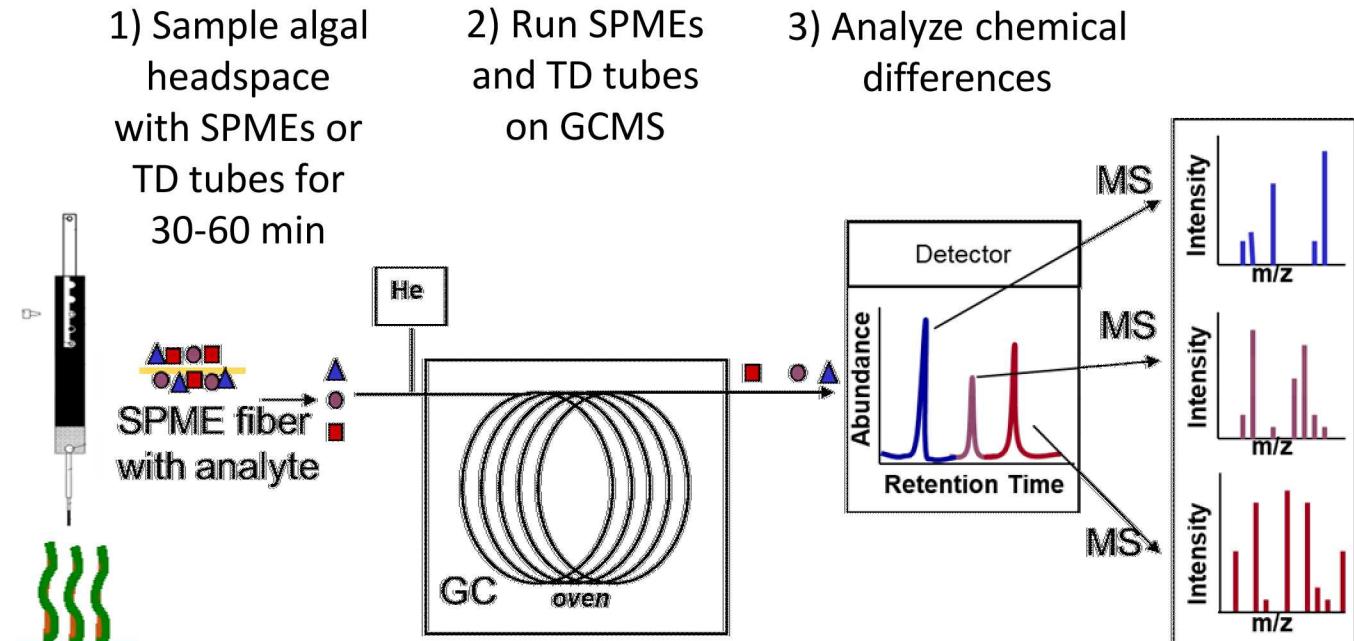
Distinguished Member of Technical Staff, Sandia National Laboratories

September 16, 2020 Algae Biomass Summit.  
The Ether

# Project Goal: Can we monitor chemical signals from algal ponds in order to determine pond health?

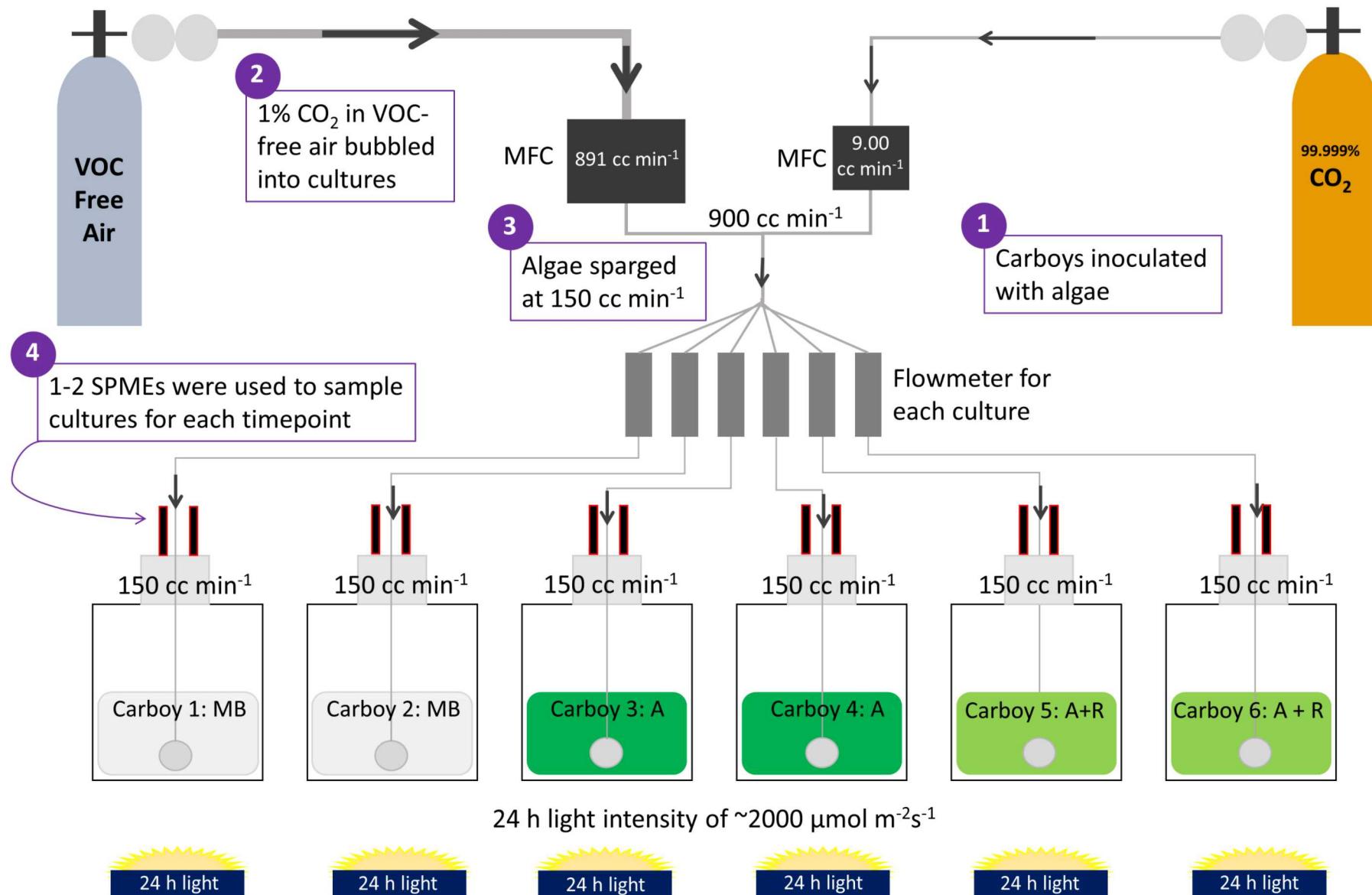


Early detection of algal grazers means earlier treatment and higher likelihood to save the pond.



**AVOCs sampling using SPMEs and Carbopack thermal desorption tubes → GCMS analysis**  
*Effort led by: Carolyn Fisher and Kristen Reese*

# We designed a system to effectively emulate an algal mass production system.



AVOCs = algal volatile organic compounds

AVOCs sampling using:

- SPMEs (solid phase microextraction fibers)
- Carbopack thermal desorption (TD) tubes
- GCMS metabolomics analysis using Agilent MassHunter and Mass Profile Professional.

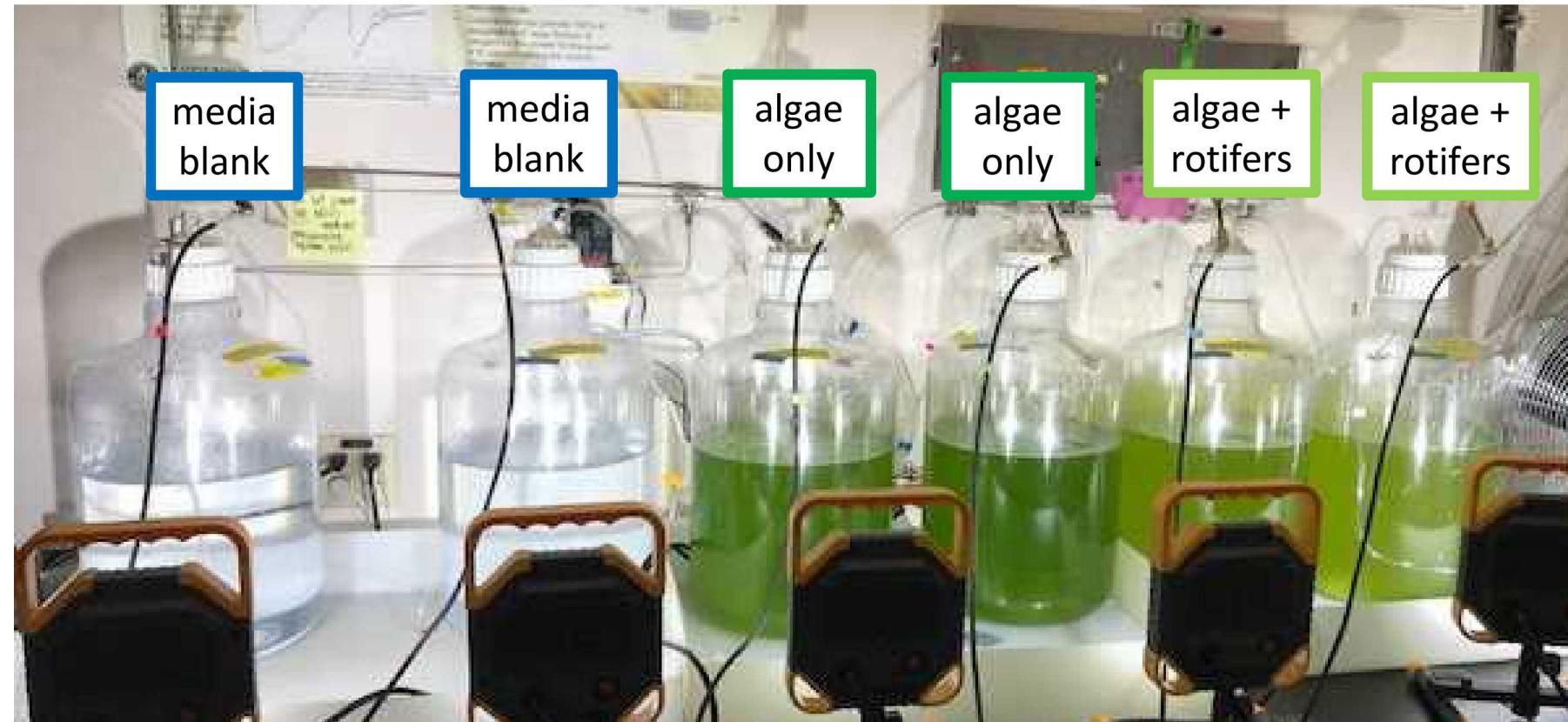
Reese KL, Fisher CL, Lane PD, Jaryenneh JD, Moorman MW, Jones AD, Frank M, Lane TW, Chemical Profiling of Volatile Organic Compounds in the Headspace of Algal Cultures as Early Biomarkers of Algal Pond Crashes, *Scientific Reports*, 9:13866 (2019). DOI: 10.1038/s41598-019-50125-z

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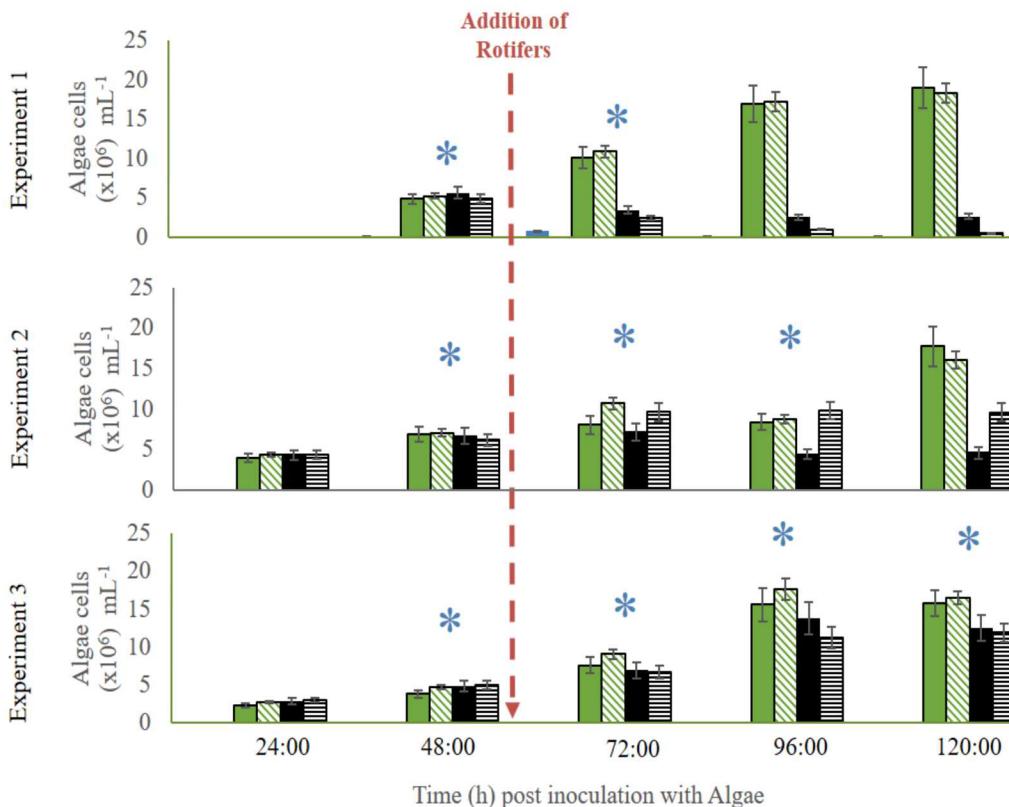
*In situ* sample collection under physiologically-relevant conditions via SPMEs and TD tubes.

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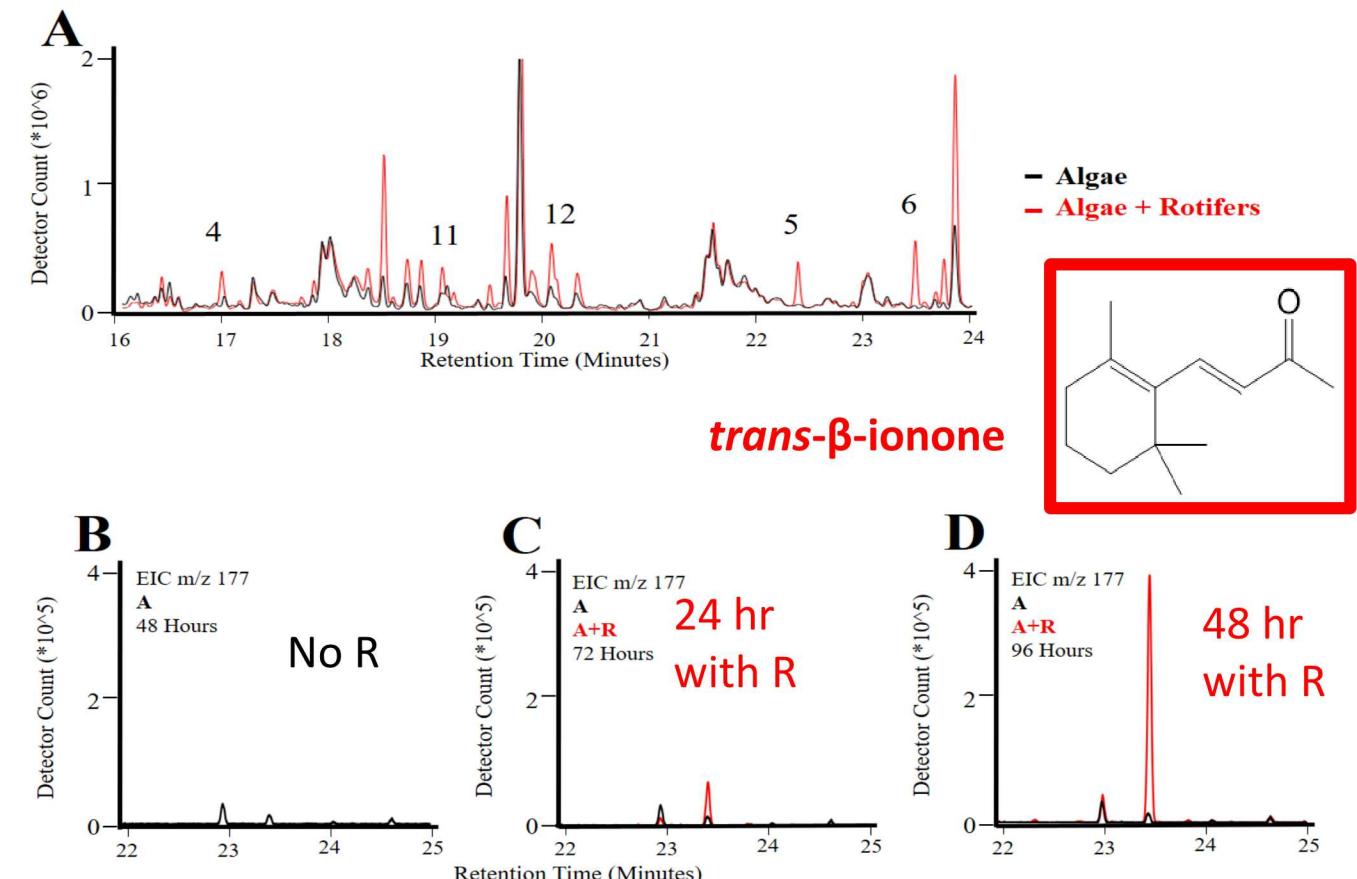
- Carboy 1: MB
- Carboy 2: MB
- Carboy 3: A
- Carboy 4: A
- Carboy 5: A+R
- Carboy 6: A+R

*Daily cell counts (M/mL) to track algal growth and productivity.*

\* SPME Sampling



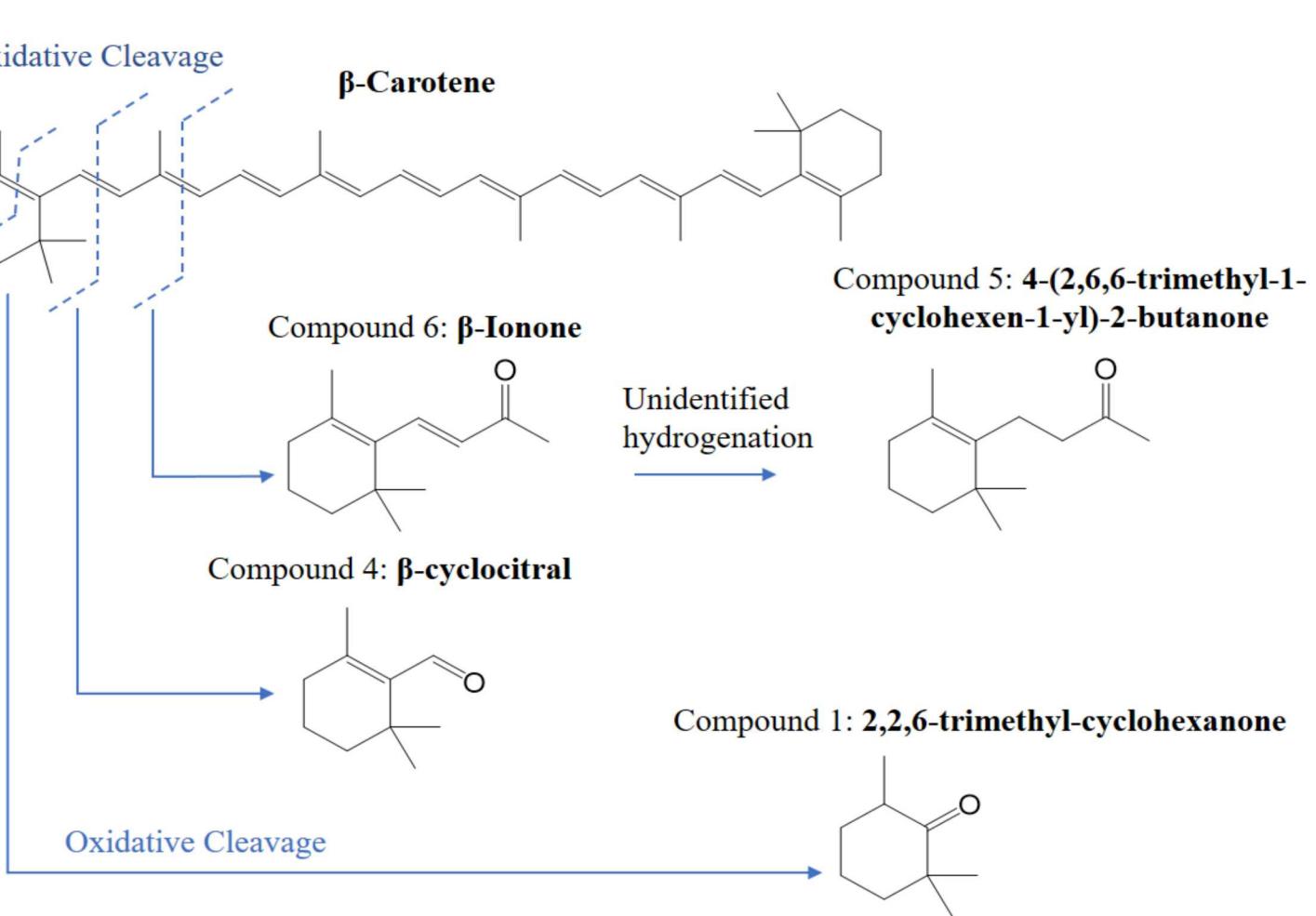
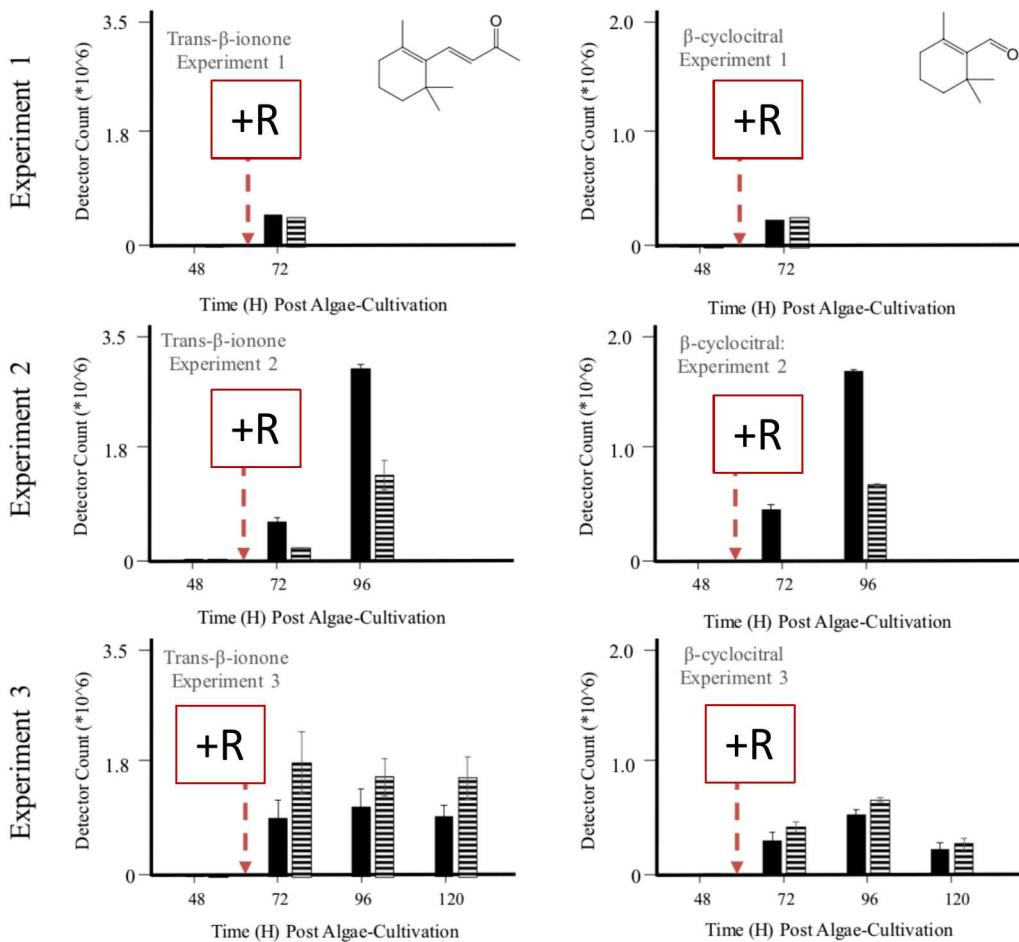
**RESULTS: Different chemical signals are detected for algae only vs. algae + rotifers cultures**



**Patent Pending:** Reese KL, Fisher CL, Lane TW, and Frank M are inventors on a 2019 pending application: Sandia Ref. No. SD14957.0/S162755, by Sandia National Laboratories, that covers the identification of several volatile organic chemical biomarkers for detecting predators in microalgae cultures, grown for biofuel production.

# In algae with rotifer samples, specific volatiles increase over time

CONCLUSION: Chemical signals are likely derived from oxidation of algal carotenoids

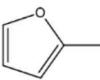


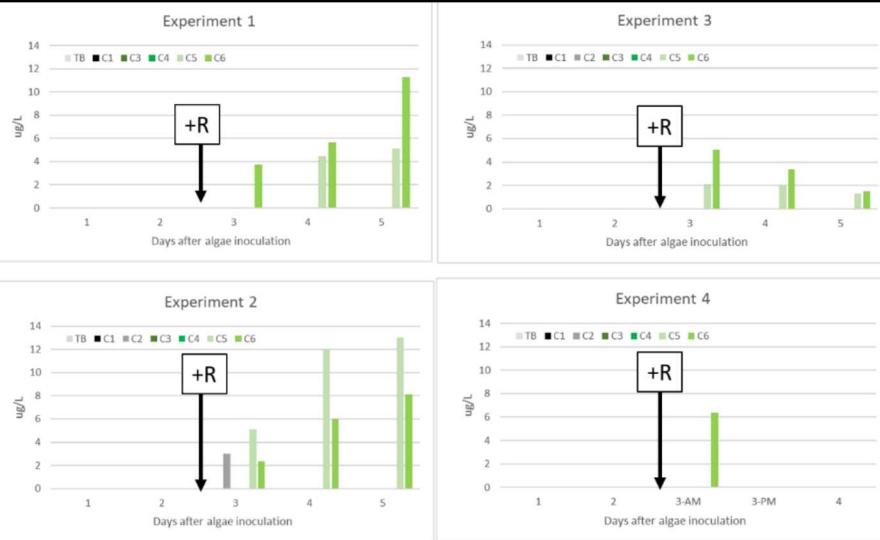
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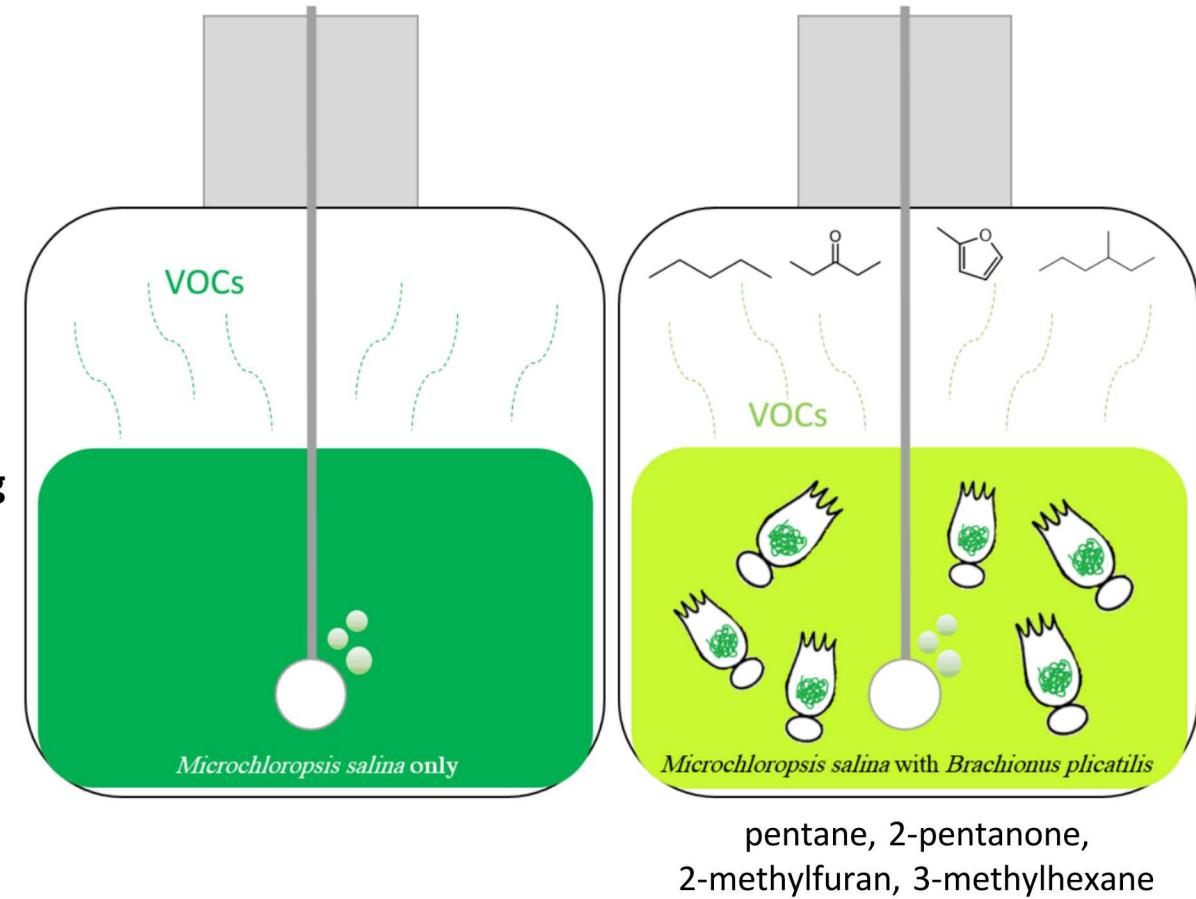
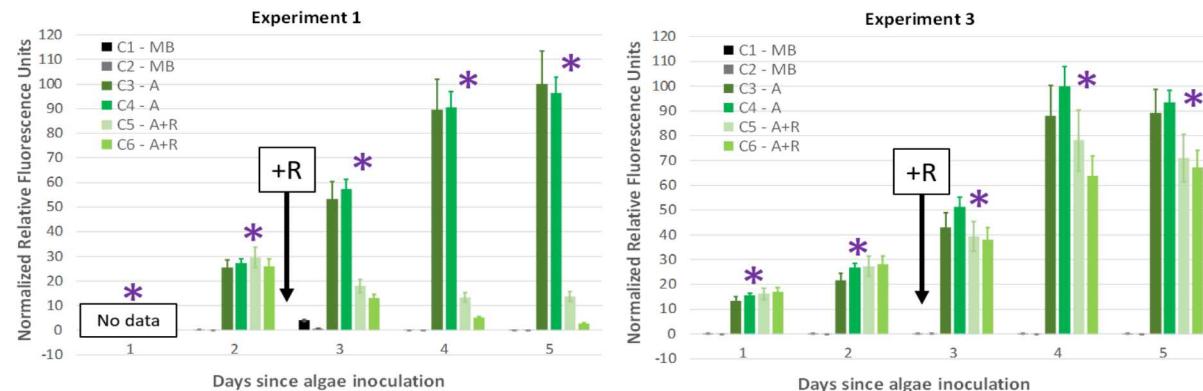
# RESULTS: Low MW VOCs detected by Carbopack B/X<sup>TM</sup> thermal desorption tubes differ for algae only versus algae + rotifer samples



**Compound 3:**  
 $m/z = 82$   
 r.t. = 10.409  
**NIST14 ID:** 2-methylfuran  
**NIST14 % :** 79.74  
**CAS:** 534-22-5  
**Molecular formula:** C<sub>5</sub>H<sub>6</sub>O  
**MW:** 82.10 g/mol  
**Structure:** 



Algal growth via chlorophyll fluorescence \*indicates Carbopack sampling

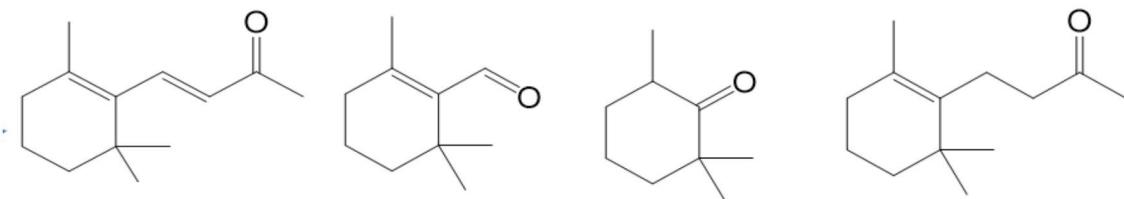


pentane, 2-pentanone,  
 2-methylfuran, 3-methylhexane

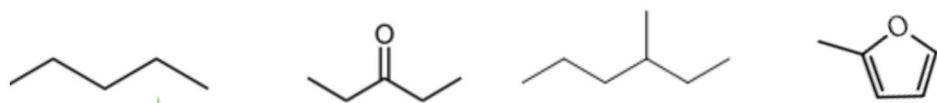
Fisher C.L., Russell M., Maddalena R., Lane, T.W. 2020. Low molecular weight volatile organic compounds (VOCs) indicate grazing by the marine rotifer *Brachionus plicatilis* on the microalgae *Microchloropsis salina*. *Metabolites* **10**, 361; doi:10.3390/metabo10090361

# Summary of the current AVOCs work:

- Several putative carotenoid oxidation products detected only from cultures of *M. salina* in the presence of active, grazing rotifers, *B. plicatilis*, by SPMEs (Reese KL, Fisher CL, *et al*, 2019)



- Low MW VOCs also identified from *M. salina* in the presence of actively grazing *B. plicatilis* using Carbopack B/X™ thermal desorption tubes (Fisher CL, *et al*, in revision 2020)

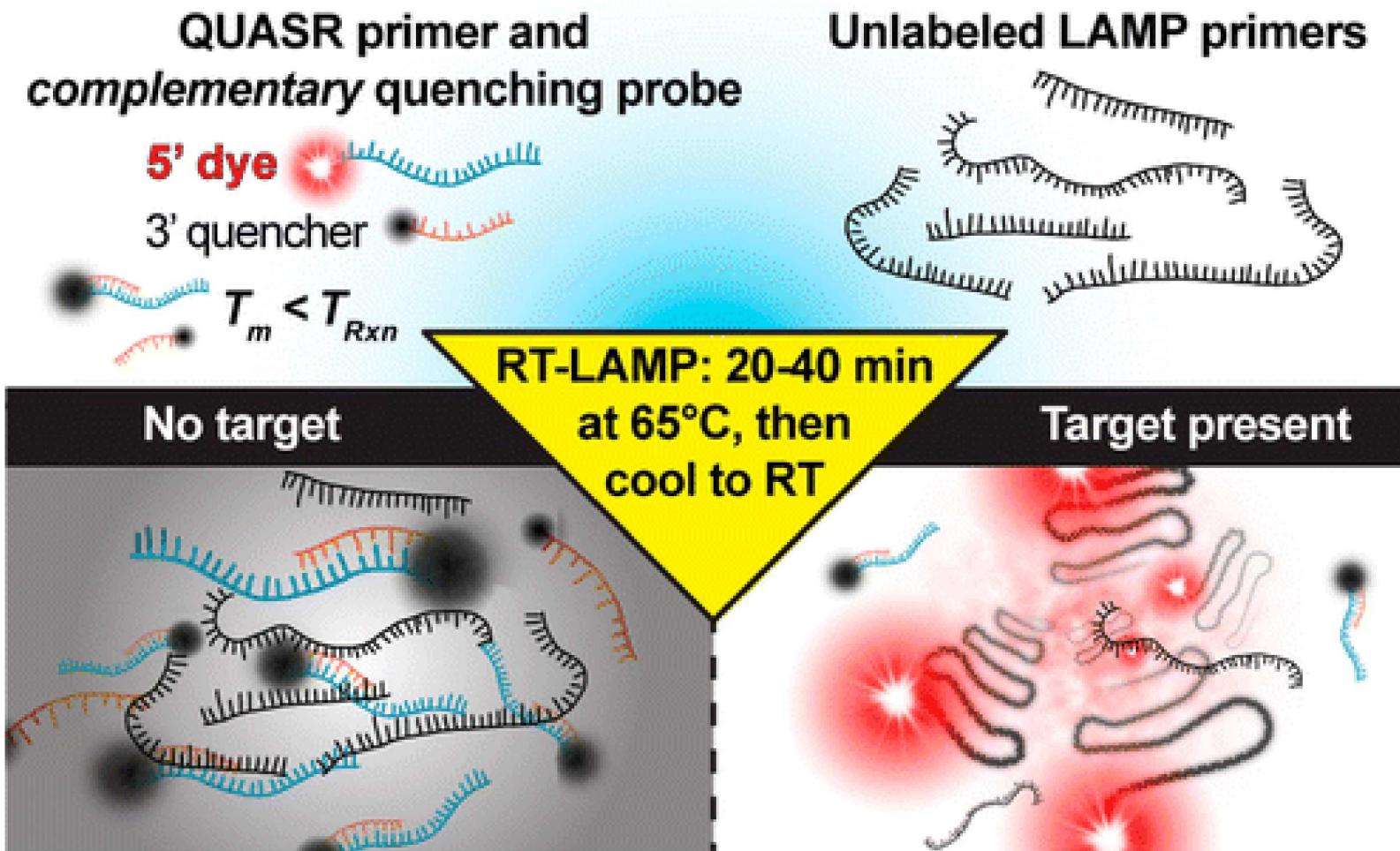


*Our success is an encouraging starting point for the discovery of volatile biomarkers of algal wounding.*

## FUTURE AVOCs work:

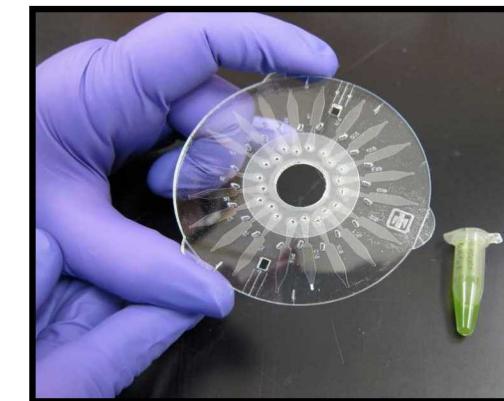
- Time course experiments
- Test additional algal strains of interest
- Test additional algal grazers, pathogens, etc
- Test different stress conditions (light, pH, salinity)
- Test different community structures (algae mono- vs. poly cultures, various bacterial cultures, etc)
- Targeted SIM-GC/MS analysis for mass algal cultures
- Collect VOCs from mass algal cultures “at scale”, such as outdoor mesocosms or raceways
- Take shorter timepoints to determine earliest detection
- Increase number of biological replicates
- Run longer experiments to determine how long chemical signals persist.

# Key Objective: Provide rapid detection of positive and negative members of the pond microbiome



Sandia Technology: Analytical Chemistry 2016, 88, 3562-3568

Pond-side Capability



Exceptional service. Exceptional results.



# PonDx QUASR Assays

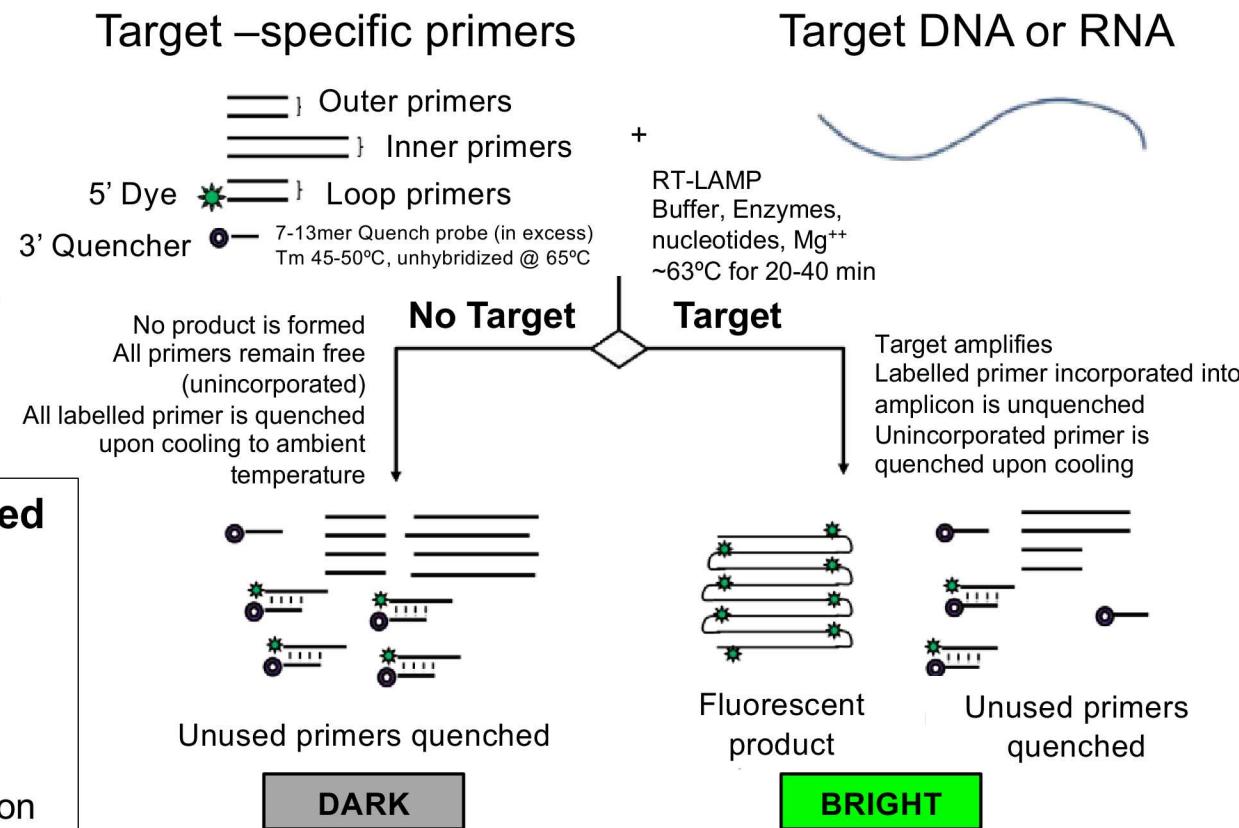
## Quenching of Unincorporated Amplification Signal Reporters

Sandia Technology: Analytical Chemistry 2016, 88, 3562-3568

- Six primers allow for enhanced target specific detection
- Assays are fast < 40 min
- Amplification of target allows for very sensitive detection
- Can be multiplexed if using a plate reader
- Can be monitored using the pond-side diagnostic equipment: PonDx

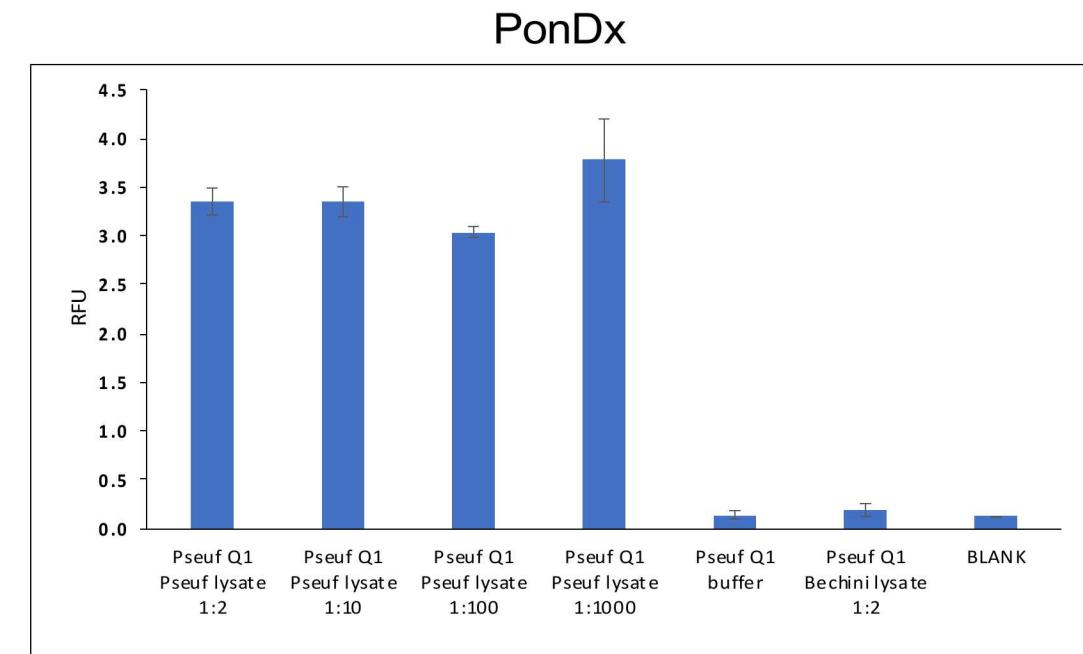
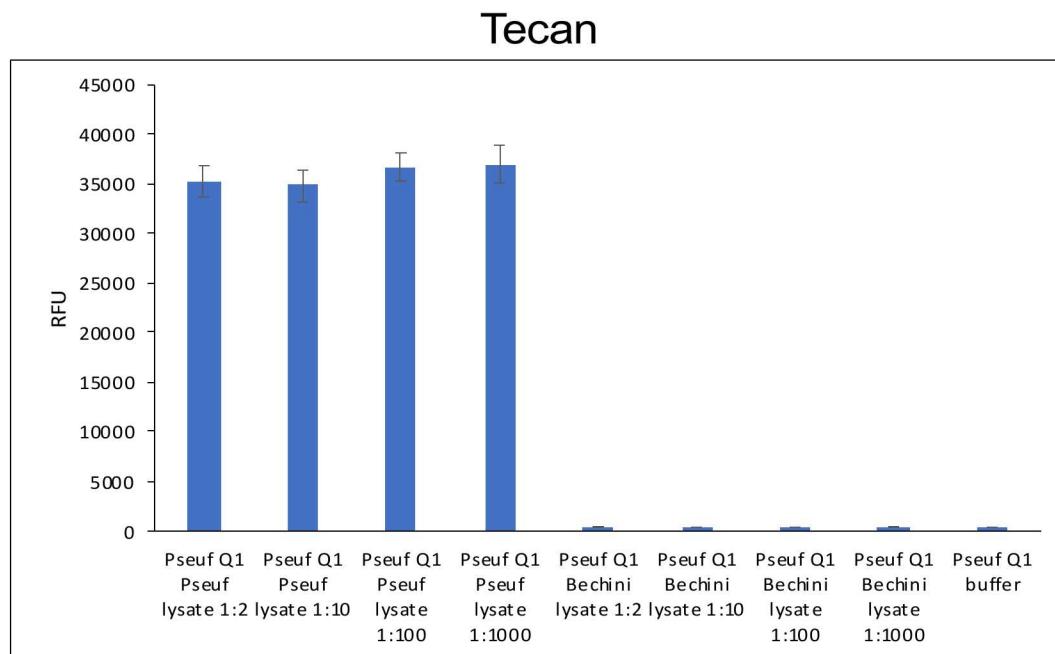
**Assay reagents are pre-mixed and provided to GAI staff so that protocol is:**

1. 9  $\mu$ l assay mix + 1  $\mu$ l pond lysate
2. Heat in plate reader or on PonDx @ 63°C, 20-40 min
3. Cool to room temperature
4. Detect fluorescence using plate reader or on PonDx box



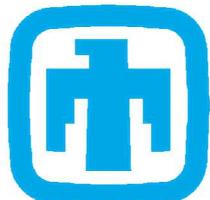
# Assays for the detection of algal pond microbiota

Assays can be performed on a plate reader with temperature controls or on the PonDx box



QUASR Probe designed to detect organism of interest: *Pseuofulvimonas gallinarii*  
Detected on Tecan plate reader Ex 532 nm Em 571 nm and PonDx box

# Thank you!!



**Sandia  
National  
Laboratories**

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Pamela Lane

Kristen Reese

James Jaryenneh

Matt Moorman

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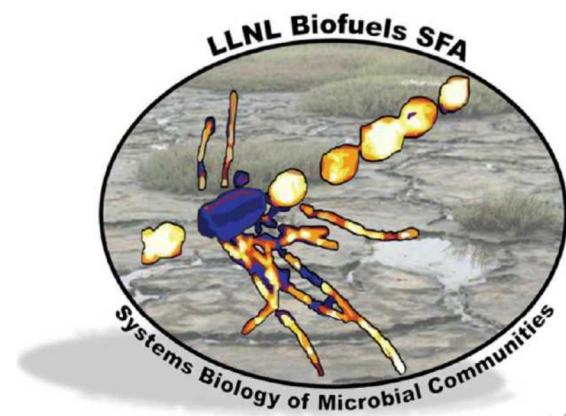
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Exceptional service in the national interest

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