

CINT

SAND2018-8094PE

The Center for Integrated Nanotechnologies

Nanomaterials

Integration

A U.S. DOE Nanoscale Science Research Center

CryoEM for fundamental study of soft/bio and inorganic nanomaterials

Sun Hae Ra Shin, Ph.D.,
Center for Integrated Nanotechnologies (CINT)
Sandia National Laboratories

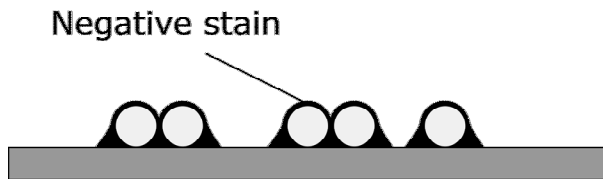
Interview Seminar
August 29th, 2018

Sandia is a multimission laboratory managed and operated by National Technology and Engineering Solutions of Sandia, LLC, a wholly owned subsidiary of Honeywell International, Inc., for the U.S. Department of Energy's National Nuclear Security Administration under contract DE-NA-0003525

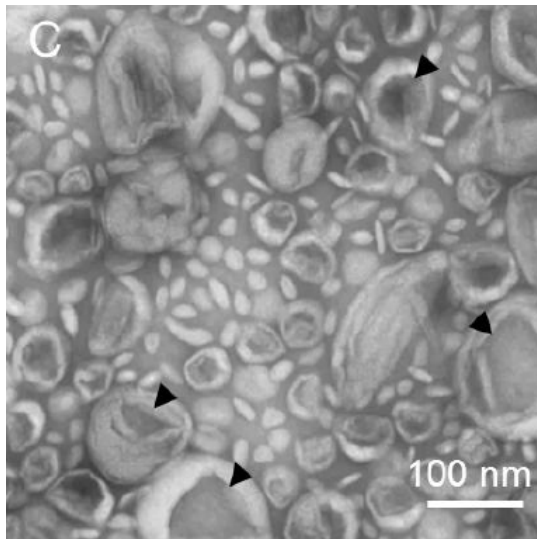


What is cryo electron microscopy (EM)?

Stained specimen
for conventional EM

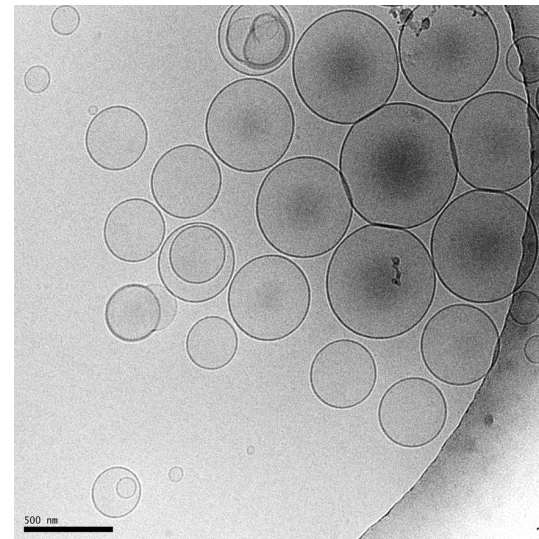
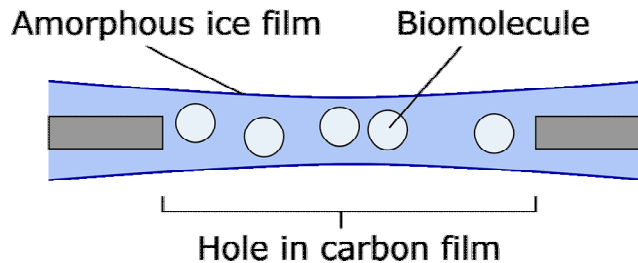


Liposome



Polymers 9, 521 (2017)

Vitrified specimen
for cryoEM



Liposome

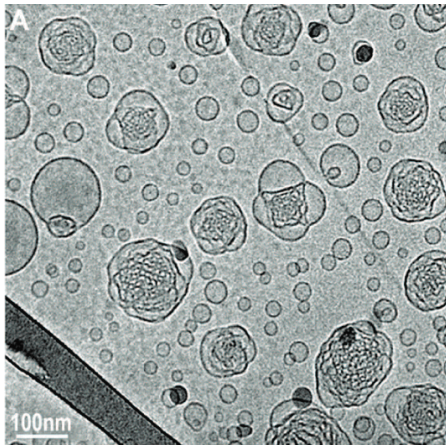
www.cicbiogune.es



What is cryoEM used for?

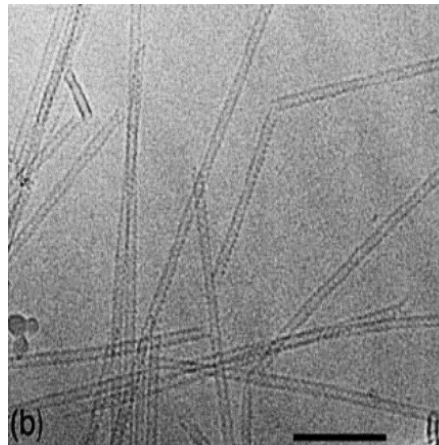
Analyzing complex, flexible structures in their native, hydrate states

Lipid superstructure



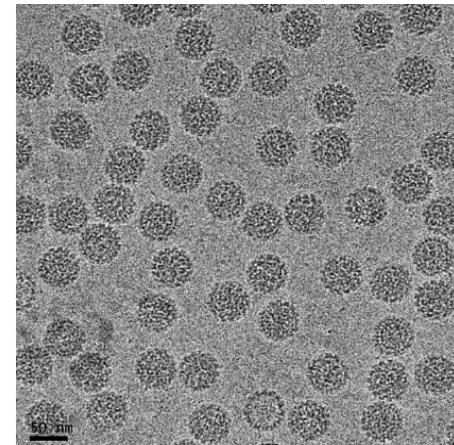
Langmuir 25, 1316-1326 (2009)

Peptide nanotube

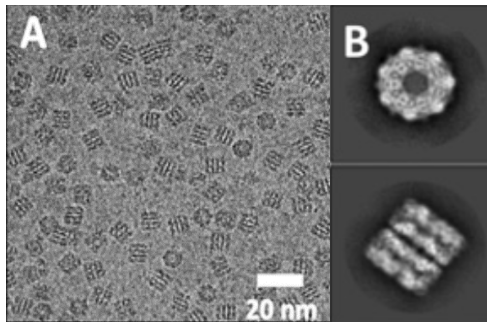


Structure 23, 280-289 (2015)

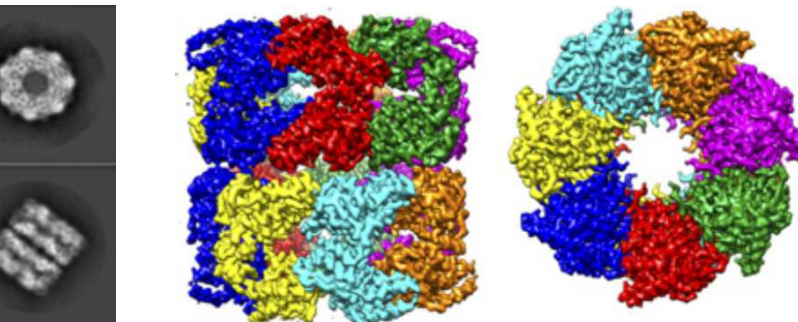
Human papillomavirus



www.phys.org



GroEL protein



PNAS 114, 8259-8264 (2017)



Overview

Part 1. Self-assembly of inorganic nanoparticles in solution

Part 2. Interaction between inorganic-organic materials

Part 3. Research plan



Part 1. Self-assembly of inorganic nanoparticles in solution

Part 2. Interaction between inorganic-organic materials

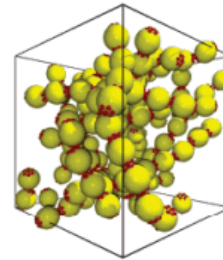
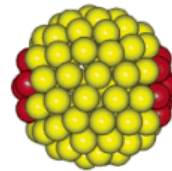
Part 3. Research plan



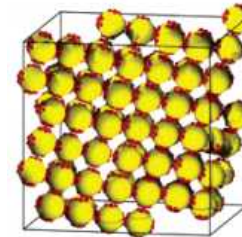
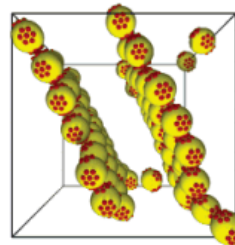
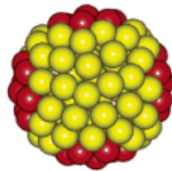
Self-assembly of patchy particles

Directed self-assembly of particles with fixed patches

$n=2$



$n=4$

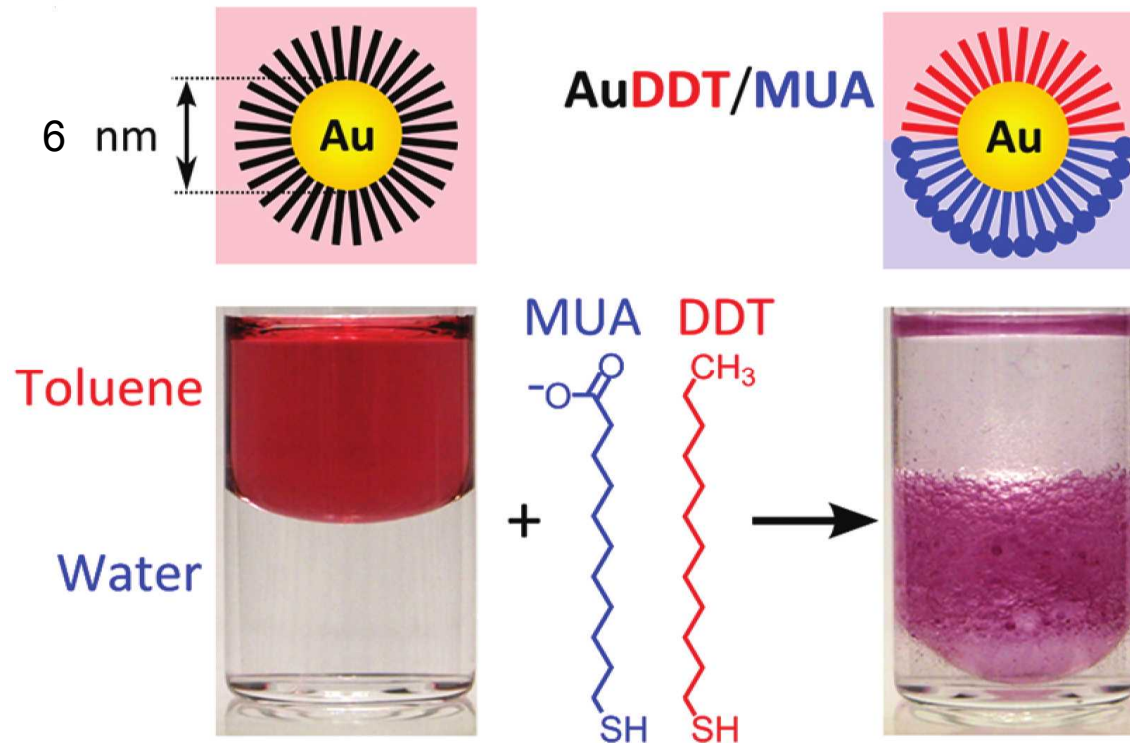


Nano Lett., 4,1407-1413 (2004)

What happen if the patches rearrange on the particles' surface?



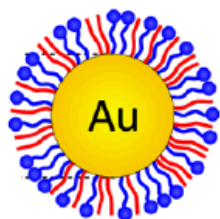
Amphiphilic nanoparticles (NPs) with adaptive surface chemistry



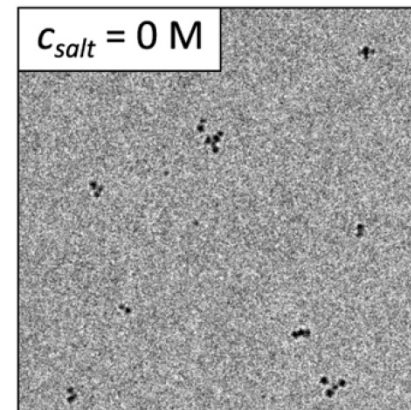
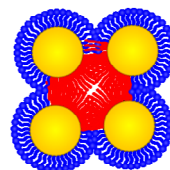
Andala, Shin et al. *ACS Nano*, 6,1044-1050 (2012)



Self-assembly of same NPs into small cluster and chain

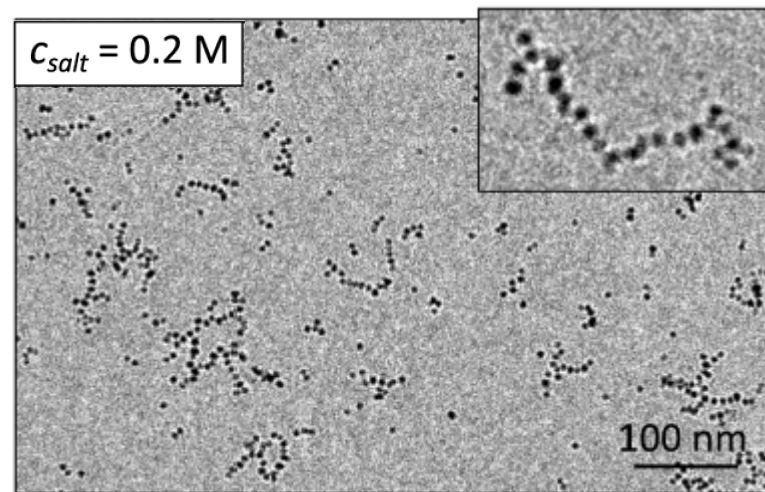
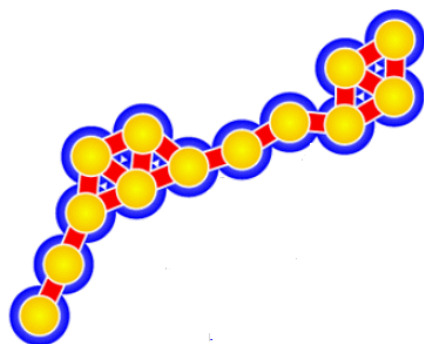


Small cluster



CryoEM

Chain-like structure



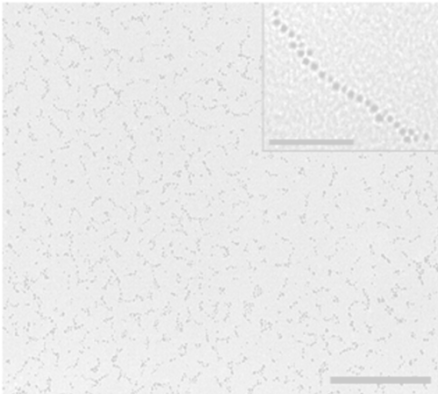
CryoEM

Lee, Shin et al. *ACS Nano*, 8,9979-9987 (2014)



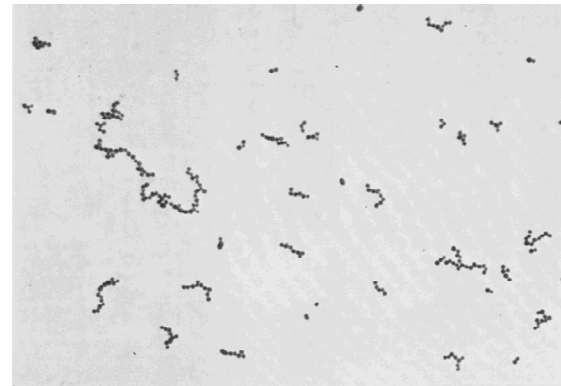
Several routes to particle chains

Crosslinking divalent NPs



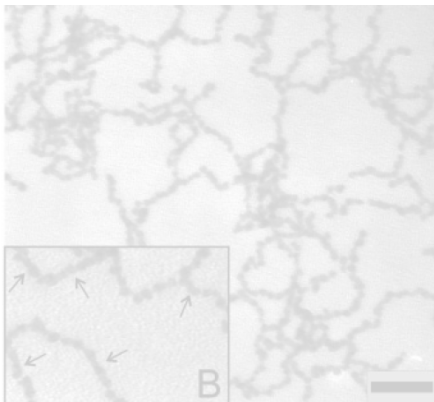
Science 315, 358-361 (2007)

Diffusion-limited aggregation



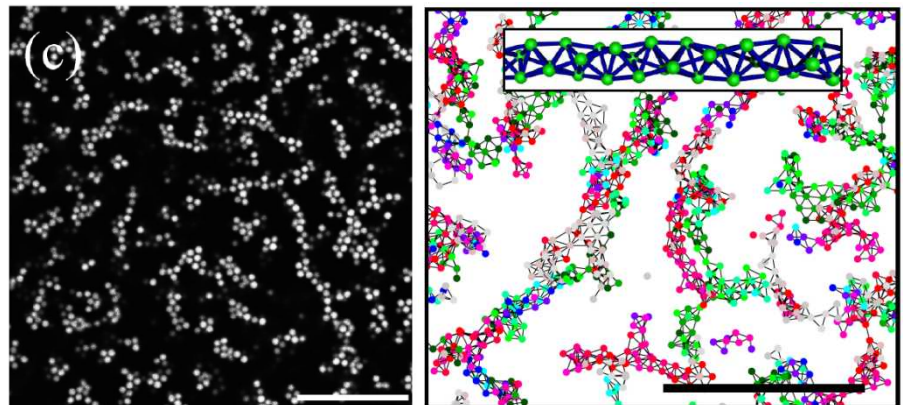
JACS. 85, 3317 -3328 (1963)

Dipolar interactions



Science 297, 237-240 (2002)

Short-ranged attraction, long-ranged repulsion



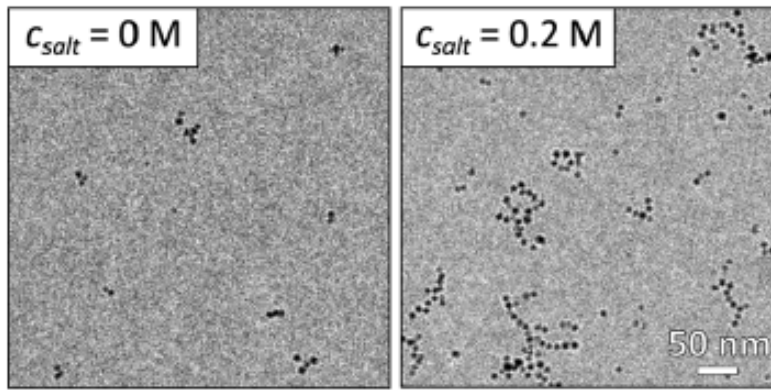
Phys. Rev. Lett. 94, 208301 (2005)



Chains form only in salty conditions

(a)

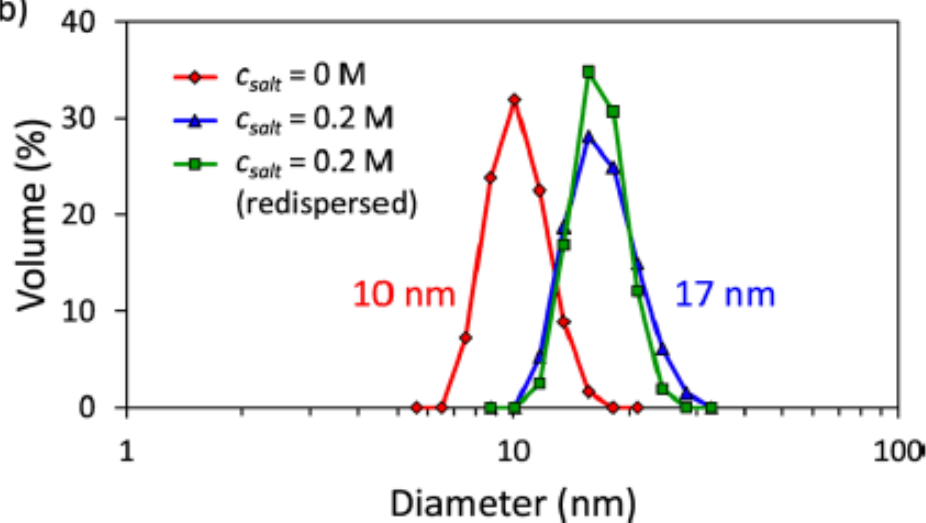
CryoEM



Short-ranged interactions

$$\kappa^{-1} = \sqrt{\frac{\epsilon\epsilon_0 k_B T}{2e^2 c_{salt}}} < 1 \text{ nm}$$

(b)

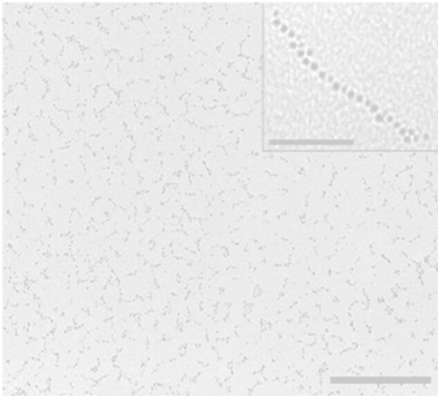


Equilibrium structures
(not kinetic aggregates)



Several routes to particle chains

Crosslinking divalent NPs



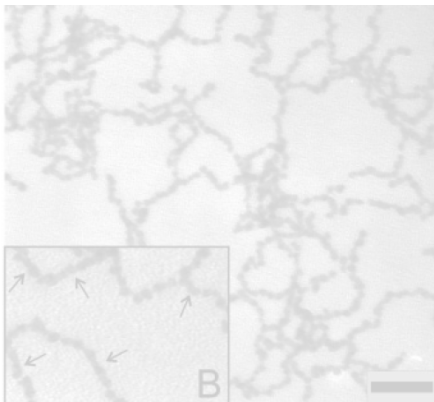
Science 315, 358-361 (2007)

Diffusion-limited aggregation



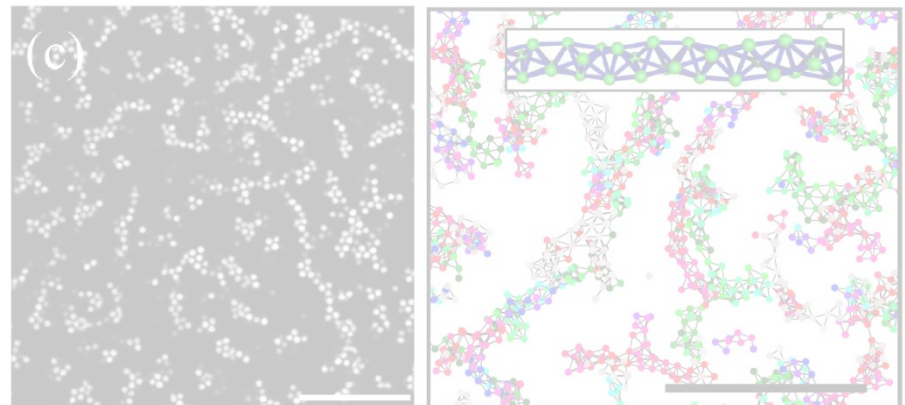
JACS. 85, 3317 -3328 (1963)

Dipolar interactions



Science 297, 237-240 (2002)

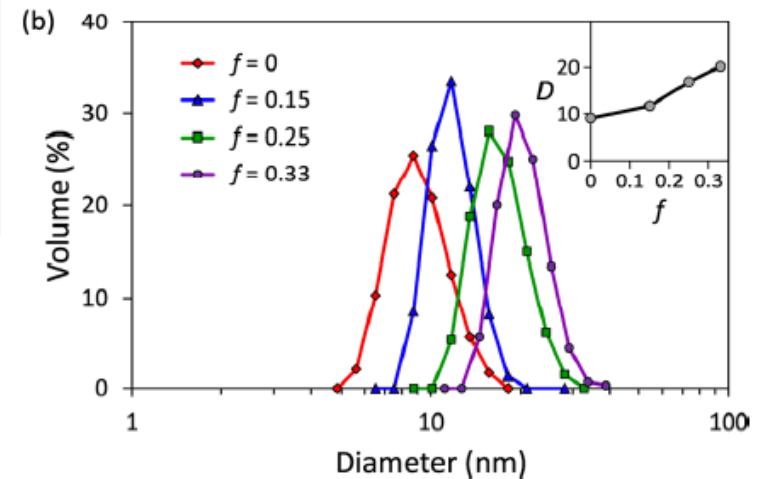
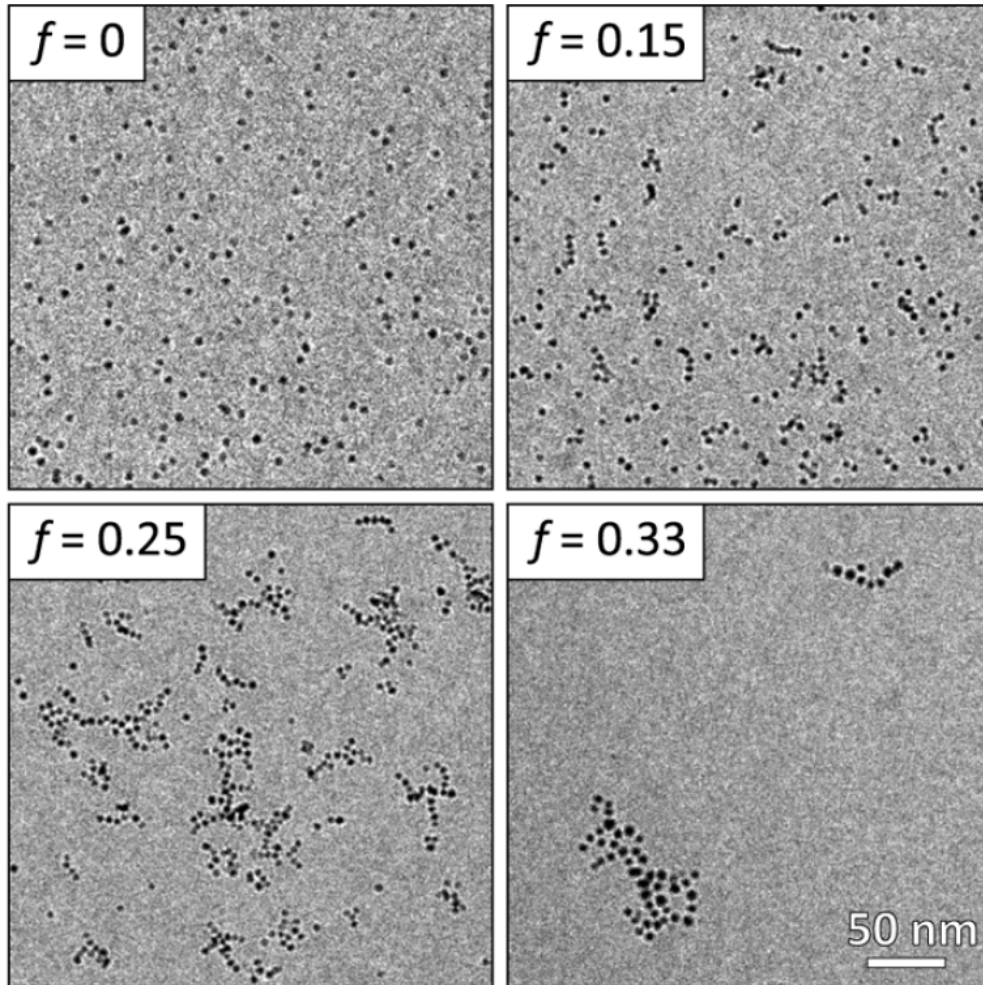
Short-ranged attraction, long-ranged repulsion



Phys. Rev. Lett. 94, 208301 (2005)



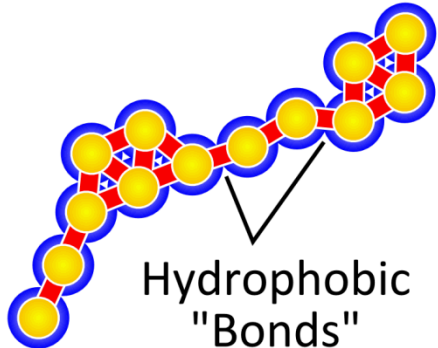
Amount of hydrophobic ligand (f)



CryoEM, $C_{salt} = 0.2$ M



Adaptive hydrophobic bonding model



- Short-ranged interactions (electrostatic and hydrophobic)
- Dynamic rearrangement of ligands

$$E_{hp} = \varepsilon_{hp} \sum_{i=1}^{N_p} H_{ii}$$

Minimize

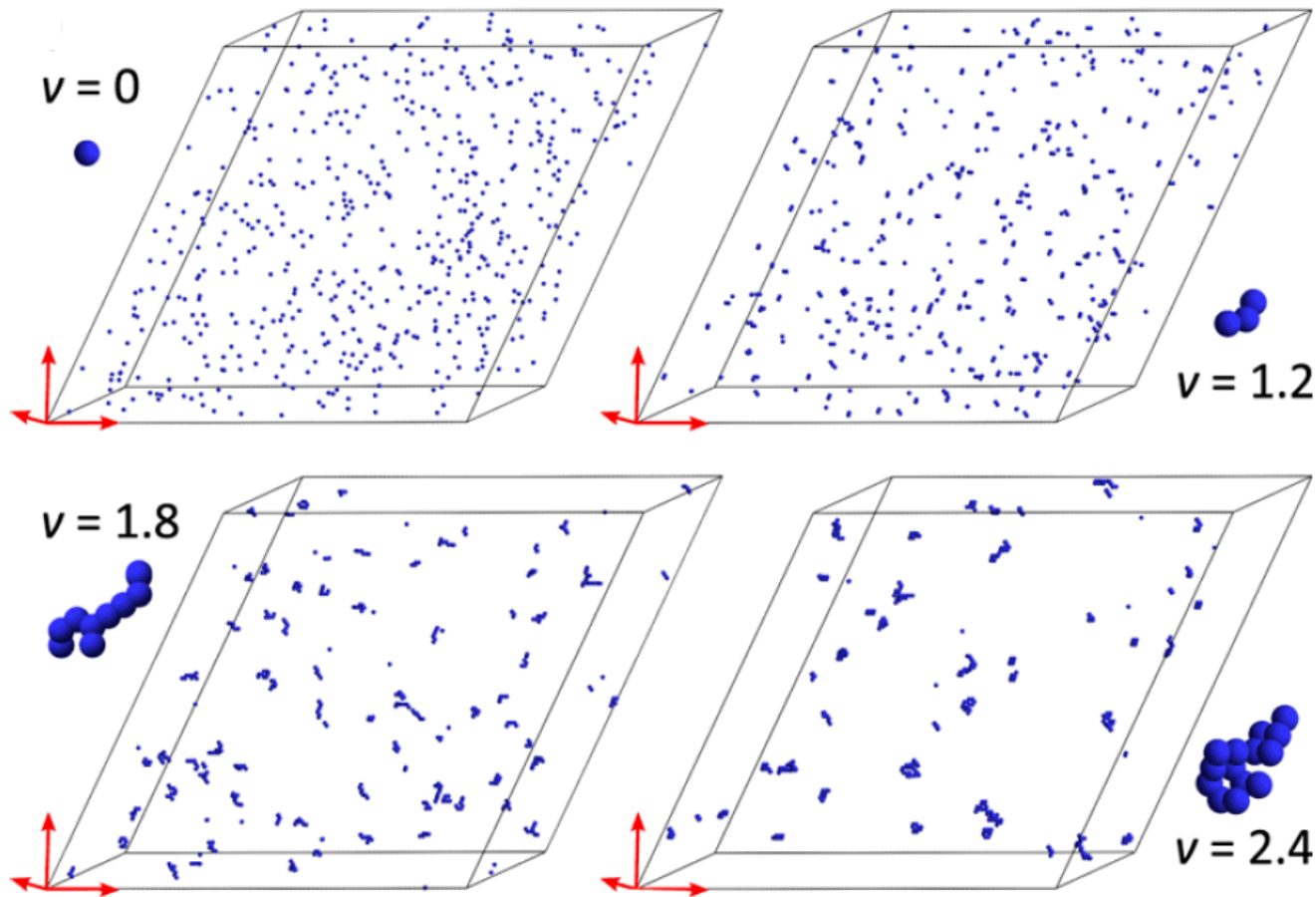
ε_{hp} Energy due to contact between a hydrophobic ligand and water

N_p Total number of particles

H_{ii} Number of “nonbonding” hydrophobic ligands



3D Monte Carlo simulation

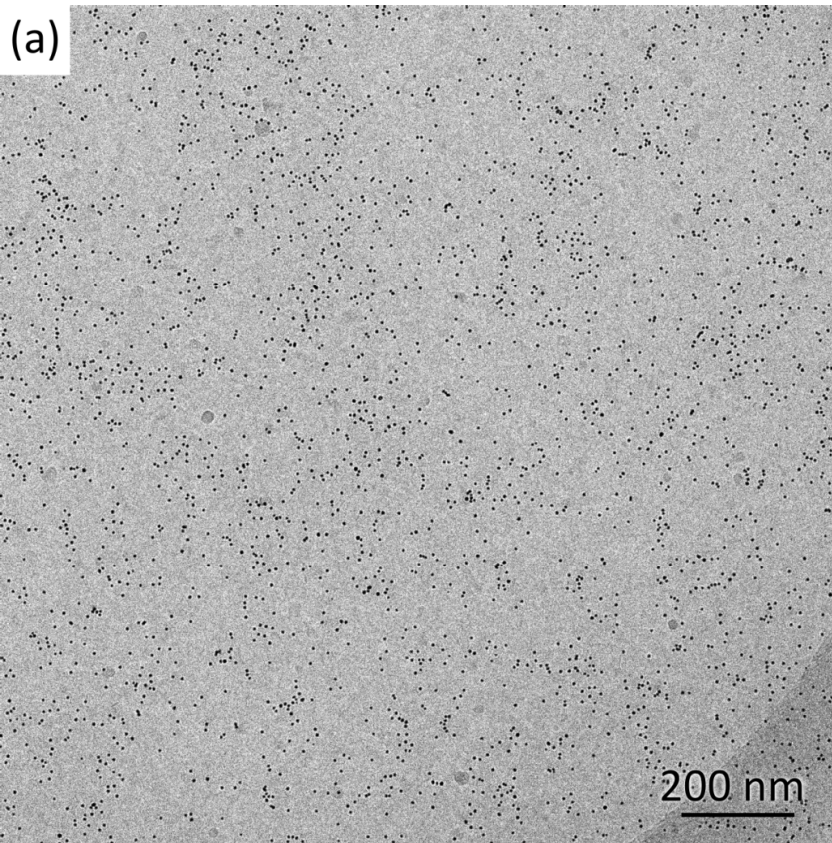


ν corresponds to f (hydrophobic coverage)

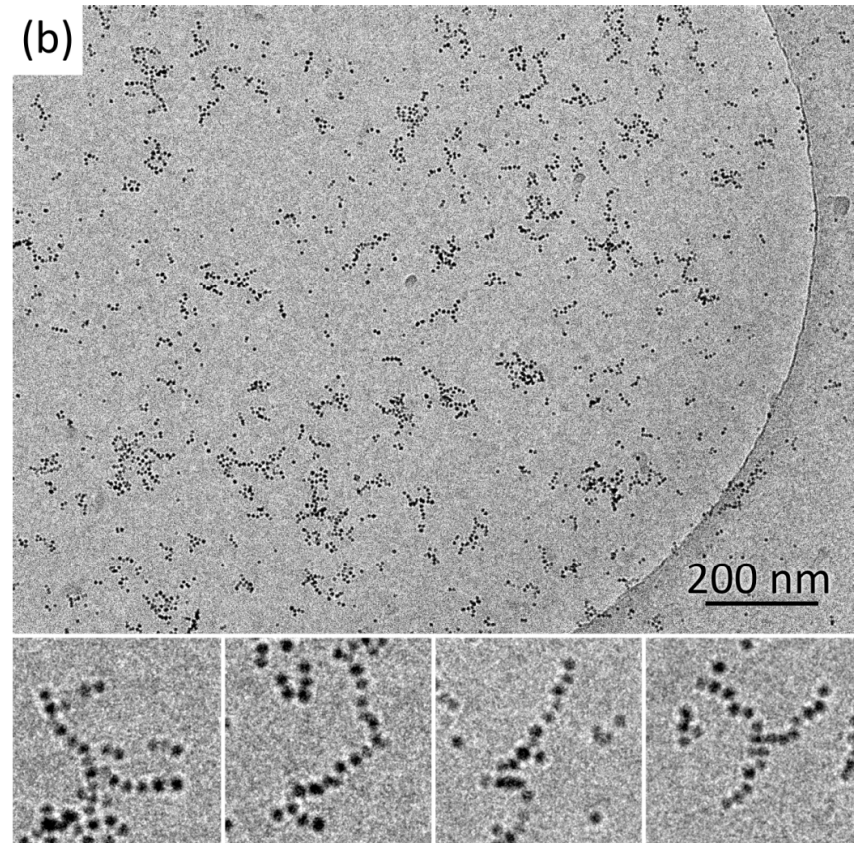


Time scale of ligand reorganization

~ 1 min



~24 h

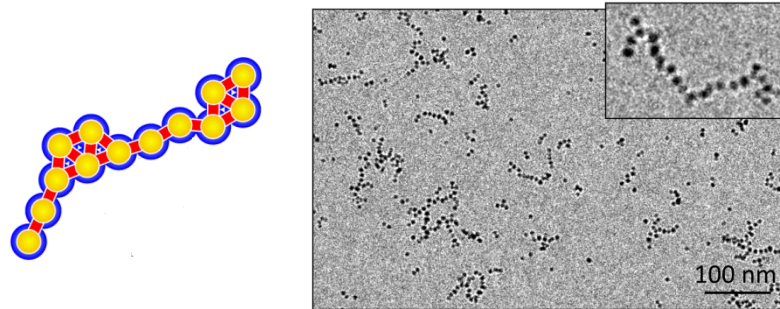


CryoEM, $C_{\text{salt}} = 0.2 \text{ M}$, $f = 0.25$



Conclusion

Part 1. Self-assembly of inorganic nanoparticles in solution



- Formation of chain structure via short-ranged electrostatic/hydrophobic interaction
- Dynamic rearrangement of ligands
- 3D simulations capture experimental trends



Part 1. Self-assembly of inorganic nanoparticles in solution

Part 2. Interaction between inorganic-organic materials

2.1. Selective binding of nanoparticles to open edges

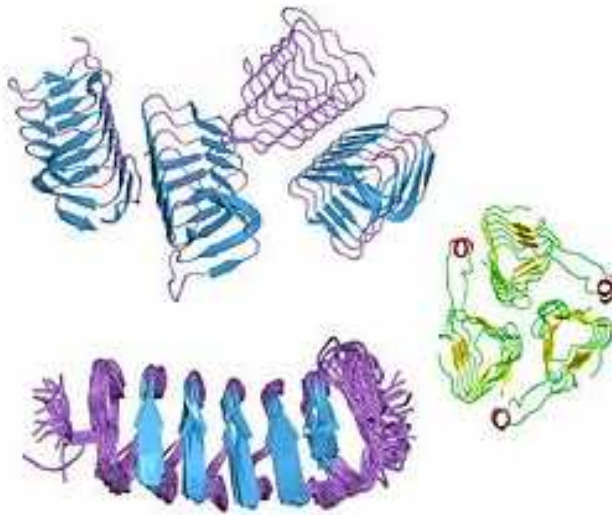
2.2. Formation of nanoparticle-supported bilayers

Part 3. Research plan

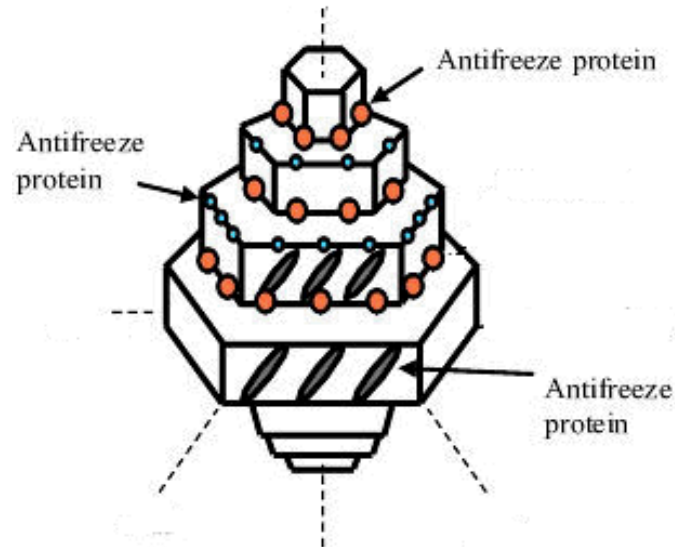




Antifreeze proteins as kinetic capping agents



Antifreeze protein



Ice crystal

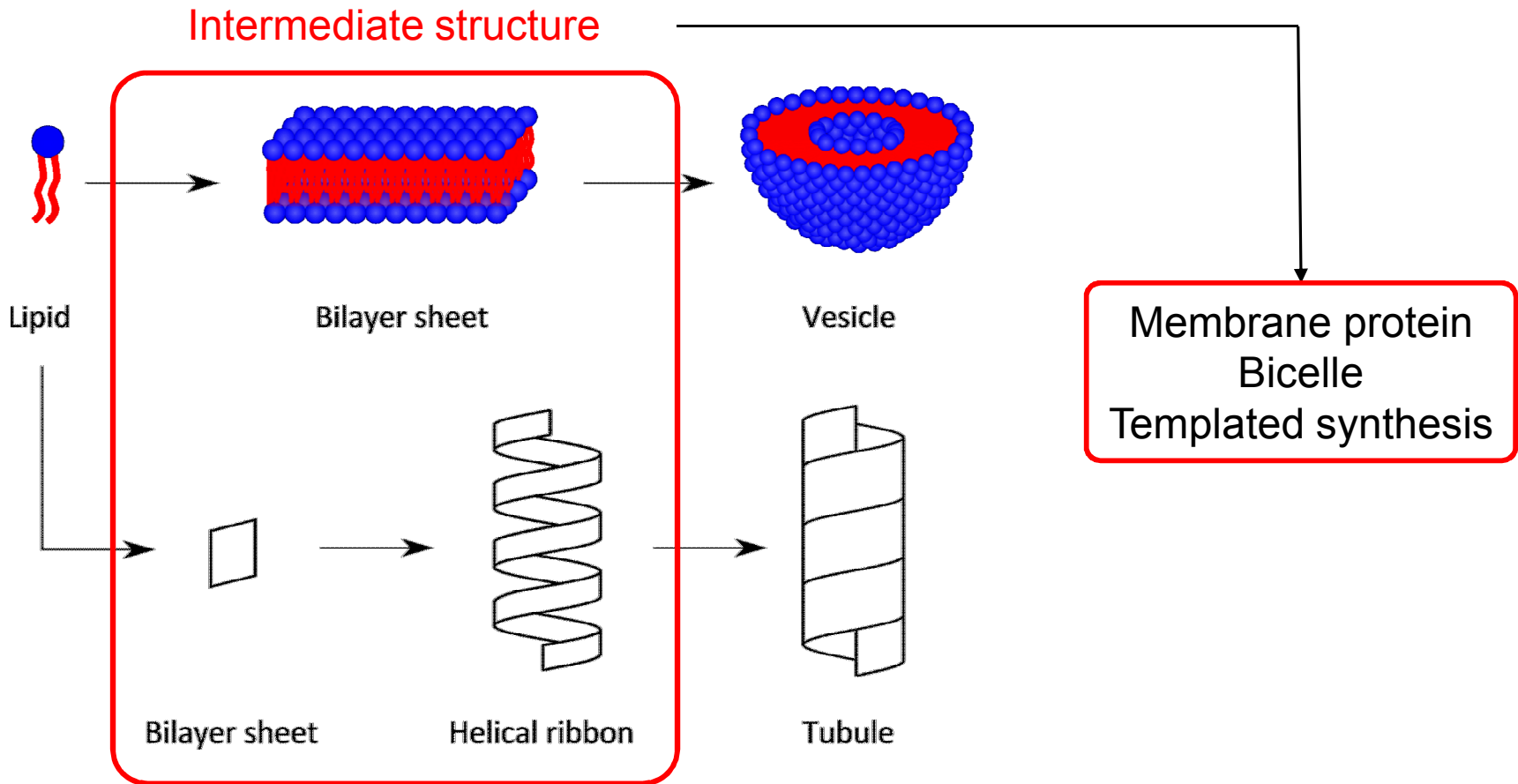


SKIM MILK, SUGAR, CORN SYRUP, POLYDEXTROSE, MALTODEXTRIN, PROPYLENE GLYCOL MONOESTERS, MONO AND DIGLYCERIDES, CELLULOSE GUM, CREAM*, CAROB BEAN GUM, GUAR GUM, NATURAL FLAVOR, CARRAGEENAN, ICE STRUCTURING PROTEIN, VITAMIN A PALMITATE, ANNATTO (FOR COLOR) * ADDS A DIETARILY INSIGNIFICANT AMOUNT OF FAT. Ingredients and Nutrition Facts are current as of 3/1/13. Please see shelf packaging for any changes. Nutrition Facts may vary in high altitude areas.

Chem. Rev., 96, 601-618 (1996)
www.wikipedia.org; www.intechopen.com

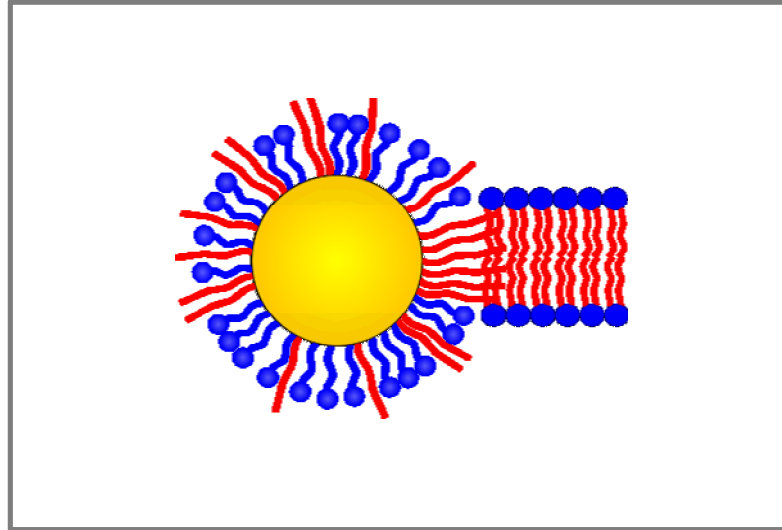


Intermediate structures with open edges





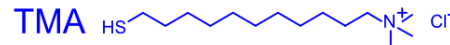
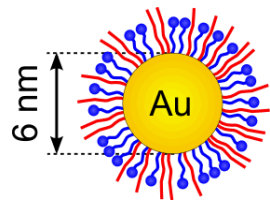
NP amphiphiles bind selectively to open edge of bilayers



- 1) Controlling growth of lipid tubules
- 2) Inhibiting the formation of gallstone precursors

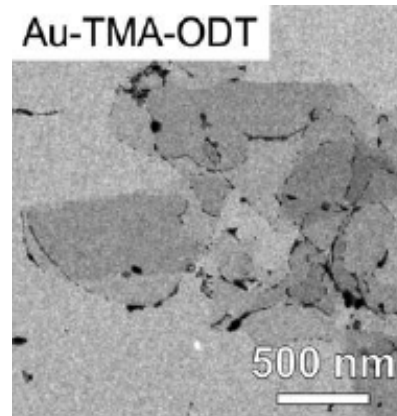
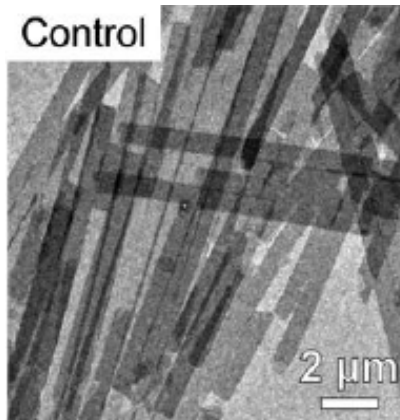


NP amphiphiles stabilizing transient assemblies

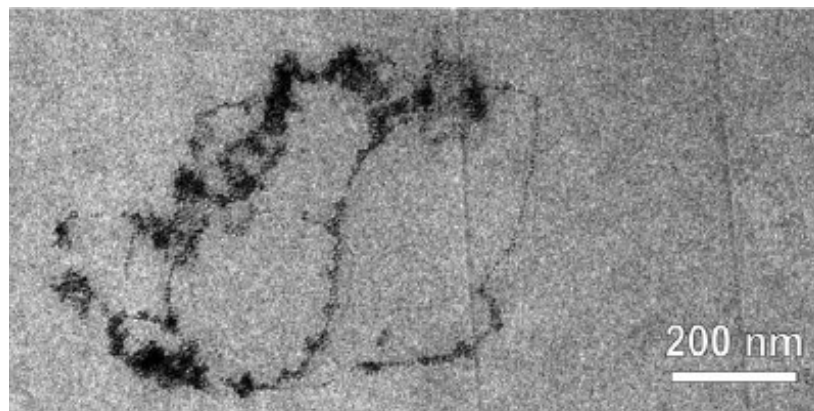


DC_{8,9}PC
lipid tubule

TEM

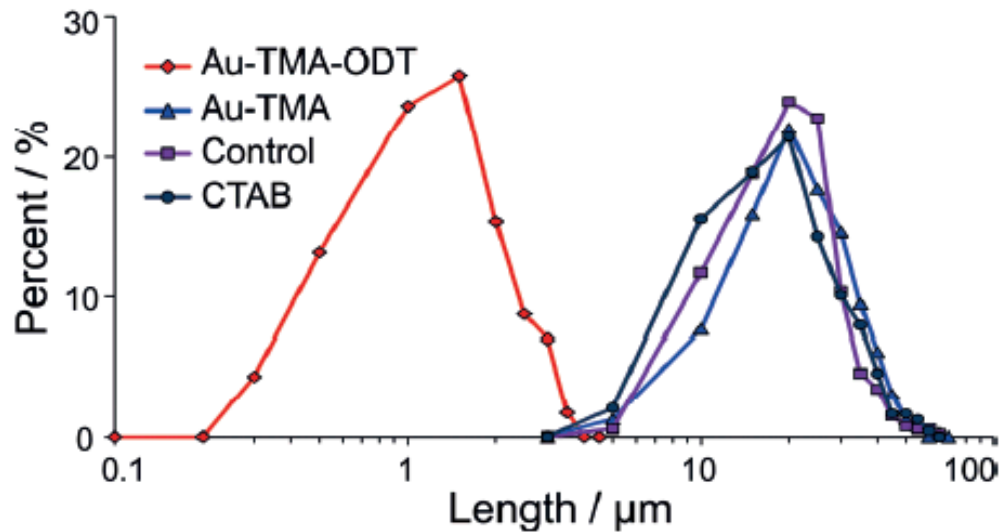


CryoEM

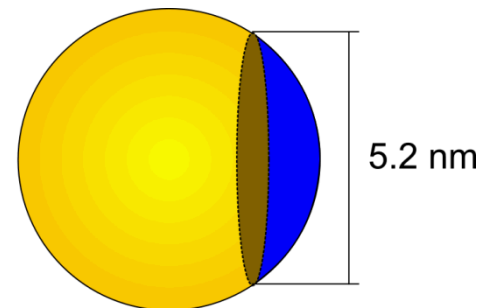
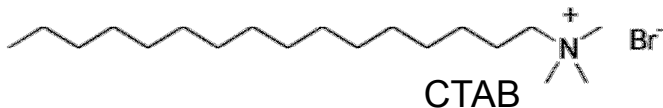


Shin et al. *Angew. Chem. Int. Ed.* 54, 10816 – 10820 (2015)

Hydrophobic interaction



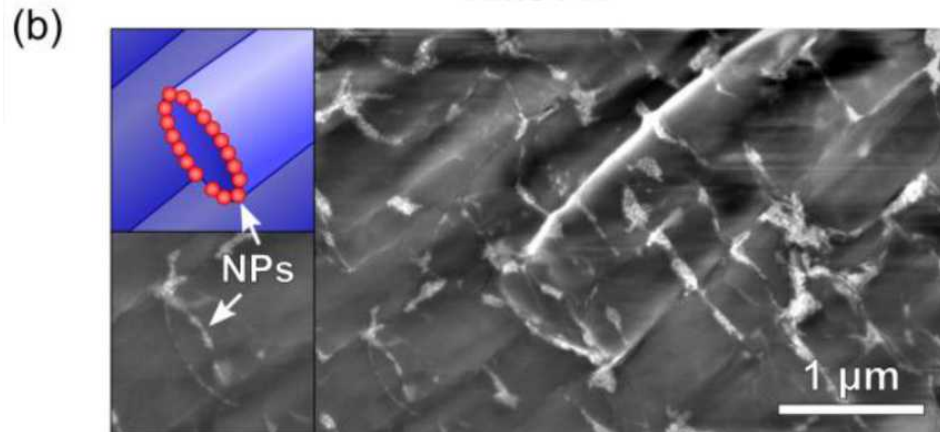
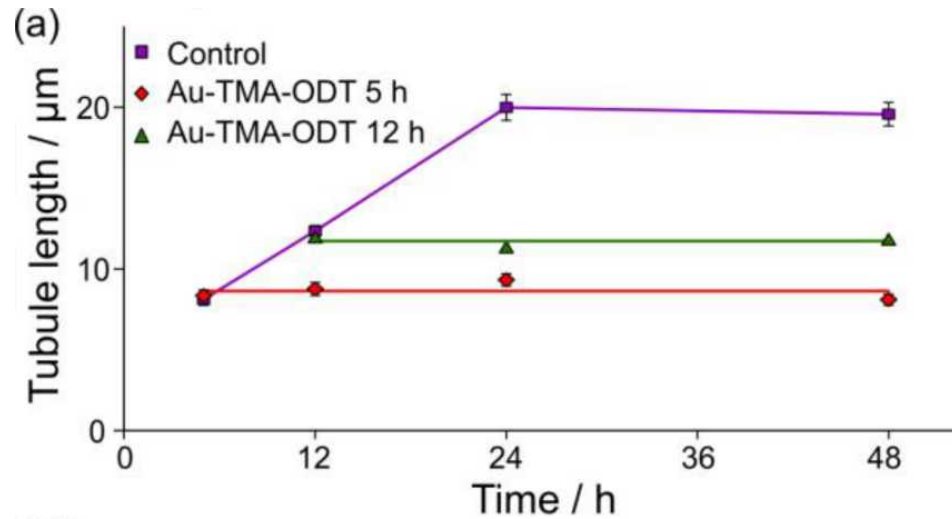
- Nanoscopic size of NP amphiphiles



TMA : ODT = 77 : 23

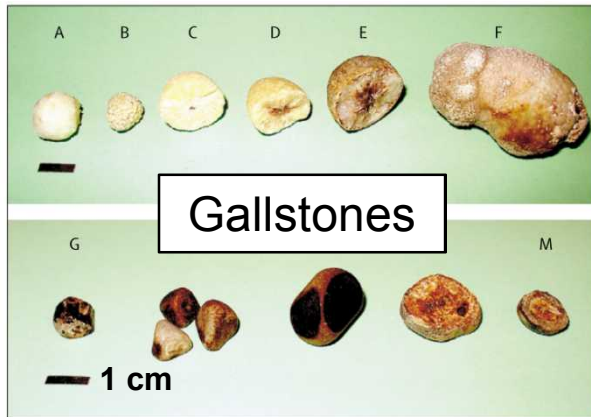


Tubule of desired length

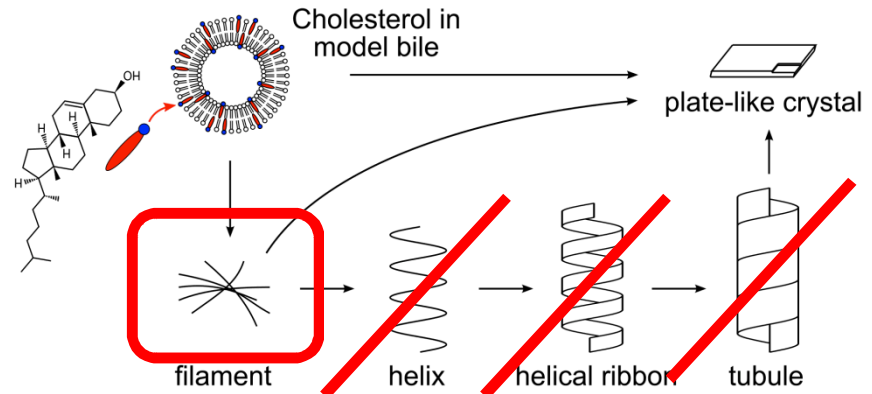


SEM

Inhibition of undesired assembly

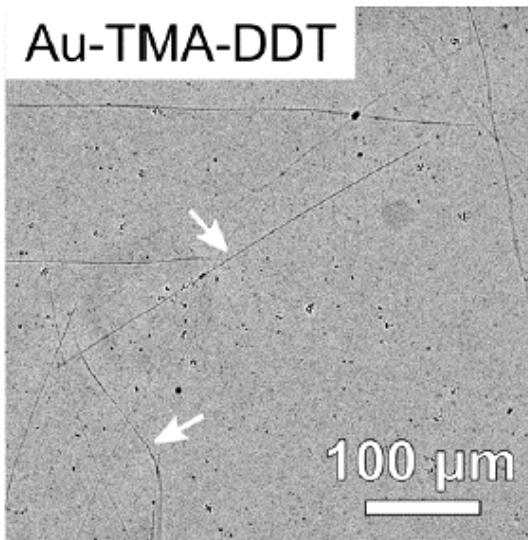
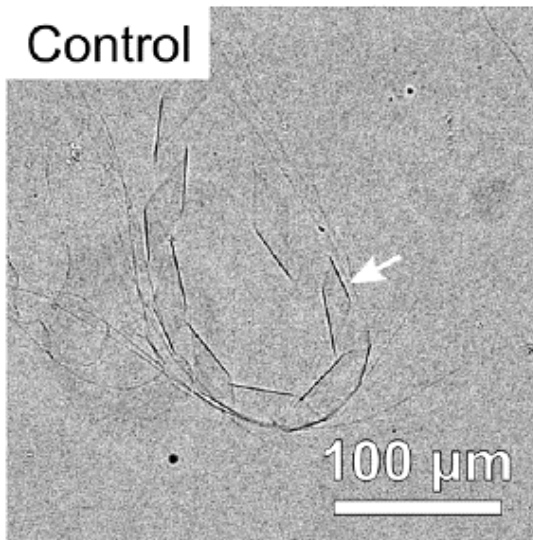


Lancet **368**, 230-239 (2012)



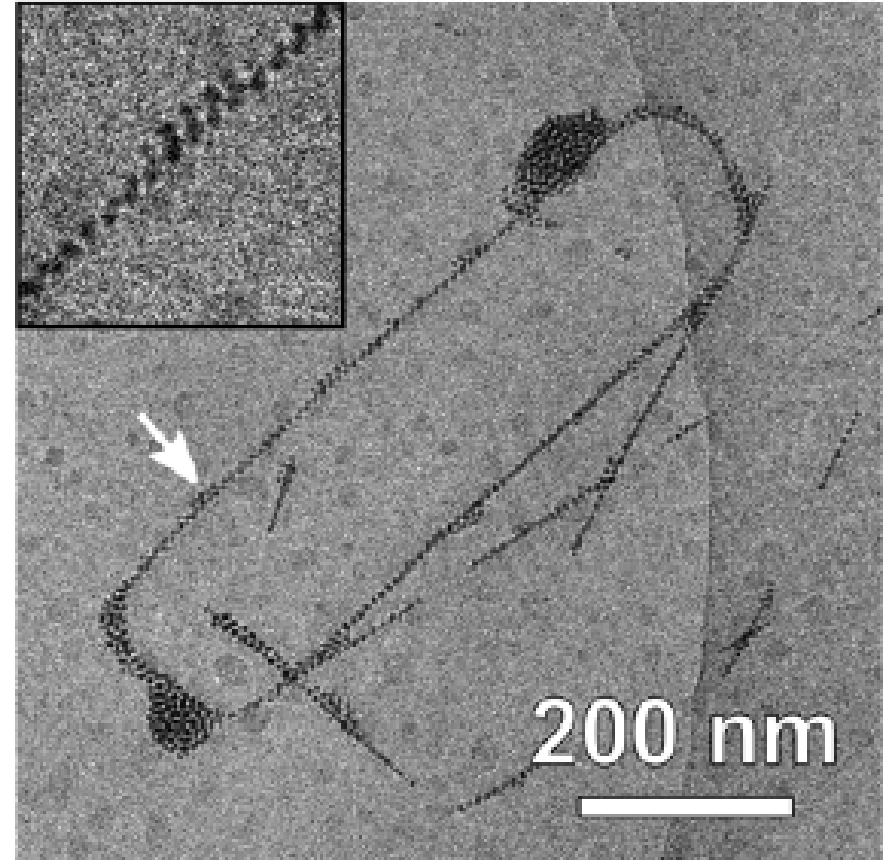
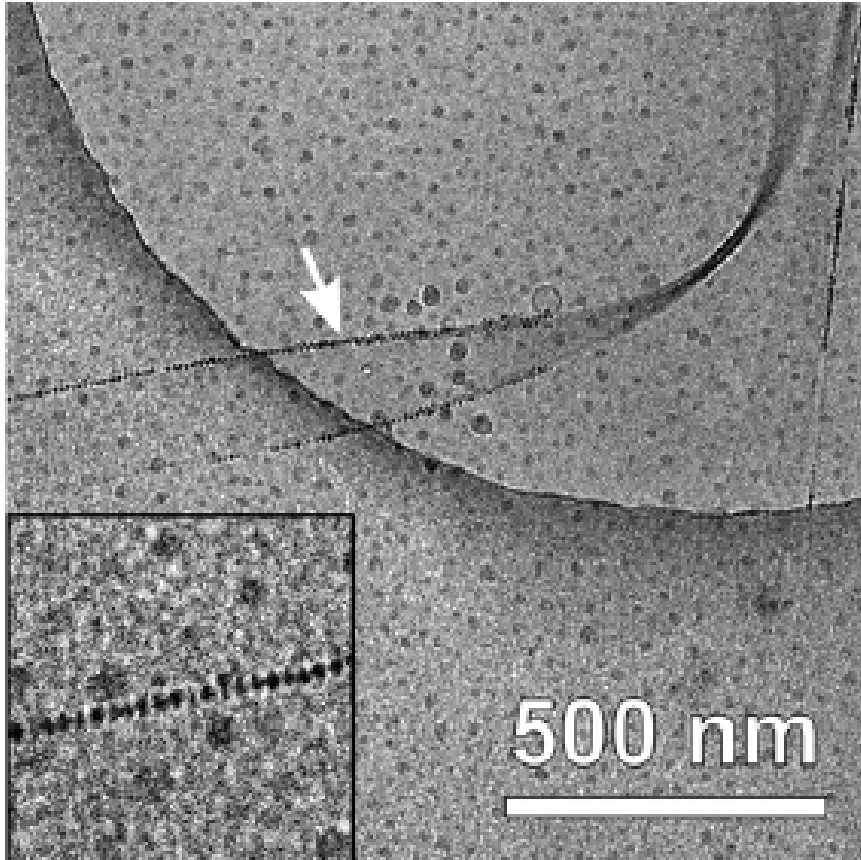
Cholesterol
helical ribbon

OM





Selective binding of NPs to cholesterol filaments (1)

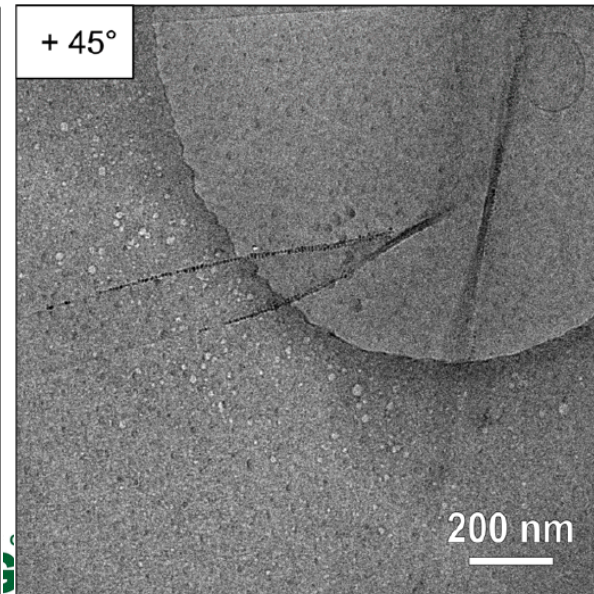
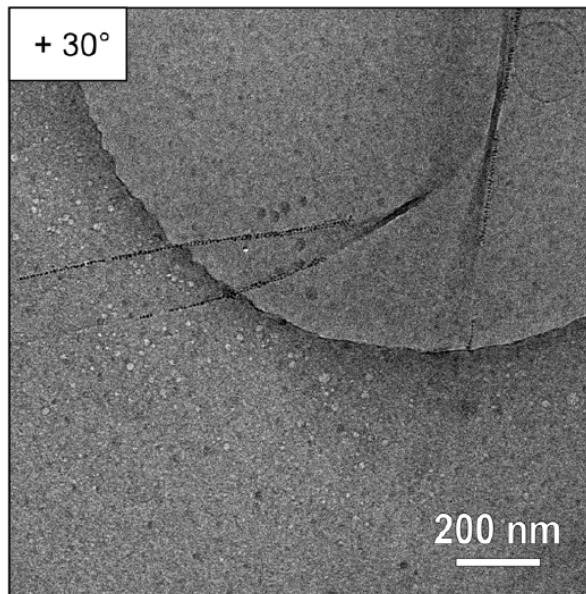
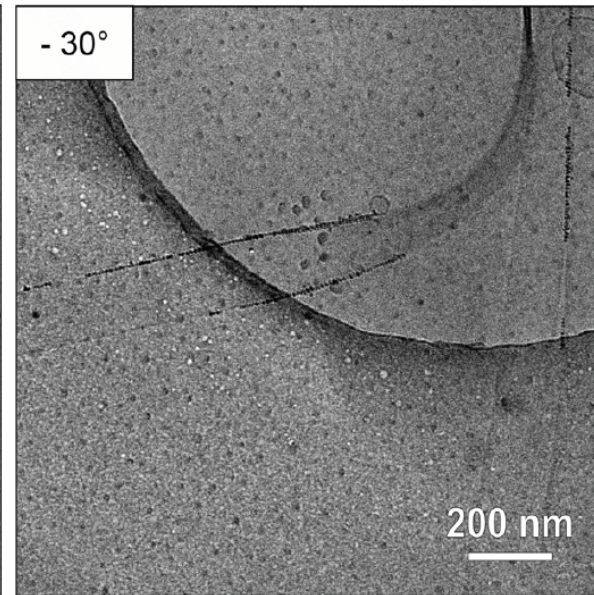
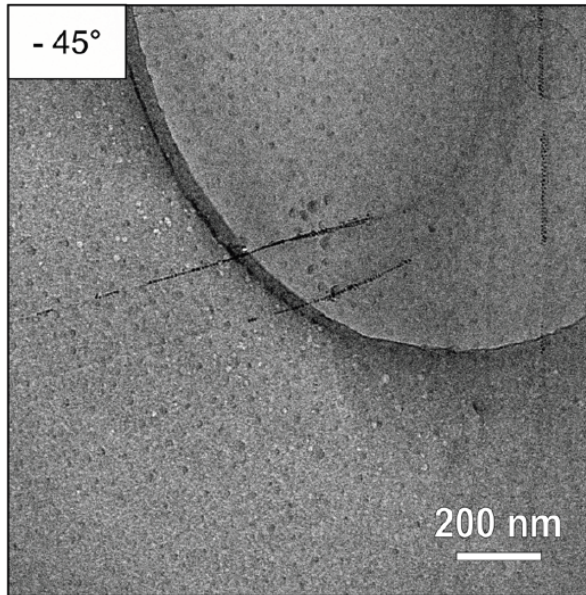


CryoEM



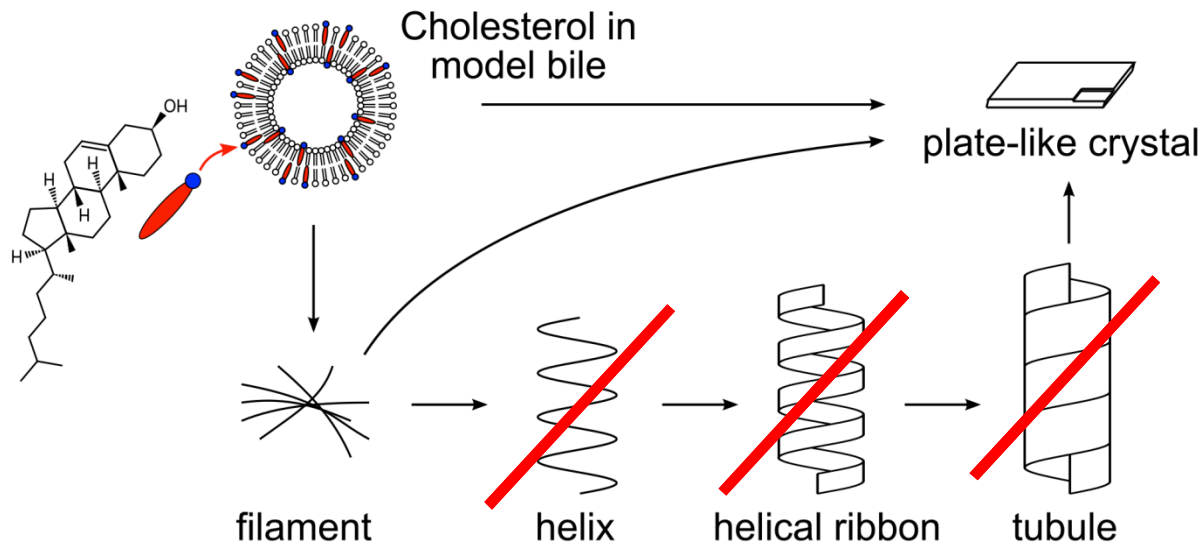
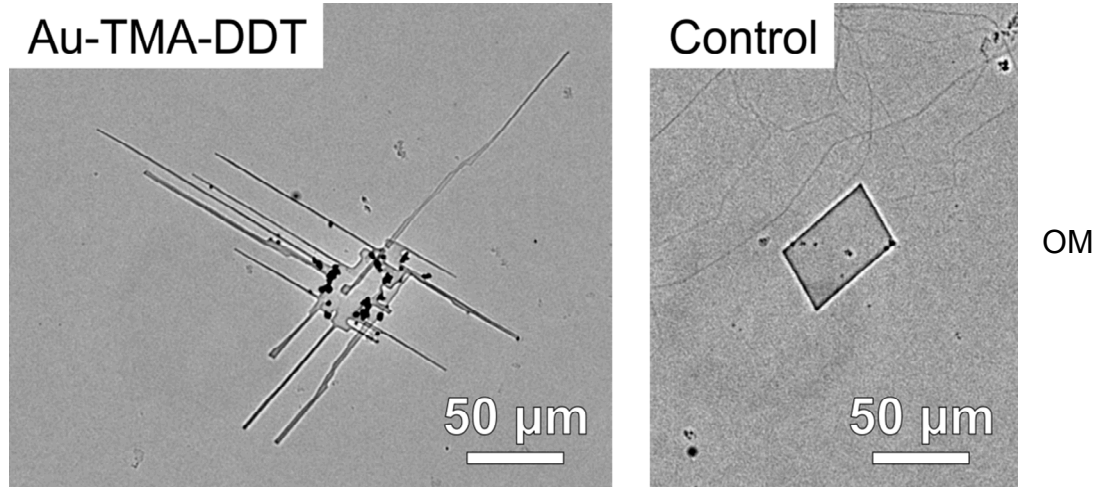
Selective binding of NPs to cholesterol filaments (2)

CryoEM with tilt angle





Incomplete inhibition of crystallization

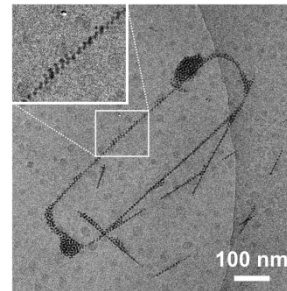
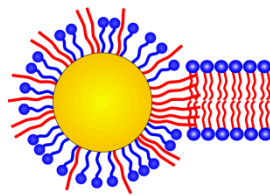




Conclusion

Part 2. Interaction between inorganic-organic materials

2.1. Selective binding of nanoparticles to open edges



- Stabilization of transient structures
- Inhibition of undesired assemblies

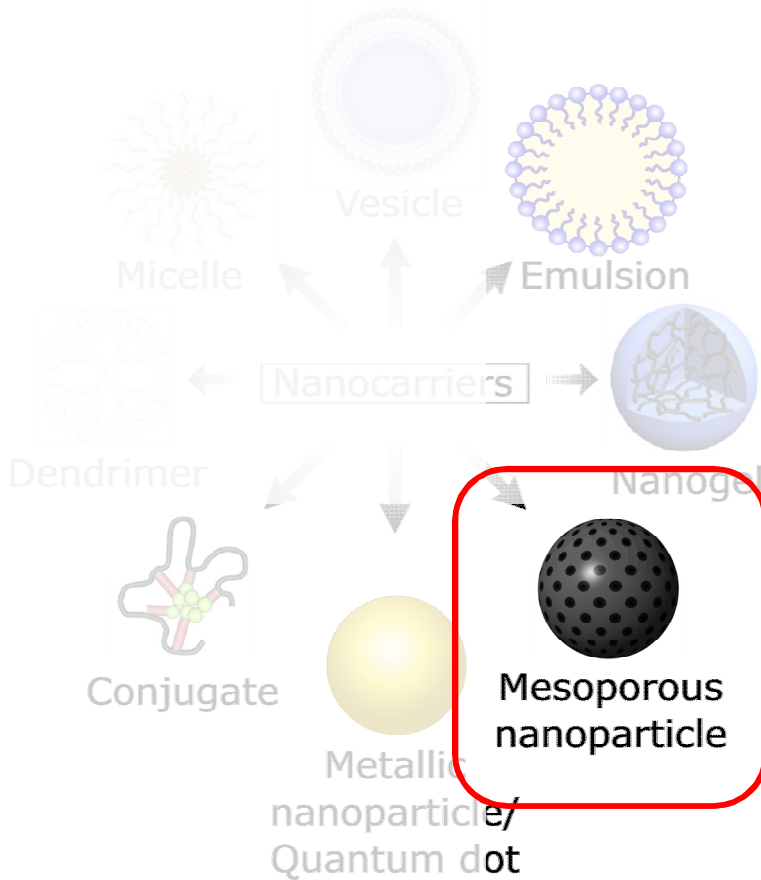
2.2. Formation of nanoparticle-supported bilayers



Smart delivery vehicles

Mesoporous silica nanoparticle (MSNP)

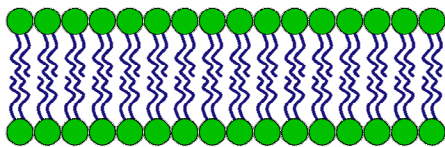
- High loading capacity
- Controllable size, shape, pore size, surface chemistry
- Biodegradable
- **Often require coatings**
 - ❑ Cargo encapsulation
 - ❑ Colloidal stability
 - ❑ Blood circulation time
 - ❑ Potential toxicity of unmodified silica



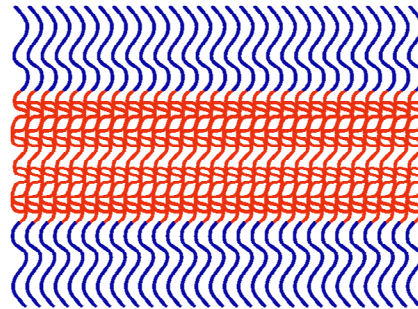


Hybrid bilayers

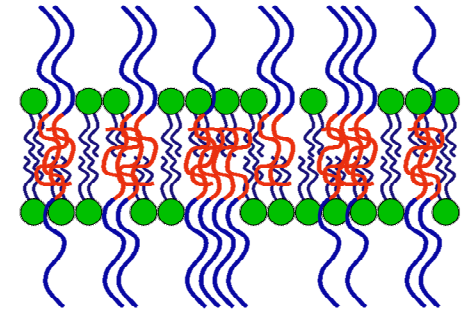
Lipid



Polymer



Hybrid bilayer



+

Fluidity
Biocompatibility

Stability
Versatility

Synergistic
membrane
properties

—

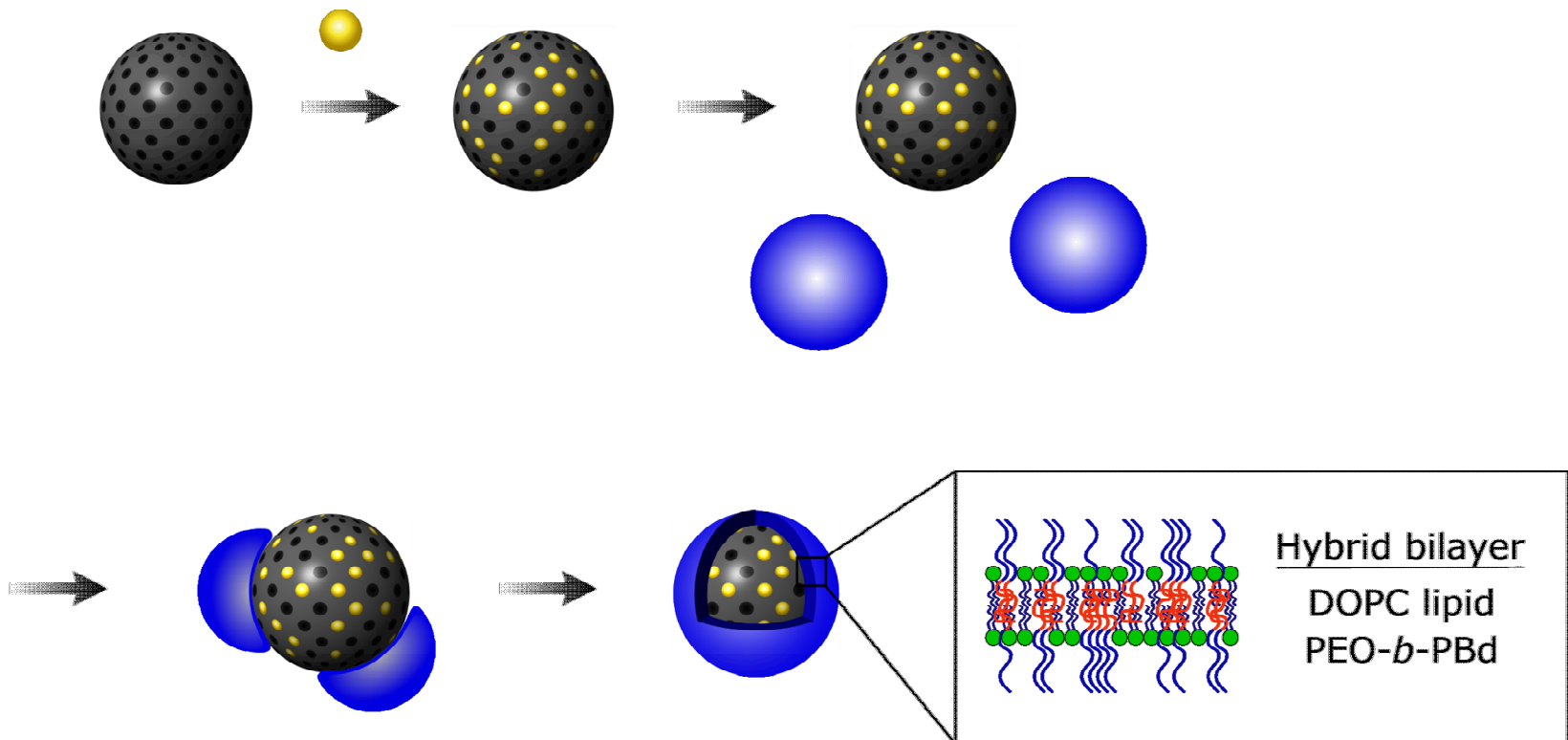
Stability

Fluidity



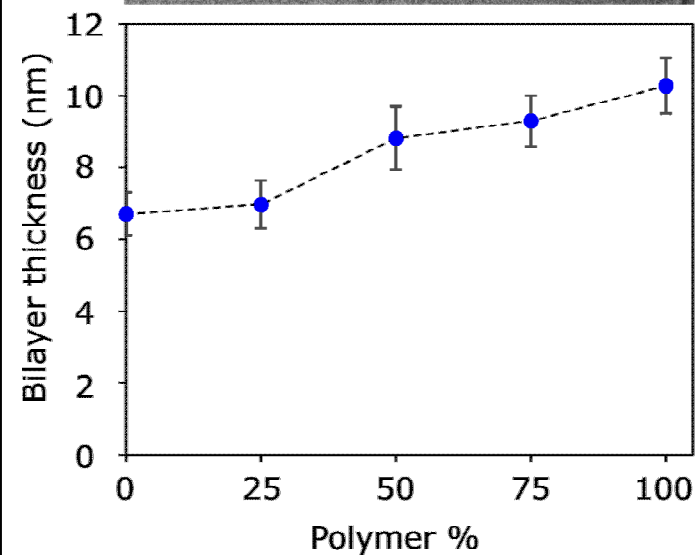
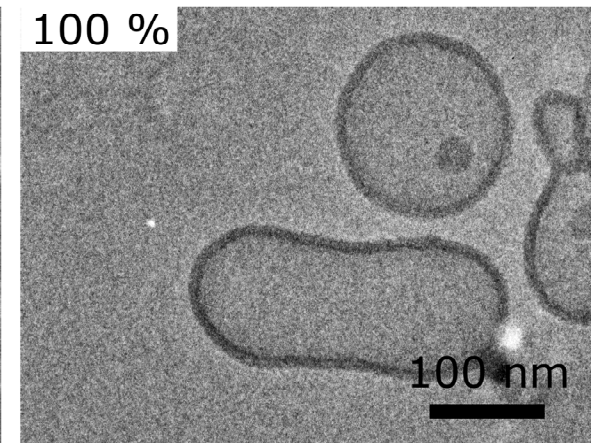
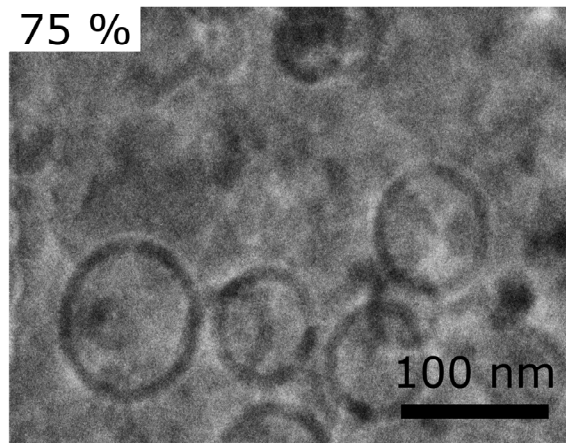
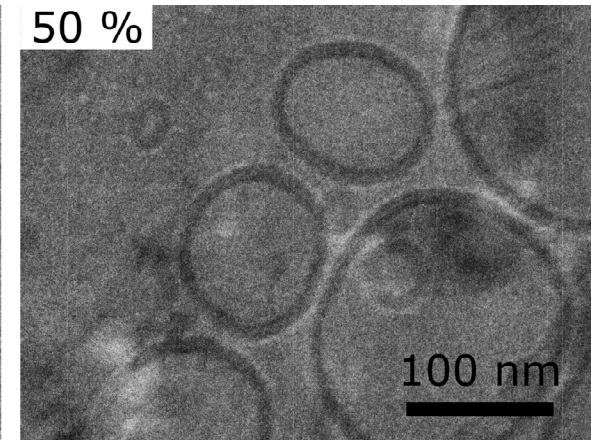
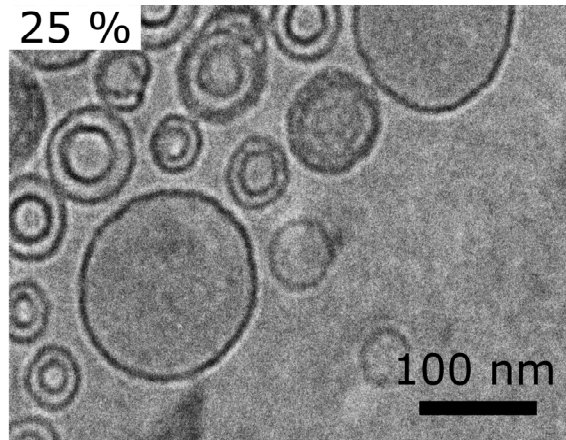
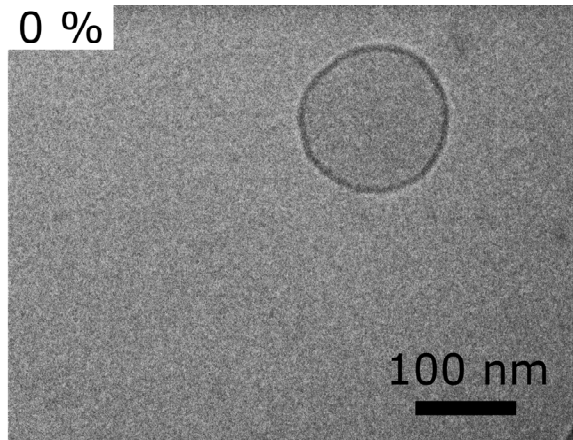
MSNP based delivery vehicles

Formation of supported hybrid bilayer on MSNP by fusion





Characterization of hybrid bilayers

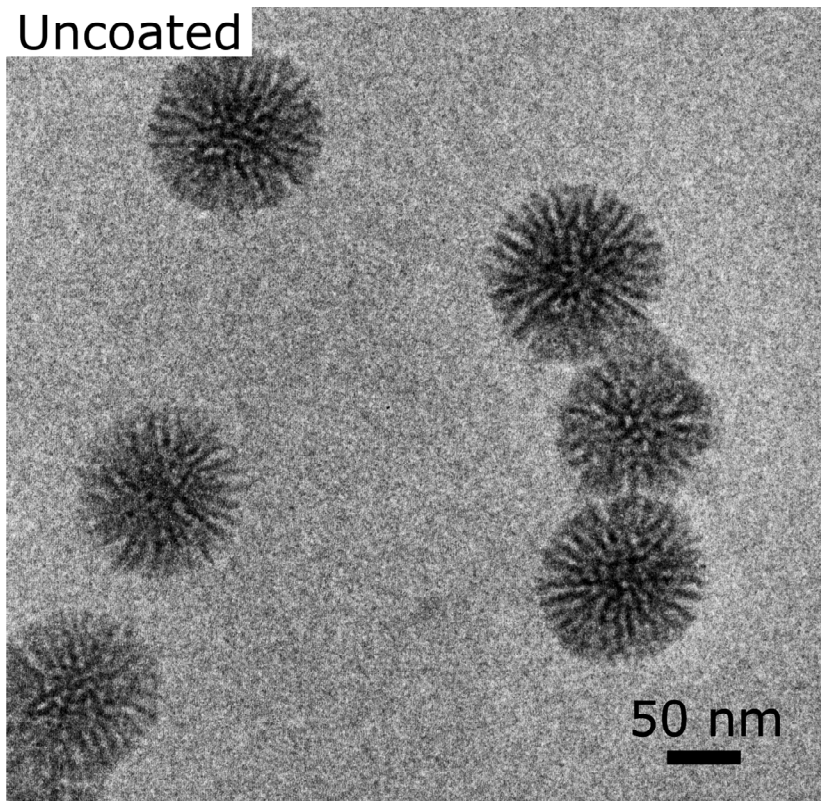


CryoEM

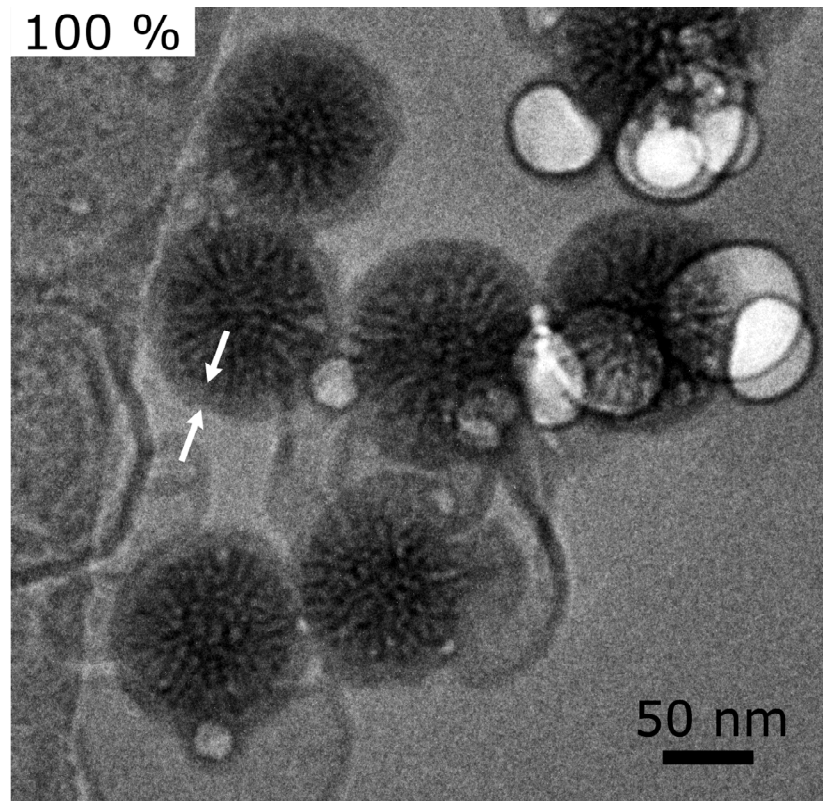


MSNP-supported hybrid bilayers

Uncoated



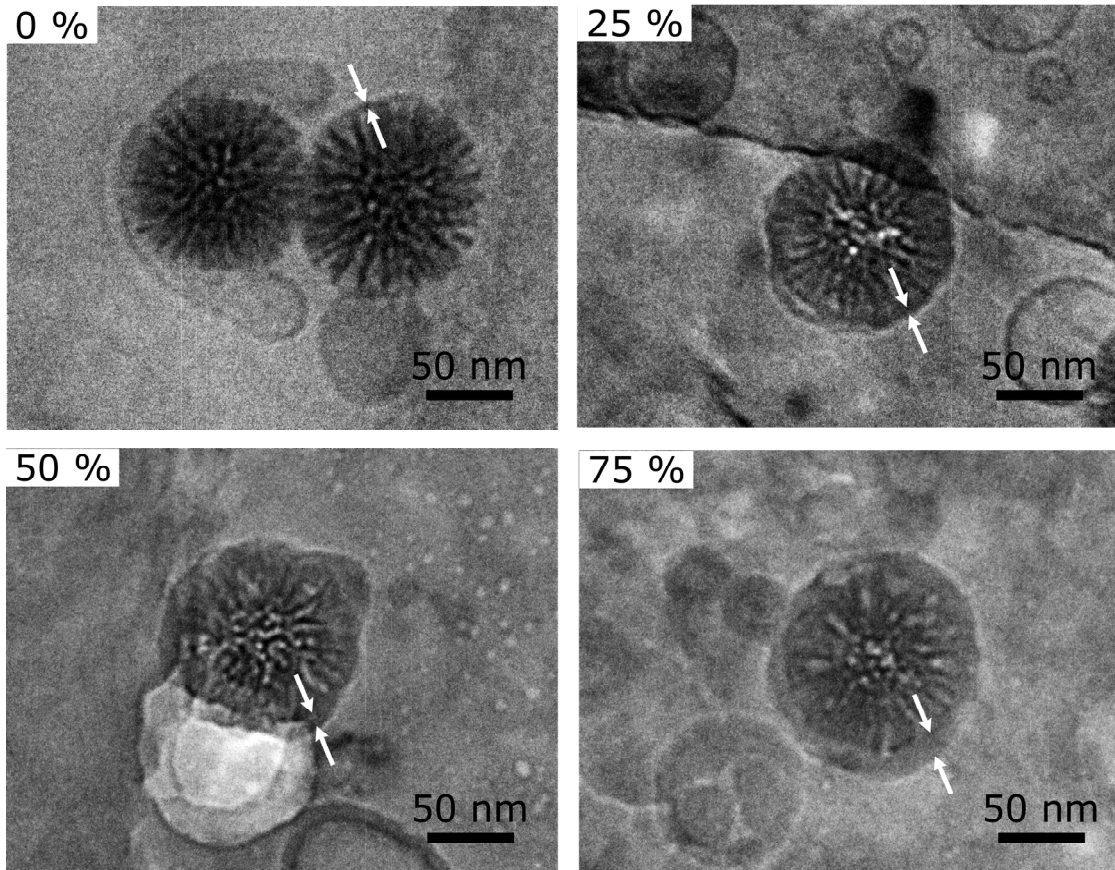
100 %



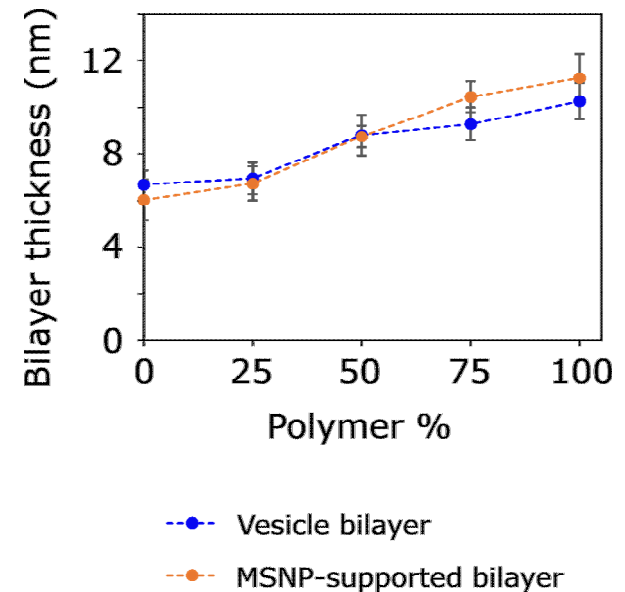
CryoEM



MSNP-supported hybrid bilayers



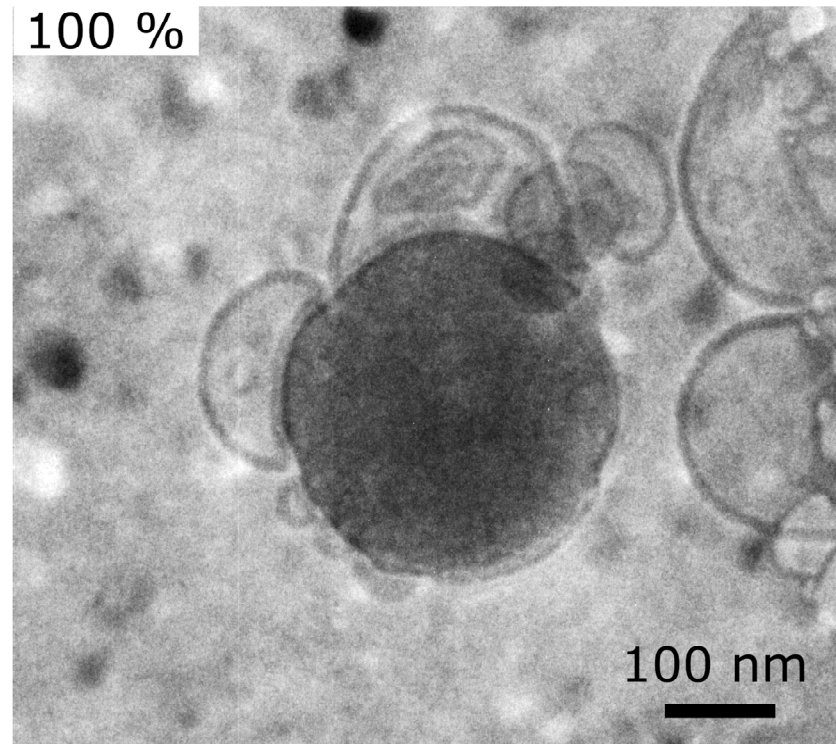
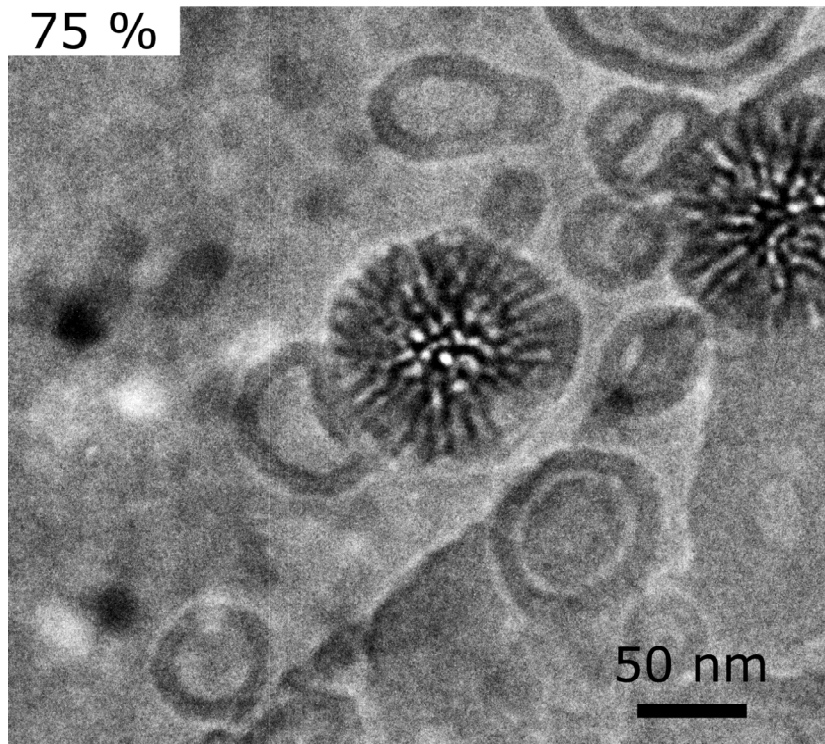
CryoEM





MSNP-supported hybrid bilayers

Adsorbed vesicles on MSNP due to robust membrane

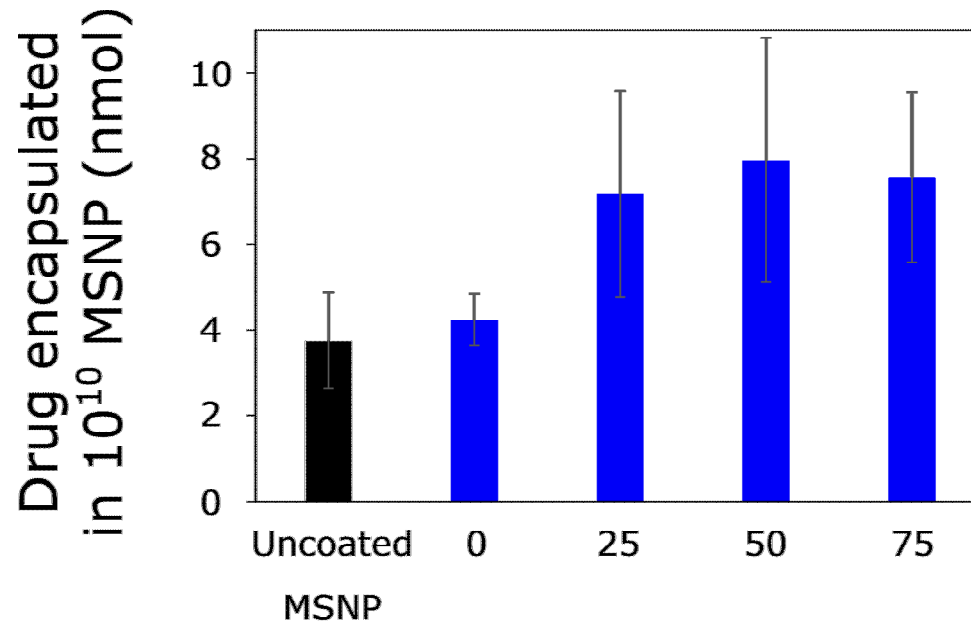


CryoEM



Encapsulation efficiency

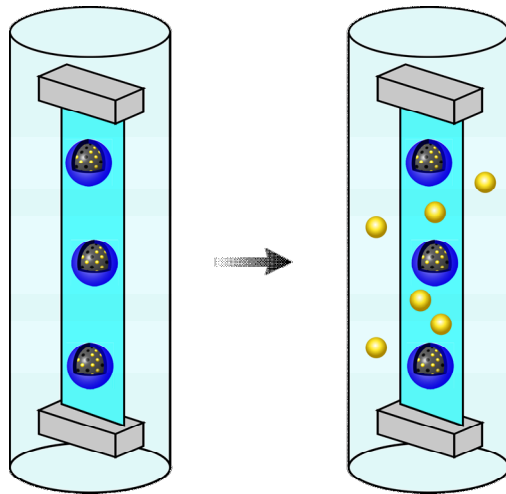
MSNPs with hybrid bilayer coating (~7.5 nmol) encapsulate more drug than MSNPs with lipid bilayer coating (4.2 nmol) or uncoated MSNP (3.7 nmol)



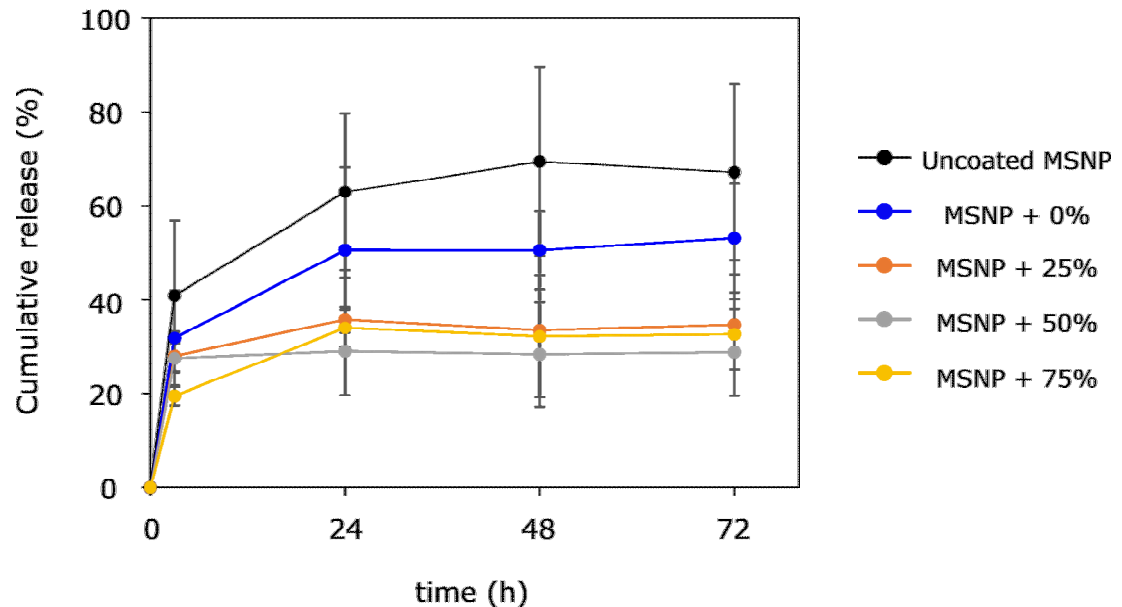
Polymer % in hybrid bilayer



In vitro drug release



Large-pore MSNP



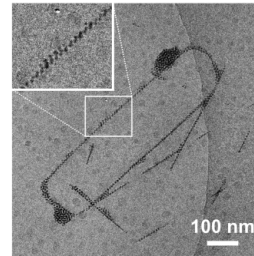
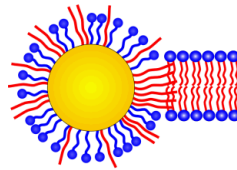
MSNPs with hybrid bilayer coating (32 %) release less drug less
MSNPs with lipid bilayer coating (53 %) or uncoated MSNP (67 %)



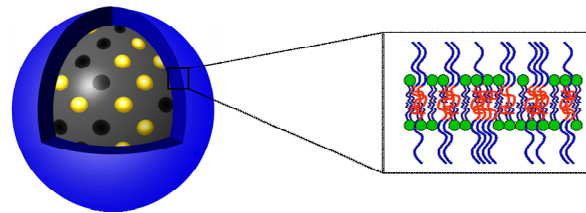
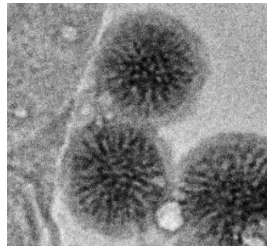
Conclusion

Part 2. Interaction between inorganic-organic materials

2.1. Selective binding of nanoparticles to open edges



2.2. Formation of nanoparticle-supported bilayers



- Supported hybrid bilayers by fusion
- Decreased permeability of hybrid bilayers



Part 1. Self-assembly of inorganic nanoparticles in solution

Part 2. Interaction between inorganic-organic materials

Part 3. Research plan

3.1. 3D visualization in synthetic architectures

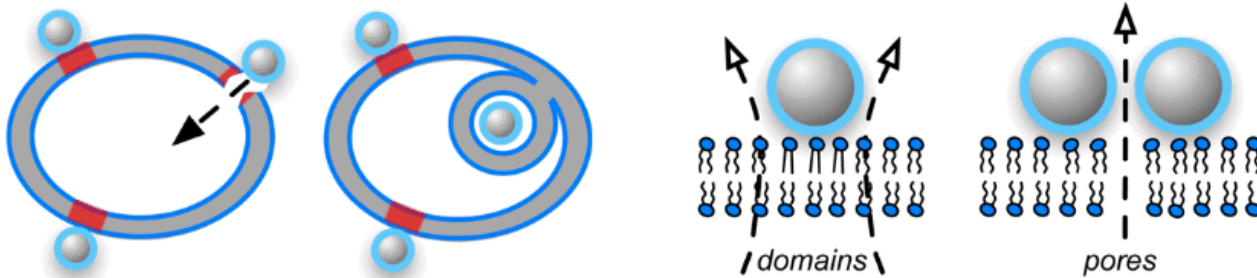
3.2. Time-resolved characterization of phase transition

3.3. *In situ* characterization at a wide range of temperature



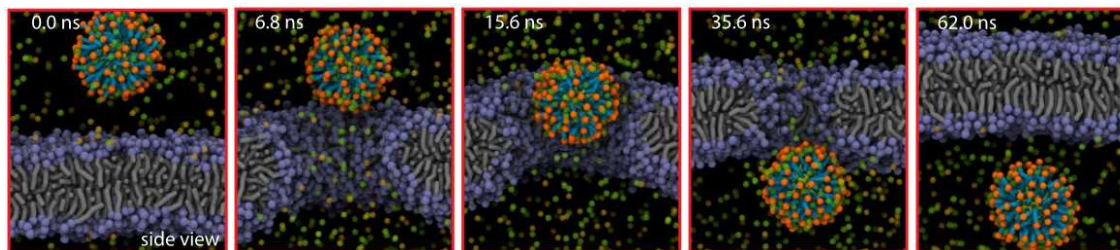
1. 3D visualization in synthetic architectures

Interaction of nanomaterials with biological interfaces



Environ.Sci.Technol. 48, 873-880 (2014)

Theoretical and computational investigations of nanomaterial-biomembrane interaction



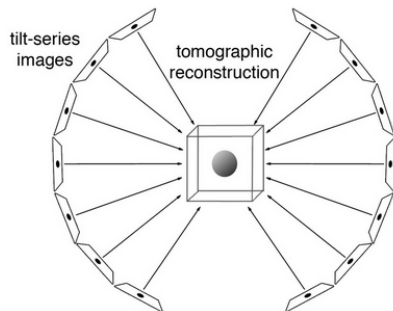
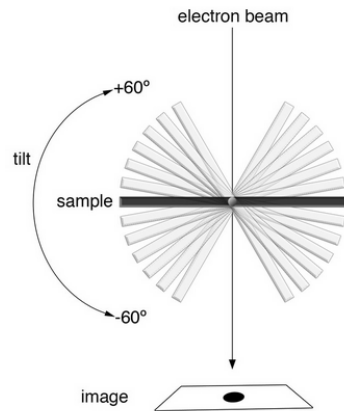
ACS Nano 7, 10799-10808 (2013)



1. 3D visualization in synthetic architectures

Cryo tomography

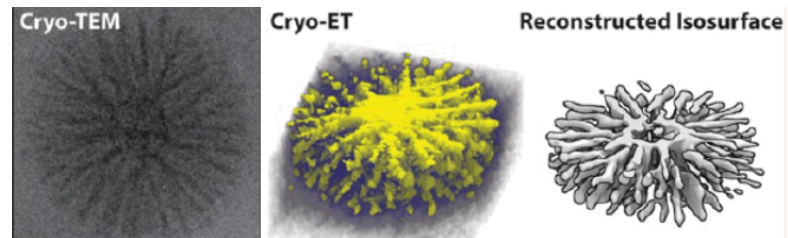
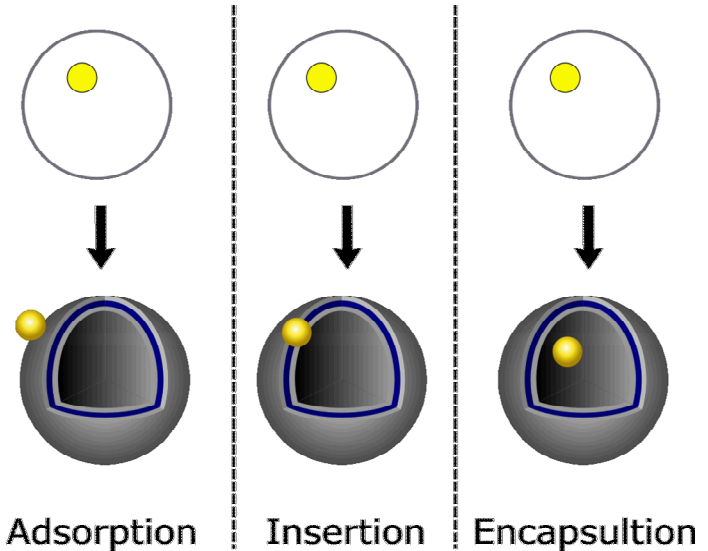
Hidden structural features revealed by tomography



www.wikipedia.org

Standard TEM
(2D)

CryoTEM with
tomography (3D)

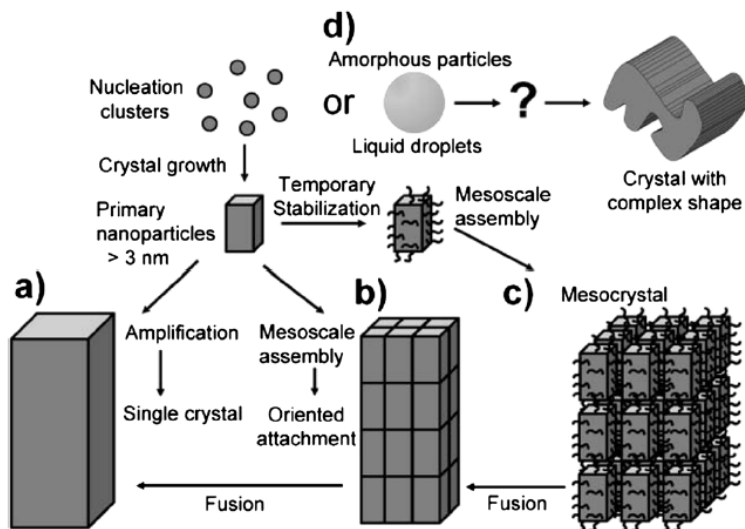


ACS Nano 8, 11330-11340 (2014)

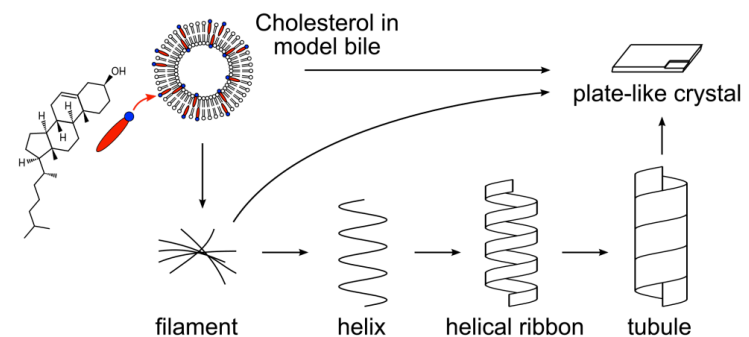


2. Time-resolved characterization of dynamic phase transition

Metastable phases during nucleation and growth of crystals



Nanoscale 4, 54-65 (2012)



Angew. Chem. Int. Ed. 54, 10816–10820 (2015)

Limitation: disordered, transient, highly hydrated intermediates

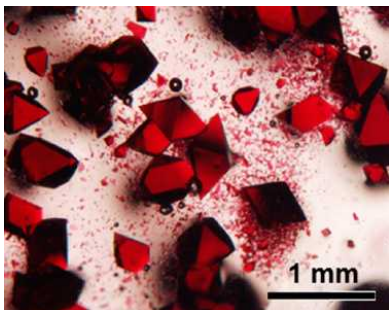


2. Time-resolved characterization of dynamic phase transition

- Detection of precursors in their native state during early stages of the precipitation/nucleation
 - Size, shape, lifetime of metastable intermediates
- Kinetic pathways of crystallization of organic/inorganic materials

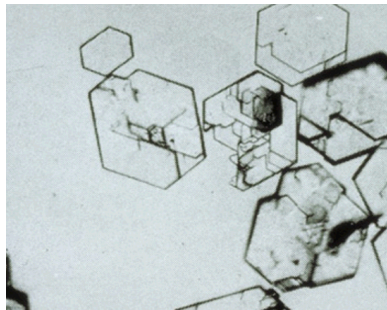
Protein crystallization

Hemoglobin



Crystals 7, 282 (2017)

Cystine



www.laboratoryinfo.com

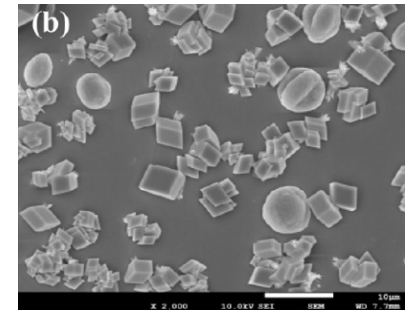
Biomineralization

Calcium phosphate



www.laboratoryinfo.com

Calcium carbonate

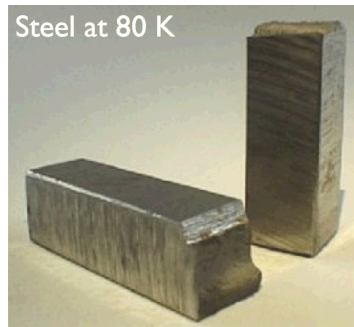


Cryst. Growth. Des. 18, 1710-1721 (2018)



3. *In situ* characterization at a wide range of temperature

Brittle Fracture



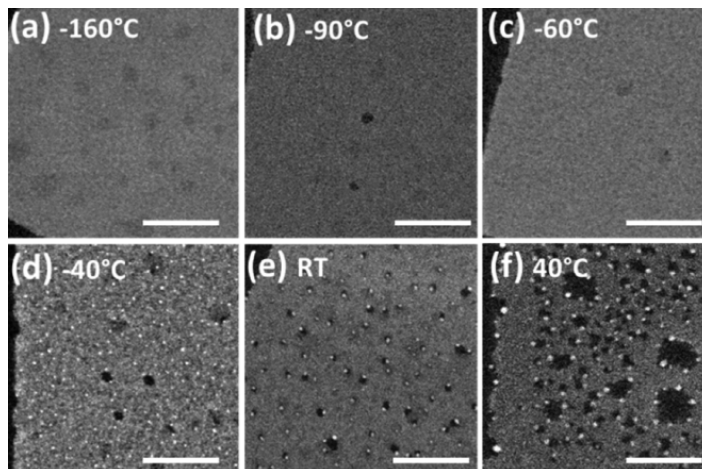
Ductile Fracture



www.civildigital.com

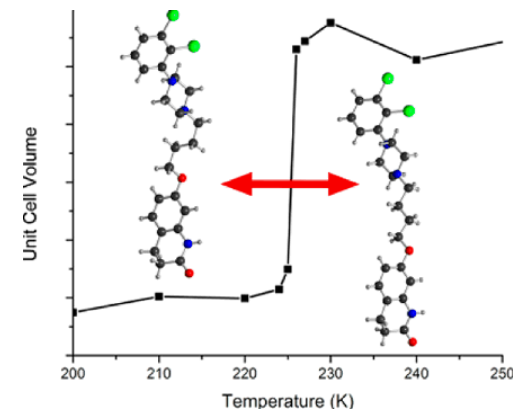
Low temperature phase transition in crystal

Nucleation of Pb NPs on CsPbBr₃ nanocrystal



ACS Nano 11, 2124-2132 (2017)

Aripiprazole Form VIII \leftrightarrow Form II



Cryst. Growth. Des. 14, 5004-5010 (2014)



3. *In situ* characterization at a wide range of temperature

CryoEM/FIB + *in situ* TEM

- Accessible temperature range of -170 °C to 300 °C
- Low dose imaging technique

Temperature-dependent studies

- Interface between hard and soft matters
- Structural phase transition in crystals
- Electron beam sensitive perovskites and metal-organic frameworks
- *In situ* nanomechanical testing



Acknowledgement

- **Principal Investigator:** Walter Paxton, Ph.D. (SNL)
Student Intern: Haley Monteith (SNL)



Paxton Monteith

- **MSNP synthesis**
Achraf Nouredine, Ph.D.; Jeffrey Brinker, Ph.D. (Univ. of New Mexico)

- **Funding**

This work was performed in part, at the Center for Integrated Nanotechnologies, an Office of Science User Facility operated for the U.S. Department of Energy (DOE) Office of Science (project number 2017BC0053). Research was supported by the Laboratory Directed Research and Development program at Sandia National Laboratories, a multi-mission laboratory managed and operated by National Technology and Engineering Solutions of Sandia, LLC, a wholly owned subsidiary of Honeywell International, Inc., for the U.S. Department of Energy's National Nuclear Security Administration under contract DE-NA-0003525.



Acknowledgement

- **Bishop research group at Penn State**

Principal Investigator: Kyle Bishop, Ph.D. (currently at Columbia Univ.)

Hee-Young Lee, Ph.D. (currently at KIT)

Aaron Drews, Ph.D. (currently at UCSD)

Aaron Chirsan

Sean Lewis



Bishop



Lee



Drews

