



**Sustainable Algal Biofuels Consortium**

*Cultivating Energy Solutions* SAND2012-1284P

# **Advanced Spectroscopic Imaging Techniques in Support of the Biomass Conversion Team**

**Howland Jones, Jerilyn Timlin and Michael Sinclair**  
**SABC Review 02/22/2012**

## **Sandia National Labs**

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## **Microscopic characterization**

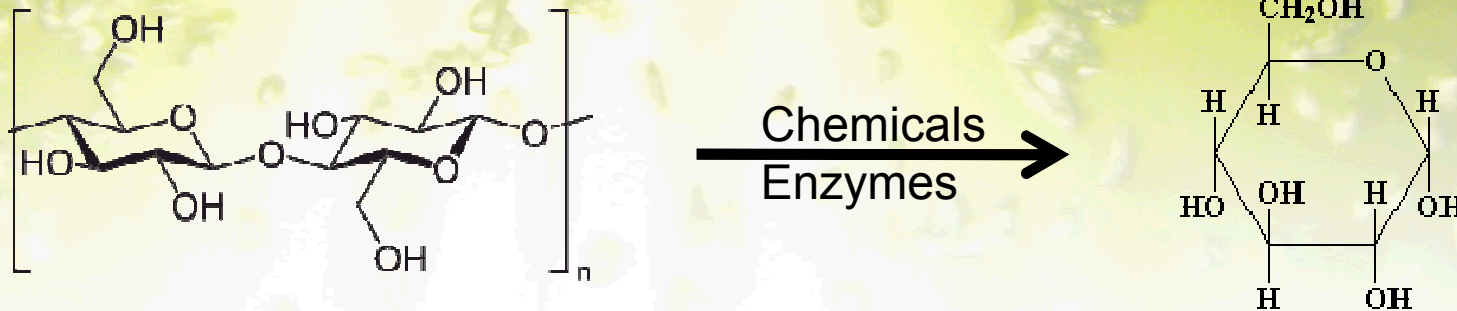
- **Study the real-time biochemical conversion of algal biomass residual and whole algal cells to fermentable sugars during chemical and enzymatic pre-treatment of cells at the microscopic level**
- **Provides unique spatial, temporal and biochemical information that is required for the support of the biochemical conversion task (1.3)**

## **Outcome from the Last Review**

- **Gary Dirks requested that we reach out and apply our advanced spectroscopic imaging techniques beyond the Ionic Liquid investigations**
  - **Acid pretreatment studies**
    - **NREL - Nick Nagel, Lieve Laurens, Stefanie Van Wychen**
  - **Enzyme conversion studies**
    - **SNL – Amy Powell, Kylea Parchert**
      - **To be presented by Amy Powell**



*Study the real-time biochemical conversion of algal biomass residual and whole algal cells to fermentable sugars during pre-treatment of cells at the microscopic level.*



Imaging provides unique spatial, temporal and biochemical information

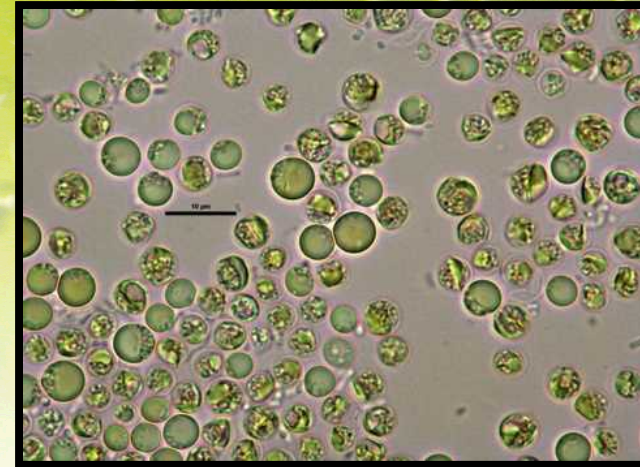
- Fluorescence Imaging
  - Endogenous Photosynthetic pigments (Chlorophyll components, carotenoid)
  - Fluorescent labeling
- Raman Imaging
  - Improved molecular specificity to measure endogenous components
    - Cellular and conversion products

# Cultivating Energy Solutions

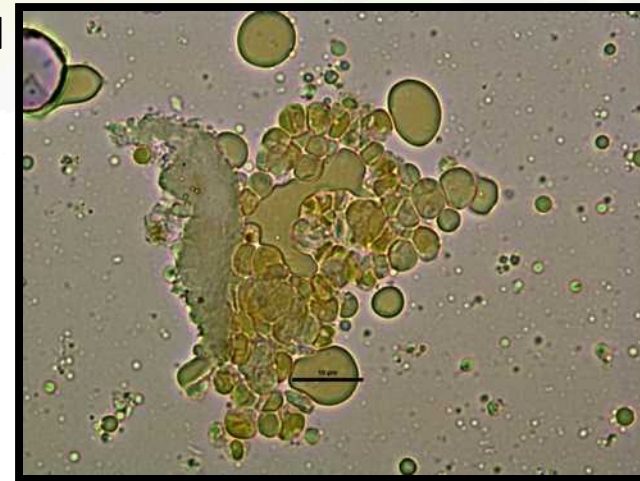
## Acid Pretreatment Studies

- Purpose and Goals
  - Understand the effect of acid pretreatment procedures (cellular level)
  - Understand aggregate formations at the high acid level
  - Determine Laser requirements to guide development of Raman line-imaging microscope
- Biomass
  - NREL supplied
  - High lipid Chlorella (HLCZ)
  - Control – untreated biomass
  - High acid – 2% H<sub>2</sub>SO<sub>4</sub>, microwave digestion (145°C for 1 minute)
- Imaging
  - Fluorescence and Raman Hyperspectral

Control

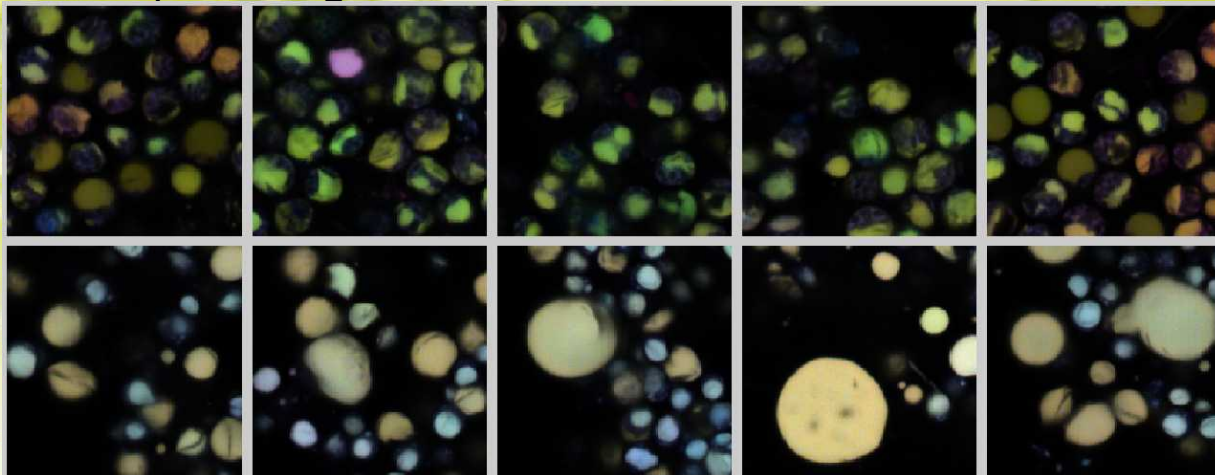


High Acid



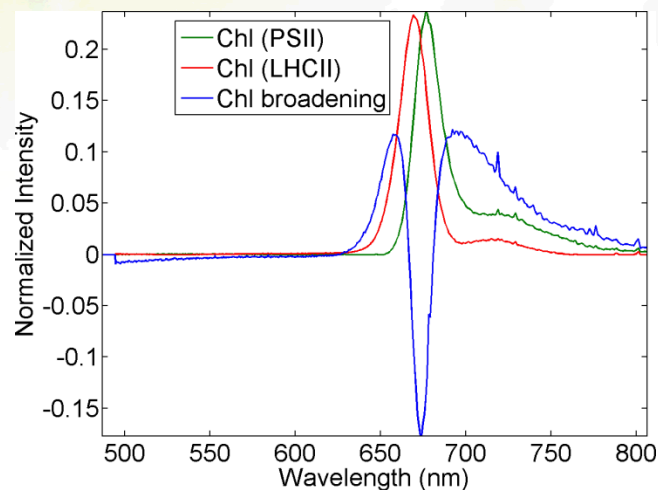


Five repeat images of each biomass



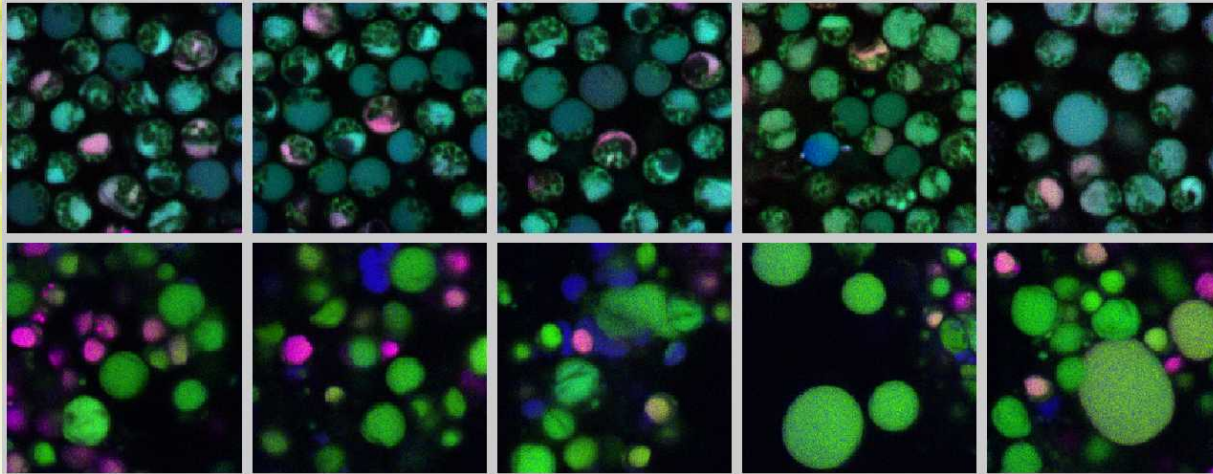
Untreated HLCZ  
Biomass

HLCZ Biomass  
2% H<sub>2</sub>SO<sub>4</sub>, 145°C, 1 min



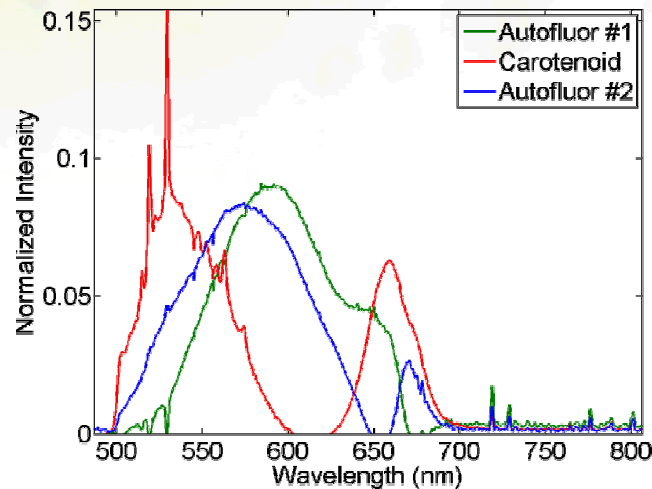
- Large differences between the untreated and treated biomass
- Approximately equal parts of chlorophyll components present in large oil droplets
- Understanding how the photosynthetic pigments degrade and change upon treatment

Five repeat images of each biomass



Untreated HLCZ  
Biomass

HLCZ Biomass  
2% H<sub>2</sub>SO<sub>4</sub>, 145°C, 1 min

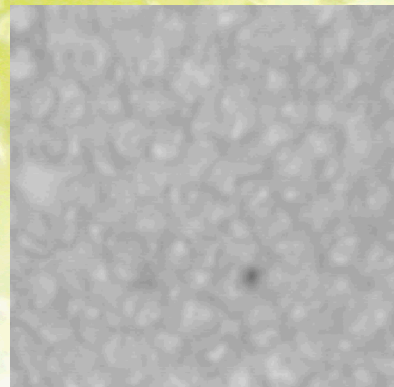


- Large differences between the untreated and treated biomass
- Decomposition components are more intense in the treated biomass
- Difficult to interpret the meaning of these results

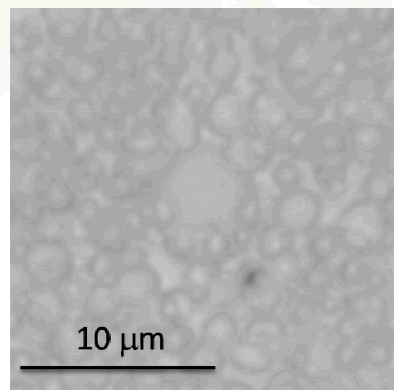


### Raman Imaging

Video Images

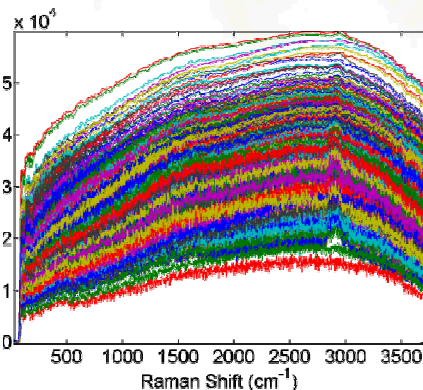
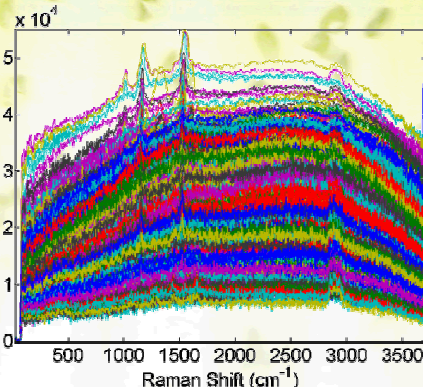


Untreated HLCZ Biomass

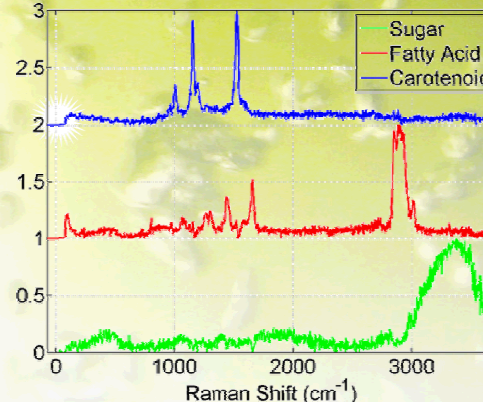


HLCZ Biomass  
2% H<sub>2</sub>SO<sub>4</sub>, 145°C, 1 min

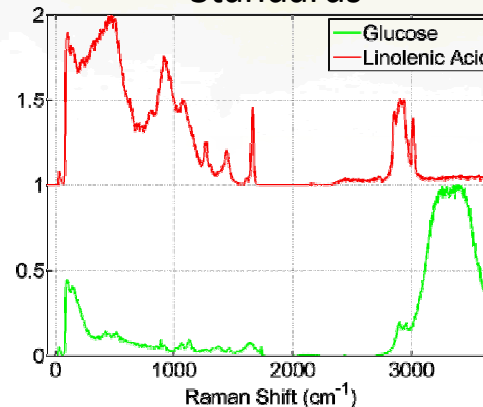
Spectral Data



MCR Pure Components



Standards



Witec Raman confocal microscope used for preliminary measurements.

14 spectral components

Measured standards confirms the presence of fatty acid deposits and free sugars in the biomass.



# Raman Imaging

### Video Images

### MCR Pure Components

### MCR Concentrations

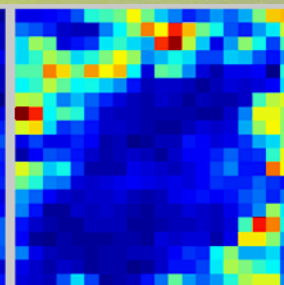
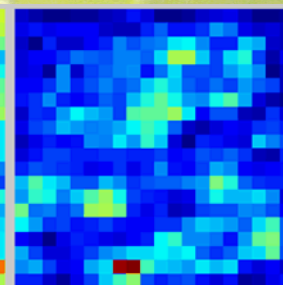
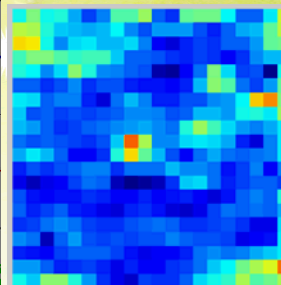
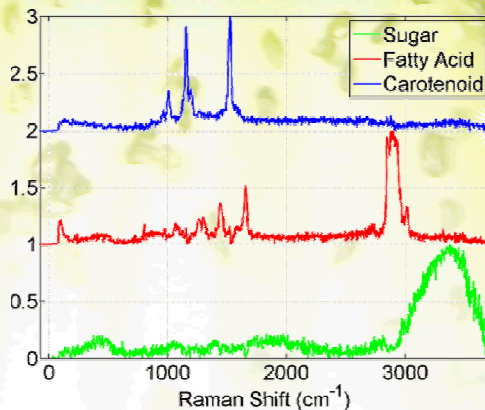
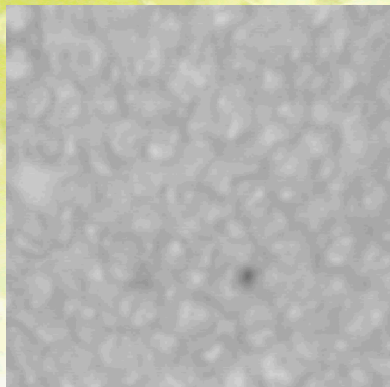
#### Free Sugars

#### Fatty Acids

#### Carotenoid

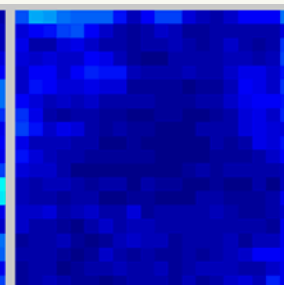
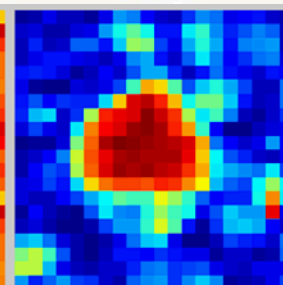
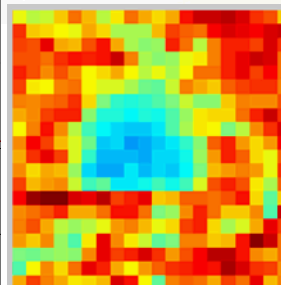
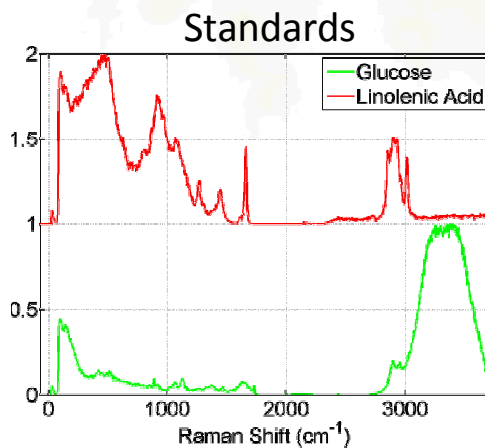
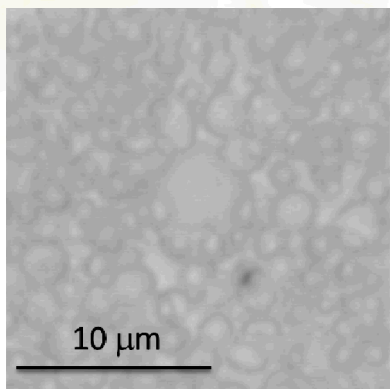
Untreated HLCZ

Biomass



HLCZ Biomass

2% H<sub>2</sub>SO<sub>4</sub>, 145°C, 1 min



# Raman Imaging

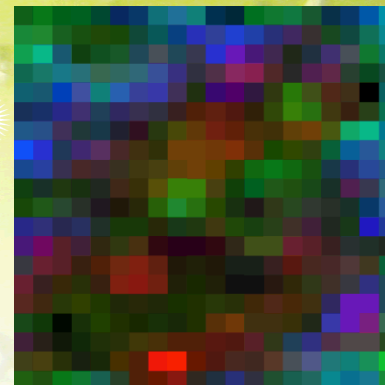
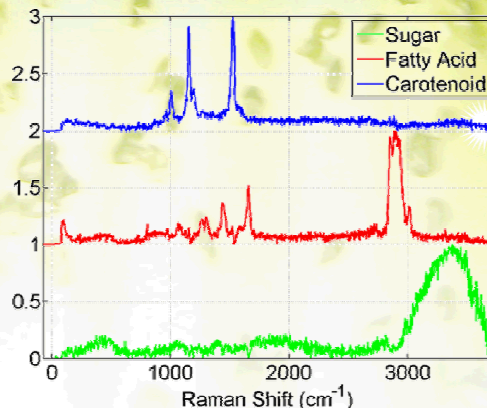
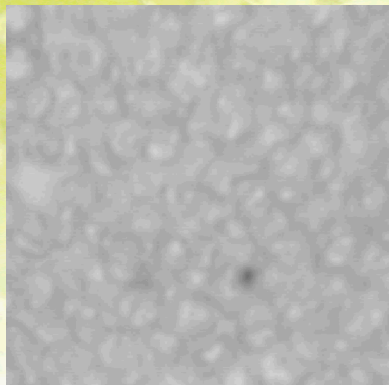
Video Images

MCR Pure Components

RGB Images

Untreated HLCZ

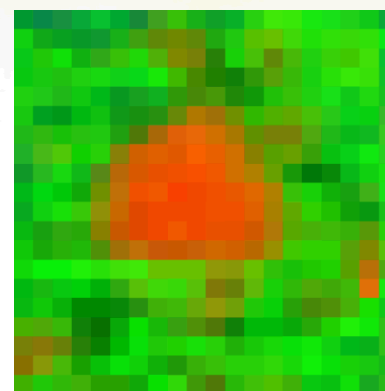
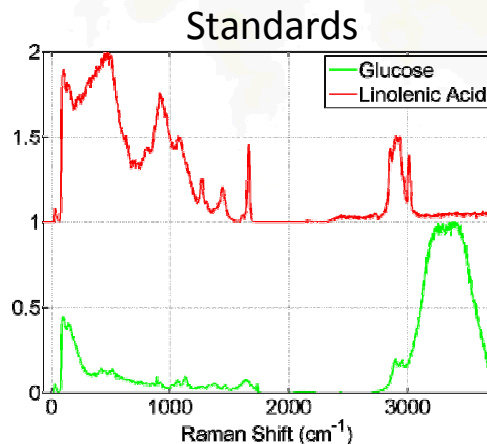
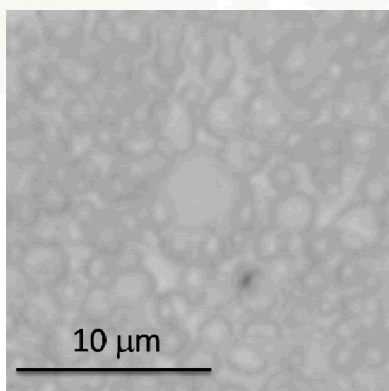
Biomass



High chemical specificity!

Poor spatial resolution is due to slow microscope mapping.

HLCZ Biomass  
2% H<sub>2</sub>SO<sub>4</sub>, 145°C, 1 min

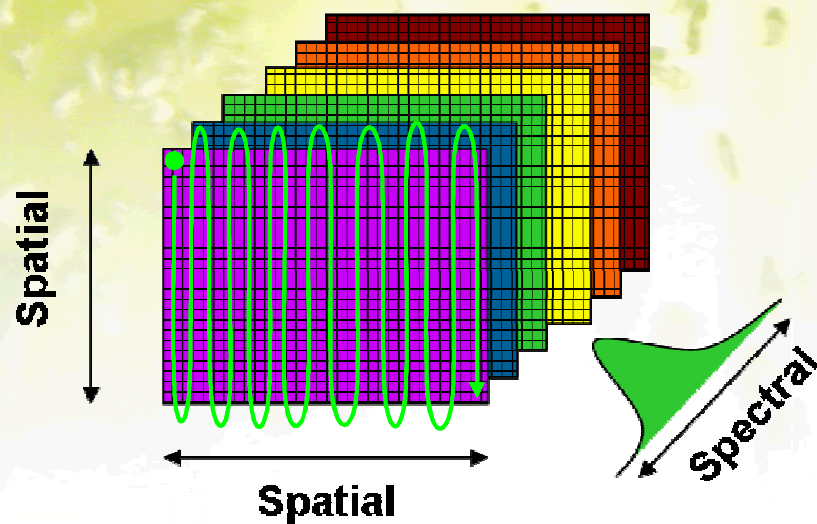


Spatial resolution can be improved either on the commercial scope or using our research scope.

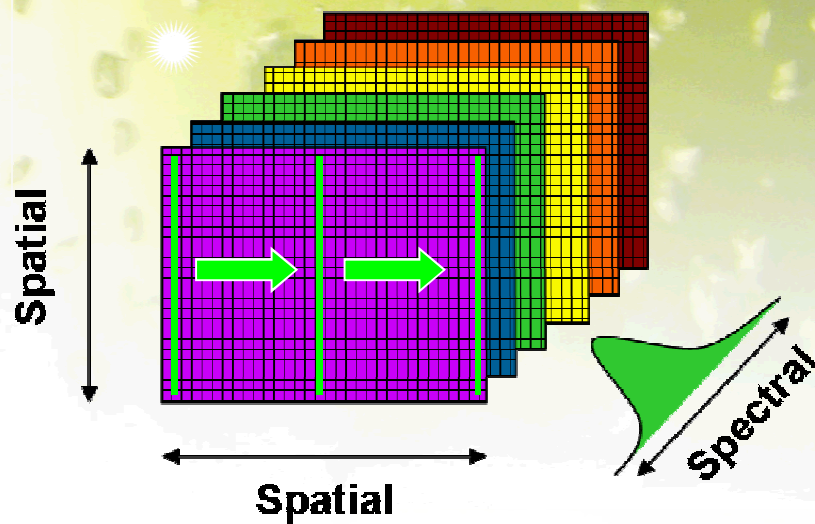


Line imaging is the key to increase our data acquisition speed!

$S/N$  is proportional to Laser Power  $\times$  Integration Time



Most commercial instruments employ spot mapping to image a sample by illuminating and collecting a spectrum at each pixel.



We augmented our current fluorescence line imager to do Raman line imaging which illuminates and collects 200 pixels simultaneously.

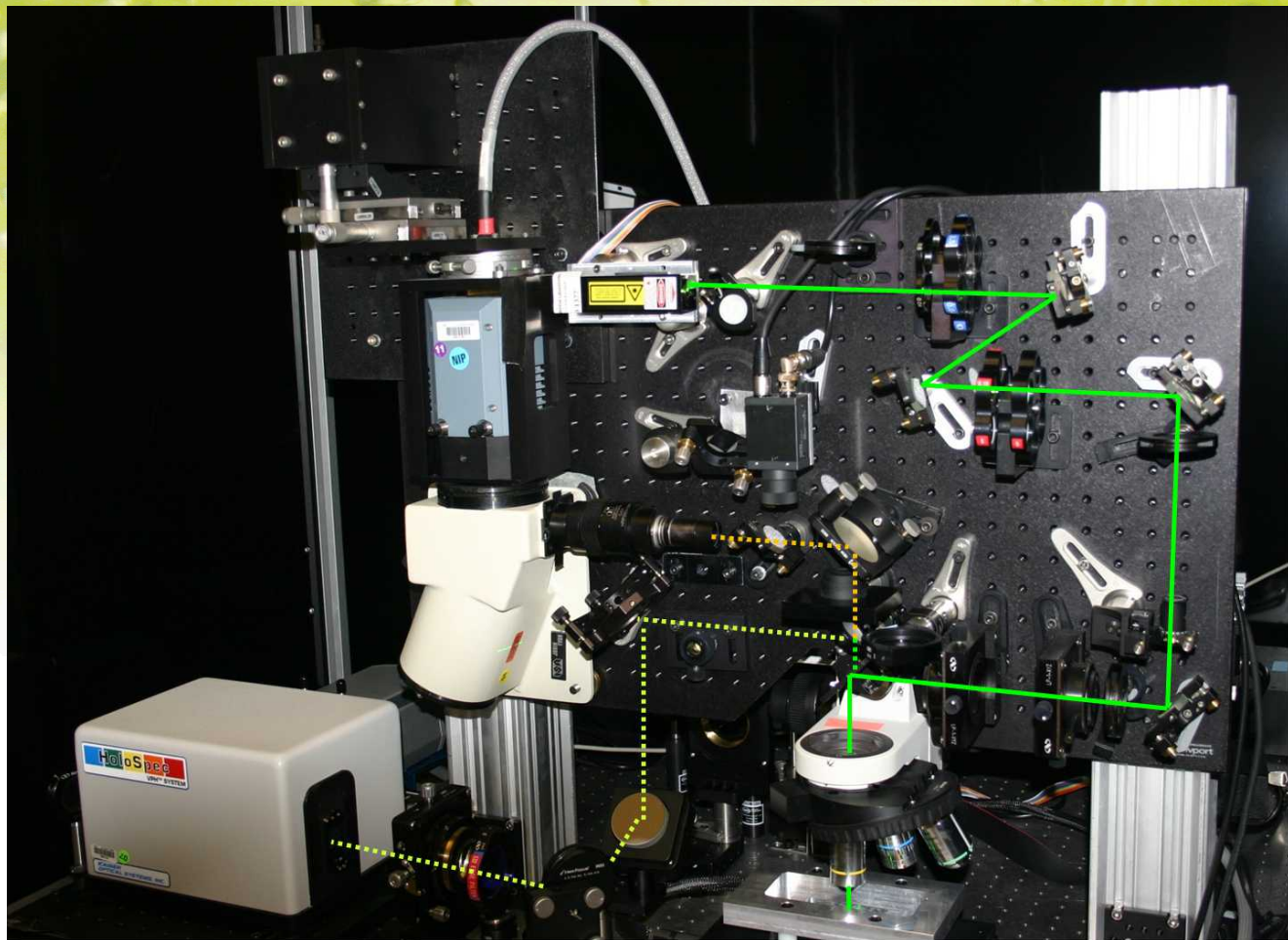
### *Cultivating Energy Solutions* Fluorescence and Raman line-imaging microscope

Optical assembly completed and adjustments are being made to improve the SNR.

3 watt, 532 nm laser is being purchased to improve the excitation of our biomass samples.

Software being modified to operate in stop and stare modality.

Initial Raman spectra have been collected.



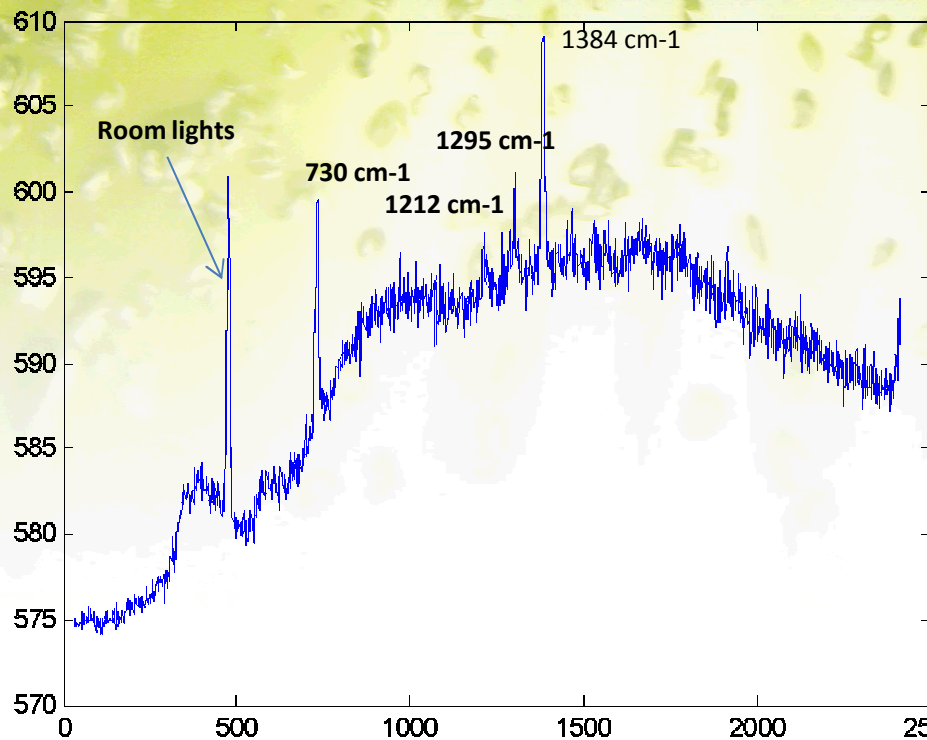




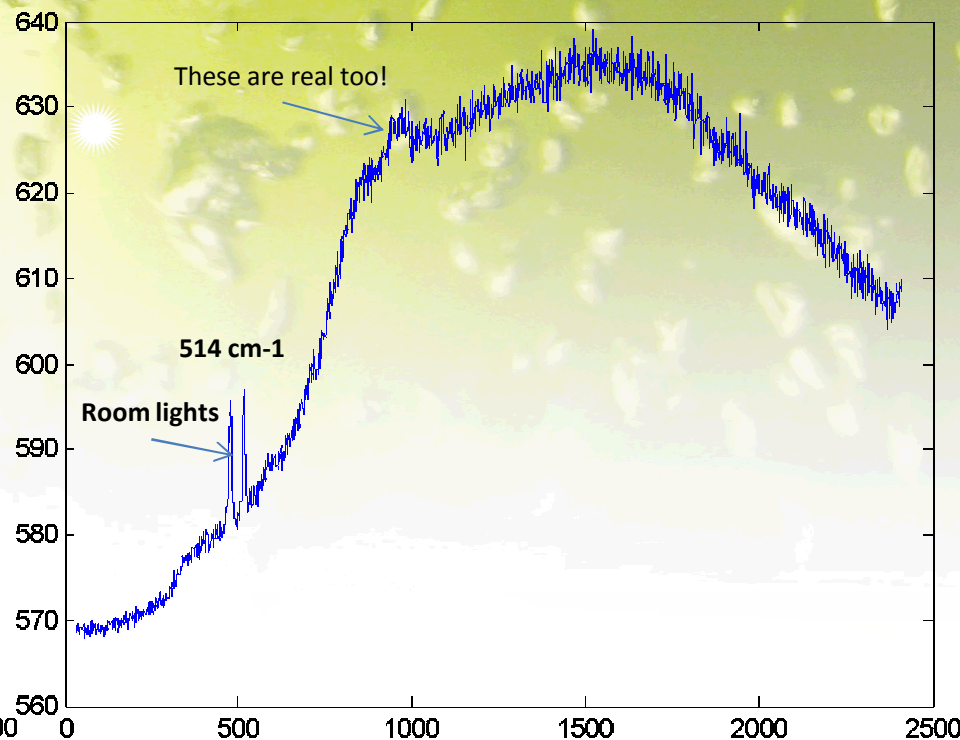
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# Cultivating Energy Solutions First Raman Spectra from Line-Imaging System

Teflon Tape



Silicon



# Conclusions / Next Steps

## • Conclusions

- Working with conversion team to improve the understanding of the conversion processes at the cellular level.
- Large differences seen between the control and the acid and enzyme pretreated biomass.
- Raman imaging provides molecular specificity
  - Increased FFA after pretreatment
  - Increased free sugars
- High-speed Raman microscopy becoming a reality!

## • Next Steps

- Continue to work with Amy Powell and NREL to provide critical conversion insights at the cellular level.
- Acid pretreatment studies
  - Test different treatment schemes
  - Conduct temporal studies
  - Provide higher spatial resolution for improved relational understanding
  - Identify additional Raman components (e.g., Protein)
- Provide Amy with Raman imaging