

A “Universal” Bio-Sample Stabilization & Preservation Medium: Simple, Low-Cost Environmental Sample Collection and Storage

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THE UNIVERSITY OF
NEW MEXICO

Far-Forward Military Need for Bio-Sample Stabilization



- Rapid in-field detectors are capable of providing preliminary analysis of biosamples
- Subsequent testing for validation or forensic analysis may be necessary
- Safe and secure collection and stabilization of biological samples would allow for:
 - ✓ Accurate identification of the biological
 - ✓ Proper treatment received for warfighters exposed to a potential biothreat
 - ✓ Vaccines received for biothreats (if available) prior to deploying forces

Photo courtesy of militarysystems-tech.com

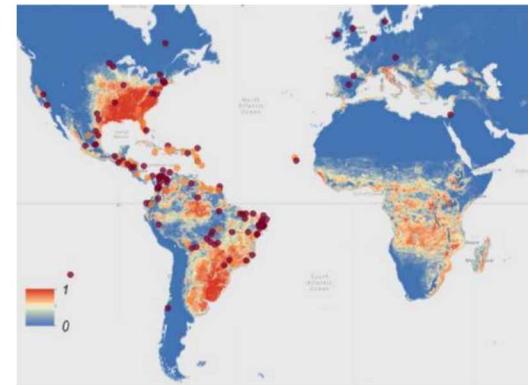
“Deadly diseases like Ebola, Marburg, and Anthrax are prevalent in Africa. These pathogens can be made into horrible weapons aimed at our troops, our friends and allies, and even the American public. This is a threat we cannot ignore.” -Senator Lugar, stated during his 2010 trip to Africa



Ebola Outbreaks 2014



Zika Outbreaks 2015-2017



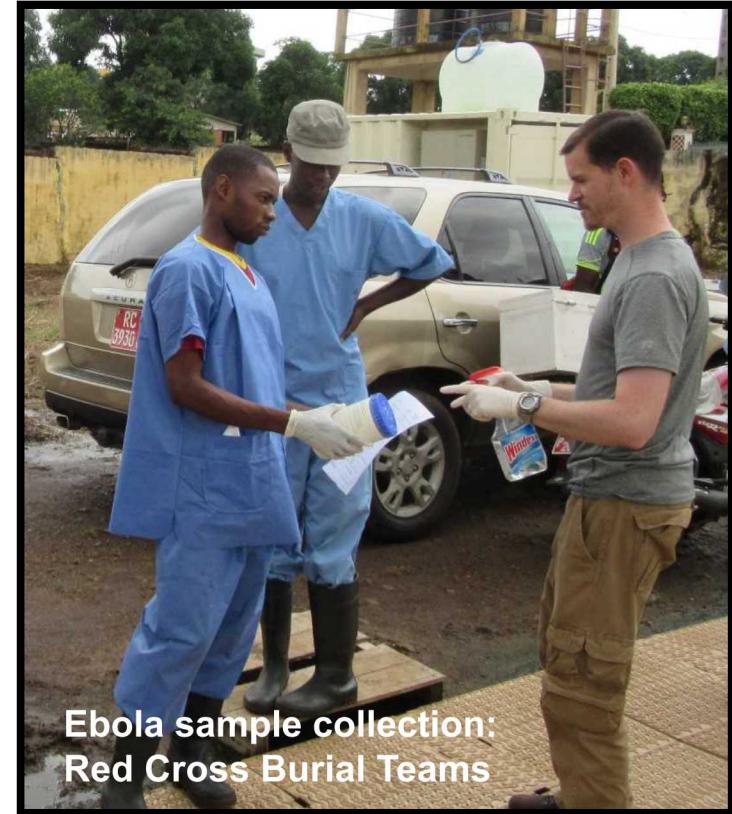
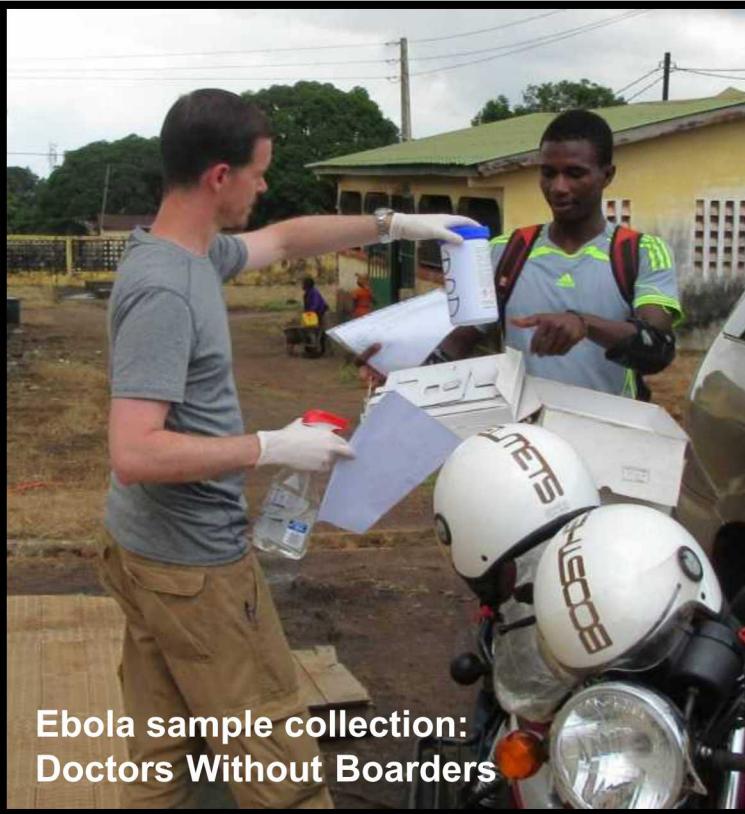
Anthrax Outbreaks 2011



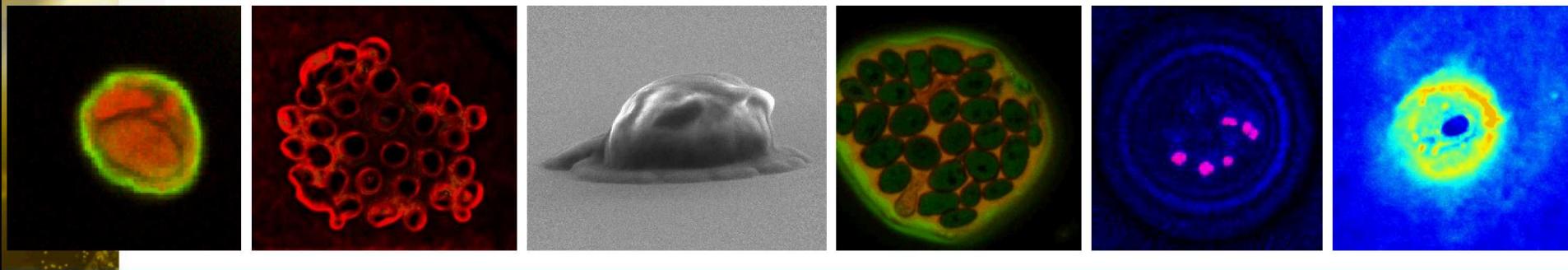
Photos courtesy of
Melissa Finley

Between 2005-2012 there were
3057 Anthrax outbreaks reported

Ebola Outbreak Response Support Conakry, Guinea



Encapsulation of Living Cells

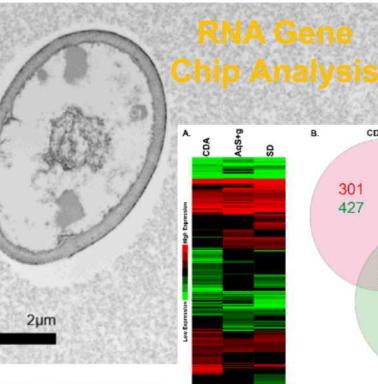
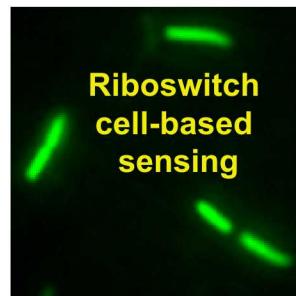
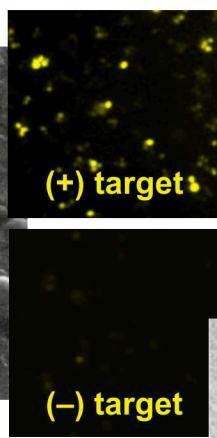
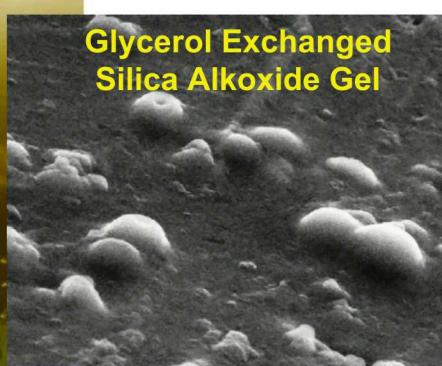


Development of Living Biocomposites – Harness unique properties innate to biomolecules and living cells via 3D immobilization within matrices that preserve cellular behavior and accessibility to cells under *ex-vivo* conditions; Provide a biocompatible interface between immobilized cells and the macro world

Encapsulation of living cells in silica matrices has attracted considerable attention as these materials are *mechanically stable, chemically and biologically inert, easily processed at room temperature, retain water with negligible swelling, resist microbial attack, and can be tailored to provide desired porosity and other material and chemical properties.*[†]

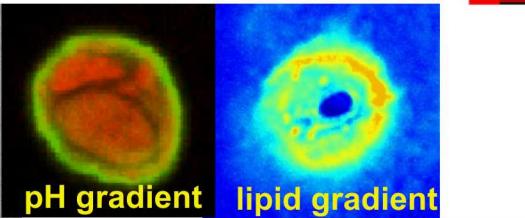
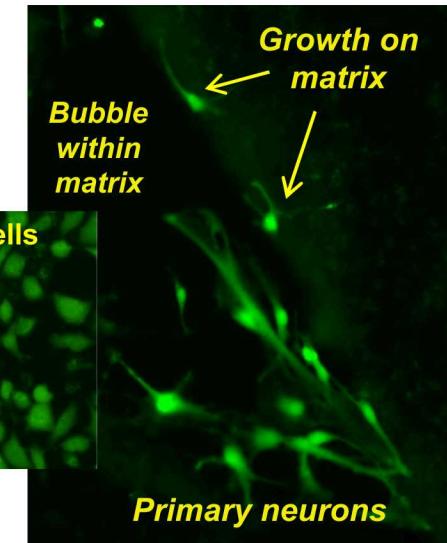
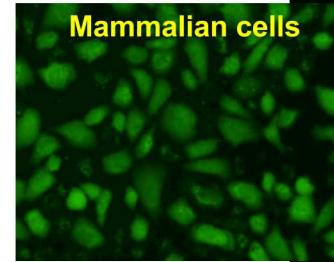
† Avnir *et al. J. Mater. Chem.* **2006**, *16*, 1013;
Nassif *et al. J. Mater. Chem.* **2003**, *13*, 203.

Technical Approach: Biocompatible 3D Silica Immobilization Strategies



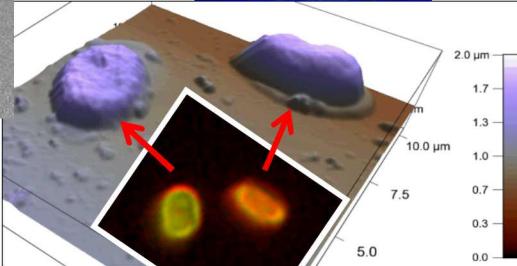
Savage et al.
ACS Biomater. Sci. Eng. 2015

Harper et al.
Chem. Mater. 2011



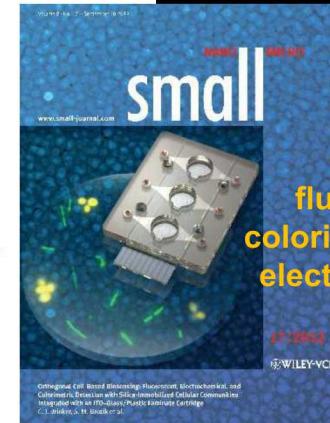
Lipid templated
mesoporous silica film

Harper et al.
ACS Nano 2010



Fazal et al.
ACS Nano
2017

Harper et al.
Small 2012



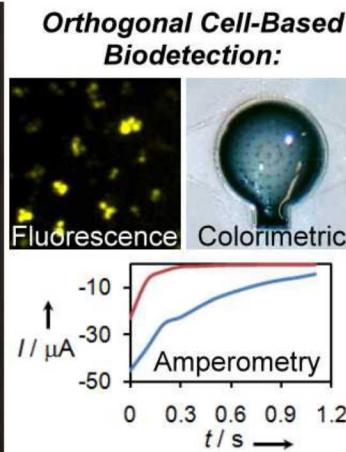
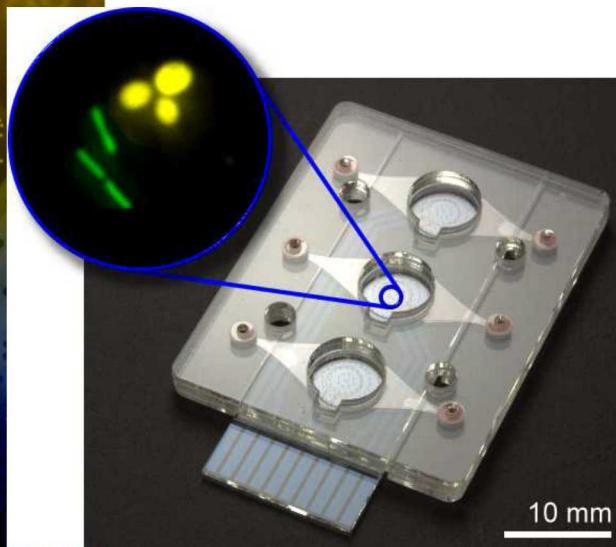
Orthogonal
fluorescence,
colorimetric,
and
electrochemical
sensing

India National Laboratories

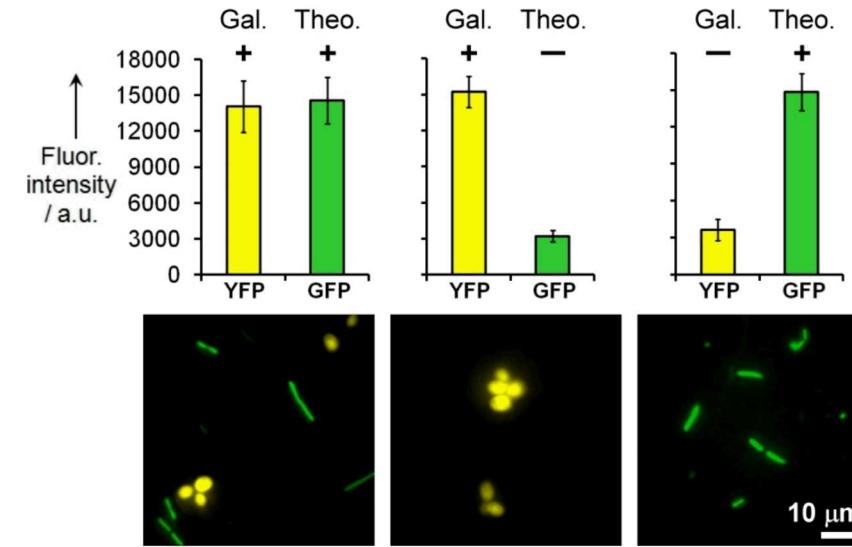
Orthogonal Cell-Based Sensing: Fluorescent, Electrochemical, and Colorimetric Detection with Silica-Immobilized Cellular Communities
H. S. Kim, C. J. Anderson, S. H. Baeck et al.

First Simultaneous Fluorescent, Electrochemical, & Colorimetric Detection via Silica Stabilized Cellular Communities

Orthogonal Fluorescent, Electrochemical, and Colorimetric Detection



Both Eukaryote and Prokaryote Cells Remain Viable and Responsive



- Multiple signals provide **complementary data, increasing confidence in output**
- Orthogonality enabled by integrating multiple cell lines

- First co-encapsulation of **eukaryote and prokaryote cells**
- Enabled multi-analyte biodetection

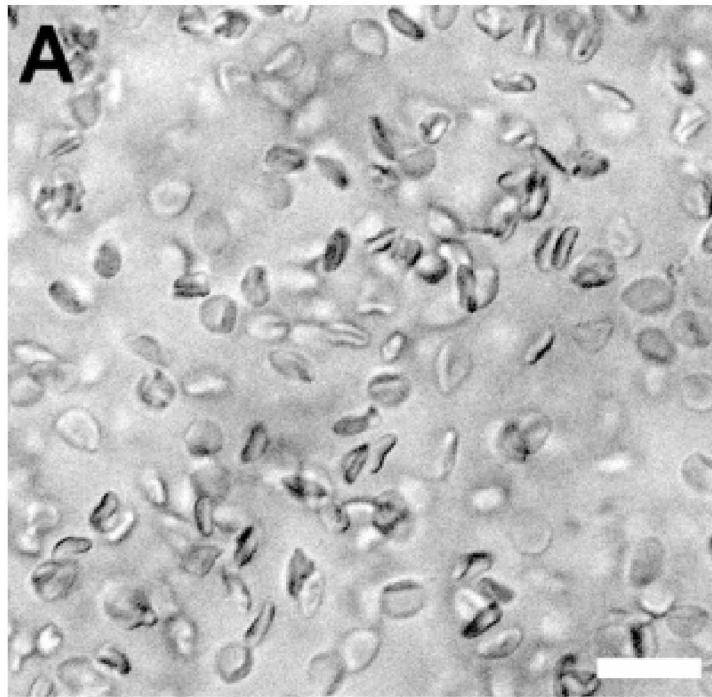
Harper et al. *Small.* **2012**, 8, 2743

First Stabilization of Human Whole Blood in a Silica Gel Monolith

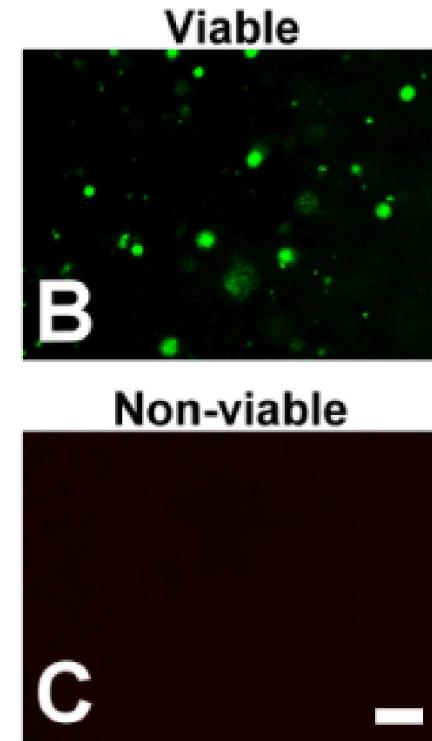
Human Whole Blood Stabilized in a Buffered Aqueous Silicate (AqS) based Inorganic Matrix



Photo of human whole blood stabilized in a AqS-based gel monolith



Bright field (black & white) image of red and white blood cells within a AqS-based matrix thin film. Scale = 20 μ m.

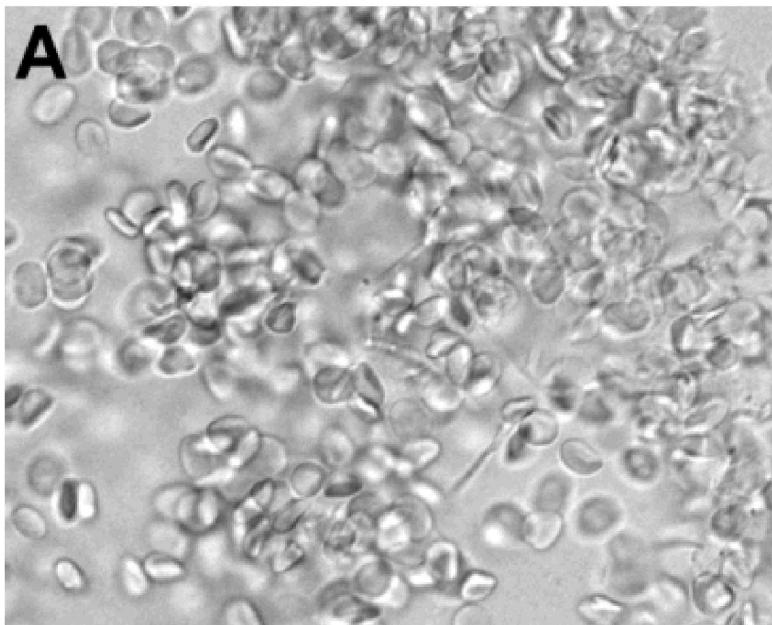


Fluorescent microscopy images of whole blood within a AqS-based matrix thin film. Scale = 50 μ m.

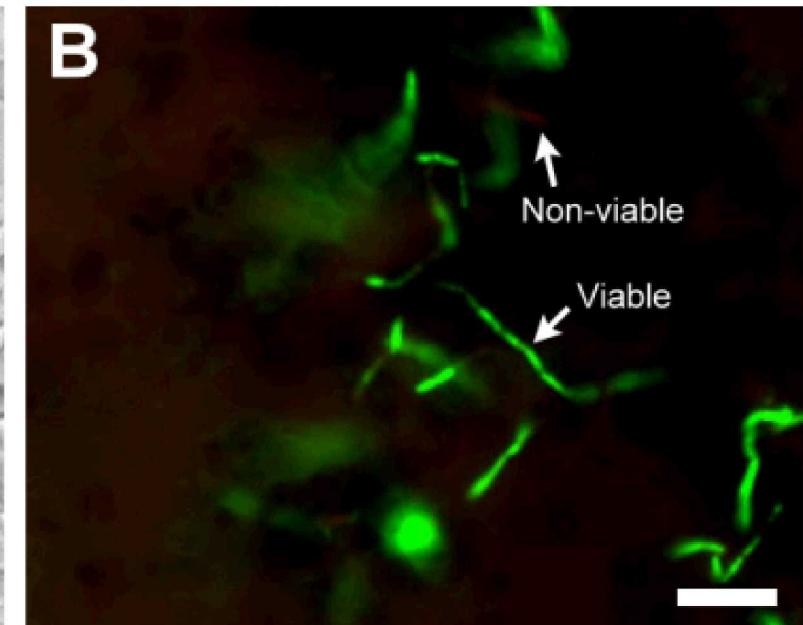
87% of white blood cells survive encapsulation and remain viable

First Co-Stabilization of Human Whole Blood & Bacteria Pathogen in Silica Matrix

Human Whole Blood and *Bacillus anthracis* (Sterne) Stabilized in a Buffered Aqueous Silicate (AqS) based Inorganic Matrix



Bright field (black & white) image of human whole blood with *B. anthracis* (vegetative) stabilized within a AqS-based matrix thin film.

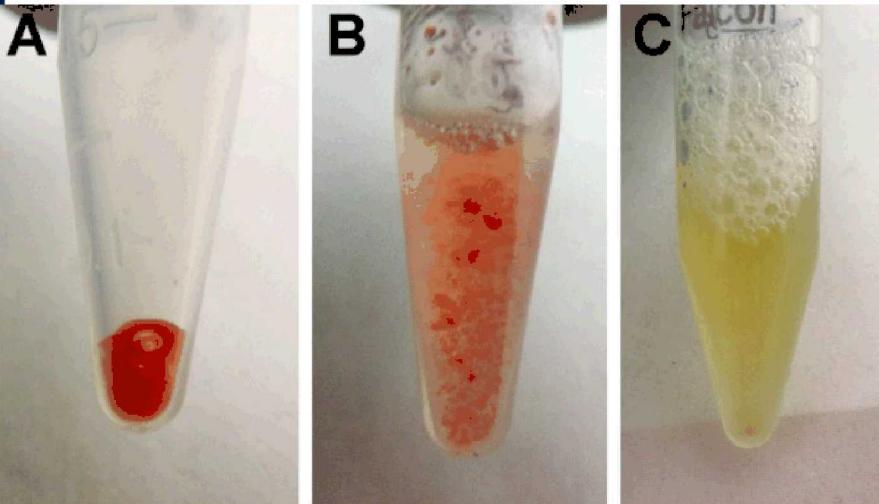


Merged fluorescence microscopy images of vital dye stained human whole blood and *B. anthracis*. Green stained bacteria and white blood cells are viable; red stained are non-viable. Scale = 20 μ m.

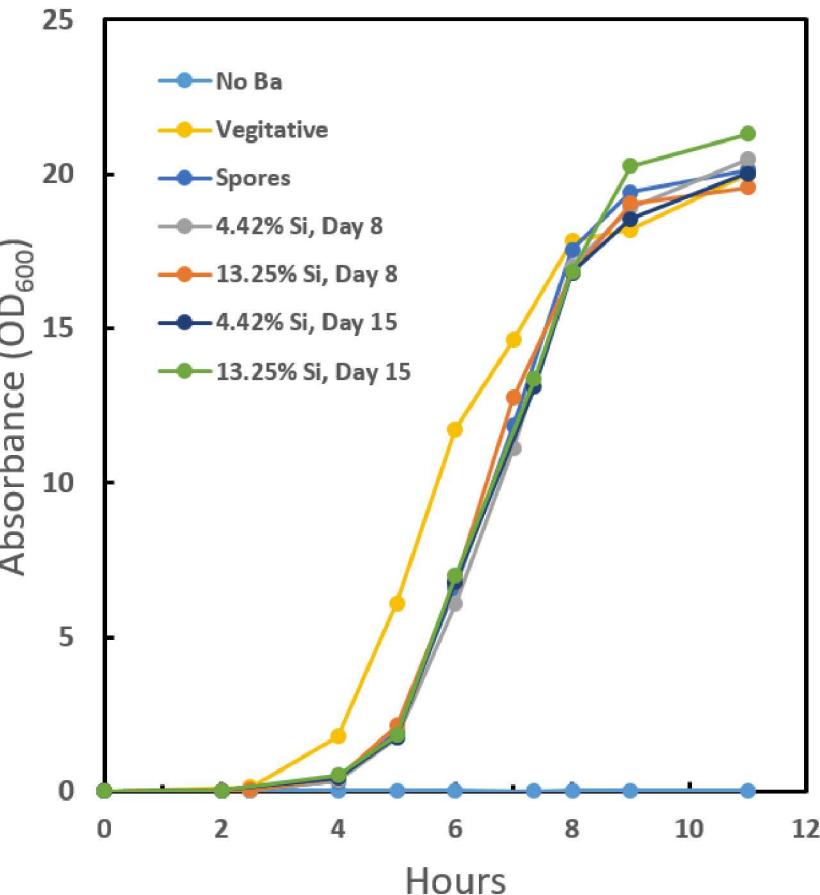
92% of *B. anthracis* cells survive encapsulation and remain viable

Extraction of Viable and Culturable *B. anthracis* from an Inorganic Silica Matrix

- Human whole blood spiked with *B. anthracis*, Sterne (vegetative), stabilized within an AqS-based silica matrix monolith
- Stored under ambient conditions for 3 days (22 C)
- Sample extracted w/ spatula, vortexed, resuspended in Heart Infusion (HI) media



Growth Curves Following Extraction of Stabilized Bio-Samples



Inoculated in 50 mL HI, 37 C incubation, 200 rpm

Culture on Solid Medium of *B. anthracis* Extracted from AqS-based Matrices

- Stabilized samples extracted with spatula, vortexed in 150 μ L of 1x PBS, pH 7. This solution was plated (100 μ L) and incubated overnight at 37 C.

+ Whole blood; + *B. anthracis*

+ Whole blood;
- *B. anthracis*

3 Days

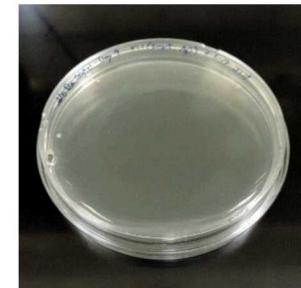
5 Days

8 Days

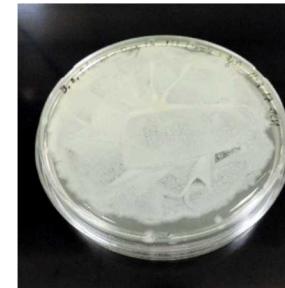
15 Days

9 Days

4.42%
Silica



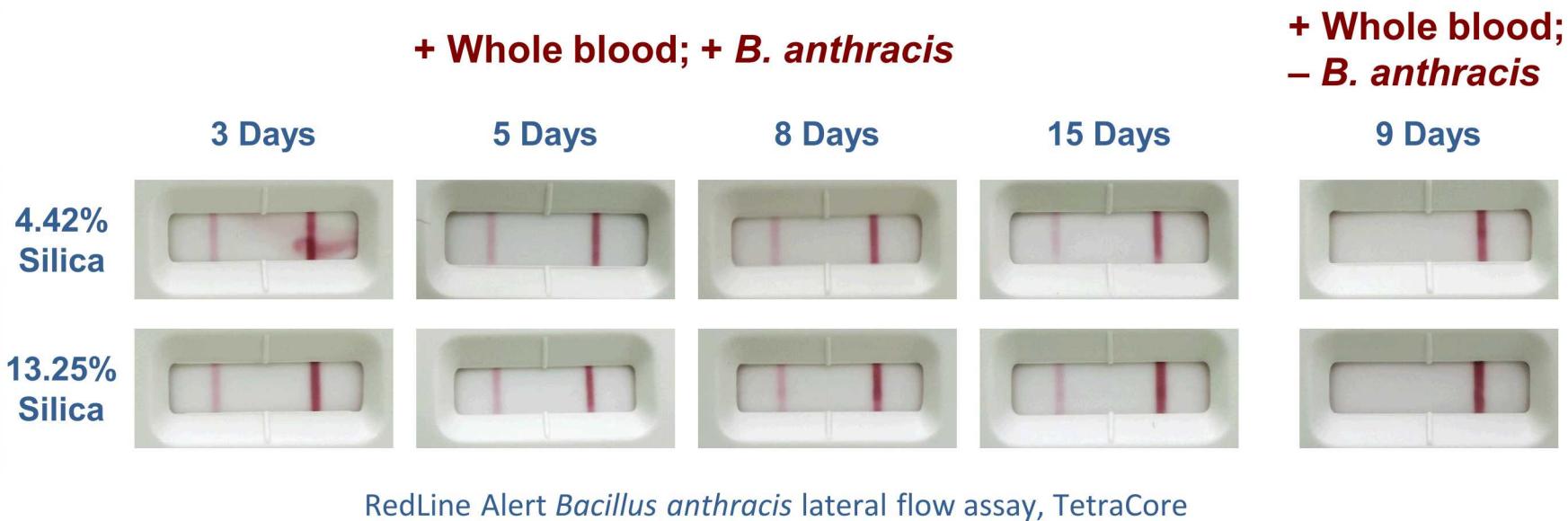
13.25%
Silica



Heart Infusion (HI) agar plates

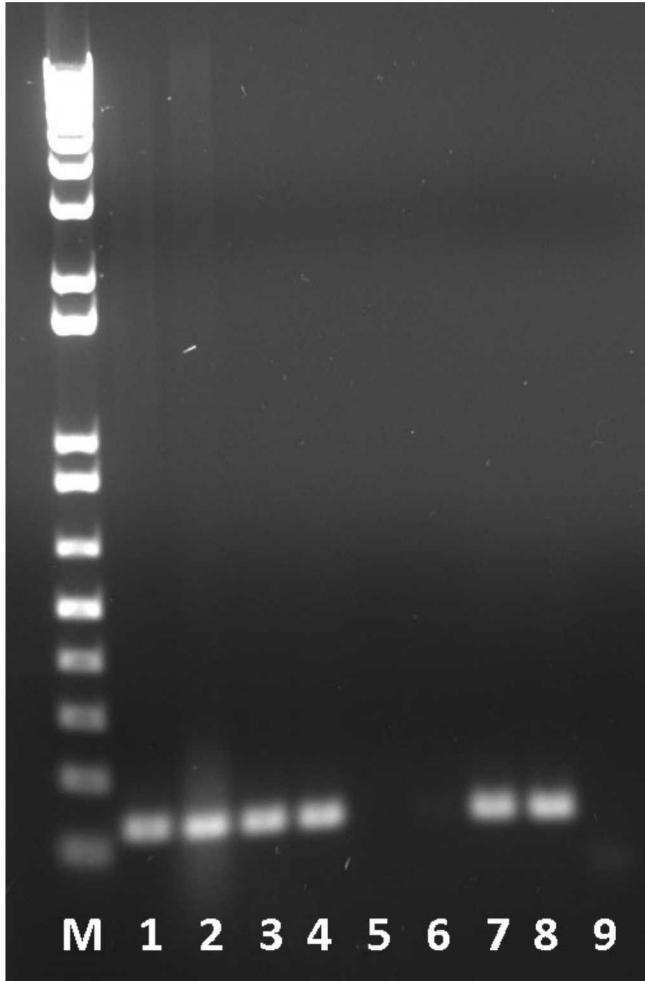
Use of Gold Standard Affinity Assay to Positively Identify Extracted *B. anthracis*

- Following overnight incubation on HI-agar, a colony was picked from the plate, resuspended in colony isolation buffer, and loaded onto a lateral flow assay.



Stabilized Samples Sent from Albuquerque to Omaha; PCR of Extracted Sample

PCR Products from Extracted Sample Identify Presence of *B. anthracis*



- Stabilized samples shipped from Sandia Labs to Univ. of Nebraska Medical Center **without ice/dry ice**; stored at ambient for 3+ weeks

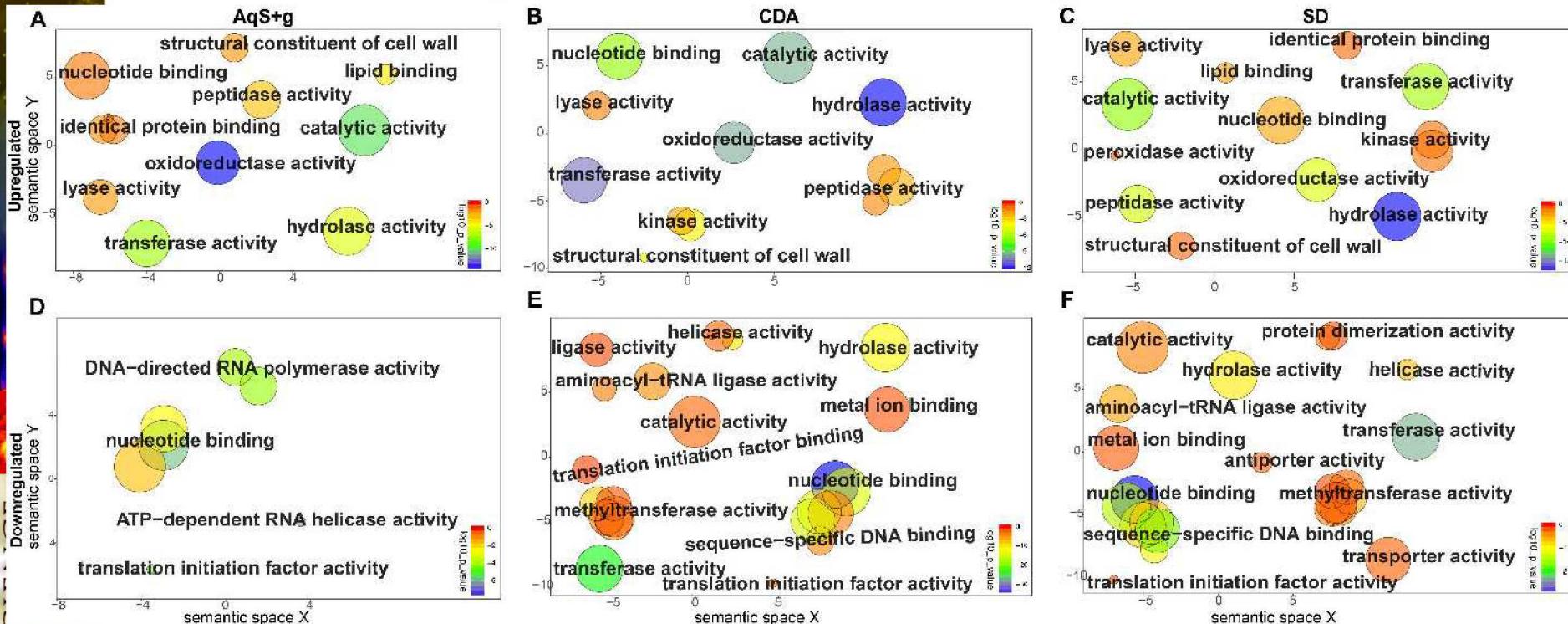
M: Marker, 1kD DNA ladder

- 1: *B. anthracis* genomic DNA (+ control)
- 2: DNA extracted from *B. anthracis* culture (+ control)
- 3: Si Stabilized + *B. anthracis*, cultured day 18, pelleted and frozen, DNA extracted on day 25; 4.42% Si
- 4: Si Stabilized + *B. anthracis*, cultured day 18, pelleted and frozen, DNA extracted on day 25; 13.25% Si
- 5: Si Stabilized – *B. anthracis*, cultured day 18, pelleted and frozen, DNA extracted on day 25; 4.42% Si
- 6: Si Stabilized – *B. anthracis*, cultured day 18, pelleted and frozen, DNA extracted on day 25; 13.25% Si
- 7: Si Stabilized + *B. anthracis*, DNA extracted directly on day 25; 4.42% Si
- 8: Si Stabilized + *B. anthracis*, DNA extracted directly on day 25; 13.25% Si
- 9: No template (– control)

Use of RNA Gene Chip Assay to Probe Metabolic State of Si Matrix Entrapped Cells

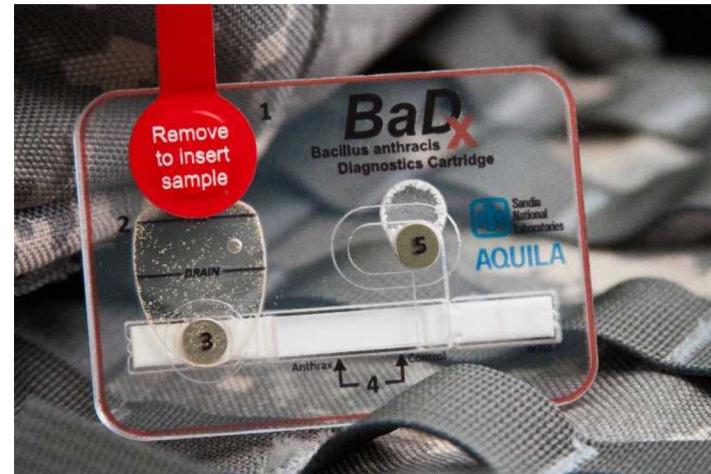
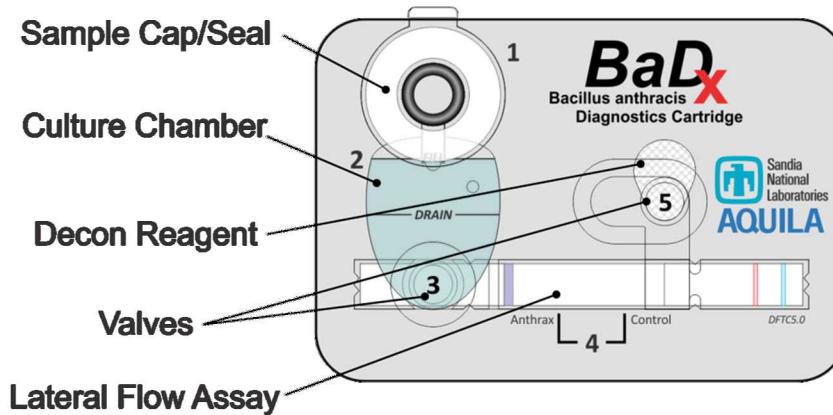
- *S. cerevisiae* cells stabilized in 3 differing silica matrices
- Cells extracted and RNA gene chip assay performed (Affymetrix) to measure gene expression profiles

Biological Processes Gene Ontology Terms



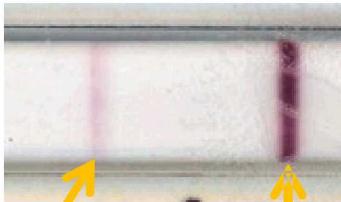
Frazal, Harper et al. ACS Nano 2017, 11, 3560

B. anthracis Diagnostics (*BaD*_X) Cartridge



Self-contained, credit card-sized “Laboratory in a Pocket” for bacterial detection, containment, and destruction

Ames (BSL 3) 100 spore inoculum



B. anthracis
detected

Assay
control

- ✓ Field deployable
 - Credit card sized
 - Rugged/robust
- ✓ No power to operate
- ✓ No instrumentation or equipment to operate/read
- ✓ *Detection as low as 1 viable spore demonstrated*

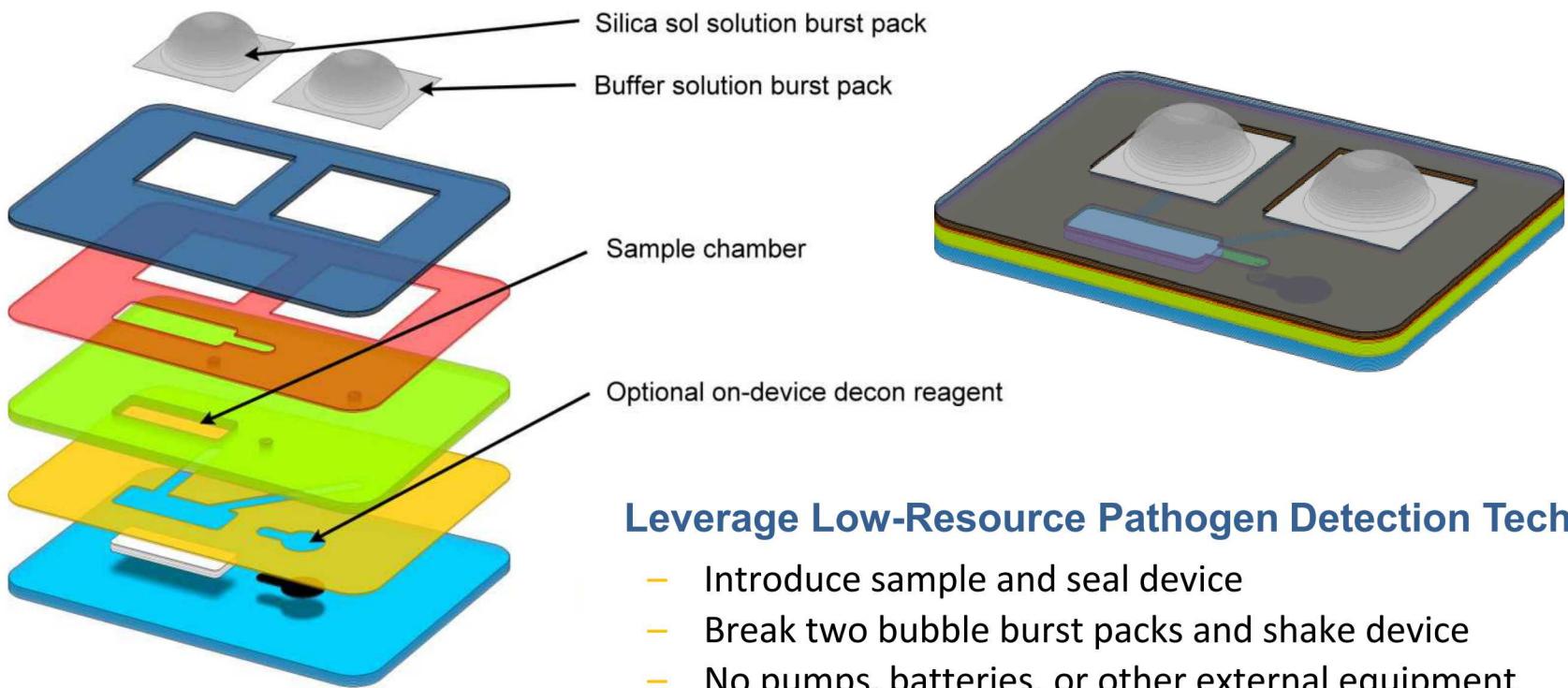
- ✓ Self-sterilizing upon assay completion
- ✓ Operable by individuals with little to no technical training

Commercially available – Aquila Technologies: \$98.99



Use of Bio-Sample Stabilization Chemistry in Far-Forward or Low-Resource Settings

Proof-of-Concept Universal Bio-Sample Stabilization Device



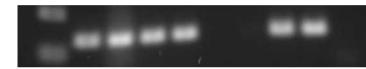
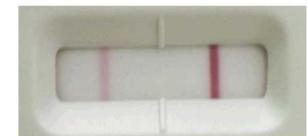
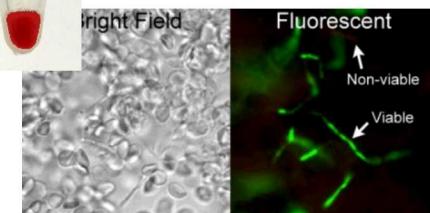
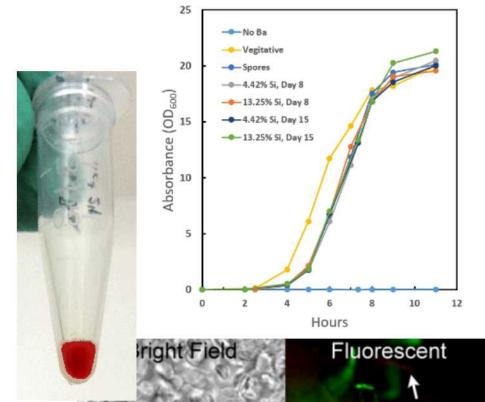
This device could serve as a back-end to current far-forward detectors, or as a stand-alone unit

Leverage Low-Resource Pathogen Detection Tech

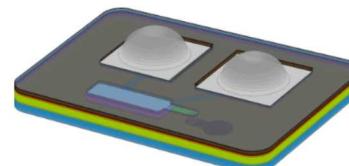
- Introduce sample and seal device
- Break two bubble burst packs and shake device
- No pumps, batteries, or other external equipment required to operate
- Low cost: < \$2 chemistry; < \$5 BOM
- High stability - chemistry stable for several years w/o cold chain

Summary & Future Work

- ***First stabilization of human whole blood in an inorganic silica matrix***
 - 87% of white blood cells remain viable post-encapsulation
- ***First co-stabilization of human whole blood and pathogen (*B. anthracis*) in an inorganic silica matrix***
 - 92% of *B. anthracis* remain viable post-encapsulation



- ***Extracted viable and culturable samples following storage under ambient conditions for 2+ weeks***
 - Positive culture on liquid & solid medium
 - Positive identification of pathogen by affinity assay (LFA)
 - Positive ID of pathogen via PCR after shipping from Albuquerque to Omaha w/o cold chain, 25 days storage
 - Extracted nucleic acid used with standard RNA gene chip



- Extend to other select agent pathogens:
 - RNA virus, DNA virus, other bacteria
- Test in other complex matrices:
 - Oral swab, mucosal swab, urine, fecal matter, tissue
- Develop hand-held, low cost device for stabilizing bio-samples

Acknowledgements

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