

# Elucidating Genetic Mechanisms of Bacterial Pathogenesis for Host-Directed Protection

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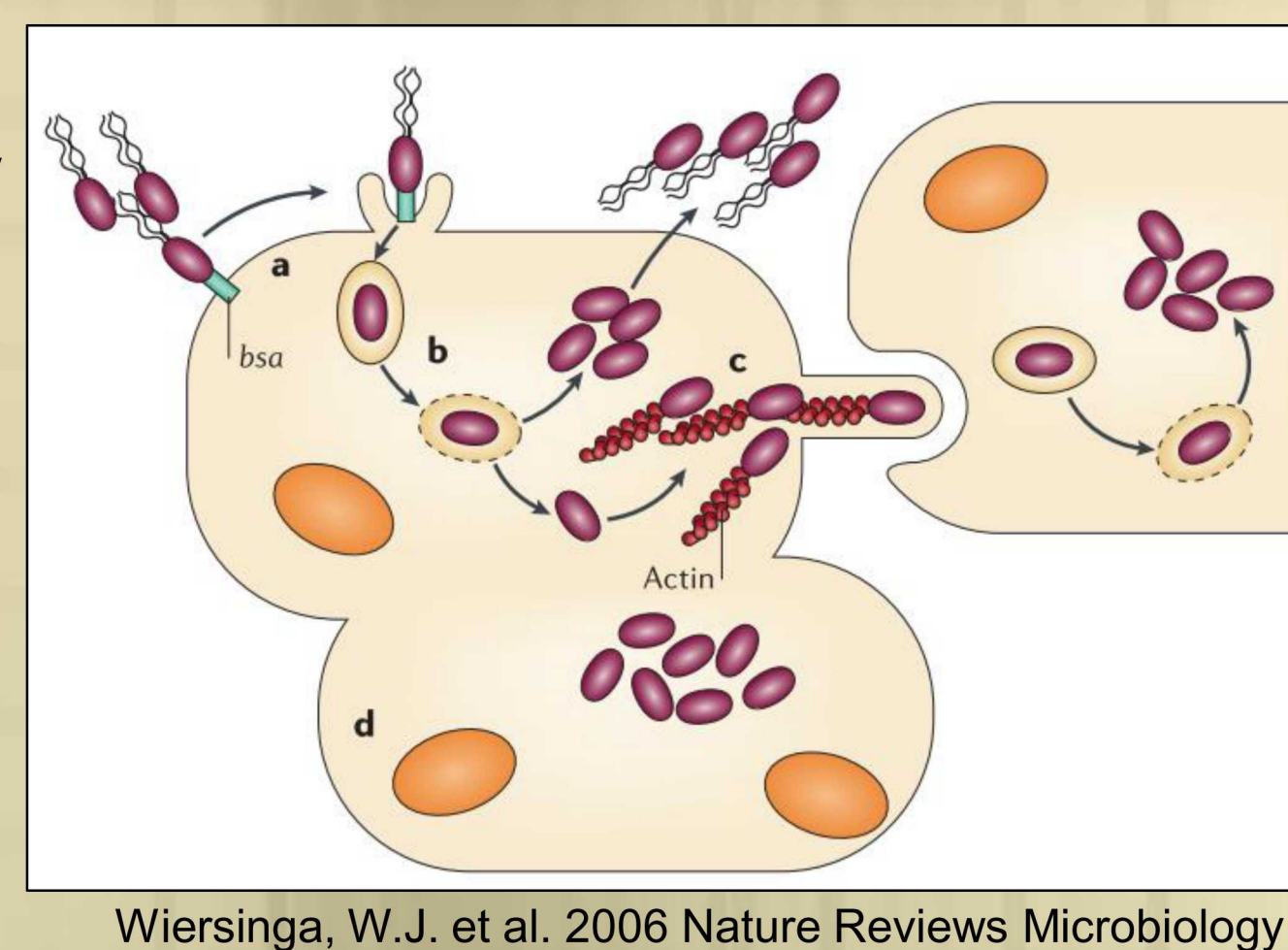
## Motivation and Approach

### Why *Burkholderia*?

- Naturally resistant to most commonly used antibiotics
- Highly virulent, especially via airway
- Readily isolated from environment
- High-risk bioterror agent

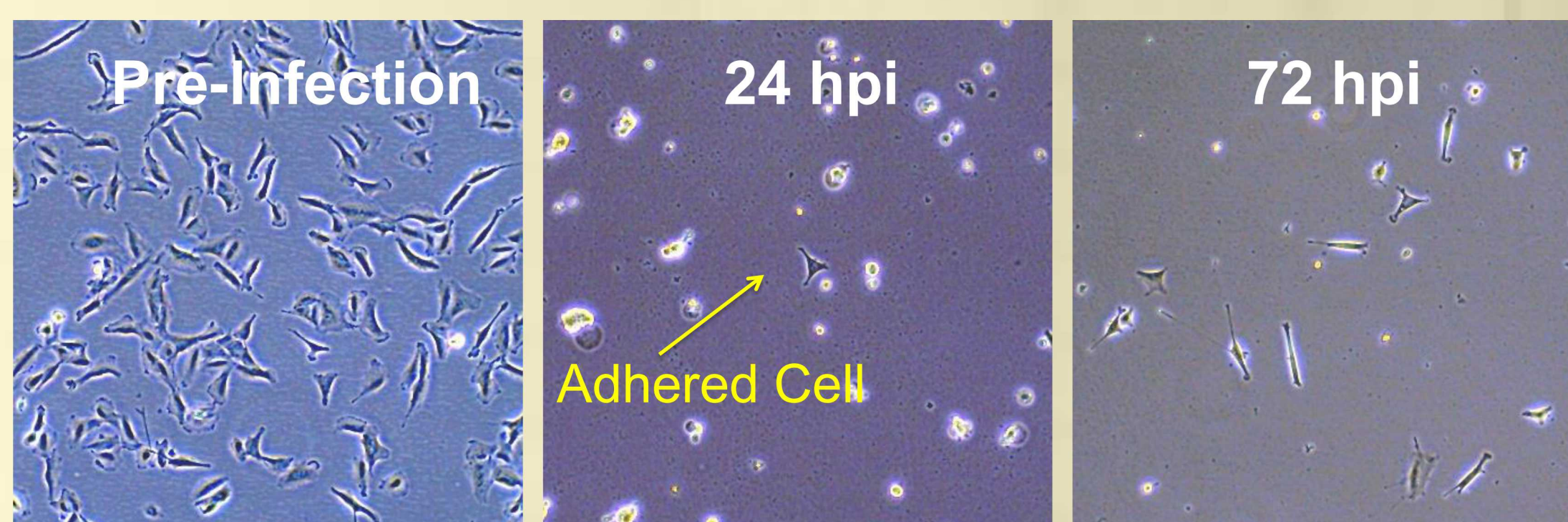
### Our approach:

- Use a Genome-Scale CRISPR-Cas9 Knockout (GeCKO) Screen to find genes that aid in host protection
- Develop assays to confirm their protective action and elucidate genetic mechanisms in pathogenesis
- Transition validated, protective genes to murine primary cell, *ex vivo* lung/airway explant, and *in vivo* validation.



Wiersinga, W.J. et al. 2006 Nature Reviews Microbiology

## Host GeCKO Screen



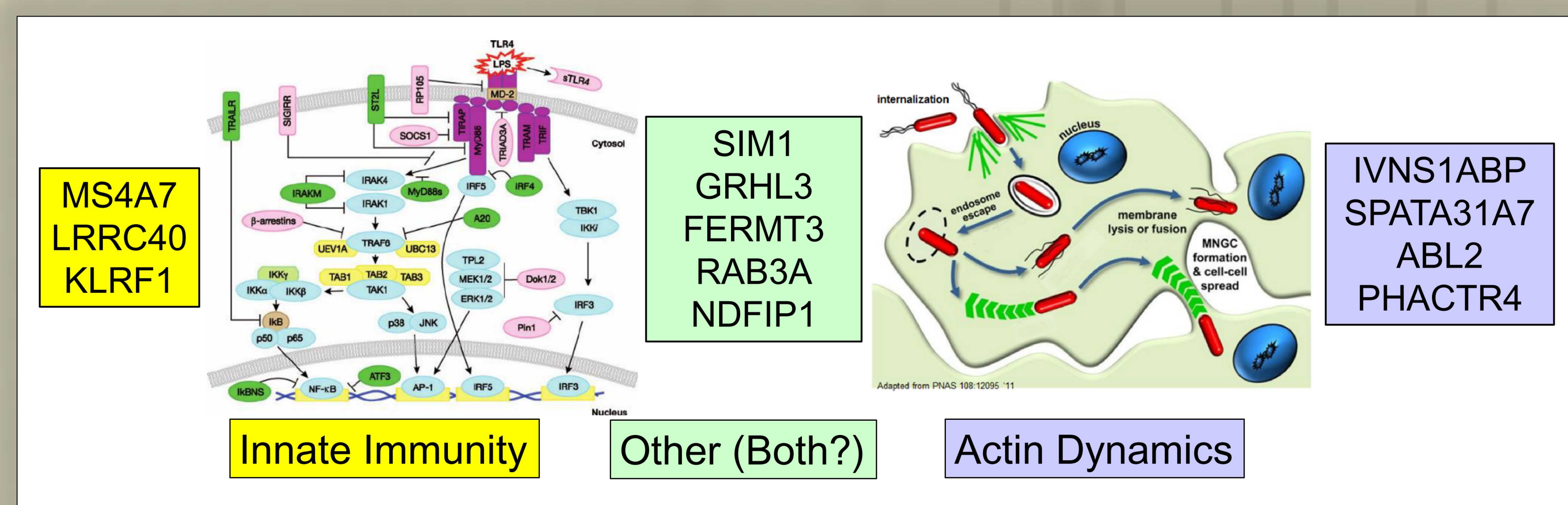
sgRNAs Hitting Protein-Coding Gene	Protein-Coding Genes	Average Number of Infections from which sgRNA was Recovered							
1	5325	3580	325	184	169	184	246	225	412
2	817	370	101	91	122	93	21	11	8
3	79	32	11	27	5	2	0	2	0
4	5	3	1	1	0	0	0	0	0
5	1	0	0	1	0	0	0	0	0
6	0	0	0	0	0	0	0	0	0

Genes carried on for assay Second priority gene targets

Identify gene knockouts that protect human airway epithelial cells from *B. thailandensis* (*Bt*)

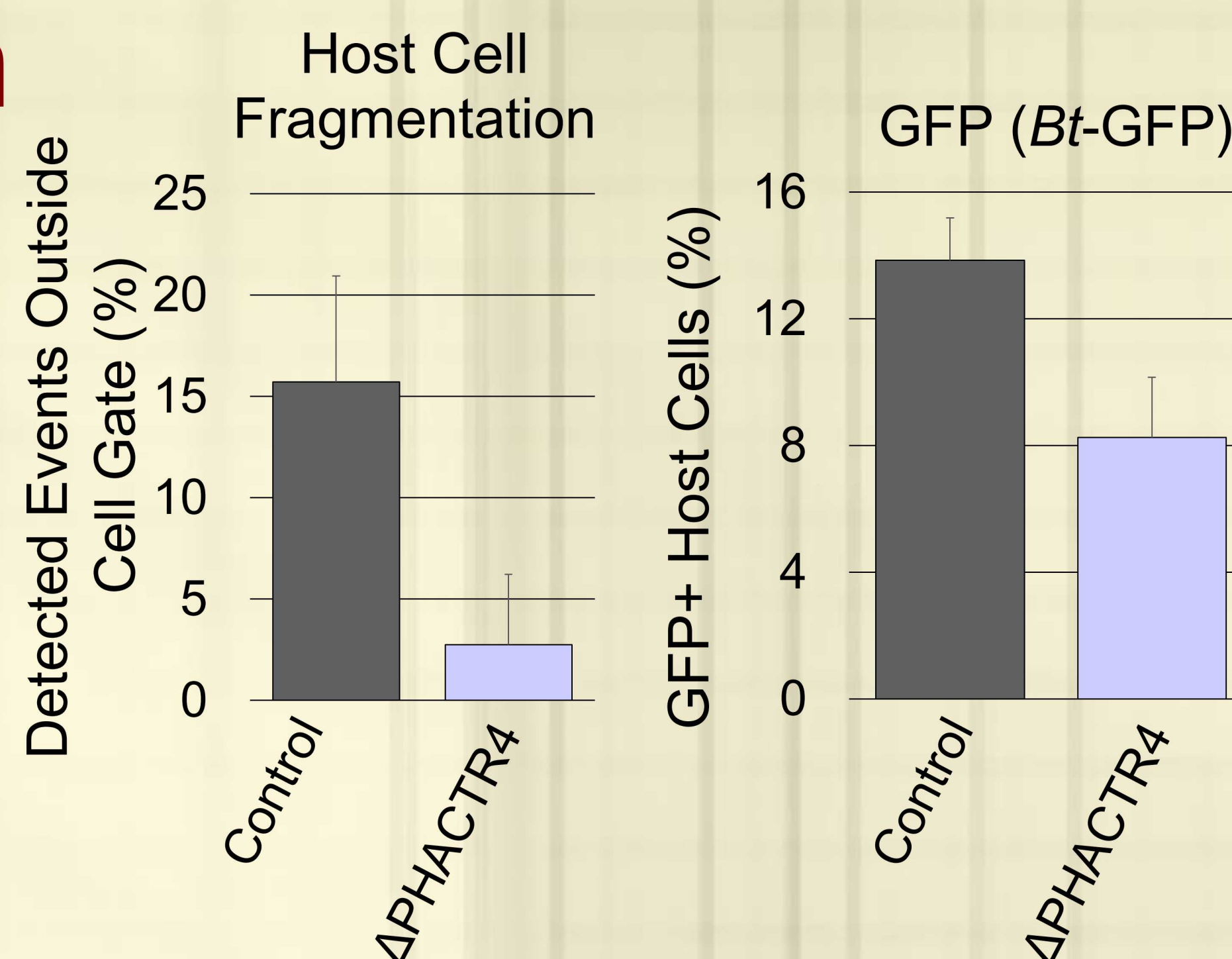
- 8 independent infections in 3 screens
- 24 hpi allow cells to recover
- 72 hpi collect for DNA

## Host Defense Targets



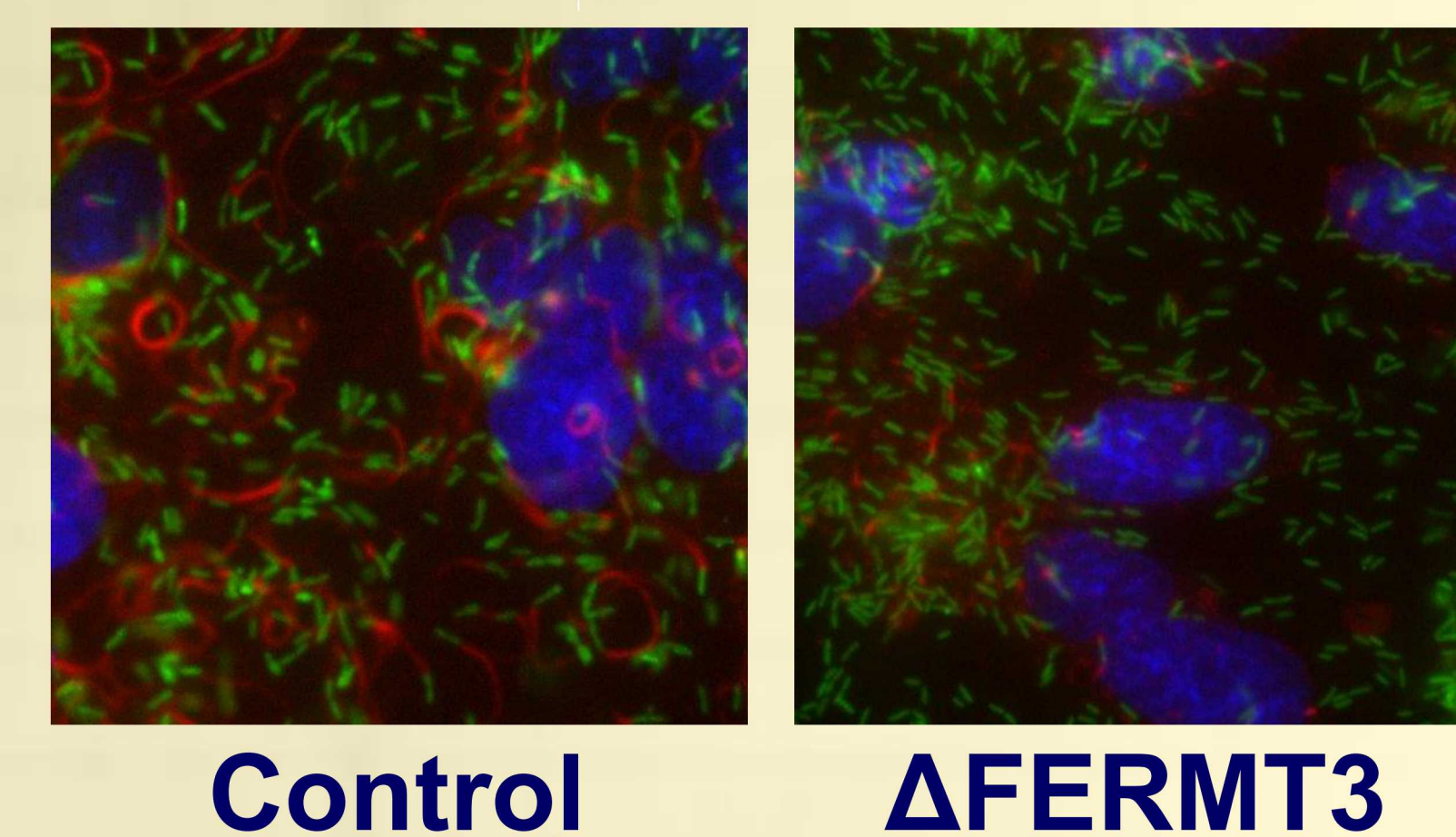
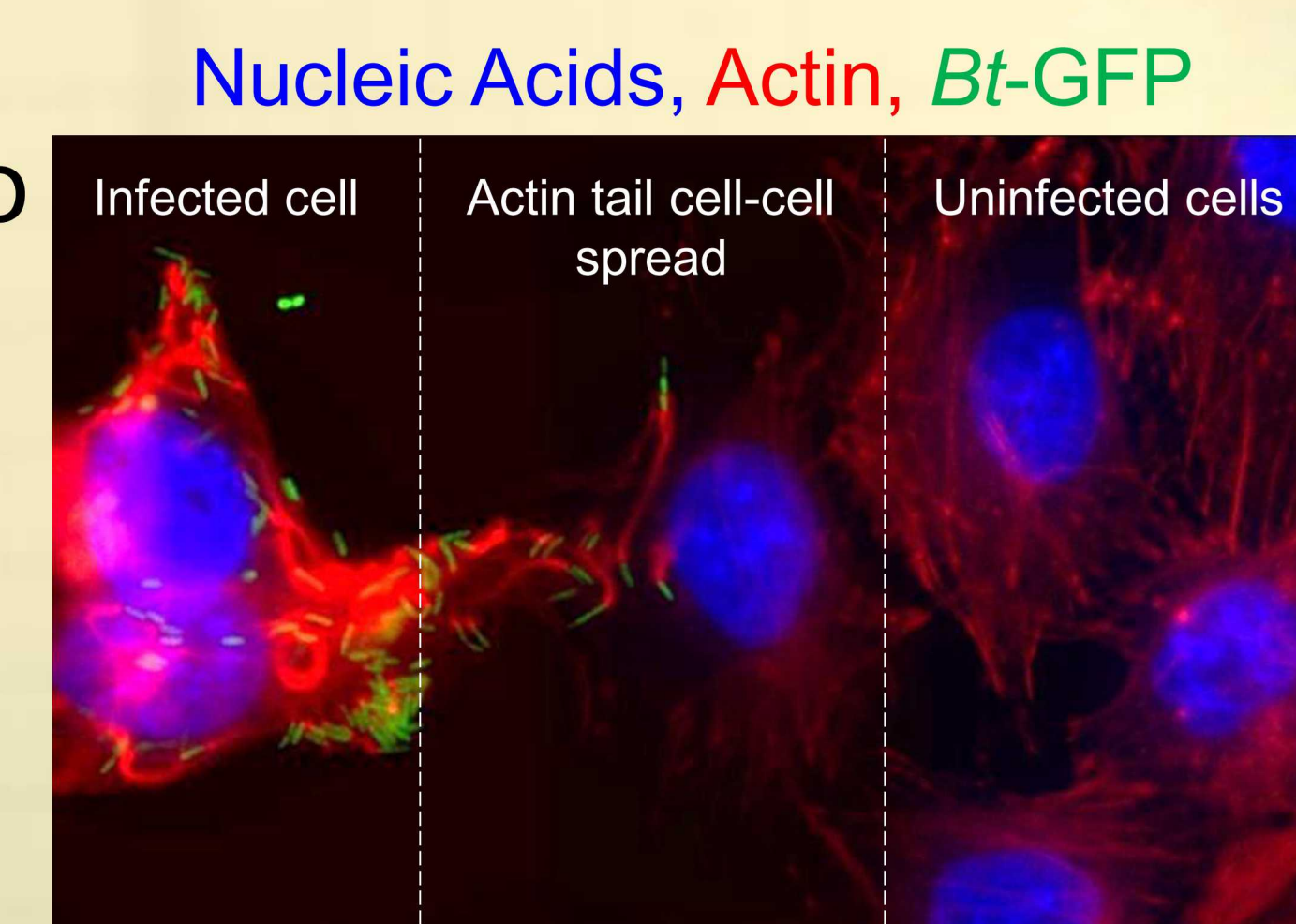
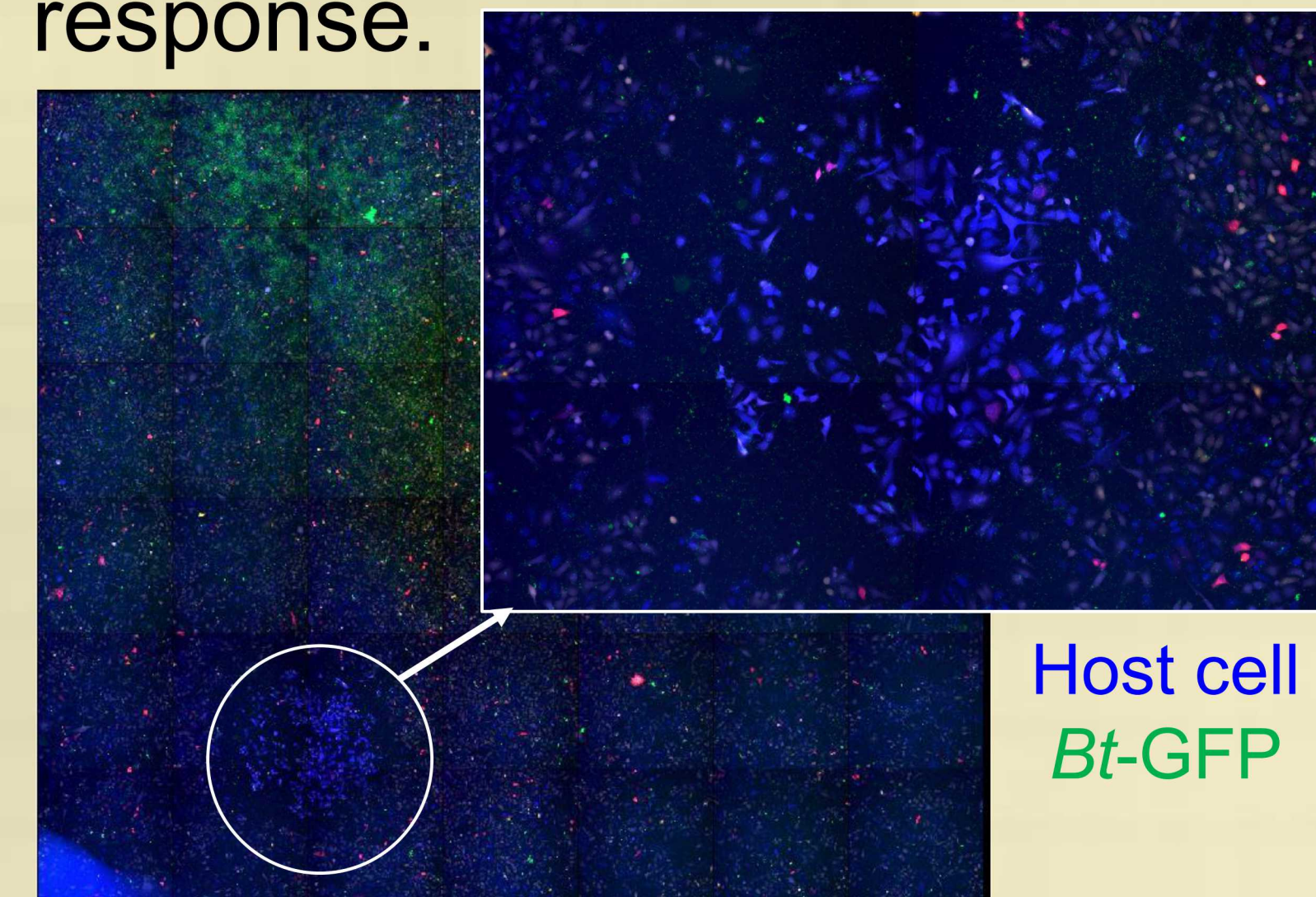
## Cell Fragmentation and Bacterial Internalization

Cell health and bacterial internalization (GFP signal) are being assayed using flow cytometry. Less fragmentation and GFP, like the PHACTR4 example, indicates greater protection by gene knockout.

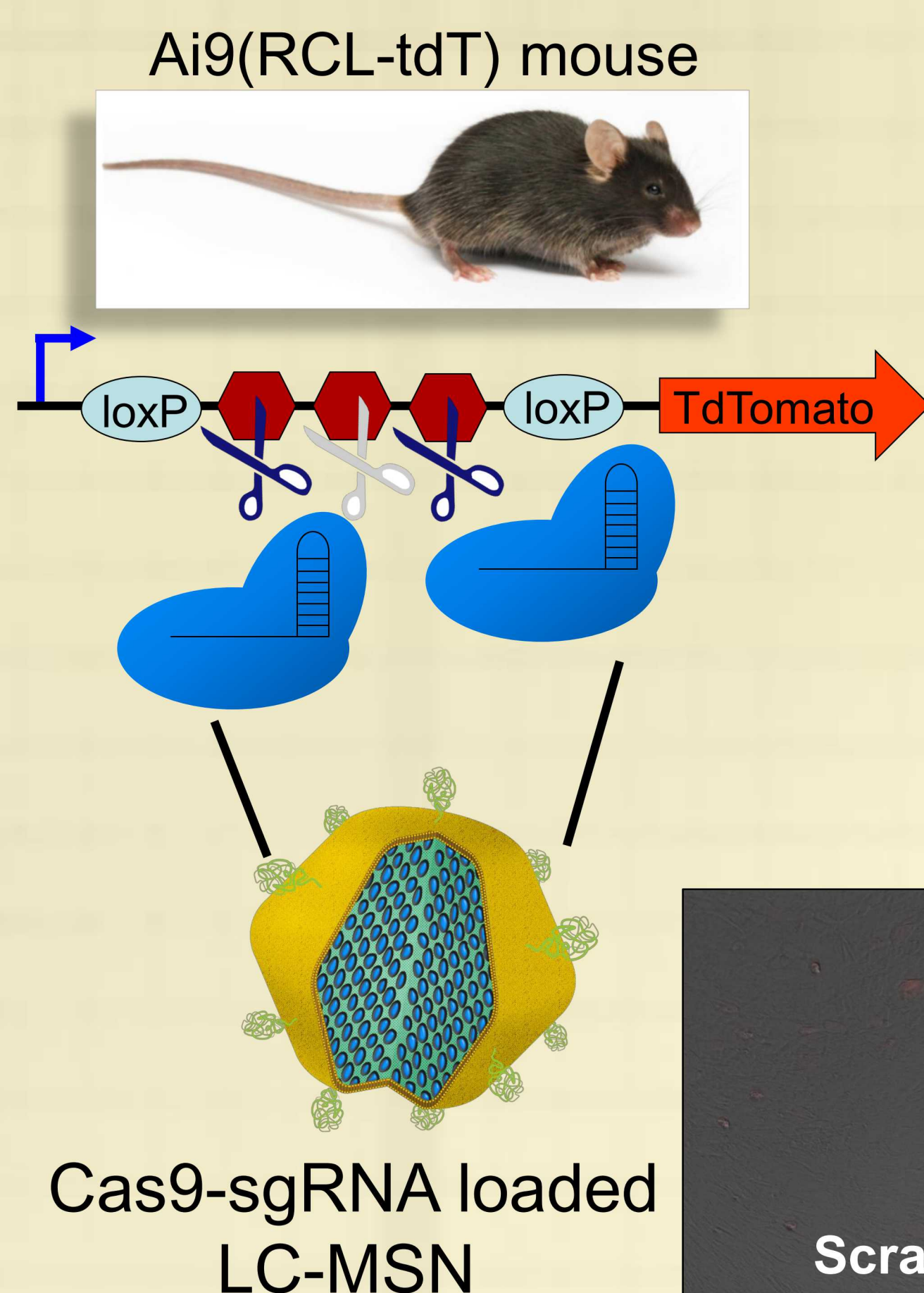


## Actin Tail motility

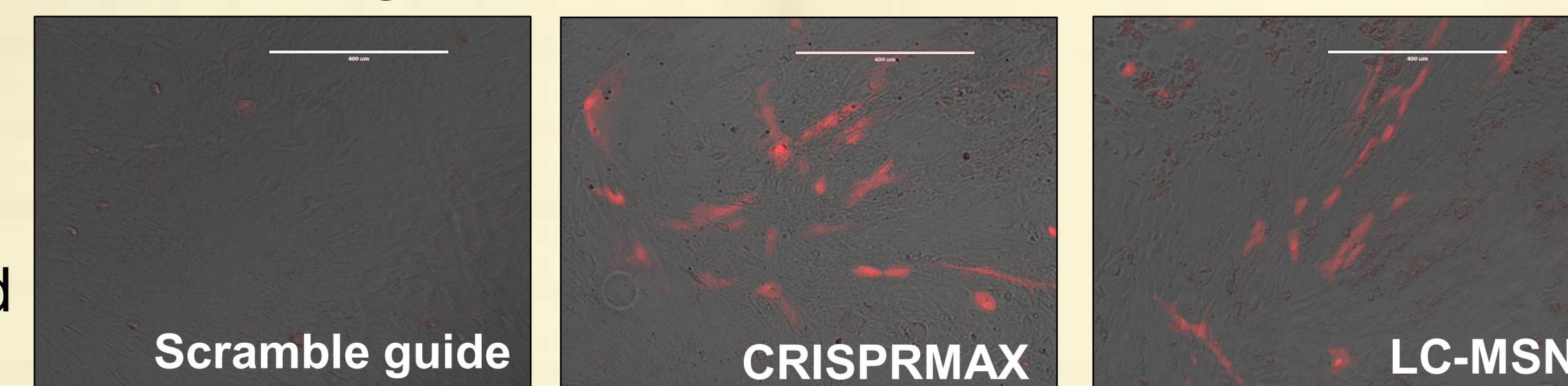
Actin tails are a signature of *Bt* pathogenesis and allow the bacteria to propel within the host and cause cell-cell spread. Presence and length of actin tails (right) and cell-cell spreading (bottom) elucidates host response.



## Model Mouse for Gene Editing

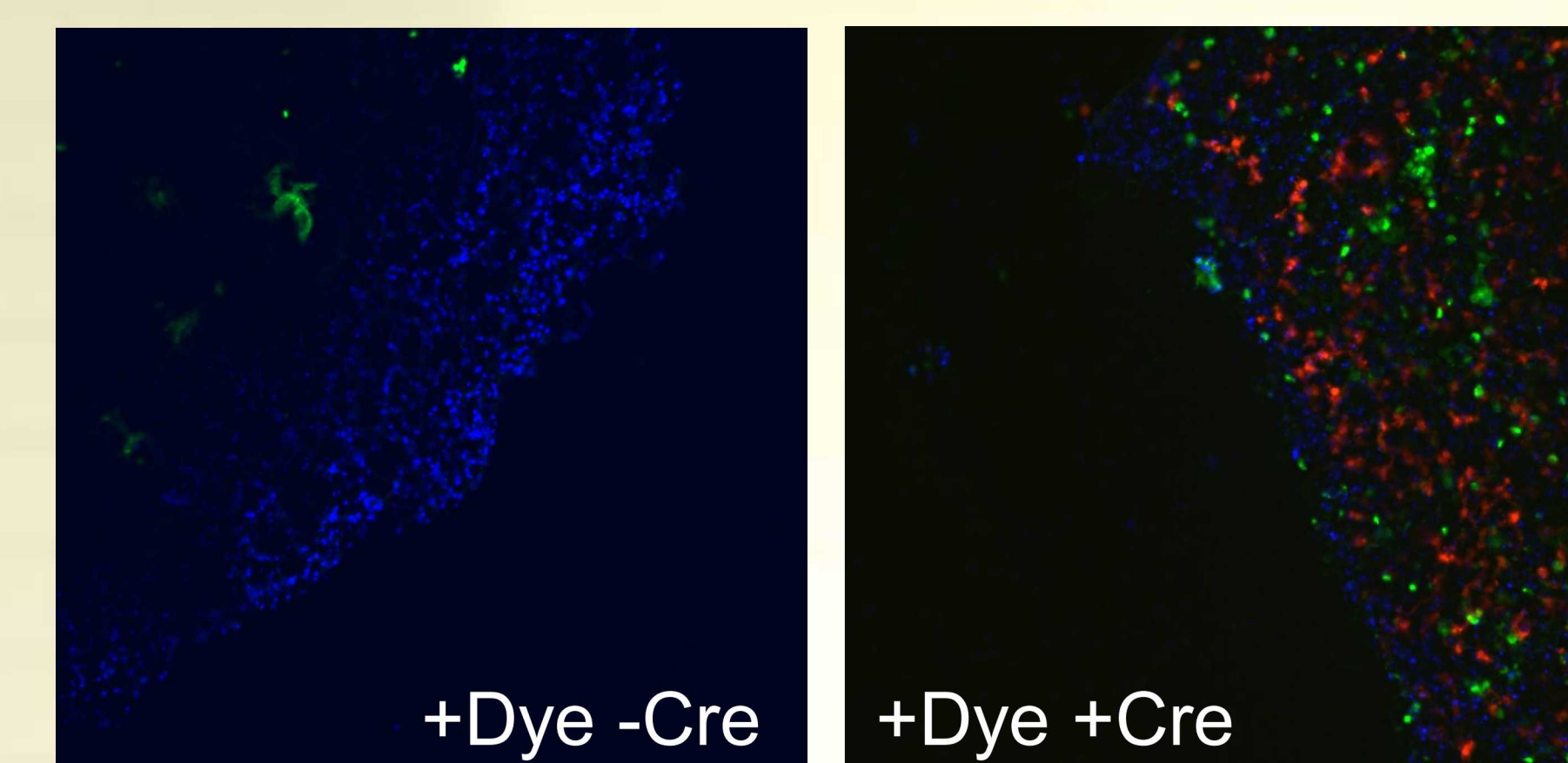
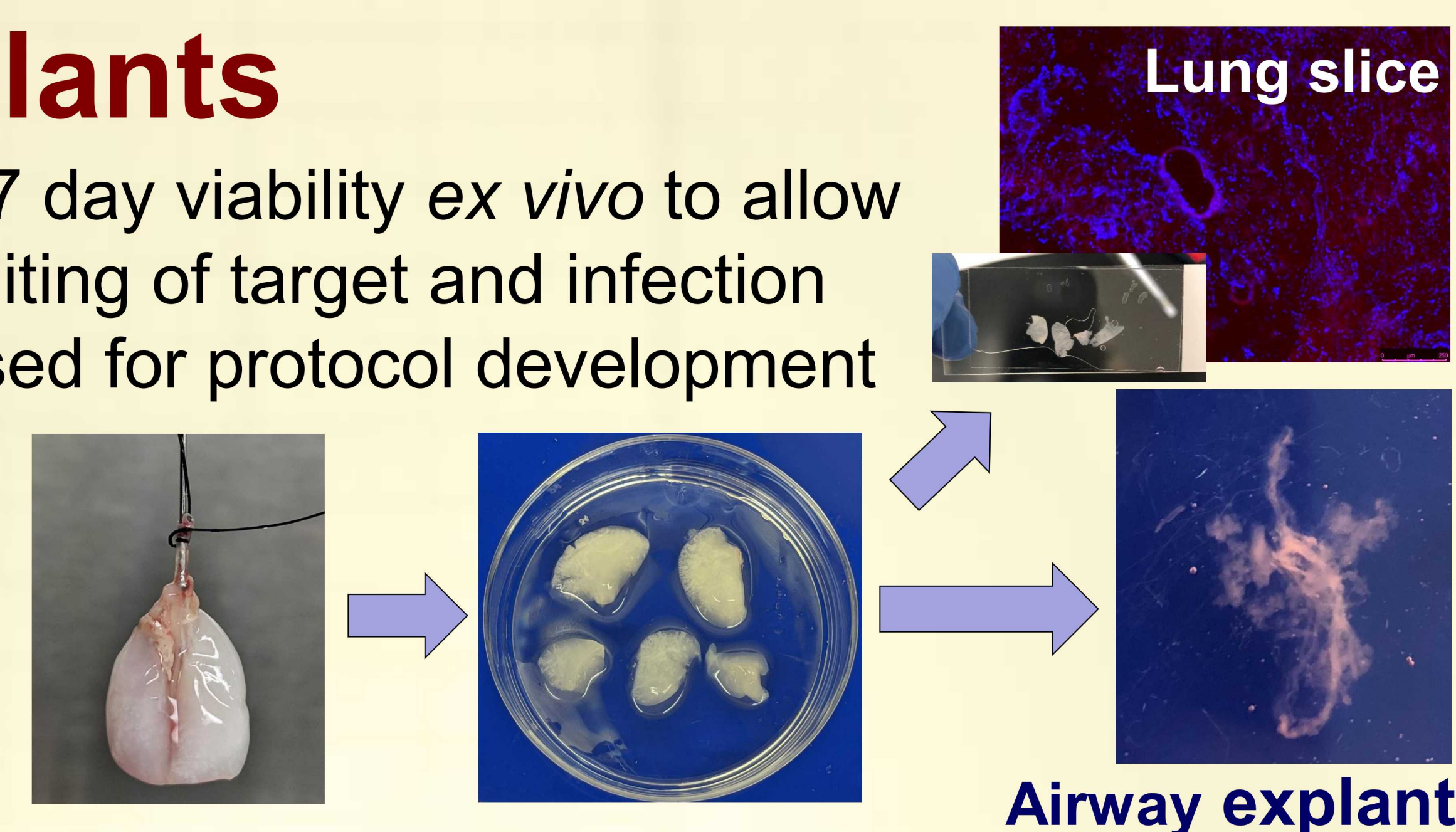


The construct in Ai9 mice can be targeted with one sgRNA that cuts twice and removes a stop cassette allowing for constitutive expression of TdTomato (red) fluorescent protein. We are using the Ai9 mouse to evaluate our protein delivery and gene editing efficiency for an easy gain of signal fluorescent readout.



## Lung Explants

Need to achieve 4-7 day viability *ex vivo* to allow for CRISPR/Cas editing of target and infection with *Bt*. Ai9 mice used for protocol development for gain of signal editing and quick protein delivery readout with Cre.



Dye (green) and cell permeable Cre recombinase delivery to Ai9 lung slices (Hoechst) shows TdTomato (red) signal indicial of Cre delivery to Ai9.

## Progress to Date and Future Plan

- Confirmed genome screen identified protective genes
- Continuing multiple assays to elucidate genetic mechanisms of *Bt* pathogenesis
- Working to down select host defense targets to best *in vivo* option for mouse studies
- Transition to CRISPRi and move towards a therapeutic