

Article

On the Short-Range Structure of Amorphous Calcium Hydrogen PhosphateBing-Qiang Lu, Natalya Garcia, Daniel M Chevrier, Peng
Zhang, Paolo Raiteri, Julian D Gale, and Denis Gebauer*Cryst. Growth Des.*, **Just Accepted Manuscript** • DOI: 10.1021/acs.cgd.9b00274 • Publication Date (Web): 02 Apr 2019Downloaded from <http://pubs.acs.org> on April 8, 2019**Just Accepted**

“Just Accepted” manuscripts have been peer-reviewed and accepted for publication. They are posted online prior to technical editing, formatting for publication and author proofing. The American Chemical Society provides “Just Accepted” as a service to the research community to expedite the dissemination of scientific material as soon as possible after acceptance. “Just Accepted” manuscripts appear in full in PDF format accompanied by an HTML abstract. “Just Accepted” manuscripts have been fully peer reviewed, but should not be considered the official version of record. They are citable by the Digital Object Identifier (DOI®). “Just Accepted” is an optional service offered to authors. Therefore, the “Just Accepted” Web site may not include all articles that will be published in the journal. After a manuscript is technically edited and formatted, it will be removed from the “Just Accepted” Web site and published as an ASAP article. Note that technical editing may introduce minor changes to the manuscript text and/or graphics which could affect content, and all legal disclaimers and ethical guidelines that apply to the journal pertain. ACS cannot be held responsible for errors or consequences arising from the use of information contained in these “Just Accepted” manuscripts.



On the Short-Range Structure of Amorphous Calcium Hydrogen Phosphate

Bing-Qiang Lu^{[a,b]*}, Natalya A. Garcia^[c], Daniel M. Chevrier^[d], Peng Zhang^[d], Paolo Raiteri^[c], Julian D. Gale^[c] and Denis Gebauer^{[b,e]*}

[a] Dr. B. Q. Lu

State Key Laboratory of High Performance Ceramics and Superfine Microstructure,

Shanghai Institute of Ceramics, Chinese Academy of Sciences

Dingxi Road 1295, Shanghai 200050, P.R. China

Email: b.q.lu@mail.sic.ac.cn

[b] Prof. Dr. D. Gebauer

Department of Chemistry, Physical Chemistry

University of Konstanz

Universitätsstrasse 10, Box 714, D-78457 Konstanz, Germany

[c] N. A. Garcia, A/Prof. P. Raiteri, Prof. J. D. Gale

Curtin Institute for Computation/The Institute for Geoscience Research (TIGeR), School of Molecular and Life

Sciences, Curtin University, PO Box U1987, Perth, Western Australia 6845, Australia.

[d] Dr. D.M. Chevrier, Prof. P. Zhang

Department of Chemistry

Dalhousie University

Halifax, NS, B3H4R2, Canada

[e] present address:

Prof. Dr. D. Gebauer

Institute of Inorganic Chemistry

University of Hanover

Callinstr. 9, D-30167 Hanover, Germany

Correspondence: gebauer@acc.uni-hannover.de (D. Gebauer); b.q.lu@mail.sic.ac.cn (B. Q. Lu)

Abstract

Calcium orthophosphates (CaPs) are the hard constituents of bones and teeth, and thus of ultimate importance to humankind, while amorphous CaPs (ACPs) may play crucial roles in CaP biomineralization. Among the various ACPs with Ca/P atomic ratios between 1.0~1.5, an established structural model exists for basic ACP (Ca/P=1.5), while those of other ACPs remain unclear. Herein, the structure of amorphous calcium hydrogen phosphate (ACHP; Ca/P=1.0) obtained via aqueous routes at near-neutral pH values, without stabilizers, was studied by experiments (mainly, TEM with ED, XRD, IR and NMR spectroscopies, as well as EXAFS) and computer simulation. Our results globally show that ACHP has a distinct short-range structure, and we propose calcium hydrogen phosphate clusters (CHPCs) as its basic unit. This model is consistent with both computer simulations, and the experimental results, where CHPCs are arranged together with water molecules to build up ACHP. We demonstrate that Posner's clusters, which are conventionally accepted to be the building unit of basic ACPs, do not represent the short-range structure of ACHP, as Posner's clusters and CHPCs are structurally distinct. This finding is important not only for the determination of the structures of diverse ACPs with varying Ca/P atomic ratios, but also for fundamental understanding of a major mineral class that is central to biomineralization in vertebrates, and, thus, humans, in particular.

Introduction

Calcium orthophosphates (CaPs) constitute a diverse family of minerals that play important roles in both geology and biology. Among them, carbonated hydroxyapatite is vital for humankind, as it is the mineral constituent of bones and teeth.^{1, 2} Due to related health issues like osteoporosis, the exploration of the mechanisms of CaP mineralization brings about the great promise to find solutions to upcoming health issues of our ageing societies,^{1, 3} but is also attractive from a fundamental science perspective.

Transient amorphous intermediates generally play important roles in biomineralization processes. This is particularly well established for the case of calcium carbonate,^{4, 5} which is the most abundant

1
2 biomineral, and may also involve liquid mineral precursors stabilized by acidic macromolecules.⁶ The
3
4 role of amorphous calcium (ortho)phosphate (ACP) in biomineralization, however, has been a matter of
5
6 considerable debate.⁷⁻¹⁴ Today, there is mounting evidence that in animal bones and teeth, ACP is a
7
8 precursor to apatites (or an important component of the complex structures),¹⁵⁻²² and liquid precursors
9
10 may also play a role in CaP biomineralization.²³⁻²⁵ HPO_4^{2-} ions were shown to be present within an
11
12 "acidic" ACP, which is thought to be essential for CaP biomineralization.²¹ Generally, the term "acidic"
13
14 ACP implies that HPO_4^{2-} ions are present in the material, yielding Ca/P atomic ratios between ~1.0-1.5
15
16 even though it is obtained at near-neutral, slightly basic pH values.
17
18
19

20
21 Synthetic ACP can transform into octacalcium phosphate (OCP), hydroxyapatite (HAP) or calcium-
22
23 deficient hydroxyapatite (CDHA),²⁶ where the Ca/P atomic ratio varies. This implies that the Ca/P
24
25 atomic ratio in amorphous intermediates may be a key factor for controlling the structure of the
26
27 subsequently formed crystalline CaPs.²⁶ Basic ACP (Ca/P=1.5) is built up from so-called Posner's
28
29 clusters,²⁷ i.e., $\text{Ca}_9(\text{PO}_4)_6$ units with a diameter of 0.95 nm, which are also a building block of
30
31 hydroxyapatite when arranged in crystallographic register, together with additional Ca^{2+} and OH^- ions.²⁷
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
It has been suggested that deviations from a Ca/P ratio of ~1.5 in ACPs are mostly due to surface
adsorbed, more soluble phases arising from non-aqueous syntheses, or the additional incorporation of
calcium ions.²⁶ This led to the conclusion that Posner's clusters may also generally represent the short-
range order in chemically different ACPs.²⁶ However, the structure of "acidic" ACPs with lower Ca/P
atomic ratios, has been less studied.^{29, 30} In the investigated examples, the (Ca/Mg)/P atomic ratio was
not lower than 1.15, and the structure of pure amorphous calcium hydrogen phosphate (ACHP; i.e., with
an invariant Ca/P atomic ratio of 1.0) has not yet been determined, although it was previously prepared
in the presence of impurities from aqueous solutions, whereas the Ca/P ratio of the obtained precipitates
was not experimentally confirmed.^{31, 32} Moreover, stabilizers likely affected the structure of the
AC(H)Ps, while some structural differences among AC(H)Ps prepared under different conditions were
also reported.^{33, 34} Indeed, HPO_4^{2-} ions were presumed to be present in pre- and post-nucleation species
during CaP mineralization in tris(hydroxymethyl)methyl aminomethane) (TRIS)-buffered saline

1 solutions at physiological conditions (pH 7.4),³⁵ although the validity of the assumptions made to arrive
2
3
4 at the claimed speciation was subsequently challenged.³⁶
5

6 While organic components and ion substitutions may influence the structure and crystallization
7
8 behavior of ACPs, studying the pure forms is a fundamental pre-requisite in order to be able to discuss
9
10 any specific roles that these foreign species may play, for instance, in biomineralization. Furthermore,
11
12 the determination of the structure of ACHP could be required for investigating those of other ACPs with
13
14 intermediate Ca/P atomic ratios, which could be conceived of as chemical intermediaries between basic
15
16 ACP and ACHP, of which only the former structure is known until now. Herein, we synthesized ACHP
17
18 via an aqueous route with a confirmed atomic ratio of Ca/P \approx 1.0. Notably, the synthesis was achieved
19
20 without the use of additional stabilizers that might affect its structural characteristics. We use basic ACP
21
22 (Ca/P=1.5) and crystalline CaPs with different Ca/P ratios as reference materials in detailed structural
23
24 analyses. This, in conjunction with computer simulations, leads us to propose a structural basic unit for
25
26 describing this amorphous form of CaP, the calcium hydrogen phosphate cluster (CHPC), which is
27
28 structurally distinct from Posner's clusters building up basic ACPs.
29
30
31
32
33
34
35

36 **Results and discussion**

37
38
39

40 AC(H)Ps with different Ca/P atomic ratios were prepared by mixing CaCl₂ and Na₃PO₄ solutions
41
42 with different pH values. ACPs prepared at pH 11.9 and pH 8.9 exhibit Ca/P ratios of 1.47 and 1.02
43
44 (according to ICP-OES, Supporting Information (SI), Experimental Section, Table S1), and are
45
46 regarded as basic ACP and ACHP, all respectively. Transmission electron microscopy (TEM) analyses
47
48 show that the AC(H)Ps consist of particles with sizes in the range of tens to hundreds of nanometers
49
50 (Figure 1). The selected area electron diffraction (SAED) patterns (insets, Figure 1) do not show distinct
51
52 reflections, proving the absence of long-range order and confirming their amorphous character.
53
54
55
56
57
58
59
60

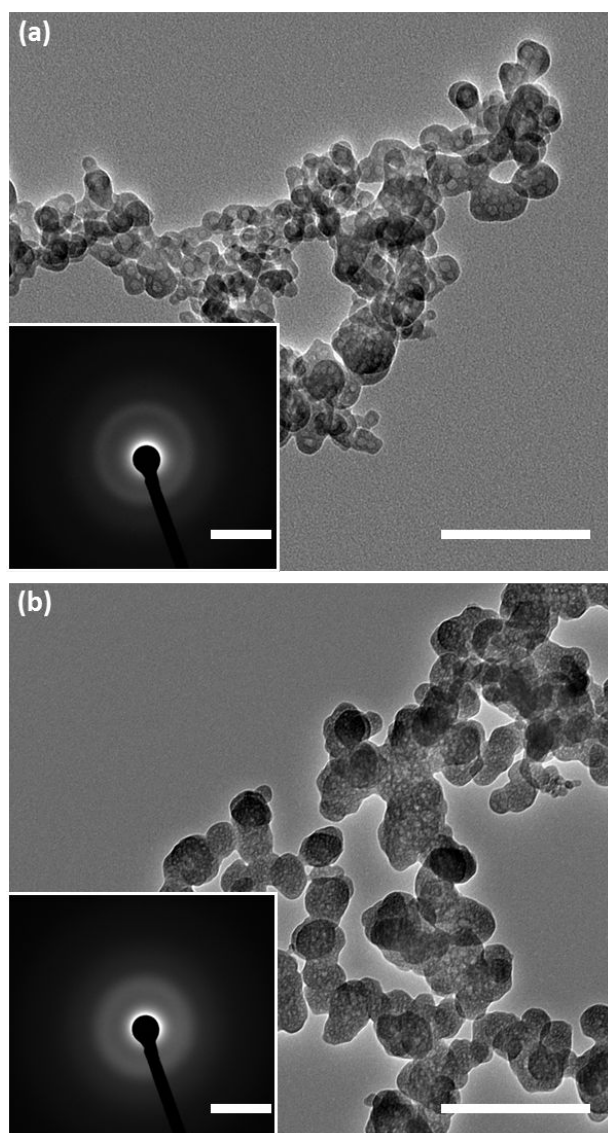


Figure 1. TEM micrographs of basic ACP (a) and ACHP (b). The insets show corresponding SAED patterns. The scale bars are 100 nm and 0.5 $1/\text{\AA}$ for the micrographs and SAED data, respectively.

The X-Ray powder diffractograms (XRD) show two broad peaks for each ACP (Figure S1), one is relatively strong at around $2\theta=25-35^\circ$, the other one is much weaker at around $2\theta=40-55^\circ$. This indicates that the basic ACP and ACHP do possess distinct short- to medium-range orders, but neither form exhibits crystalline long-range order. Notably, the maximum position of the stronger peak varies slightly in between basic ACP and ACHP, shifting to lower 2θ values for ACHP by ca. 1° (Figure 2a), already indicating differences in the structural characteristics of basic ACP and ACHP. These were further explored by means of Fourier Transform Infrared Spectroscopy (FTIR) analyses (SI section S1 and Figure 2b). The single ν_3 band (1047 cm^{-1}) in the FTIR spectrum of basic ACP corresponds to the

1
2 asymmetric stretch vibration of P-O (in PO_4), and is split into ν_6' and ν_6'' bands in the spectrum of
3
4 ACHP due to the presence of HPO_4^{2-} ions (Figure S2b). At the same time, the ν_4 bands, corresponding
5
6 to the O-P-O bending vibration, shift from 567 cm^{-1} (basic ACP) to 543 cm^{-1} (ACHP). This indicates
7
8 that the lengths of the P-O bonds, and/or the symmetry of the surrounding short-range orders in ACP
9
10 and ACHP are indeed different. The ν_3' bands, corresponding to the P-OH (HPO_4) stretching mode, are
11
12 clearly seen in the spectrum of ACHP, which corroborates that HPO_4^{2-} ions are structural constituents,
13
14 and confirms the substitution of PO_4^{3-} by HPO_4^{2-} when compared with the spectrum of basic ACP. FTIR
15
16 spectral differences between dicalcium phosphate dihydrate (DCPD, $\text{CaHPO}_4 \cdot 2\text{H}_2\text{O}$) and ACHP also
17
18 show that the hydrogen bonding of neighboring hydrogen phosphates in ACHP is distinct, whereas
19
20 structural relations with neither dicalcium phosphate (DCP, CaHPO_4) nor octacalcium phosphate (OCP,
21
22 $\text{Ca}_8\text{H}_2(\text{PO}_4)_6 \cdot 5\text{H}_2\text{O}$) are evident (SI section S1 and Figure S2, S3). For more detailed discussions of the
23
24 IR spectra, see the SI, section S1.

25
26
27
28
29 Further insight into the structural role of HPO_4^{2-} ions in ACHP was obtained by ^1H magic angle
30
31 spinning (MAS) NMR spectroscopy (SI, Figure S4-S6). The broad resonance of HPO_4^{2-} ions (7-14
32
33 ppm) is observed in case of ACHP, but not basic ACP (SI, Figure S4). The broad resonance at ~ 4 ppm
34
35 arises from chemisorbed or structural H_2O , and is seen in the MAS NMR spectra of both amorphous
36
37 forms (SI, Figure S4). ^1H MAS NMR spectra of other crystalline CaPs (SI, Figure S5) are all different
38
39 from that of ACHP. However, comparison of the ^1H MAS NMR spectra of ACHP and DCPD (SI,
40
41 Figure S6) shows that the respective chemical environments of the protons of HPO_4^{2-} and H_2O are
42
43 similar.
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

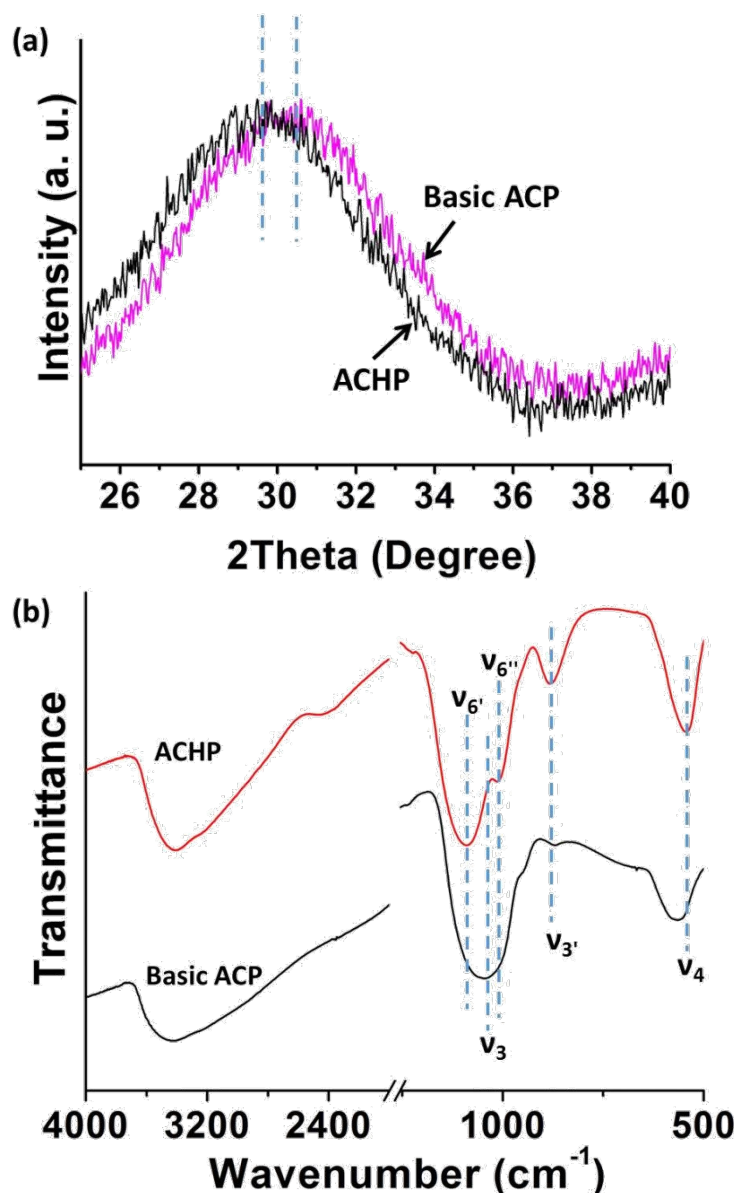


Figure 2. XRD patterns (a) and FTIR spectra of basic ACP and ACHP. The XRD patterns with full 2θ range corresponding to (a) can be found in the SI, Figure S1. The dashed vertical lines are guides for the eye to illustrate the shift of the peaks. “ v_i ” represent different vibrational modes i of the phosphate ion. The band assignments follow Bailey et al.³⁷.

³¹P MAS NMR spectra (Figure 3) show that basic ACP and ACHP display single, broad Gaussian-shaped resonances in the range of -15~15 ppm, whereas the maximum position is centered at 0.32 and 2.2 ppm for ACHP and basic ACP, respectively. For crystalline CaPs, only HAP and DCPD exhibit single resonances (SI Figure S7). The peak maximum positions of basic ACP (2.2 ppm) and HAP (1.9 ppm) agree within experimental accuracy (SI, Figure S8), as is the case for ACHP (0.32 ppm) and

DCPD (0.40 ppm) (SI, Figure S9). This indicates that the P chemical environment of ACHP is, on average, similar to that of DCPD, but no other crystalline CaP. This corroborates the notion already suggested by the corresponding ^1H MAS NMR spectra.

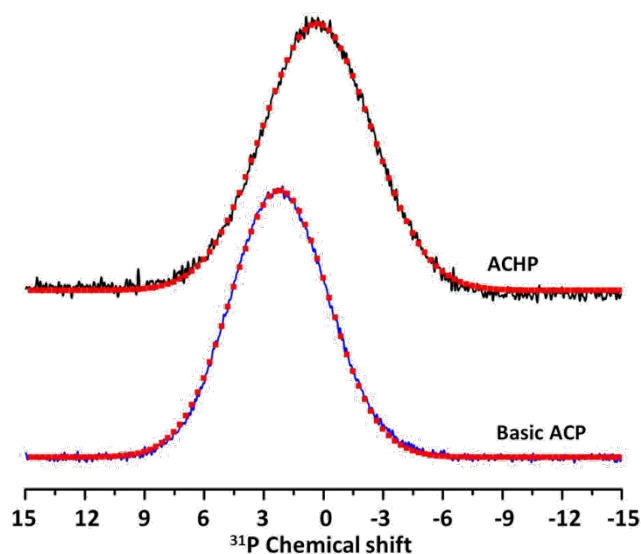


Figure 3. ^{31}P MAS NMR spectra of different ACPs. ^1H de-coupling was applied. $\text{NH}_4\text{H}_2\text{PO}_4$ was used as the references (set as 0 ppm). Solid curves: original spectra; dotted curves: Gaussians from the fitting of the original spectra.

To further shed light on the structural configurations in basic ACP, ACHP and different CaPs, Ca K-edge x-ray absorption near edge structure (XANES) and extended x-ray absorption fine structure (EXAFS) analyses were performed (Figure 4 and SI, Figure S10). The Ca K-edge XANES spectra of DCPD, HAP, basic ACP and ACHP (Figure 4a) show a pre-edge peak between 4037-4041 eV, and a white line (the most intense peak following the absorption edge) in the energy range of 4045-4055 eV. The maxima positions of the pre-edge peaks do not show any significant differences, while their normalized intensities vary (Figure 4b). The intensity of the pre-edge feature is similar for basic ACP and ACHP, and lies in between that of DCPD and HAP. This particular feature relates to Ca $1s \rightarrow$ Ca $3d/\text{O } 2p$ transitions. Thus, the higher pre-edge intensity of HAP could originate from the distinct combination of highly coordinated Ca-O sites. As for the white line features, the two ACPs are essentially identical in regard to both the peak maximum position (4049.7 eV) and the intensity (Figure 4c). The white line position of DCPD (4050.0 eV) is close to that of the ACPs, while that of HAP is

1
2 split into two peaks at 4048.9 and 4051.2 eV. The split in the spectrum of HAP is due to the presence of
3
4 two unique crystallographic sites for Ca.^{38, 39} The relative intensity of this main absorption feature
5
6 corresponds well with the Ca-O scattering intensity in the EXAFS spectra (i.e., higher white line
7
8 intensity will result from more Ca-O coordination, see the SI, Figure S11, Table S2, and section S2, for
9
10 details). These results indicate that the short-range environments of the Ca atoms in the basic ACP and
11
12 ACHP are very similar in regard to their Ca-O local structure (Ca-P or Ca-Ca are not visible for basic
13
14 ACP and ACHP due to their amorphous character). The similarity of Ca environments between basic
15
16 ACP and ACHP is consistent with previous EXAFS measurements,^{29, 30, 40} although the “acidic” ACP
17
18 studied therein had a Ca/P atomic ratio significantly larger than 1. For “acidic” ACP, by investigating
19
20 casein-stabilized micelles containing CaP with X-Ray Absorption Spectroscopy, Holt et al. proposed
21
22 that the O atoms from H₂O and HPO₄²⁻ coordinate to the Ca ion to form the first shell of the short-range
23
24 order.⁴¹ However, this is also the case for the Posner’s cluster model, where water can contribute to the
25
26 coordination to Ca ions, since 8 of 9 Ca ions in a Posner’s cluster are located at the surface.
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

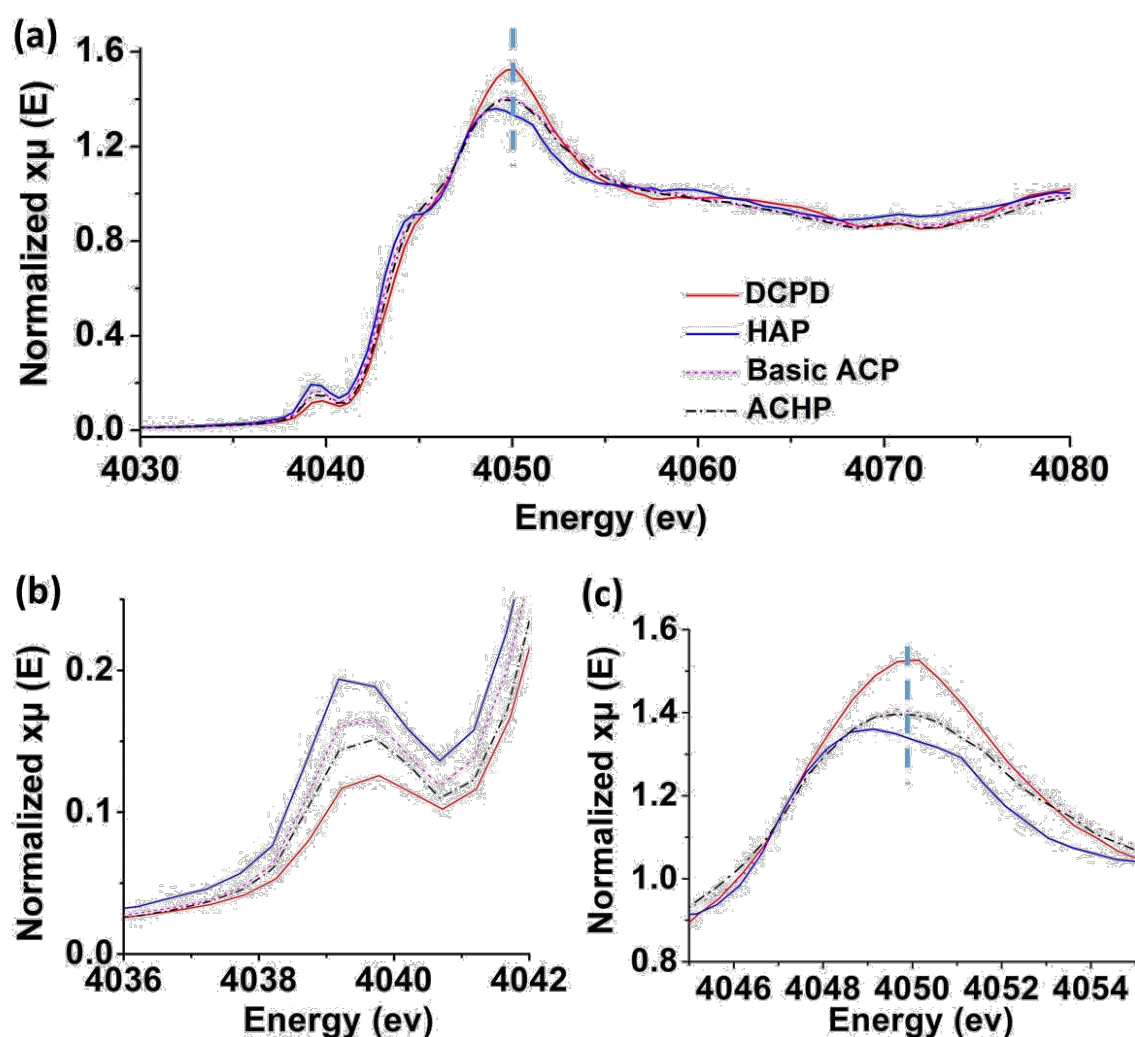
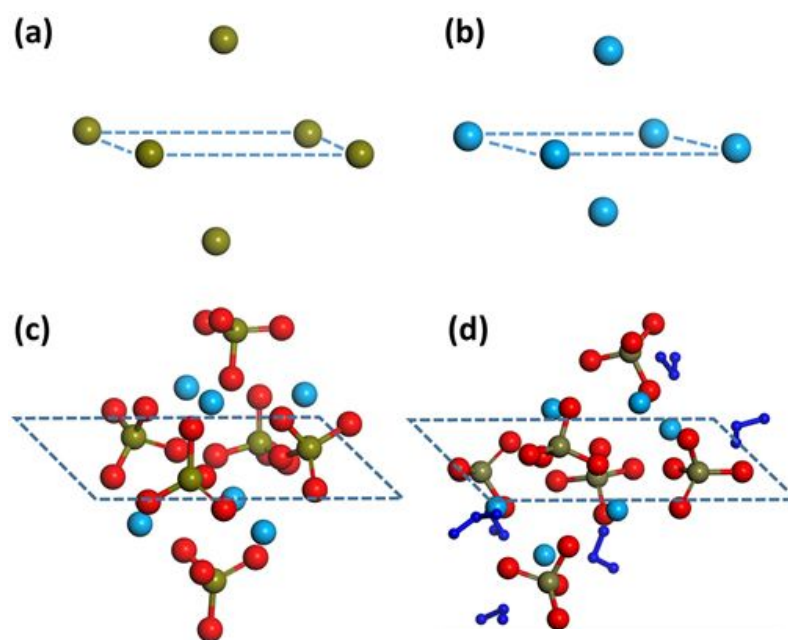


Figure 4. Normalized Ca K-edge XANES (a-c) spectra of DCPD, HAP, basic ACP and ACHP. (b) and (c) show magnified regions of the spectra in the energy range of the pre-edge, 4036-4042 eV (b) and white line, 4045-4055 eV (c).

In order to investigate the structure of the possible building unit of ACHP, assuming a Ca:P ratio of 1:1, we monitored the stability of various clusters in an aqueous environment using computer simulations (Experimental Section for selection rules and simulation details; SI, Figure S12, S13, Table S3, and section S3 for detailed discussion). The neutral $(\text{CaHPO}_4)_6$ cluster (CHPC) was among the most stable of the clusters, remaining together for the full duration of the simulations (20 ns), and exhibited dynamic switching between two main configurations (SI, Figure S13). While $[\text{Ca}_6(\text{HPO}_4)_4(\text{PO}_4)_2]^{-2}$ and $[\text{Ca}_6(\text{HPO}_4)_5(\text{PO}_4)]^{-1}$ clusters were also comparatively stable during the simulation times studied, the aggregation of a negatively charged cluster yielding ACHP is assumed to be less favorable. We note

1
2 that a definitive determination of the free energy landscape of clusters in aqueous solution is beyond the
3
4 scope of the present work, which deals with the solid-state building unit.
5

6 Studying a single CHPC in vacuum, it evolves into a structure shown in the Figure 5a-c. The addition
7
8 of 6 H₂O molecules coordinating with the CHPC (the H₂O/P ratio is 1.0, SI section S4), the structure
9
10 remains essentially unchanged, though the waters do not distribute evenly across the calcium ions and
11
12 the structure becomes skewed (Figure 5d). Comparing this CHPC with a Posner's cluster in vacuum (SI,
13
14 Figure S14), similar to the one proposed by Treboux et al⁴², the configuration of CHPC differs greatly
15
16 from that of the Posner's cluster with or without H₂O coordination (see details in SI, section S3).
17
18
19
20
21
22
23
24



25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46 **Figure 5.** Configurations of CHPC in vacuum taken from simulation. (a-c) The arrangement of P (a),
47
48 Ca (b) and all atoms (c). (d) CHPC with H₂O molecules. The dashed quadrangles indicate the
49
50 (disordered) rectangles formed by P atoms (a, c, d) and Ca atoms (b). Ca is shown in cyan, P in tan, O in
51
52 red; H of HPO₄ is not shown; H₂O are shown in blue.
53
54
55

56
57 A structural model of ACP solid is further obtained by computer simulation (Figure 6). While the
58
59 studies of cluster stability in water indicate probable structures, assuming the 1:1 Ca:P ratio proposed
60

1 experimentally, it is also useful to compare the properties of an amorphous model built from clusters
2 with this ratio to an amorphous solid built from Posner's clusters, which have a Ca:P ratio of 9:6. Hence
3
4
5
6 amorphous models were created from both species. The densities of the resulting amorphous materials
7
8
9 were calculated to be $2.09 \pm 0.03 \text{ g/cm}^3$ for $\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$ and $2.13 \pm 0.03 \text{ g/cm}^3$ for
10
11 $\text{Ca}_9(\text{PO}_4)_6(\text{H}_2\text{O})_{12}$ (SI, Table S4, Figure S16, S17). As expected, these are significantly lower than the
12
13 experimental values for crystalline phases, such as $\text{Ca}_5(\text{OH})(\text{PO}_4)_3$ and CaHPO_4 which are 3.16^{43} and
14
15 2.92 g/cm^3 ,⁴⁴ respectively, due to the presence of stoichiometric amounts of water. The calcium
16
17 coordination number by water for the amorphous solid built from hydrated Posner's clusters was
18
19 slightly lower than the solid built from $\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$ (6.76 ± 0.03 vs 7.32 ± 0.09 , respectively,) (SI,
20
21 Table S4), reflecting the more closed, spherical structure of the Posner's cluster and the larger amount
22
23 of stoichiometric water. It follows that the phosphate coordination number by calcium for the
24
25 amorphous solid built from hydrated Posner's cluster was higher than the solid built from
26
27 $\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$ (6.02 ± 0.03 vs. 4.59 ± 0.08) (SI, Table S4), due to the higher amount of
28
29 stoichiometric Ca to P. We also note that longer 100 ns simulations in both cases do not result in
30
31 significantly different CNs. Using an analysis of the coordination number of individual Ca sites with
32
33 water oxygen we can identify the packing structure (SI, Table S5). In both basic ACP and ACHP the
34
35 amount of bound water suggests that the cluster packing includes interstitial water bound to Ca. We
36
37 expect that the amount of unbound Ca will be higher in ACHP due to the lower stoichiometric ratio of
38
39 water but the magnitude of the increase (55% versus basic ACP's 33%) suggests that there is a different
40
41 packing pattern. However, the water oxygen environment indicates that there is more bulk-like water in
42
43 basic ACP than ACHP (SI, Table S5).
44
45
46
47
48
49

50 For the plane wave density functional theory (DFT) calculations, it was necessary to construct small
51
52 unit cells to model the amorphous structures. In the case of the structures based on hydrated Posner's
53
54 clusters and on a composition of $\text{CaHPO}_4 \cdot \text{H}_2\text{O}$, the unit cells both contained 300 atoms, which
55
56 correspond to four Posner or five $(\text{CaHPO}_4)_6$ clusters plus either 48 or 30 water molecules, respectively.
57
58 After relaxation at the PBE-D2 level of theory, the densities were 2.19 and 2.22 g/cm^3 , also respectively.
59
60

Both densities are similar to those for the larger force field simulations quoted above. One unexpected feature of the amorphous $\text{CaHPO}_4\cdot\text{H}_2\text{O}$ material was that a proton transfer reaction occurred spontaneously, resulting in a disproportionation of 2 HPO_4^{2-} anions into H_2PO_4^- and PO_4^{3-} . Attempts to redistribute the protons to different sites in order to attain the expected composition were unsuccessful, suggesting that the tendency to form a specific protonation state depends on the local environment of the PO_4 tetrahedron within the amorphous material. The same behavior was observed for two independently generated models of this amorphous phase.

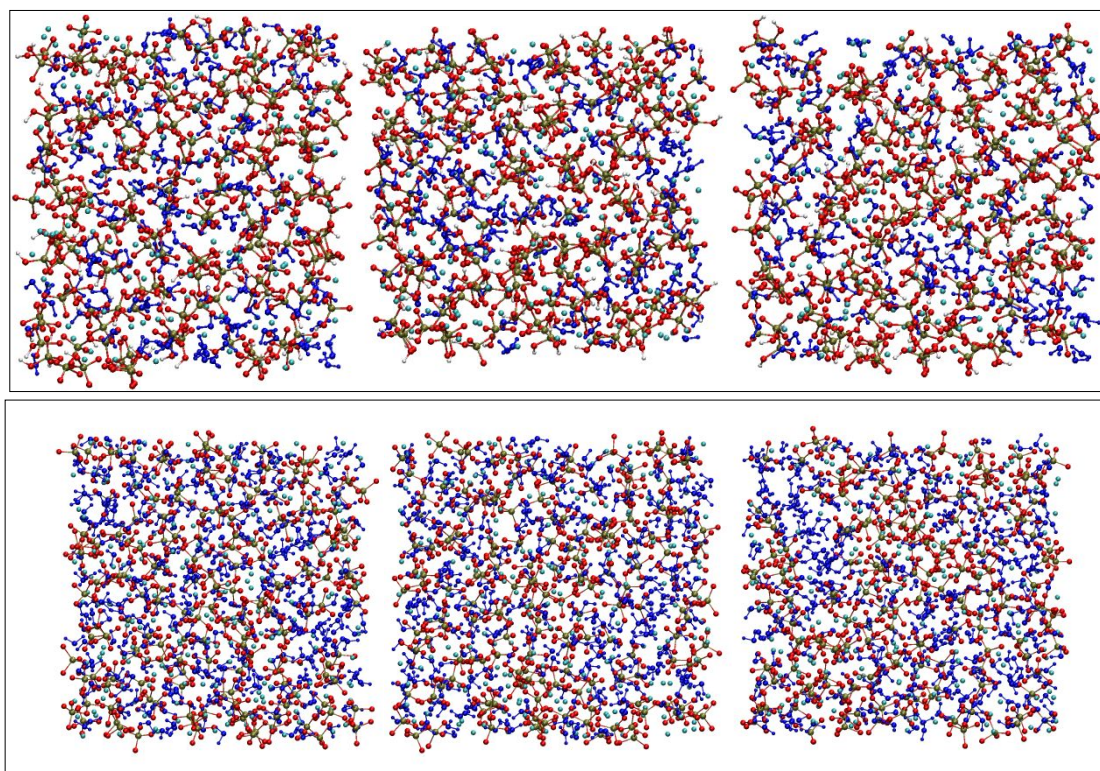
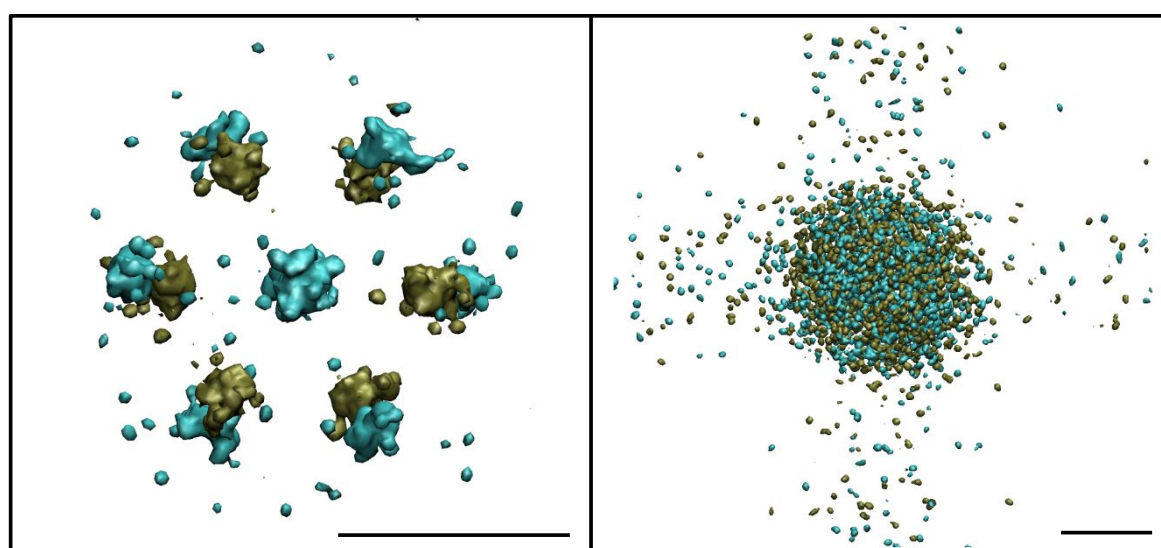


Figure 6. Two amorphous models built from $\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$ clusters (top row) and $\text{Ca}_9(\text{PO}_4)_6(\text{H}_2\text{O})_{12}$ clusters (bottom row) shown along each of the three orthogonal axes. Here Ca is cyan; phosphate O is red, H is white and P is tan; H and O of water are shown in blue to highlight the approximately uniform dispersal of water, phosphate ions and calcium ions.

Previous experimental and computational evidence showed that Posner's clusters are the building units of basic ACP, which agrees with our simulations. In the calculated density maps of the clusters within the amorphous solid (Figure 6), the Posner's clusters maintain their geometry over the course of

1
2 the simulations (Figure 7, left). There is some increased diffusivity as evidenced by the slight smearing
3
4 of the atomic positions but the overall geometry observed in vacuum and in aqueous solution, where a
5
6 central Ca is surrounded by 6 PO₄ and 8 Ca atoms with a cluster diameter of ~9 Å, is maintained.
7
8 However, our simulations of ACHP suggests that the CHPCs, with a larger average diameter (~10 Å),
9
10 are more distorted within the solid and in many cases exchange ions and molecules between units, with
11
12 no single stable cluster geometry (Figure 7, right). Thus, though the existence of CHPC in ACHP is
13
14 confirmed here, it does not specifically suggest that ACHP is constructed via addition of clusters with a
15
16 uniform configuration.
17
18
19



20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38 **Figure 7.** Average 3D atomic density maps of (left) the Ca₉(PO₄)₆(H₂O)₆ cluster and (right) the
39
40 Ca₆(HPO₄)₆(H₂O)₆ cluster within the amorphous solid. The iso-density surfaces have been colored
41
42 based on atom type where phosphorus is tan and calcium is cyan; oxygen and hydrogen have been
43
44 omitted for clarity. In both cases scale bars are 5 Å. Individual clusters were selected from the solid
45
46 based on their initial geometry and superimposed using a method described in S1.
47
48
49

50 **Conclusions**

51
52
53 In conclusion, our data shows that the structure of ACHP is distinct from basic ACP and cannot be
54
55 described by Posner's clusters, nor disordered/Ca-deficient Posner's clusters. We thus propose a novel
56
57 structural model for ACHP (Figures 6-7) based upon simulation. Similar to basic ACPs, we propose that
58
59 ACHP consists of CHPCs (Figure 7) that are aggregated with H₂O in the interstices to build up the
60

1
2 structure of ACHP. Unlike the Posner's clusters, however, the CHPC has lower symmetry and is more
3
4 susceptible to structural change on formation of an amorphous solid. The chemisorbed, structural H₂O
5
6 molecules coordinating to Ca ions or even exhibiting hydrogen-bonding interactions with the HPO₄²⁻
7
8 ions are arranged around the CHPC to build up ACHP. This structure is globally consistent with the
9
10 experimental analyses as follows:
11

12
13 (1) The Ca environments are similar for basic ACP and ACHP in the short- to medium-range. From the
14
15 simulations, the Ca-O coordination number (first shell) is similar throughout calcium sites in the
16
17 amorphous solids at 7.32 ± 0.09 for ACHP and 6.76 ± 0.04 for basic ACP.
18
19

20
21 (2) The P short-range environments differ between basic ACP and ACHP. In addition to the complete
22
23 substitution of phosphate ions by hydrogen phosphate ions, the interactions between the Ca and HPO₄
24
25 (PO₄) ions change due to rather distinct Ca arrangements around P. This is seen in the calculated P-Ca
26
27 coordination number, which is lower for ACHP than for basic ACP (4.62 ± 0.03 and 6.02 ± 0.04 ,
28
29 respectively). The different bonding configurations give rise to the distinct FTIR spectral features
30
31 observed experimentally.
32
33

34
35 (3) The chemical environment of the P atoms in ACHP and DCPD is similar according to ³¹P MAS
36
37 NMR spectra, which is due to identical Ca/P atomic ratios within homogeneous phases.
38

39
40 (4) Comparison of the computed (SI, Figure S15) and experimental (SI, Figure S4) proton NMR spectra
41
42 reveals that the calculated spectrum supports the assignment of the broad feature at high chemical shifts
43
44 to clusters of CaHPO₄ in ACHP, and that the experimental data is inconsistent with an amorphous solid
45
46 composed of Posner-like clusters (for further discussions, see SI, section S5).
47

48
49 (5) HPO₄²⁻ ions transform into P₂O₇⁴⁻ ions in the second step of TGA measurements (SI, Figures S18-21,
50
51 and section S4), as opposed to DCPD. Because the CHPCs are not connected within a long-range order
52
53 crystallographic arrangement, and nearly all the HPO₄²⁻ ions are at the surface of the clusters close to
54
55 water molecules within the structure of ACHP, the HPO₄²⁻ ions are less stable than in DCPD upon
56
57 heating.
58
59
60

1
2 All of this reveals that ACHP is structurally distinct from all other known CaPs, and its short-range
3 environment can be described by means of CHPCs. This suggests that at least ACHP cannot be seen as
4 a proto-crystalline form^{4, 45, 46} that directly relates to a certain crystalline CaP, although its Ca/P atomic
5 ratio agrees with that of DCPD. As the structures of both basic ACP and ACHP are now established, it
6 should be investigated whether or not other ACPs with varying Ca/P atomic ratios ($1 < \text{Ca/P} < 1.5$) are
7 intermediaries of the two forms, from a structural point of view. In addition, these pure forms also
8 provide the fundamental point of reference for investigating specific roles that organic components and
9 ion substitutions might play, for instance, in biomineralization. Certainly, ACHP can serve as an
10 amorphous intermediate for the formation of different crystalline forms of CaP (SI, Figures S22, S23),
11 and its crystallization behavior will have to be studied in detail, in the future. Last, but not least, we note
12 that the current solid-state characterization and structural model must not be extrapolated to the solution
13 state, where pre-nucleation clusters occur, which have very likely a different structure and speciation
14 than the CHPC building units of AC(H)P³⁶.
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33

34 **Experimental Section**

37 **Preparation of AC(H)Ps**

38 1.67 mmol $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$ was dissolved in 3 mL N_2 degassed deionized water (solution A).
39 $\text{Na}_3\text{PO}_4 \cdot 12\text{H}_2\text{O}$ and Na_2HPO_4 was dissolved in 3 mL N_2 degassed deionized water (solution B), in which
40 the total molar amounts of $\text{Na}_3\text{PO}_4 \cdot 12\text{H}_2\text{O}$ and Na_2HPO_4 were kept at together 1.0 mmol, while the
41 molar ratio was varied to adjust the pH of the solution. The actual molar ratio of $\text{Na}_3\text{PO}_4/\text{NaH}_2\text{PO}_4$, the
42 pH of solution B and the Ca/P atomic ratio of the obtained AC(H)Ps are summarized in the SI, Table S1.
43 Under vigorous magnetic stirring and N_2 protection, solution B was quickly added into solution A.
44 Immediately, white precipitates of AC(H)P were formed, and 40 mL methanol was added to quench the
45 reaction. Then the obtained mixture was transferred into 2 centrifuge tubes (50 mL) with 20 mL
46 methanol, centrifuged at 9000 rpm for 0.5 min, washed with methanol 3 times, and once with acetone.
47 The white powder obtained was dried by blowing N_2 into the tubes at ambient temperature. Basic ACP
48 was prepared at ambient temperature.
49
50
51
52
53
54
55
56
57
58
59
60

1 The reference samples HAP, DCPD, DCP, alpha-TCP and beta-TCP were purchased from Sigma-
2 Aldrich; OCP was synthesized following a previously reported procedure.⁴⁷

3
4
5
6 The chemical compositions and structure of the AC(H)Ps were determined by Inductively Coupled
7 Plasma Optical Emission Spectrometry (ICP-OES), Thermogravimetric Analyses (TGA), Transmission
8 Electron Microscopy (TEM), The X-ray powder Diffraction (XRD), Fourier Transform Infrared
9 Spectroscopy (FTIR), Nuclear Magnetic Resonance (NMR), Synchrotron radiation, etc., and the
10 corresponding experimental details are present in the SI.

19 **Simulation of the CHPC Structure**

21 Initial clusters were chosen to maintain the Ca:P ratio of 1:1 with 6 calcium atoms. Thus for
22 $\text{Ca}_6(\text{H}_2\text{PO}_4)_a(\text{HPO}_4)_b(\text{PO}_4)_c$ $a+b+c=6$ and $0 \leq a,b,c \leq 6$. Additionally, it is unlikely that $\text{H}_2\text{PO}_4^{-1}$ and
23 PO_4^{-3} would exist in the same cluster due to the favorability of a hydrogen transferring from $\text{H}_2\text{PO}_4^{-1}$ to
24 the more negative PO_4^{-3} . Thus, for clusters where $a \geq b$ then c was required to be $\leq b$ and vice versa,
25 such that for $c \geq b$ then $a \leq b$. Clusters with an overall charge greater than or equal to ± 2 were not
26 examined. These constraints limit the composition of the system to 11 possible clusters (Table S3). For
27 each cluster, an MD simulation was performed in the NVT ensemble in vacuum in a 50 Å box for 5 ns
28 at 3,000 K with angular momentum removed. The trajectory was reordered by potential energy and 10
29 configurations were chosen with differing potential energies. These configurations were then added to a
30 pre-equilibrated 50 Å water box and MD simulations at 300 K were performed. All molecular dynamics
31 simulations employed the recent force field parameters of Demichelis et al⁴⁸ and were performed using
32 the LAMMPS code using a time step of 1 fs.

33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49 For consistency, new configurations for Posner's cluster ($\text{Ca}_9(\text{PO}_4)_6$) were also considered, using a
50 starting configuration with S6 symmetry described by Treboux.⁴² Simulations of the hydrated and non-
51 hydrated cluster at 300K were performed and retained essentially the same configuration (Figure S14).
52 Even at high energy (3,000K) there were no configurations with significantly different potential
53 energies from the starting configuration (an example frame is shown in Figure S14). Thus, only one
54
55
56
57
58
59
60

1 configuration was added to a pre-equilibrated 50 Å water box and an MD simulation at 300 K was
2 performed for 20 ns, using the same parameters as for the other clusters.
3
4

5
6 To explore the structure of CHPC in vacuum we also performed MD simulations of the hydrated and
7 non-hydrated CHPC [$\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$ and $\text{Ca}_6(\text{HPO}_4)_6$] in vacuum at 300K. These simulations were
8 also performed in the NVT ensemble in a 50 Å by 50 Å by 50 Å box for 5 ns using the LAMMPS code
9 again with a time step of 1 fs.
10
11
12
13
14

15 To further confirm that the $\text{Ca}_6(\text{HPO}_4)_6$ cluster represents a viable structure, an *ab initio* molecular
16 dynamics (AIMD) simulation was performed on this species in a cubic box of length 17.688 Å
17 containing 170 water molecules (552 atoms). The initial configuration was obtained by equilibration
18 using the same force field model as above. AIMD used the Gaussian-augmented plane-wave
19 approach,⁴⁹ as embodied in the Quickstep module of the CP2K code.⁵⁰ Here a TZ2P basis set was used
20 for the valence orbitals of all elements except Ca, where a DZVP basis was chosen, along with
21 Goedecker-Teter-Hutter pseudopotentials⁵¹ to represent the effective potential due to the core electrons
22 and nucleus. A cut-off of 400 Ry was used for the auxiliary basis set for the electron density. The
23 exchange-correlation was chosen to be BLYP with D3 dispersion corrections⁵² using the Becke-Johnson
24 screening function at short-range.⁵³ The dynamics used a time step of 0.5 fs and an elevated temperature
25 of 330 K, since these conditions have been previously shown to correct for the over-structuring of water
26 within GGA-based density functional theory. AIMD was run for > 50 ps and the structure analyzed.
27 Although this timescale is necessarily short with respect to water exchange rates in such systems, no
28 significant change occurred in the cluster structure or hydrolysis state of any of the HPO_4^{2-} ions,
29 confirming that the cluster model is at least a local minimum on the DFT energy landscape.
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50

51 **Simulation of Amorphous Models**

52 To create models for the amorphous material, 27 of the hydrated clusters $\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$ or
53 $\text{Ca}_9(\text{PO}_4)_6(\text{H}_2\text{O})_{12}$, were arranged on a grid. These cluster geometries were selected from the last frame
54 of the aqueous NVT simulation described in the previous section. A 10 ns NPT run was used to
55 equilibrate the system, which was then followed by 100 ns of NVT molecular dynamics. To verify that
56
57
58
59
60

1 the models were not an artifact of all the clusters being initially oriented in the same way, the model
2 was replicated 8 times with alternative orientations. The 27 clusters were both randomly oriented and
3
4 was replicated 8 times with alternative orientations. The 27 clusters were both randomly oriented and
5
6 placed in a cubic box of length 27 Å for $\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$ and 40 Å for $\text{Ca}_9(\text{PO}_4)_6(\text{H}_2\text{O})_{12}$. For basic
7
8 ACP NPT runs ranging from 2-7 ns were sufficient to equilibrate the system. However, for ACHP
9
10 pockets of vacuum were observed to persist. Thus, a manual shrink was used to ensure that the density
11
12 was at least 2 g/cm³ for $\text{Ca}_6(\text{HPO}_4)_6(\text{H}_2\text{O})_6$. This box was energy minimized, relaxed and then run with
13
14 molecular dynamics using an NPT ensemble for > 3 ns. After the cell parameters had equilibrated, NVT
15
16 molecular dynamics was run for ~30 ns for each amorphous model. The average of these NVT
17
18 simulations was used to calculate the radial distribution functions. Specific simulation times are
19
20 reported in Table S4. To compare the cluster structure within ACHP and basic ACP the trajectories of
21
22 the 27 initial clusters were cut from the amorphous solids and superimposed using the central Ca in
23
24 Posner's cluster and an arbitrary Ca followed by the center of mass in CHPC as a point of reference.
25
26 MDAnalysis⁵⁴ was used with the fast QCP algorithm to calculate the root mean square distance between
27
28 the initial cluster and the cluster within the amorphous solid⁵⁵ and the rotation matrices that minimize
29
30 the RMSD⁵⁶ and 3D atomic density maps were calculated using these aligned trajectories (see Figure
31
32 S18). It was necessary to superimpose via both a Ca as well as center of mass for CHPC to ensure that
33
34 the cluster was not cut across the periodic boundary conditions as well as being centrally aligned.
35
36
37
38
39
40
41 In order to compute the solid-state NMR spectra for the above two possible types of amorphous
42
43 structure, further calculations were performed based on Kohn-Sham density functional theory. Here
44
45 plane-wave calculations were conducted using the CASTEP code⁵⁷ with a kinetic energy cut-off of 800
46
47 eV and ultrasoft pseudopotentials. The PBE exchange-correlation functional was used in conjunction
48
49 with dispersion corrections from the work of Grimme et al.⁵⁸ Given that both amorphous materials are
50
51 wide gap insulators and that the smallest cell dimension was greater than 13 Å, the Brillouin zone was
52
53 sampled only at the gamma point. Both amorphous structures were relaxed to constant pressure under
54
55 orthorhombic cell constraints such that all forces were below 0.05 eV/Å and stresses below 0.25 GPa.
56
57
58
59
60 Once optimized, the chemical shifts were computed using an approach based on the gauge-including

1 projector augmented wave (GIPAW) approach of Pickard and Mauri.^{59, 60} In order to provide reference
2 data for the chemical shift, an ordered model for hydroxyapatite was optimized using the same settings,
3
4 data for the chemical shift, an ordered model for hydroxyapatite was optimized using the same settings,
5
6 except for the use of a 4x4x4 Monkhorst-Pack mesh for the sampling of reciprocal space due to the
7
8 smaller unit cell dimensions. Experimental data for the ¹H and ³¹P chemical shifts were then taken from
9
10 the work of Cho et al.⁶¹
11
12
13
14
15

16 Acknowledgements

17
18
19 BQL acknowledges financial support by the CSC-DAAD postdoc program for a postdoctoral stay at the
20
21 University of Konstanz. DG is a Research Fellow of the Zukunftskolleg of the University of Konstanz.
22
23 DMC is supported by the NSERC CGS-Alexander Graham Bell scholarship. PZ acknowledges the
24
25 NSERC Discovery Grant for funding. The APS Sector 20 facilities are supported by the US DOE-Basic
26
27 Energy Sciences (Contract No. DE-AC02-06CH11357), the CLS and its funding partners, the
28
29 University of Washington, and the APS. Use of the APS, an Office of Science User Facility operated
30
31 for the US DOE Office of Science by Argonne National Laboratory, was supported by the US DOE
32
33 under Contract No. DE-AC02-06CH11357. NAG, PR and JDG thank the Australian Research Council
34
35 for funding, as well as the Pawsey Supercomputer Centre and NCI for resources. We thank Uli Haunz
36
37 for help with the NMR measurements; Michael Steyer for ICP-OES measurements; Jennifer Knaus for
38
39 TGA measurements; Eduard Wiedenbeck for XRD measurements.
40
41
42
43
44
45

46 **Keywords:** Amorphous materials • Structure elucidation • Biomineralization • Calcium
47 (ortho)phosphates • Posner's clusters
48
49
50
51

52 Notes and References

- 53 1. Dorozhkin, S. V.; Epple, M., Biological and medical significance of calcium phosphates. *Angew. Chem. Int. Ed.* **2002**, *41*, 3130-3146.
- 54 2. Lowenstam, H. A.; Weiner, S., *On Biomineralization*. Oxford University Press: New York, 1989.
- 55 3. Habraken, W.; Habibovic, P.; Epple, M.; Bohner, M., Calcium phosphates in biomedical applications: materials for the future? *Mater. Today* **2016**, *19*, 69-87.
- 56 4. Cartwright, J. H. E.; Checa, A. G.; Gale, J. D.; Gebauer, D.; Sainz-Diaz, C. I., Calcium Carbonate

- 1
2 Polyamorphism and Its Role in Biomineralization: How Many Amorphous Calcium Carbonates Are
3 There? *Angew. Chem. Int. Ed.* **2012**, *51*, 11960-11970.
- 4 5. Addadi, L.; Raz, S.; Weiner, S., Taking advantage of disorder: Amorphous calcium carbonate and
5 its roles in biomineralization. *Adv. Mater.* **2003**, *15*, 959-970.
- 6 6. Gower, L. B., Biomimetic Model Systems for Investigating the Amorphous Precursor Pathway and
7 Its Role in Biomineralization. *Chem. Rev.* **2008**, *108*, 4551-4627.
- 8 7. Glimcher, M. J.; Bonar, L. C.; Grynepas, M. D.; Landis, W. J.; Roufosse, A. H., Recent Studies of
9 Bone-Mineral - Is the Amorphous Calcium-Phosphate Theory Valid. *J. Cryst. Growth* **1981**, *53*, 100-
10 119.
- 11 8. Weiner, S.; Sagi, I.; Addadi, L., Choosing the crystallization path less traveled. *Science* **2005**, *309*,
12 1027-1028.
- 13 9. Crane, N. J.; Popescu, V.; Morris, M. D.; Steenhuis, P.; Ignelzi, M. A., Raman spectroscopic
14 evidence for octacalcium phosphate and other transient mineral species deposited during
15 intramembranous mineralization. *Bone* **2006**, *39*, 434-442.
- 16 10. Suvorova, E. I.; Petrenko, P. P.; Buffat, P. A., Scanning and transmission electron microscopy for
17 evaluation of order/disorder in bone structure. *Scanning* **2007**, *29*, 162-170.
- 18 11. Rey, C.; Combes, C.; Drouet, C.; Glimcher, M. J., Bone mineral: update on chemical composition
19 and structure. *Osteoporos. Int.* **2009**, *20*, 1013-1021.
- 20 12. Nudelman, F.; Pieterse, K.; George, A.; Bomans, P. H. H.; Friedrich, H.; Brylka, L. J.; Hilbers, P.
21 A. J.; de With, G.; Sommerdijk, N. A. J. M., The role of collagen in bone apatite formation in the
22 presence of hydroxyapatite nucleation inhibitors. *Nat. Mater.* **2010**, *9*, 1004-1009.
- 23 13. Nitiputri, K.; Ramasse, Q. M.; Autefage, H.; McGilvery, C. M.; Boonrungsiman, S.; Evans, N. D.;
24 Stevens, M. M.; Porter, A. E., Nanoanalytical Electron Microscopy Reveals a Sequential Mineralization
25 Process Involving Carbonate-Containing Amorphous Precursors. *Acs Nano* **2016**, *10*, 6826-6835.
- 26 14. Yamashita, K.; Horisaka, Y.; Satomura, K.; Takagi, T., Analysis of minerals on initial calcification
27 induced by bone matrix gelatin. *Jpn. J. Oral Biol.* **1991**, *33*, 166.
- 28 15. Gordon, L. M.; Cohen, M. J.; MacRenaris, K. W.; Pasteris, J. D.; Seda, T., Amorphous
29 intergranular phases control the properties of rodent tooth enamel. *Science* **2015**, *347*, 746-750.
- 30 16. Tertuliano, O. A.; Greer, J. R., The nanocomposite nature of bone drives its strength and damage
31 resistance. *Nat. Mater.* **2016**, *15*, 1195-1202.
- 32 17. Zhao, J.; Liu, Y.; Sun, W. B.; Yang, X. B., First detection, characterization, and application of
33 amorphous calcium phosphate in dentistry. *J. Dent. Sci.* **2012**, *7*, 316-323.
- 34 18. Mahamid, J.; Aichmayer, B.; Shimoni, E.; Ziblat, R.; Li, C. H.; Siegel, S.; Paris, O.; Fratzl, P.;
35 Weiner, S.; Addadi, L., Mapping amorphous calcium phosphate transformation into crystalline mineral
36 from the cell to the bone in zebrafish fin rays. *Proc. Natl. Acad. Sci. U. S. A.* **2010**, *107*, 6316-6321.
- 37 19. Lowenstam, H. A.; Weiner, S., Transformation of Amorphous Calcium-Phosphate to Crystalline
38 Dahllite in the Radular Teeth of Chitons. *Science* **1985**, *227*, 51-53.
- 39 20. Olszta, M. J.; Cheng, X. G.; Jee, S. S.; Kumar, R.; Kim, Y. Y.; Kaufman, M. J.; Douglas, E. P.;
40 Gower, L. B., Bone structure and formation: A new perspective. *Mater. Sci. Eng. R Rep.* **2007**, *58*, 77-
41 116.
- 42 21. Akiva, A.; Kerschnitzki, M.; Pinkas, I.; Wagermaie, W.; Yaniv, K.; Fratzl, P.; Addadi, L.; Weiner,
43 S., Mineral Formation in the Larval Zebrafish Tail Bone Occurs via an Acidic Disordered Calcium
44 Phosphate Phase. *J. Am. Chem. Soc.* **2016**, *138*, 14481-14487.
- 45 22. La Fontaine, A.; Zavgorodniy, A.; Liu, H.; Zheng, R.; Swain, M.; Cairney, J., Atomic-scale
46 compositional mapping reveals Mg-rich amorphous calcium phosphate in human dental enamel. *Sci.*
47 *Adv.* **2016**, *2*, e1601145.
- 48 23. Olszta, M. J.; Douglas, E. P.; Gower, L. B., Scanning Electron Microscopic Analysis of the
49 Mineralization of Type I Collagen via a Polymer-Induced Liquid-Precursor (PILP) Process. *Calcif.*
50 *Tissue Int.* **2003**, *72*, 583-591.
- 51 24. Bleek, K.; AndreasTaubert, New developments in polymer-controlled, bioinspired calcium
52 phosphate mineralization from aqueous solution. *Acta Biomater.* **2013**, *9*, 6283-6321.
- 53 25. Jiao, K.; Niu, L. N.; Ma, C. F.; Huang, X. Q.; Pei, D. D.; Luo, T.; Huang, Q.; Chen, J. H.; Tay, F.

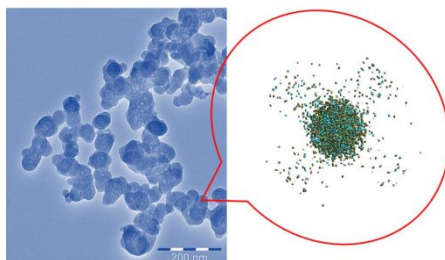
- R., Complementarity and Uncertainty in Intrafibrillar Mineralization of Collagen. *Adv. Funct. Mater.* **2016**, *26*, 6858-6875.
26. Dorozhkin, S. V., Amorphous calcium (ortho)phosphates. *Acta Biomater.* **2010**, *6*, 4457-4475.
27. Betts, F.; Posner, A. S., An X-ray radial distribution study of amorphous calcium phosphate. *Mater. Res. Bull.* **1974**, *9*, 353-360.
28. Posner, A. S.; Betts, F., Synthetic Amorphous Calcium-Phosphate and Its Relation to Bone-Mineral Structure. *Acc. Chem. Res.* **1975**, *8*, 273-281.
29. Holt, C.; Vankemenade, M. J. J. M.; Harries, J. E.; Nelson, L. S.; Bailey, R. T.; Hukins, D. W. L.; Hasnain, S. S.; Debruyne, P. L., Preparation of Amorphous Calcium-Magnesium Phosphates at Ph-7 and Characterization by X-Ray Absorption and Fourier-Transform Infrared-Spectroscopy. *J. Cryst. Growth* **1988**, *92*, 239-252.
30. Holt, C.; Vankemenade, M. J. J. M.; Nelson, L. S.; Sawyer, L.; Harries, J. E.; Bailey, R. T.; Hukins, D. W. L., Composition and Structure of Micellar Calcium-Phosphate. *J. Dairy Res.* **1989**, *56*, 411-416.
31. Zyman, Z.; Epple, M.; Goncharenko, A.; Rokhmistrov, D.; Prymak, O.; Loza, K., Thermally induced crystallization and phase evolution in powders derived from amorphous calcium phosphate precipitates with a Ca/P ratio of 1:1. *J. Cryst. Growth* **2016**, *450*, 190-196.
32. Zyman, Z.; Goncharenko, A.; Rokhmistrov, D., Kinetics and mechanisms of the transformation of precipitated amorphous calcium phosphate with a Ca/P ratio of 1:1 to calcium pyrophosphates. *J. Cryst. Growth* **2017**, *478*, 117-122.
33. Layrolle, P.; Lebugle, A., Characterization and Reactivity of Nanosized Calcium Phosphates Prepared in Anhydrous Ethanol. *Chem. Mater.* **1994**, *6*, 1996-2004.
34. Holt, C.; Vankemenade, M. J. J. M.; Nelson, L. S.; Hukins, D. W. L.; Bailey, R. T.; Harries, J. E.; Hasnain, S. S.; Debruyne, P. L., Amorphous Calcium Phosphates Prepared at Ph 6.5 and 6.0. *Mater. Res. Bull.* **1989**, *24*, 55-62.
35. Habraken, W. J. E. M.; Tao, J. H.; Brylka, L. J.; Friedrich, H.; Bertinetti, L.; Schenk, A. S.; Verch, A.; Dmitrovic, V.; Bomans, P. H. H.; Frederik, P. M.; Laven, J.; van der Schoot, P.; Aichmayer, B.; de With, G.; DeYoreo, J. J.; Sommerdijk, N. A. J. M., Ion-association complexes unite classical and non-classical theories for the biomimetic nucleation of calcium phosphate. *Nat. Comm.* **2013**, *4*, Art. No. 1507.
36. Gebauer, D.; Kellermeier, M.; Gale, J. D.; Bergstrom, L.; Colfen, H., Pre-nucleation clusters as solute precursors in crystallisation. *Chem. Soc. Rev.* **2014**, *43*, 2348-2371.
37. Bailey, R. T.; Holt, C., Fourier transform infrared spectroscopy and characterisation of biological calcium phosphates. 1989; pp 93-120.
38. Eichert, D.; Salome, M.; Banu, M.; Susini, J.; Rey, C., Preliminary characterization of calcium chemical environment in apatitic and non-apatitic calcium phosphates of biological interest by X-ray absorption spectroscopy. *Spectrochim. Acta, Part B* **2005**, *60*, 850-858.
39. Shpak, A. P.; Karbovskii, V. L.; Trachevskii, V. V., Peculiarities of the electronic structure of the ultra disperse calcium hydroxyapatite. *J. Electron. Spectrosc. Relat. Phenom.* **1998**, *88*, 973-976.
40. Nelson, L. S.; Holt, C.; Harries, J. E.; Hukins, D. W. L., Amorphous calcium phosphates of different composition give very similar EXAFS spectra. *Physica B* **1989**, *158*, 105-106.
41. Holt, C.; Hukins, D. W. L., Structural Analysis of the Environment of Calcium Ions in Crystalline and Amorphous Calcium Phosphates by X-Ray Absorption Spectroscopy and a Hypothesis Concerning the Biological Function of the Casein Micelle. *Int. Dairy J.* **1991**, *1*, 151-165.
42. Treboux, G.; Layrolle, P.; Kanzaki, N.; Onuma, K.; Ito, A., Existence of Posner's cluster in vacuum. *J. Phys. Chem. A* **2000**, *104*, 5111-5114.
43. Hughes, J. M.; Cameron, M.; Crowley, K. D., Structural Variations in Natural F, Oh, and Cl Apatites. *Am. Mineral.* **1989**, *74*, 870-876.
44. Catti, M.; Ferraris, G.; Filhol, A., Hydrogen bonding in the crystalline state. CaHPO₄ (monetite), $\overline{P1}$ or P1? A novel neutron diffraction study. *Acta Crystallogr. Sect. B* **1977**, *33*, 1223-1229.
45. Gebauer, D.; Gunawidjaja, P. N.; Ko, J. Y. P.; Bacsik, Z.; Aziz, B.; Liu, L. J.; Hu, Y. F.; Bergstrom, L.; Tai, C. W.; Sham, T. K.; Eden, M.; Hedin, N., Proto-Calcite and Proto-Vaterite in

- 1
2 Amorphous Calcium Carbonates. *Angew. Chem. Int. Ed.* **2010**, *49*, 8889-8891.
- 3 46. Farhadi-Khouzani, M.; Chevrier, D. M.; Zhang, P.; Hedin, N.; Gebauer, D., Water as the Key to
4 Proto-Aragonite Amorphous CaCO₃. *Angew. Chem. Int. Ed.* **2016**, *55*, 8117-8120.
- 5 47. Legeros, R. Z., Preparation of Octacalcium Phosphate (Ocp) - a Direct Fast Method. *Calcif. Tissue*
6 *Int.* **1985**, *37*, 194-197.
- 7 48. Demichelis, R.; Garcia, N. A.; Raiteri, P.; Innocenti Malini, R.; Freeman, C. L.; Harding, J. H.;
8 Gale, J. D., Simulation of Calcium Phosphate Species in Aqueous Solution: Force Field Derivation. *J.*
9 *Phys. Chem. B* **2018**, *122*, 1471-1483.
- 10 49. Lippert, G.; Hutter, J.; Parrinello, M., The Gaussian and augmented-plane-wave density functional
11 method for ab initio molecular dynamics simulations. *Theor. Chem. Acc.* **1999**, *103*, 124-140.
- 12 50. VandeVondele, J.; Krack, M.; Mohamed, F.; Parrinello, M.; Chassaing, T.; Hutter, J., Quickstep:
13 Fast and accurate density functional calculations using a mixed Gaussian and plane waves approach.
14 *Comput. Phys. Commun.* **2005**, *167*, 103-128.
- 15 51. Goedecker, S.; Teter, M.; Hutter, J., Separable dual-space Gaussian pseudopotentials. *Phys. Rev. B*
16 **1996**, *54*, 1703-1710.
- 17 52. Grimme, S.; Antony, J.; Ehrlich, S.; Krieg, H., A consistent and accurate ab initio parametrization
18 of density functional dispersion correction (DFT-D) for the 94 elements H-Pu. *J. Chem. Phys.* **2010**,
19 *132*, 154104.
- 20 53. Chamorro, E.; De Proft, F.; Geerlings, P., Hardness and softness reactivity kernels within the spin-
21 polarized density-functional theory. *J. Chem. Phys.* **2005**, *123*, 154104.
- 22 54. Michaud-Agrawal, N.; Denning, E. J.; Woolf, T. B.; Beckstein, O., Software News and Updates
23 MDAnalysis: A Toolkit for the Analysis of Molecular Dynamics Simulations. *J. Comput. Chem.* **2011**,
24 *32*, 2319-2327.
- 25 55. Theobald, D. L., Rapid calculation of RMSDs using a quaternion-based characteristic polynomial.
26 *Acta Crystallogr., Sect. A* **2005**, *61*, 478-480.
- 27 56. Liu, P.; Agrafiotis, D. K.; Theobald, D. L., Fast Determination of the Optimal Rotational Matrix for
28 Macromolecular Superpositions. *J. Comput. Chem.* **2010**, *31*, 1561-1563.
- 29 57. Clark Stewart, J.; Segall Matthew, D.; Pickard Chris, J.; Hasnip Phil, J.; Probert Matt, I. J.; Refson,
30 K.; Payne Mike, C., First principles methods using CASTEP. *Z. Für Krist. - Cryst. Mater.* **2005**, *220*,
31 567.
- 32 58. Grimme, S., Semiempirical GGA - type density functional constructed with a long - range
33 dispersion correction. *J. Comput. Chem.* **2006**, *27*, 1787-1799.
- 34 59. Pickard, C. J.; Mauri, F., All-electron magnetic response with pseudopotentials: NMR chemical
35 shifts. *Phys. Rev. B* **2001**, *63*, 245101.
- 36 60. Yates, J. R.; Pickard, C. J.; Mauri, F., Calculation of NMR chemical shifts for extended systems
37 using ultrasoft pseudopotentials. *Phys. Rev. B* **2007**, *76*, 024401.
- 38 61. Cho, G.; Wu, Y.; Ackerman, J. L., Detection of Hydroxyl Ions in Bone Mineral by Solid-State
39 NMR Spectroscopy. *Science* **2003**, *300*, 1123-1127.
- 40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2 FOR TABLE OF CONTENTS USE ONLY
3

4 **On the Short-Range Structure of Amorphous Calcium Hydrogen Phosphate**

5
6 Bing-Qiang Lu, Natalya A. Garcia, Daniel M. Chevrier, Peng Zhang, Paolo Raiteri, Julian D. Gale and
7
8 Denis Gebauer
9



20 We propose a new model for the short-range structure of amorphous calcium hydrogen phosphate,
21 which is distinct from Posner's cluster-based models. This is important for the determination of the
22 structures of diverse ACPs with varying Ca/P atomic ratios, and also for fundamental understanding of
23 a major mineral class that is central to biomineralization in vertebrates.
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60