

Characterizing the performance of the MinION for real-time detection

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Nanopore technology – Fieldable 3rd generation sequencing

- Oxford Nanopore Technology (ONT)'s MinION uses electrical impedance to “read” DNA as it transits through arrayed nanopores on disposable chip
- Major advantages
 - Long reads
 - Real-time data
 - Interactive sequencing (ReadUntil)
 - Field-ready, small footprint

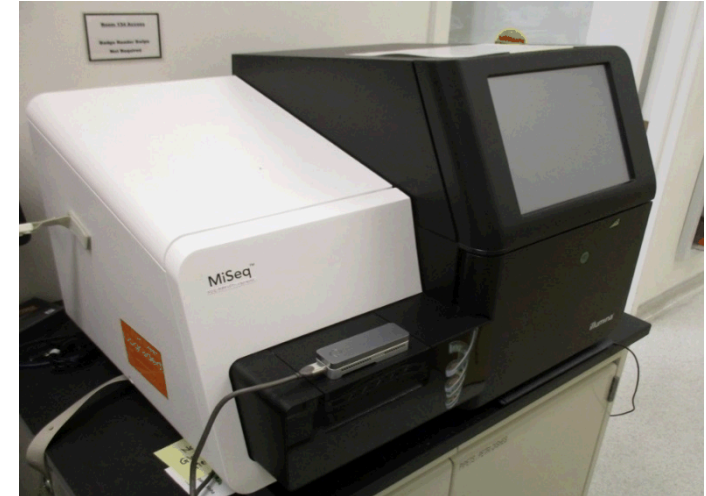


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Nanopore technology – Fieldable 3rd generation sequencing

- What can this sequencer address in the field of real-time detection?
 - Co-occurring genes (eg resistance, toxins), unique splice variants etc
 - Needle in a haystack – small amounts of pathogen DNA in large background samples
- What needs to be done?
 - Understanding how error rate affects ability to detect mutations (statistical confidence of calls)
 - Benchmark error (stochastic vs deterministic, factors affecting)
 - Develop experimental and bioinformatic pipelines that allow us to generate large amounts of accurate data

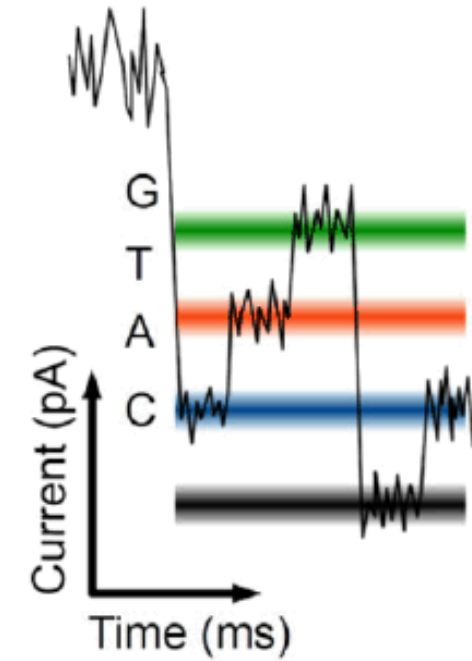
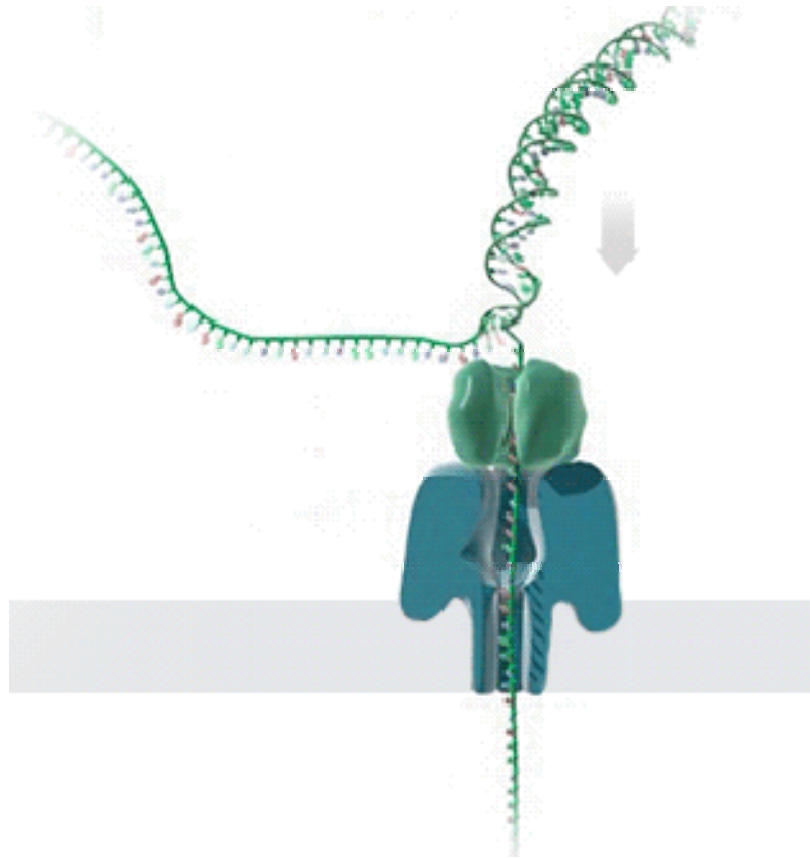


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nanoporetech.com

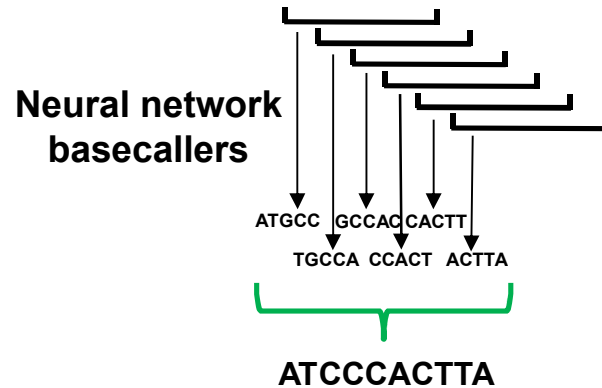
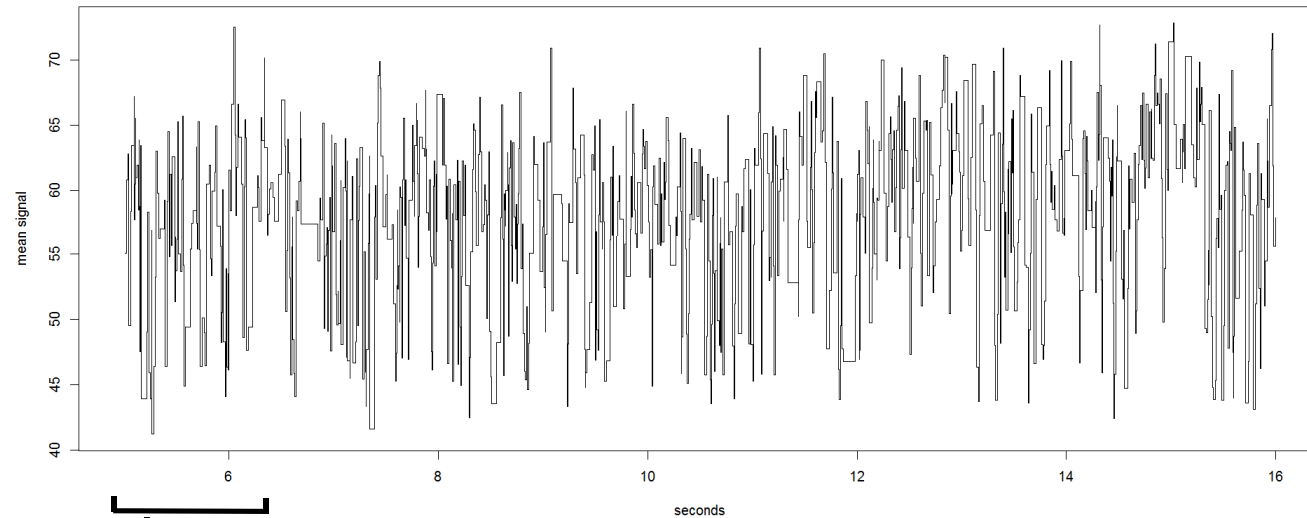
Nanopore sequencing technology



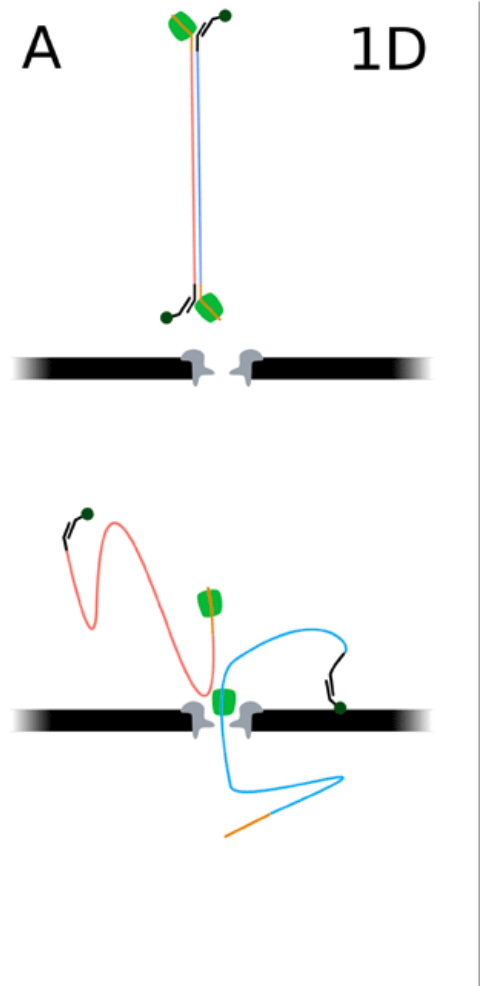
Steinbock and Radenovic 2016

MinION data analysis

Raw and/or event-based “squiggle” plot generated in real time during sequencing



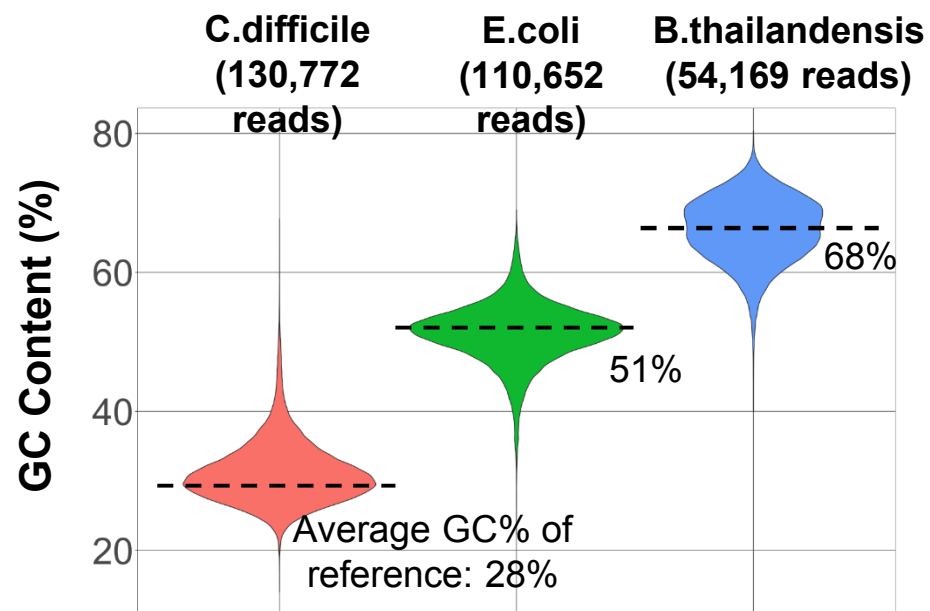
1D vs 2D vs 1D²



de Lannoy et al (F1000 Research 2017)

Understanding error sources in nanopore sequencing

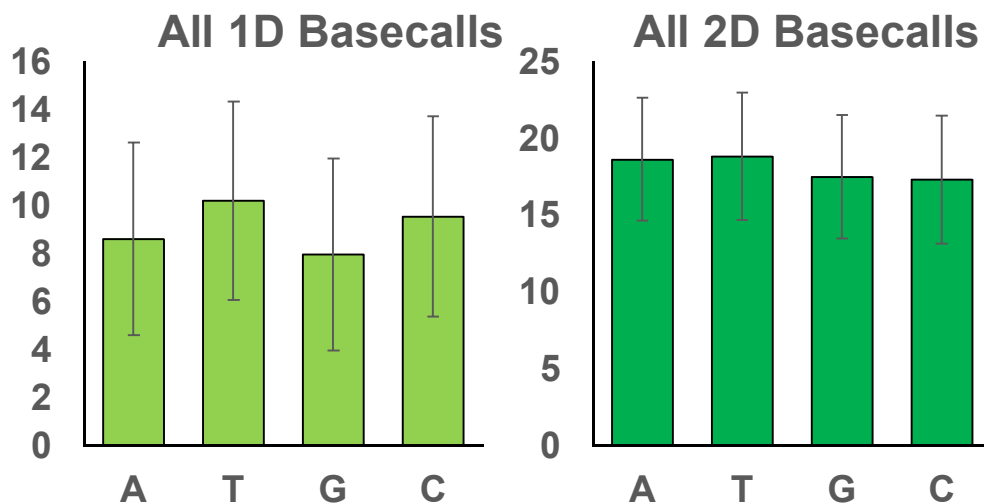
GC content of underlying sample is overall reproducible



Krishnakumar et al (Scientific Reports 2018)

Understanding error sources in nanopore sequencing

There are systematic base-specific biases

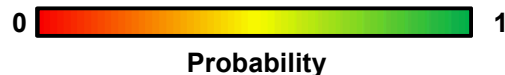


Understanding error sources in nanopore sequencing

There is strong context-specific dependence of base quality

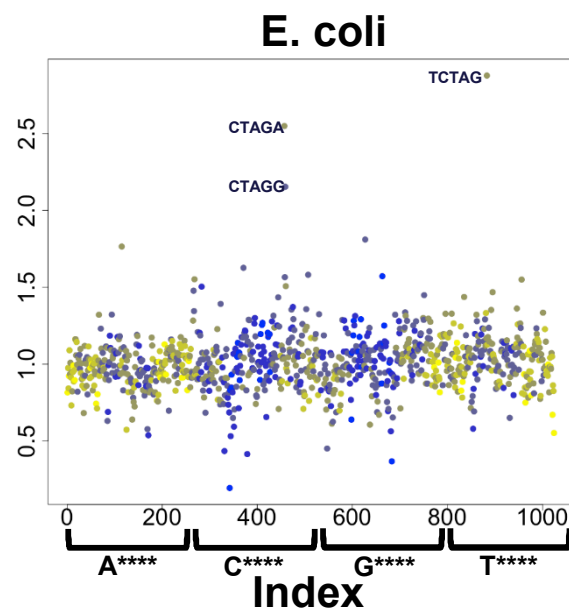
Q score of next base call will be

E. coli (2D)		1	2	3	4	5	6	7	8	9	10
Given Current Base Call Quality Score Decile	Lowest - 1	0.46	0.19	0.19	0.07	0.04	0.02	0.01	0.01	0.01	0.00
	2	0.25	0.22	0.27	0.11	0.06	0.04	0.02	0.01	0.01	0.00
	3	0.13	0.15	0.31	0.16	0.11	0.06	0.04	0.03	0.01	0.01
	4	0.07	0.08	0.22	0.21	0.17	0.11	0.06	0.05	0.03	0.01
	5	0.04	0.04	0.14	0.16	0.21	0.16	0.10	0.08	0.04	0.02
	6	0.02	0.03	0.08	0.11	0.17	0.21	0.16	0.13	0.07	0.03
	7	0.01	0.02	0.05	0.07	0.11	0.17	0.20	0.20	0.12	0.04
	8	0.01	0.01	0.03	0.04	0.07	0.11	0.16	0.27	0.21	0.08
	9	0.00	0.00	0.02	0.02	0.04	0.06	0.09	0.20	0.35	0.20
	Highest - 10	0.00	0.00	0.01	0.01	0.02	0.03	0.04	0.09	0.24	0.57



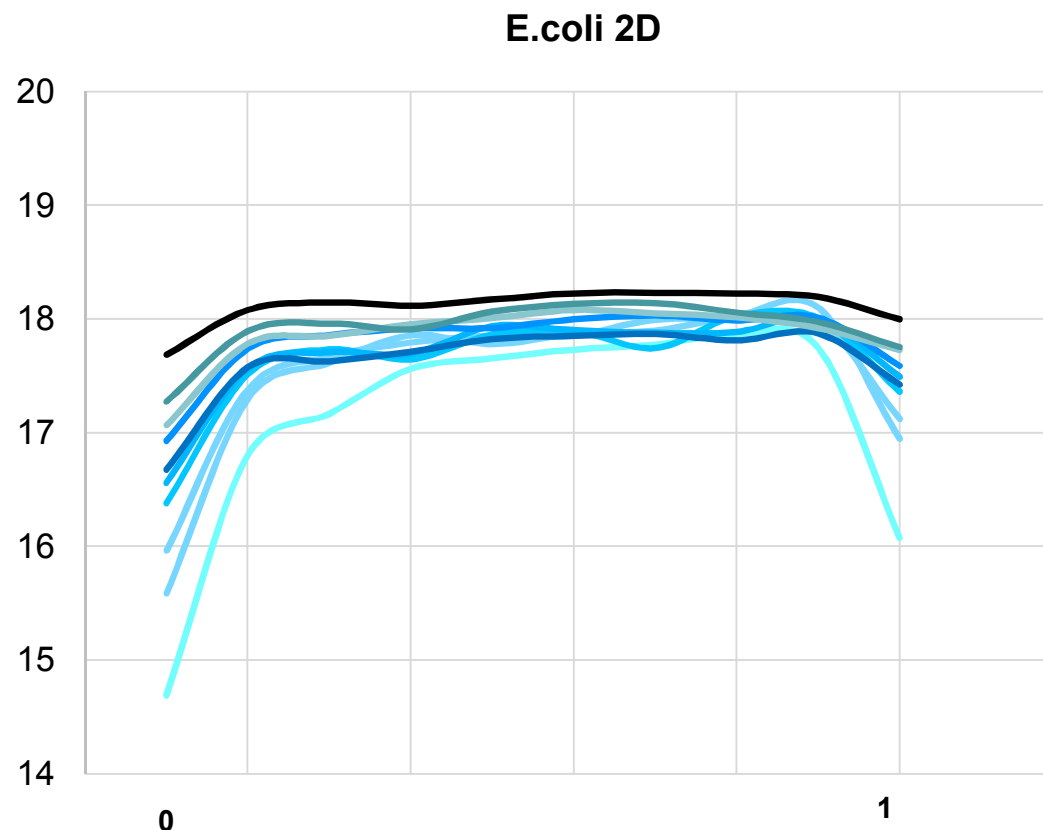
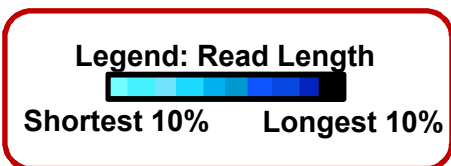
Understanding error sources in nanopore sequencing

There are context-specific k -mer biases



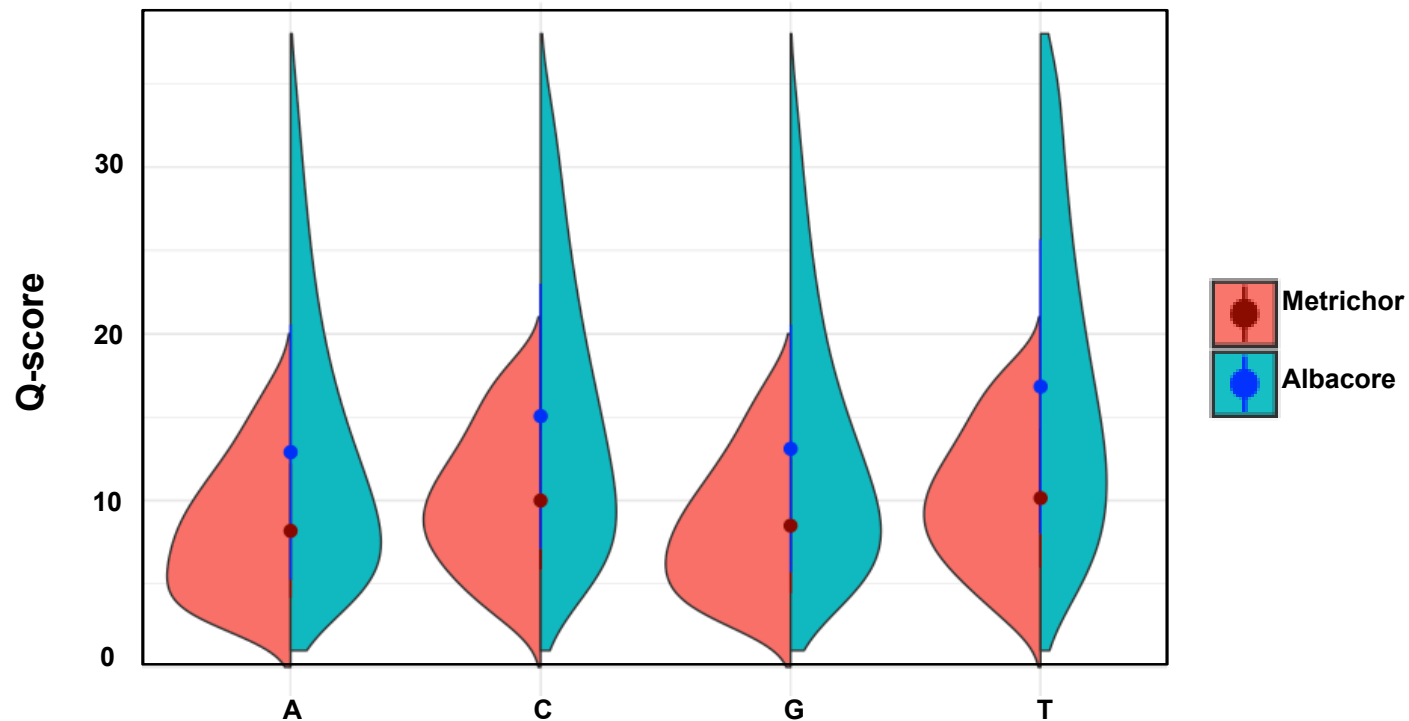
Understanding error sources in nanopore sequencing

There are positional biases, length-dependent biases along the read



Evolving basecallers

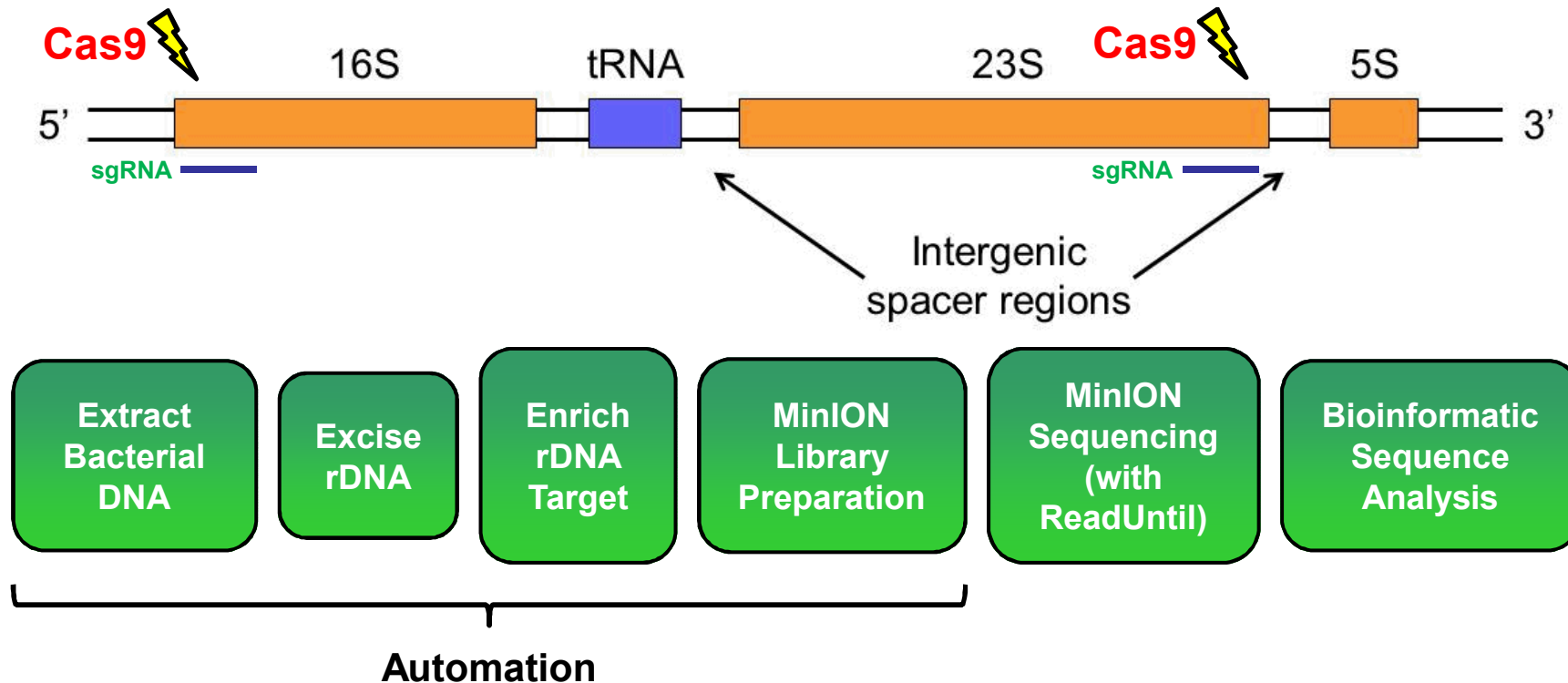
Metrichor vs Albacore



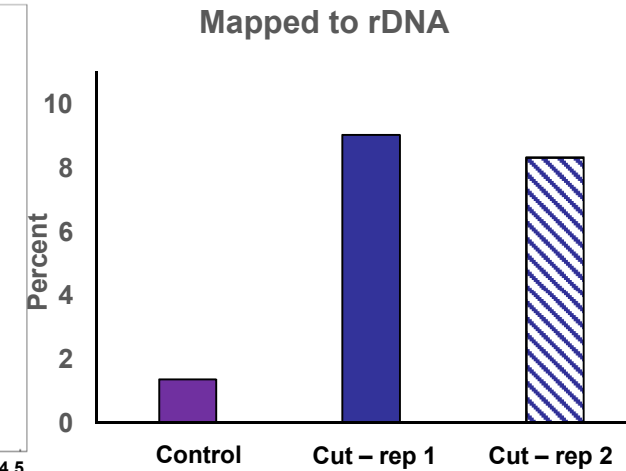
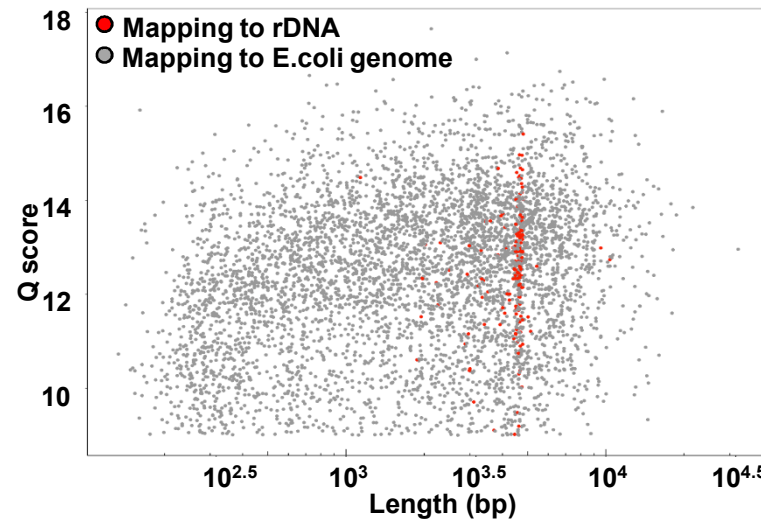
We utilize this information to identify genomic regions/sequences that are optimal for high-confidence detection

Real-time automated pathogen identification by enhanced ribotyping (RAPIER)

- Goal: Rapid, universal, amplification-free, pathogen ID & characterization suitable for clinical, fieldable, & low-resource settings
- Universally conserved bacterial rDNA locus for species typing
- Long reads span the entire locus



Cas9-based selection of rDNA

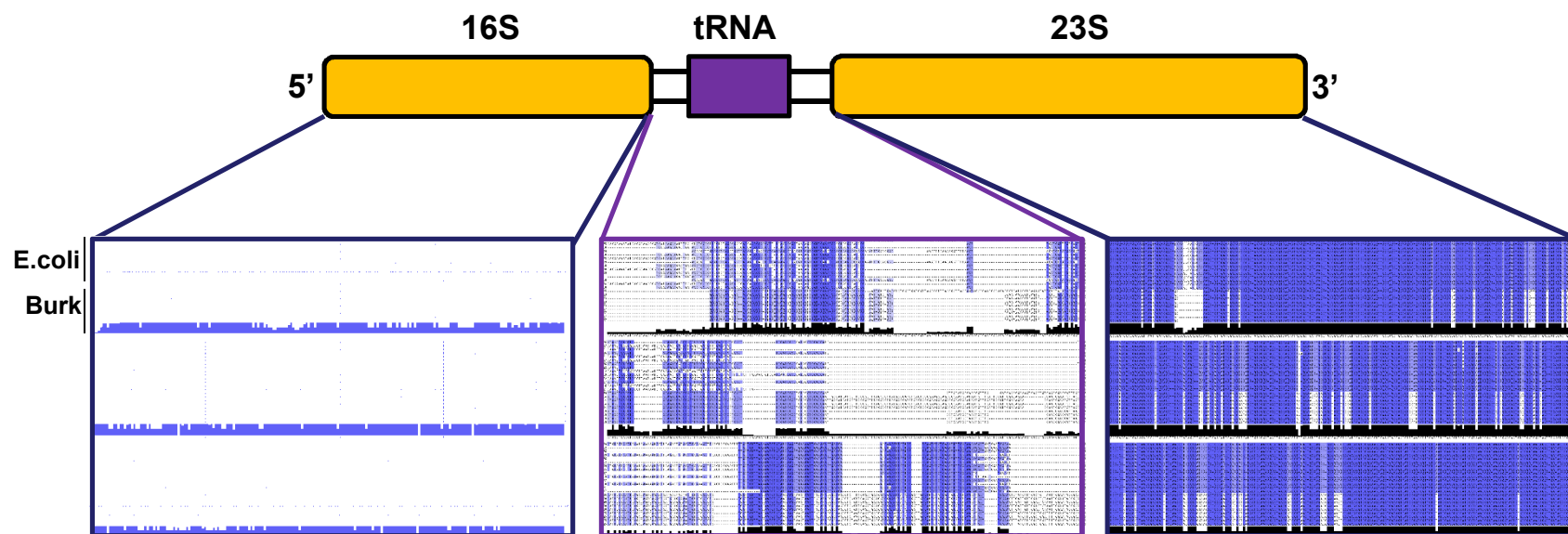


Cutting out the rDNA locus enriches its sequencing

Sorting bacterial species from error-prone rDNA reads - pipeline

QC AND BASE CALLING

CRISPR-generated rDNA reads from minION → Q-score filtering → Local basecalling → Read correction



Sorting bacterial species from error-prone rDNA reads - pipeline

QC AND BASE CALLING

CRISPR-generated rDNA reads from minION → Q-score filtering → Local basecalling → Read correction

ALIGNMENT 1 – 16S AND 23S FAMILY SORTING



ALIGNMENT 2 – INTERGENIC AND tRNA SUBGROUP SORTING



Future Perspectives

Knowledge of error behavior allows for smart selection of target

Experimental enrichment targets (eg. Cas9) can enhance probability of detection

Developing models to make rapid diagnostic calls is made possible by

- 1) the groundwork laid by the above**
- 2) implementing real-time selective sequencing (Read Until)**
- 3) empirical testing of real-world samples**

Mike Bartsch: Real-time selection can search for ‘needles in a haystack’

Acknowledgements

Mike Bartsch (PI)

Sara Bird

Steve Branda

Harrison Edwards

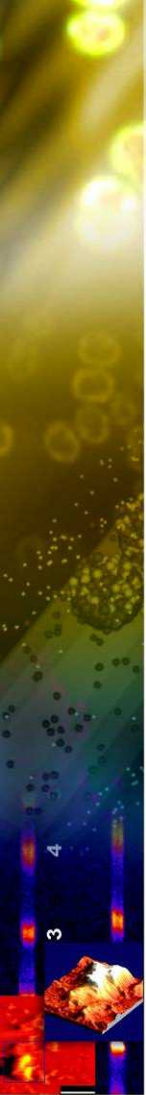
Hari Jayamohan

Ken Patel

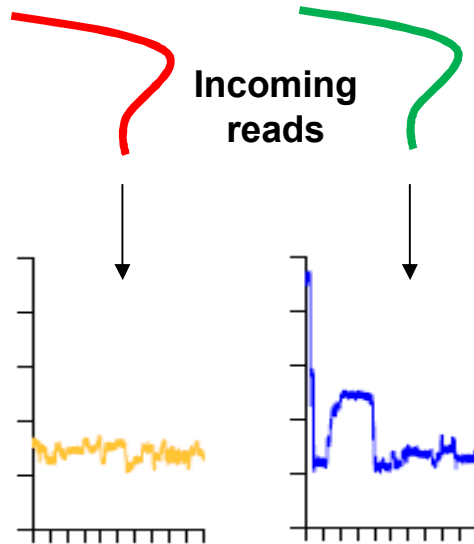
Joe Schoeniger

Anupama Sinha

Funding – Sandia LDRD



RUBRIC – Read Until with Basecall- and Reference-Informed Criteria



Real time
basecall

TTCATAG

REJECT

TTAGATC

SEQUENCE

Real time
alignment

TTAATAAGGGTTAGATCG
CTTCTTTTCTAAGGCTAG
CTATTGTTTCTTTCTTTC

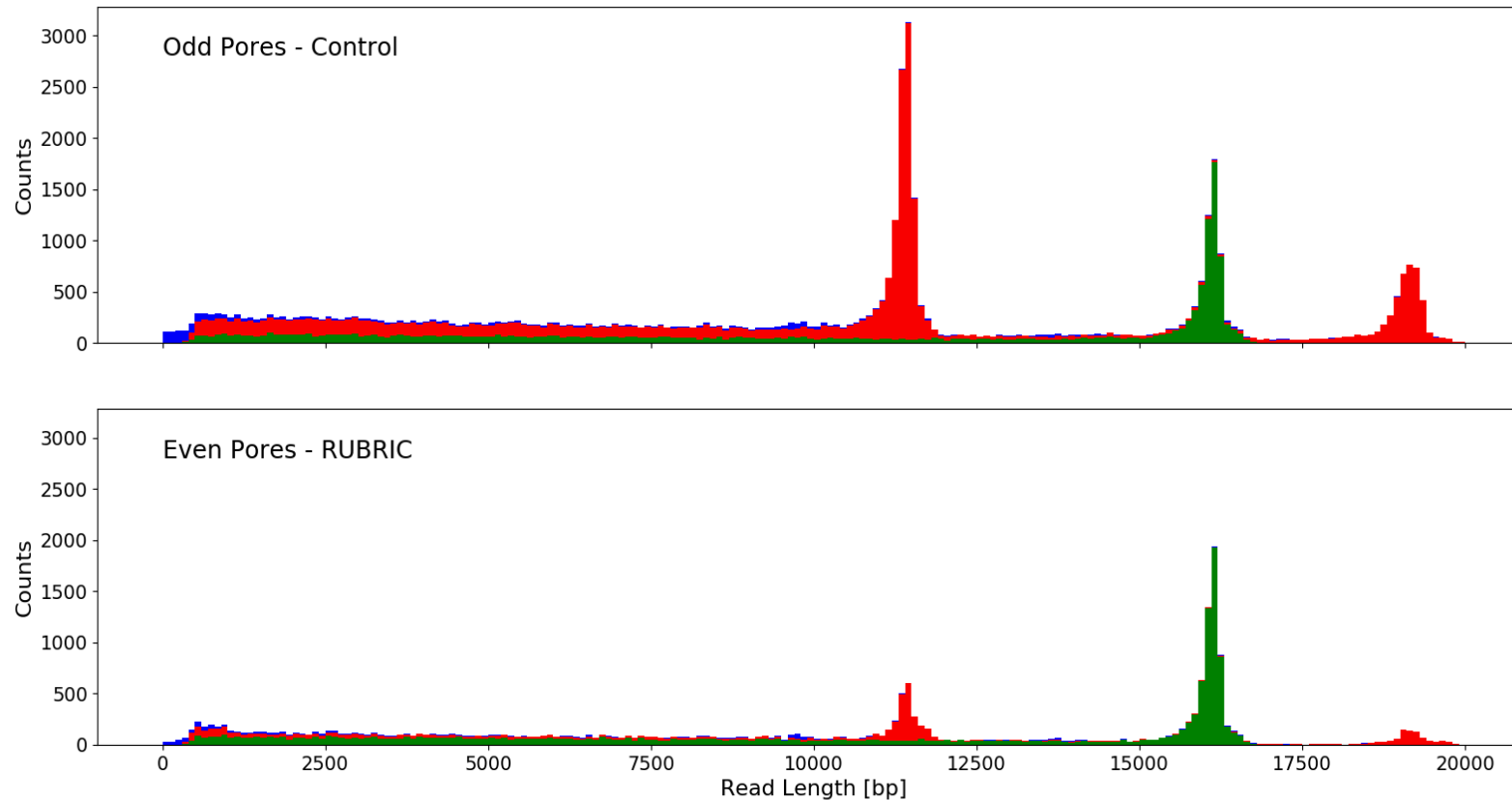
Sandia RUBRIC

RUBRIC – Read Until with Basecall- and Reference-Informed Criteria

**Bacteriophage
Lambda**



Eag1 sites



Long read sequencing technology

	Long read (eg. MinION)	Short read (eg. MiSeq)
LENGTH	Upto 100s of kb • Assembly of structurally challenging or repetitive regions • Splice variants • 3D structure	Upto 600 bp
AMPLIFICATION REQUIRED?	No	Yes
REAL TIME OUTPUT	Yes	No
SELECTIVE SEQUENCING	Yes	No
THROUGHPUT	0.88 Gb/hr	0.25 Gb/hr
COST	\$17/Gb	\$53/Gb
ACCURACY	~85 – 95%	~99.99%

Figure 3

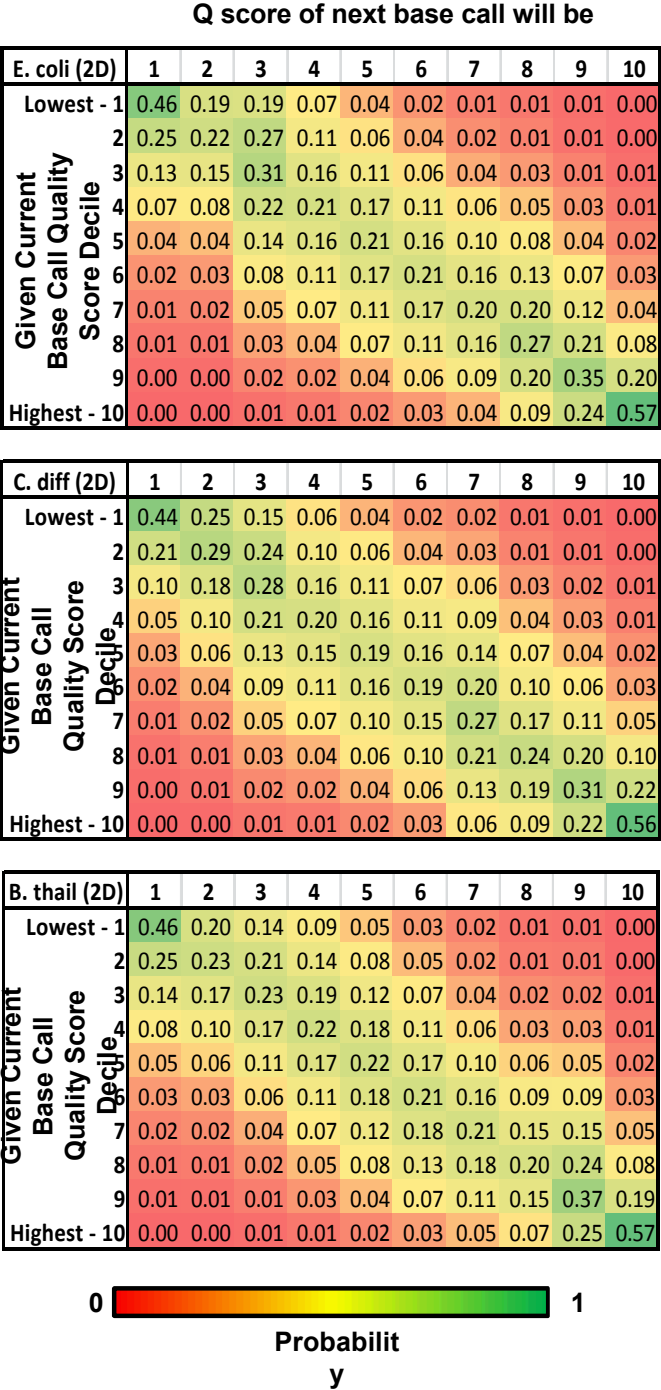


Figure 4

<i>E.coli</i>		1D Run				2D Run															
						2D template				2D complement				1D reads				2D reads			
		A	C	T	G	A	C	T	G	A	C	T	G	A	C	T	G	A	C	T	G
Quality Score Decile	Lowest - 1	0.37	0.16	0.15	0.33	0.37	0.16	0.15	0.32	0.34	0.15	0.15	0.36	0.36	0.17	0.17	0.33	0.28	0.24	0.22	0.27
	2	0.32	0.19	0.16	0.33	0.31	0.19	0.17	0.33	0.32	0.18	0.15	0.35	0.31	0.20	0.20	0.33	0.25	0.26	0.23	0.26
	3	0.27	0.22	0.20	0.31	0.28	0.22	0.19	0.32	0.28	0.21	0.18	0.33	0.28	0.22	0.22	0.31	0.23	0.27	0.24	0.25
	4	0.24	0.25	0.23	0.28	0.25	0.24	0.21	0.30					0.26	0.23	0.23	0.29	0.23	0.27	0.25	0.25
	5	0.23	0.26	0.25	0.26	0.23	0.26	0.24	0.27	0.26	0.23	0.21	0.30	0.24	0.25	0.25	0.27	0.23	0.27	0.25	0.25
	6	0.22	0.27	0.26	0.25	0.22	0.28	0.26	0.25	0.24	0.25	0.24	0.27	0.22	0.27	0.27	0.25	0.23	0.26	0.25	0.25
	7	0.21	0.28	0.27	0.24	0.21	0.28	0.27	0.24	0.23	0.27	0.26	0.24	0.21	0.28	0.28	0.23	0.24	0.25	0.25	0.25
	8	0.20	0.28	0.29	0.23	0.20	0.29	0.28	0.23	0.21	0.29	0.29	0.20	0.20	0.29	0.29	0.22	0.24	0.25	0.25	0.25
	9	0.19	0.29	0.30	0.22	0.19	0.30	0.29	0.22	0.20	0.31	0.31	0.17	0.19	0.30	0.30	0.21	0.25	0.24	0.26	0.25
	Highest - 10	0.15	0.33	0.33	0.19	0.15	0.34	0.32	0.19	0.18	0.33	0.37	0.12	0.15	0.34	0.34	0.18	0.25	0.24	0.25	0.25

<i>C.diff</i>		2D template				2D complement				2D reads			
		A	C	T	G	A	C	T	G	A	C	T	G
Quality Score Decile	Lowest - 1	0.34	0.19	0.14	0.33	0.29	0.16	0.15	0.40	0.28	0.24	0.23	0.25
	2	0.36	0.18	0.19	0.27	0.35	0.16	0.17	0.33	0.31	0.19	0.31	0.19
	3	0.35	0.17	0.27	0.22	0.36	0.16	0.22	0.27	0.32	0.17	0.35	0.16
	4	0.34	0.16	0.33	0.18					0.33	0.16	0.36	0.15
	5	0.33	0.16	0.36	0.15	0.36	0.16	0.27	0.21	0.34	0.15	0.37	0.14
	6	0.33	0.16	0.38	0.14	0.35	0.16	0.32	0.17	0.35	0.14	0.38	0.13
	7	0.32	0.15	0.41	0.12	0.34	0.15	0.37	0.13	0.36	0.13	0.38	0.13
	8	0.32	0.15	0.43	0.10	0.33	0.16	0.42	0.10	0.37	0.12	0.39	0.12
	9	0.32	0.15	0.45	0.09	0.32	0.16	0.47	0.06	0.38	0.12	0.39	0.12
	Highest - 10	0.31	0.13	0.51	0.05	0.27	0.14	0.57	0.02	0.38	0.11	0.39	0.12

<i>B.thail</i>		2D template				2D complement				2D reads			
		A	C	T	G	A	C	T	G	A	C	T	G
Quality Score Decile	Lowest - 1	0.39	0.14	0.15	0.32	0.35	0.15	0.13	0.37	0.26	0.25	0.21	0.29
	2	0.27	0.20	0.14	0.39					0.19	0.32	0.18	0.31
	3	0.19	0.26	0.16	0.39	0.29	0.20	0.12	0.39	0.16	0.35	0.17	0.32
	4	0.15	0.31	0.17	0.37	0.24	0.25	0.13	0.38	0.14	0.36	0.17	0.33
	5	0.13	0.34	0.17	0.36	0.20	0.30	0.16	0.34	0.14	0.36	0.17	0.34
	6	0.12	0.37	0.17	0.35	0.17	0.35	0.18	0.30	0.14	0.35	0.16	0.35
	7	0.11	0.38	0.17	0.34					0.14	0.35	0.16	0.35
	8	0.09	0.40	0.17	0.33	0.14	0.40	0.20	0.26	0.14	0.34	0.16	0.36
	9	0.08	0.42	0.17	0.32	0.12	0.44	0.22	0.22	0.14	0.34	0.16	0.36
	Highest - 10	0.05	0.49	0.17	0.29	0.10	0.47	0.26	0.17	0.15	0.33	0.15	0.37

Figure 5

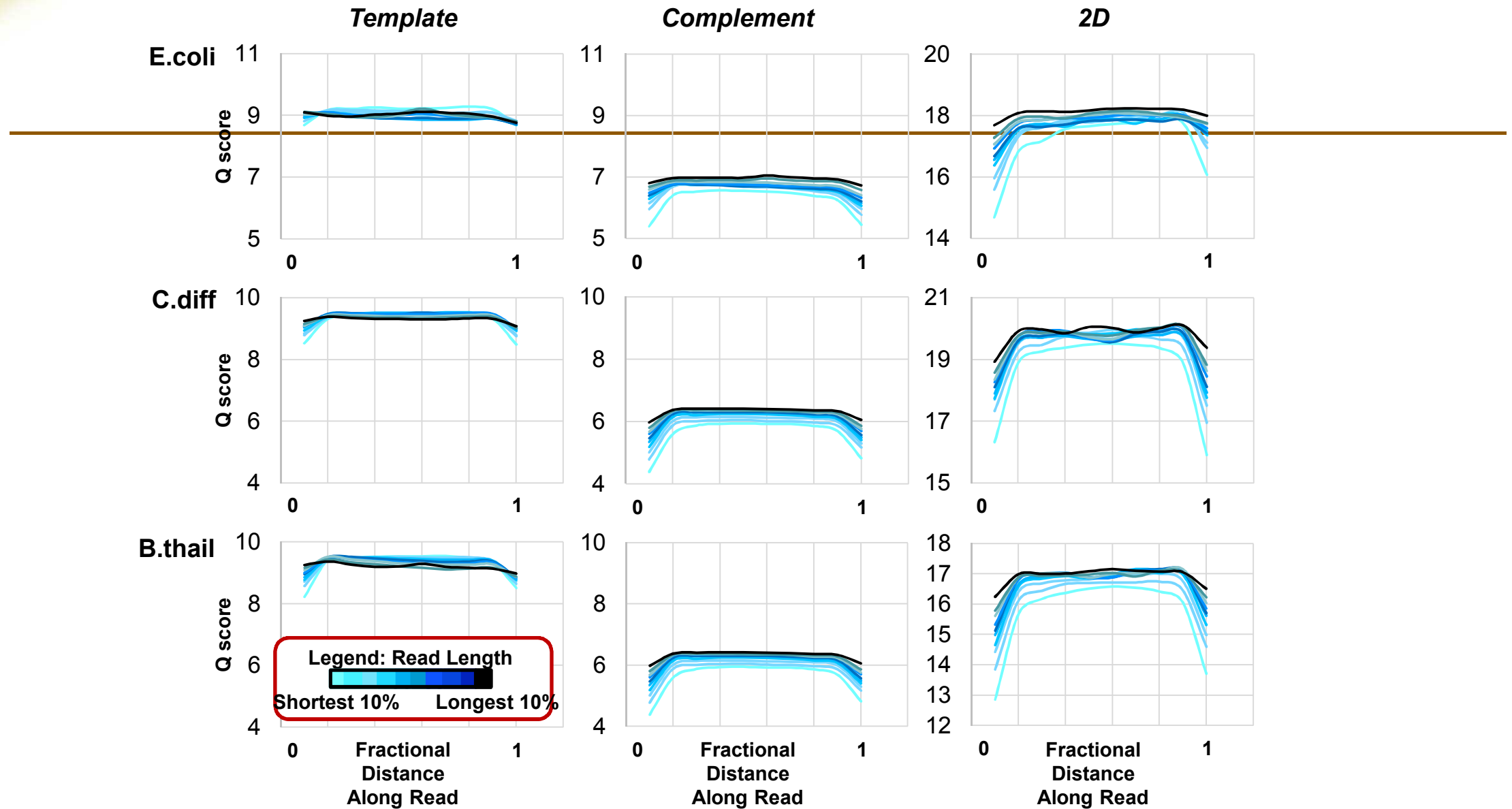


Figure 7

