

Visualization of Photosystem II

SAND2014-16650PE

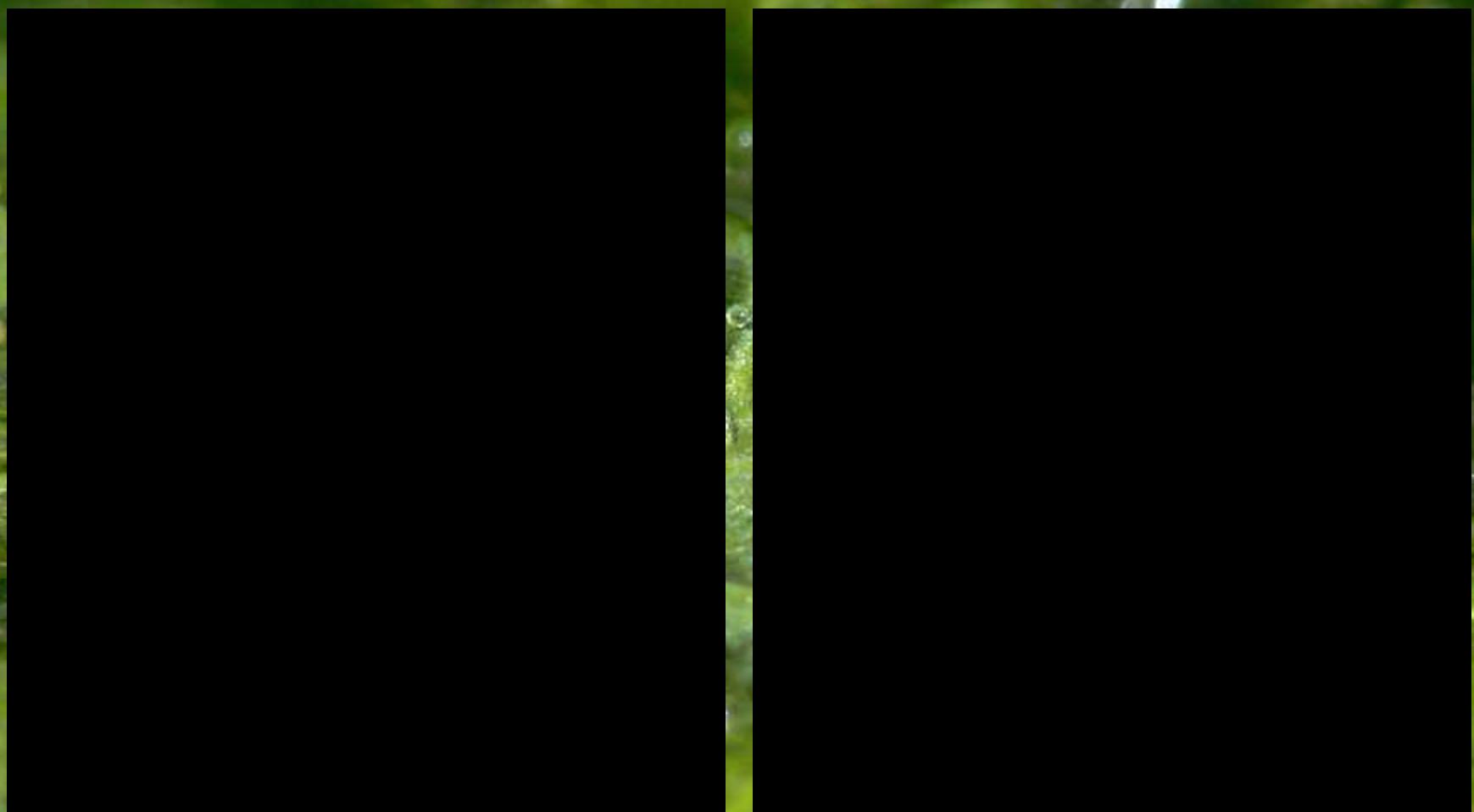
Distribution in Encapsulated *Chlamydomona reinhardtii*

Aysha McClory

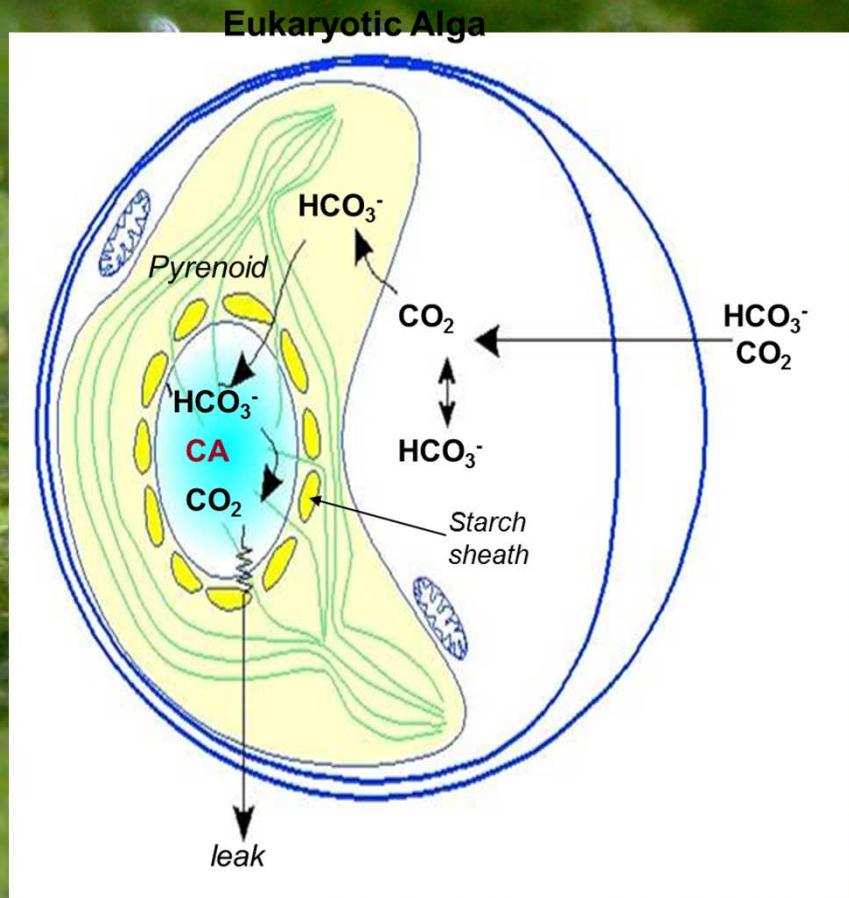
7002 WT
Amelia 1B
8/8/14 TMA

7002.I.6-22
Amelia 271B
5/8/14 TMA

Light: The Drive of Photosystem II



C. hlamydomonas rheinhardtii



Hyperspectral Microscopy



*.hsf files were converted to s3d files using batch script that reads in data with Lidke code, then calls Creates3d.m. Expansion factor was utilized to improve precision.

*.s3d files were opened individually in ShowMe3D.exe and checked for integrity/quality.

Sets of *.s3d files that were to be compared were combined into a composite data set using <Preprocess> function in ImageMCR.

MCR was run on resulting composite files, excluding any individual traces known to contain saturated pixels (more on this later - Slide 4).

Cell Concentration Hemocytometer

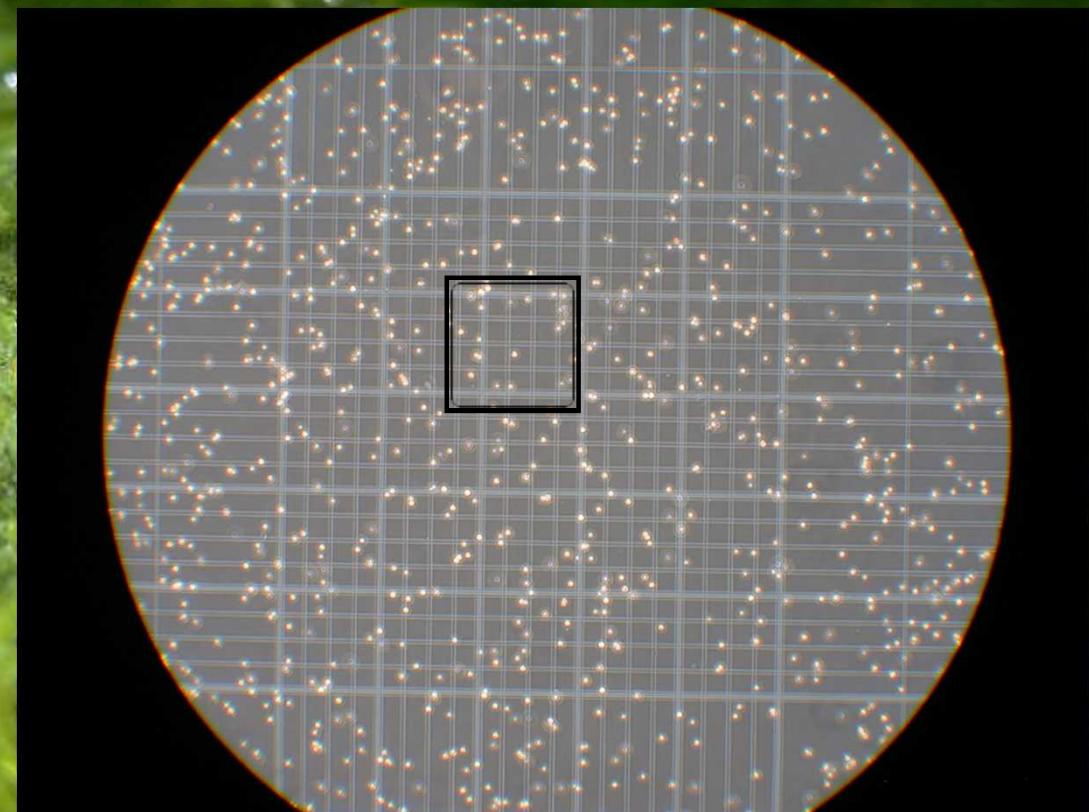
$$C = \Sigma (N/V)$$

0.04 square mm

N is cells in squares

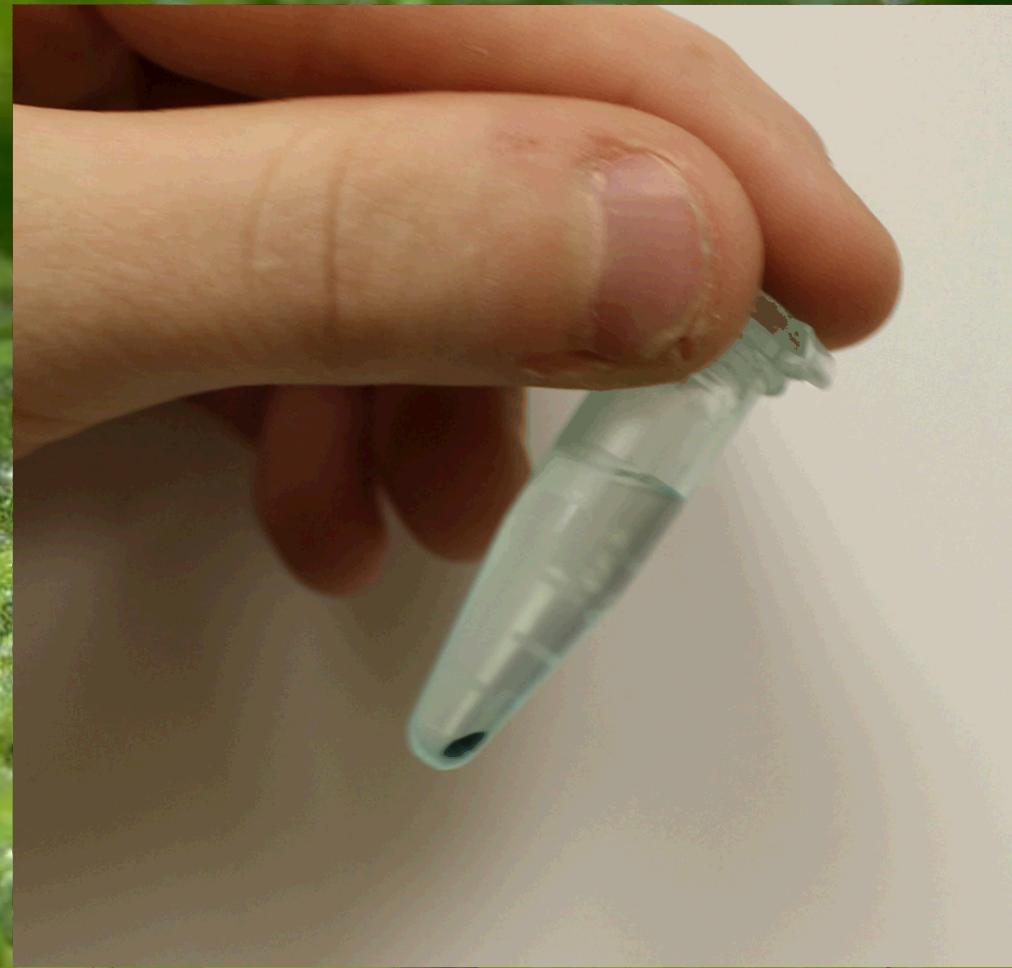
V is volume

calculated per #
square counted



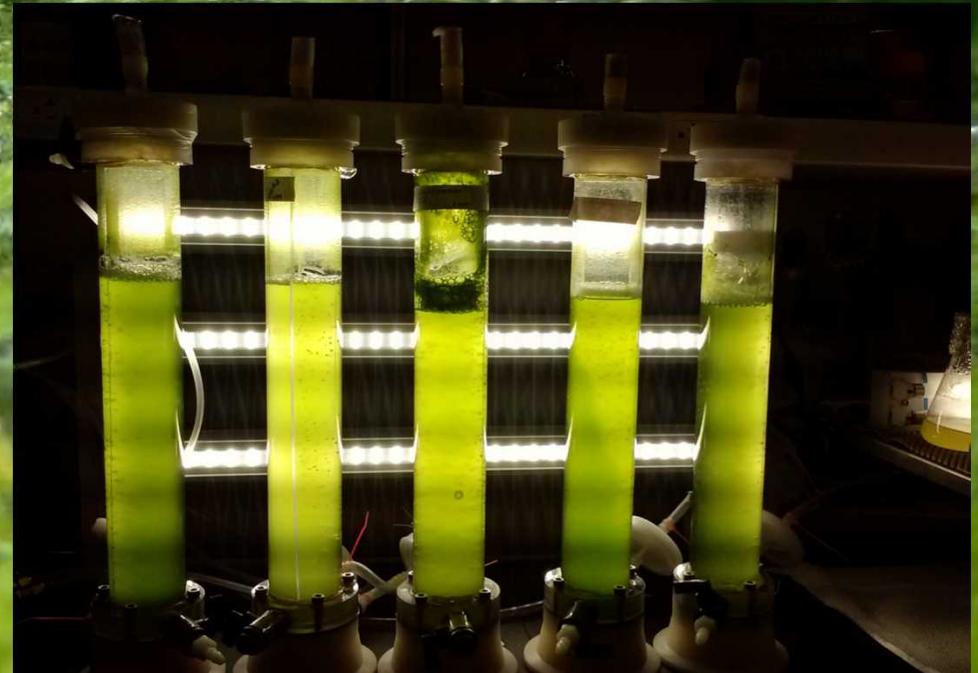
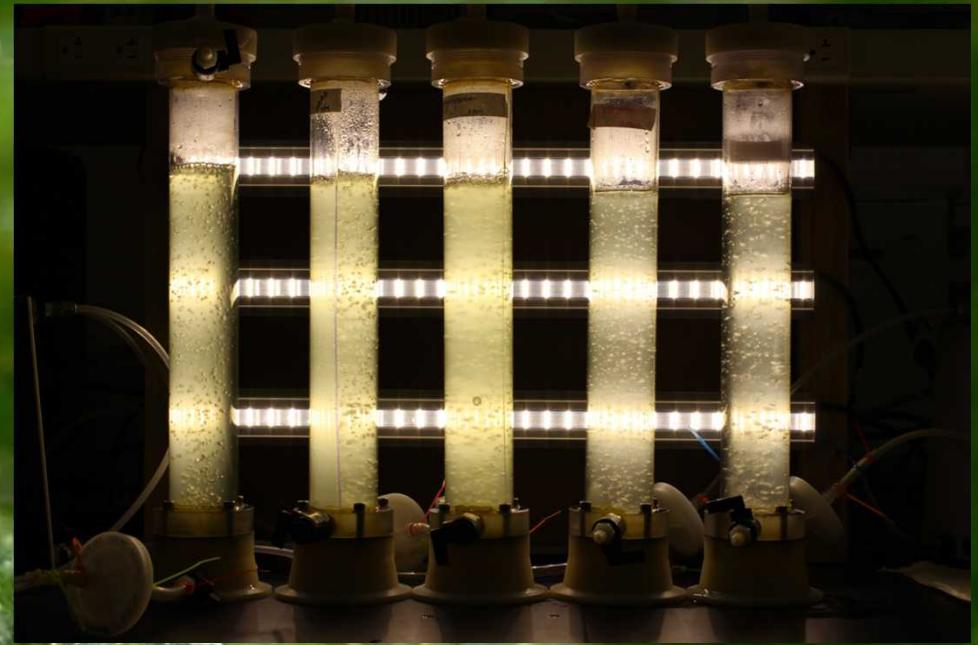
Chlorophyll assay

using Centrifuge to separate cells from media then freeze to shock chlorophyll out from cells and vortexing (spinning) methanol to extract chlorophyll from cells. Again later centrifuge down to remove cells body from chlorophyll and methanol. use methanol as zero, use spectrometer at 750, 665.2, and 650 wavelength





Culturing with Bioreactor



Sample	750	665.2	652
1	-0.1381	0.4889	0.1845
2	-0.1649	0.4489	0.1620
3	0.0311	0.6747	0.3727

0.006708552 mA/mL

Side	A	B	C	D	E
1	28	30	23	25	36
2	25	28	32	30	22

Ecapsulation





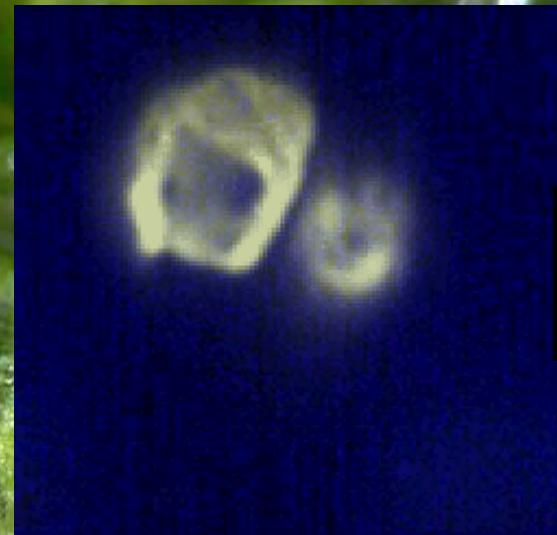
High Light
Thick



High Light
Thin

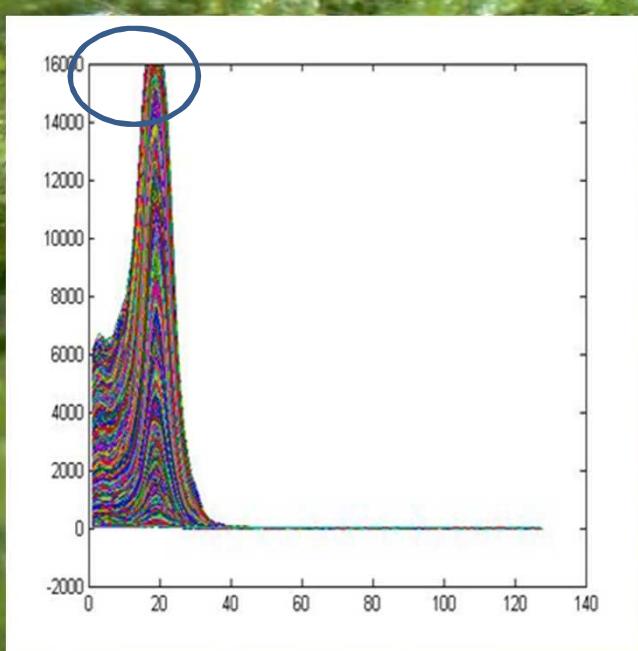
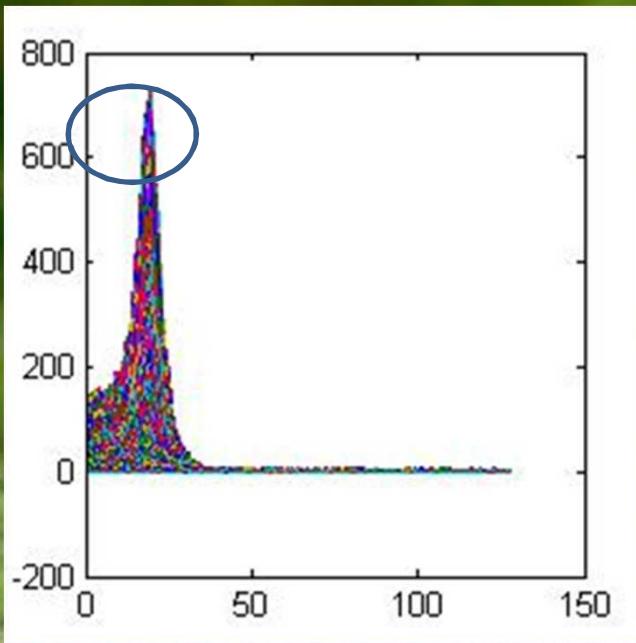


Low Light
Thick



Low Light
Thin

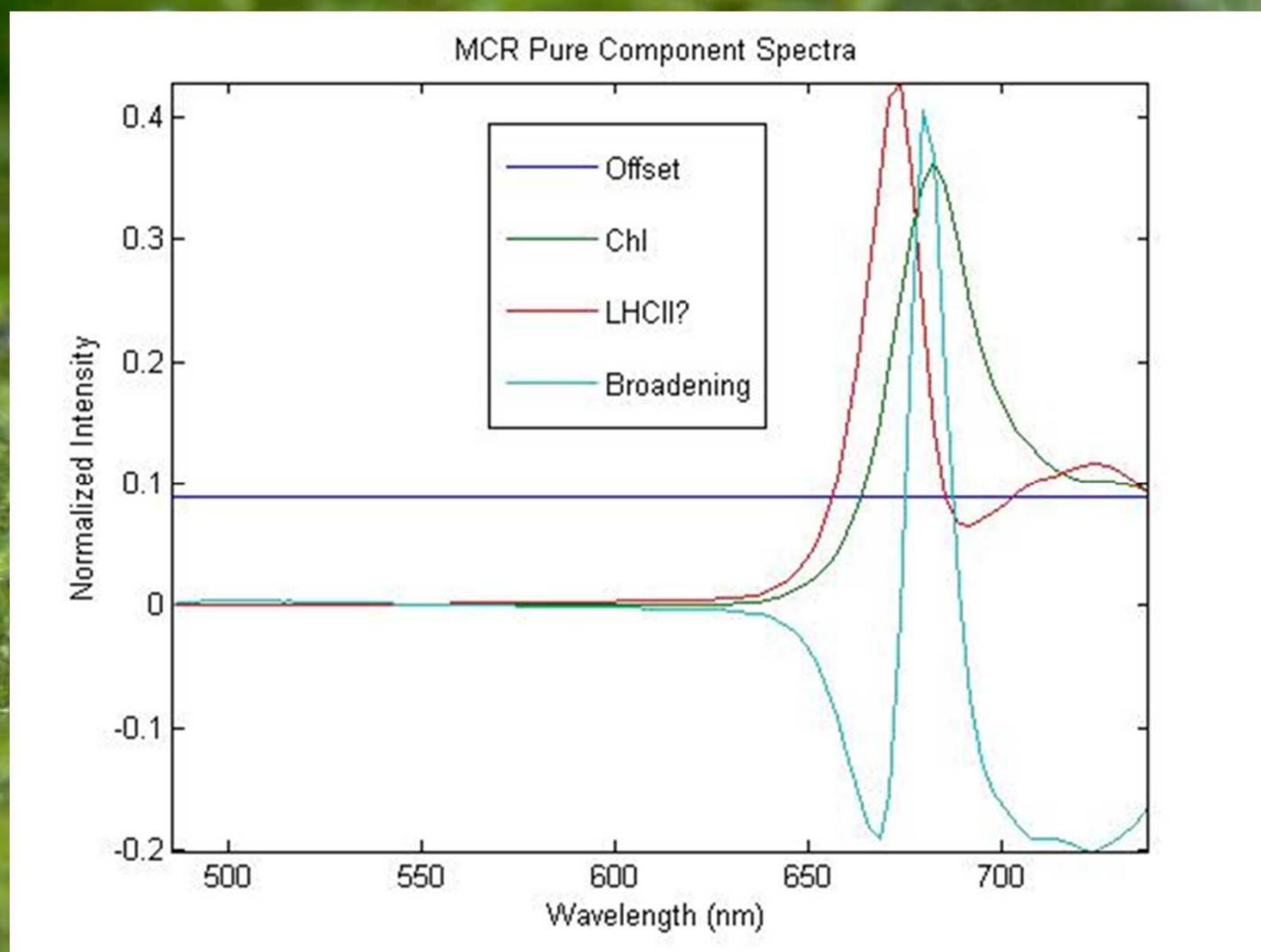


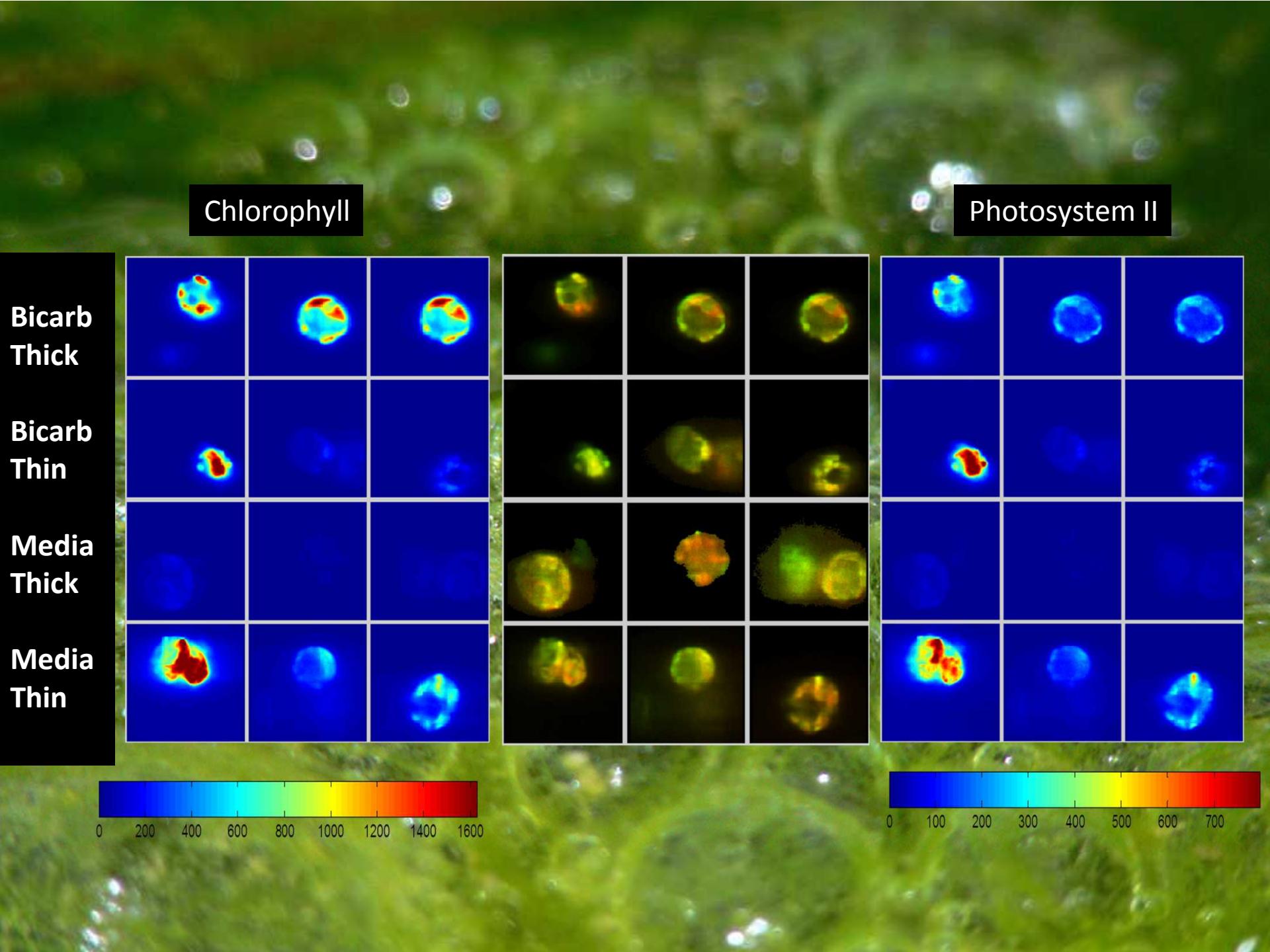


- The baseline does not appear to be a linear offset as we are used to. Instead it seems to slope upward toward the red wavelengths. Note: the data is often quite a bit below zero.
- This could be adequately accounted for if we had some dark current images of the same size, but these were not present.
- Instead a linear offset was modelled and allowed to be negative to improve the results.

- Many of the files exhibited the classic "rolled-over peak" indicative of detector nonlinearity. This was verified by opening raw *.hs1 files in hyperview (keith's software).
- This leads to strong broadening effect on data, and in most files is too severe to perform MCR analysis.

- The signal intensity varied greatly from cell to cell even within similar conditions
- Occasionally this was due to the cell being dead
- Sometimes it looked like cell wasn't well focused?
- Was the laser power or integration time adjusted from image to image? If so this is not recommended.







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QUESTIONS?