

Lipid Bilayers on Silica Beads

A Study of the Fluidity and Diffusion of Lipids on Silica-Aluminum Beads using
MAS and HRMAS NMR



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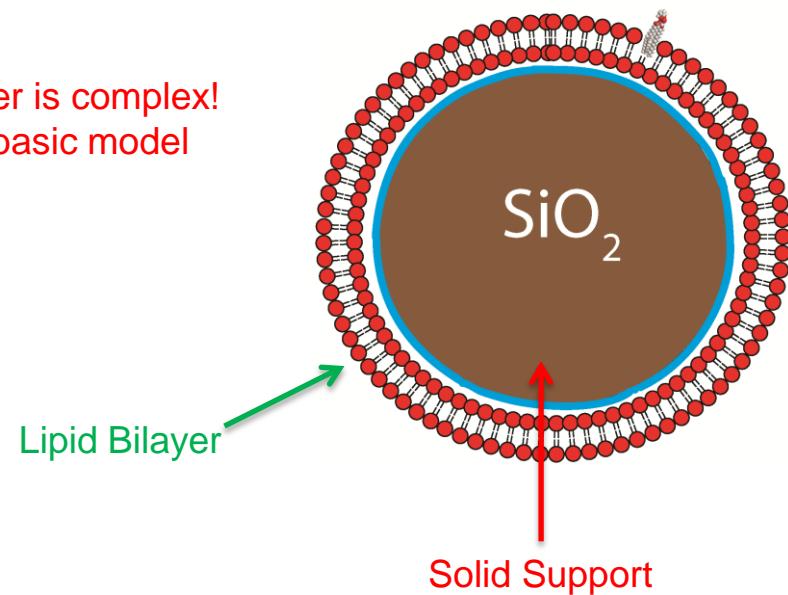
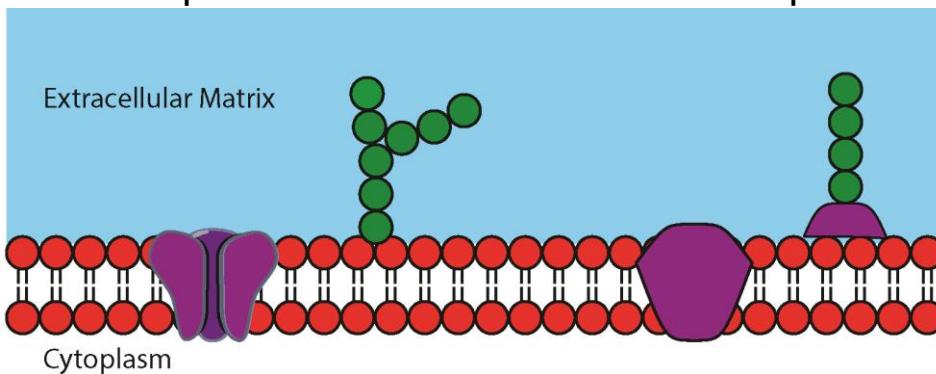
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Importance of Lipid Bilayers

Basic unit of the cell membrane

- Raft formation
- Ion and molecule transportation
- Incorporation of membrane proteins
- Participation in intra- and extracellular processes

Lipid bilayer is complex!
Start with basic model



Biotechnological Applications¹:

- The design of phantom cells
 - Study the role of cell adhesion in immunological recognition processes
 - Study the interplay of specific key-lock forces between receptors and universal forces (dynamic repulsion forces)
- Design of smart biosensors
 - Bilayer used as a thin electrical insulator
 - Bilayer used as a matrix for the incorporation of receptors (i.e. lipid-coupled antigens or antibodies)
 - Electrical monitoring of ligand binding
 - Gold-covered or $\text{SnO}_4/\text{InO}_4$ - covered supports or metal-oxide-semiconductor field-effect transistors (MOSFETs)

Supported Bilayers on Solid Substrates

Solid substrates allow lipid bilayer investigation that is difficult in bulk solution

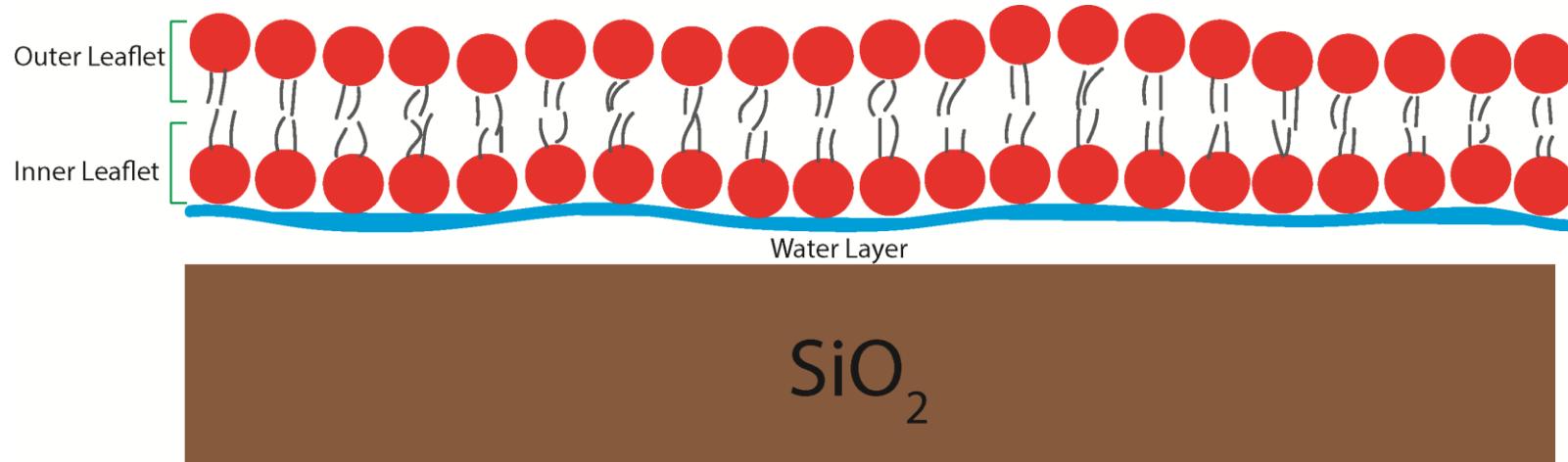
- Biological functionality must be preserved (i.e. lateral fluidity)
- Silica (SiO_2) provides the necessary balance of adhesion, repulsion, and hydration forces for fluidity

Lipid Investigation:

- Membrane fluidity
- Membrane structure disorder
- Diffusion constants of lipid bilayer
- Diffusion constant of water layer
- Physical properties of bilayer and solid substrate

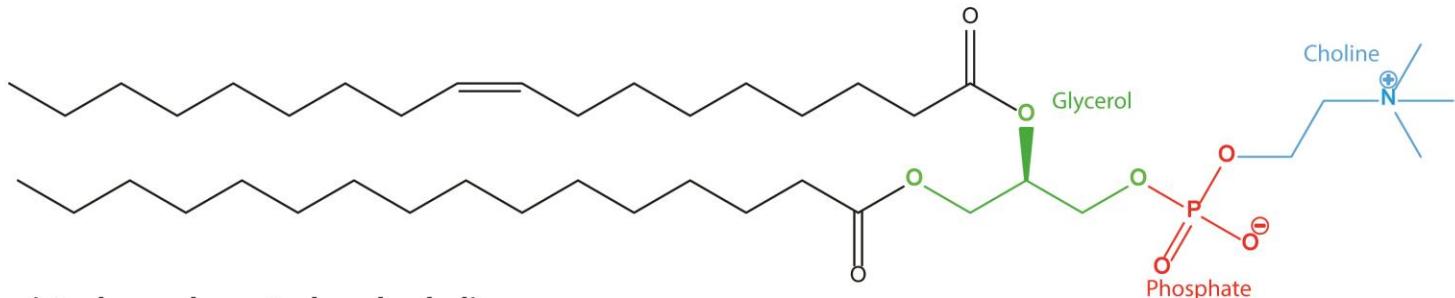
Solid Supports:

- SiO_2 with varying aluminum concentrations
- Porous vs. nonporous
- Spherical vs. non-spherical

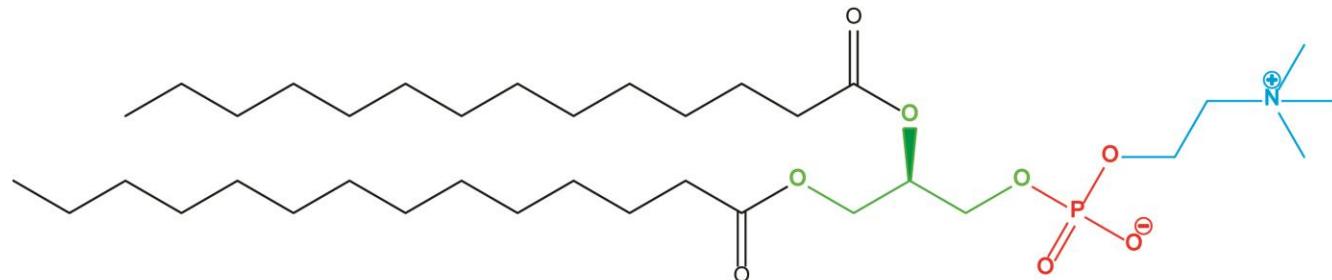


Investigated Lipids

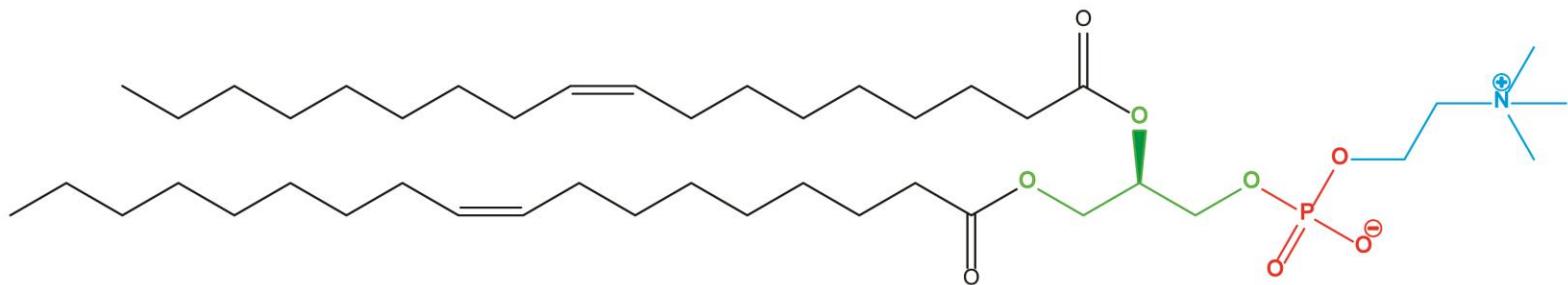
POPC = 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine



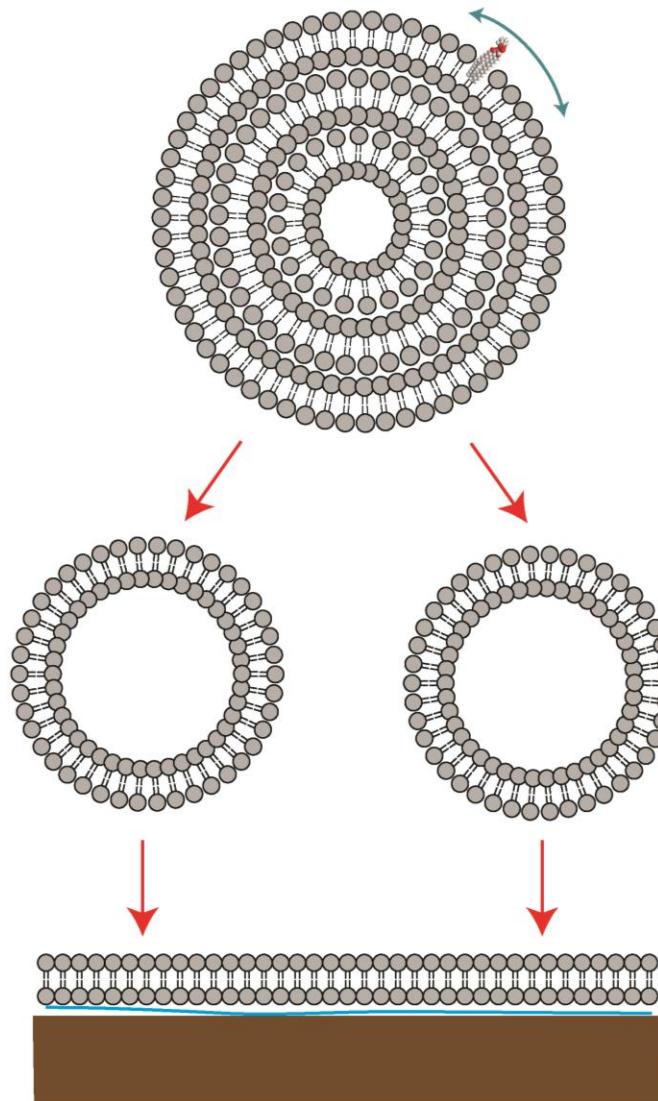
DMPC = 1,2-dimyristoyl-sn-glycero-3-phosphocholine



DOPC = 1,2-dioleoyl-sn-glycero-3-phosphatidylcholine



MLV and SLB Preparation



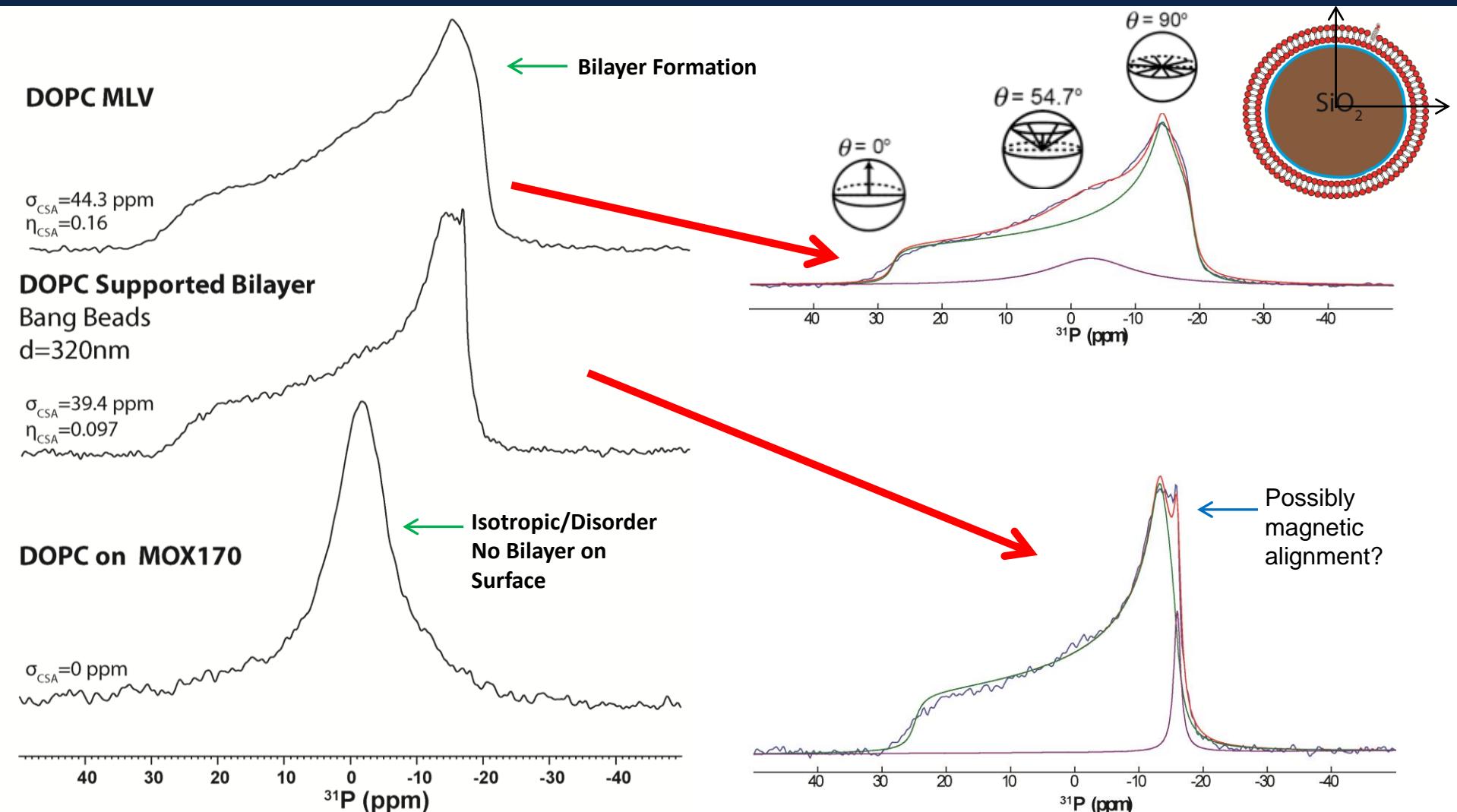
Multilamellar Vesicle (MLV):

- Lipid film formed from rotary evaporation with chloroform
- Dispersed in Nanopure DI water
- Vortexed, freeze-thaw cycles to form MLV dispersion

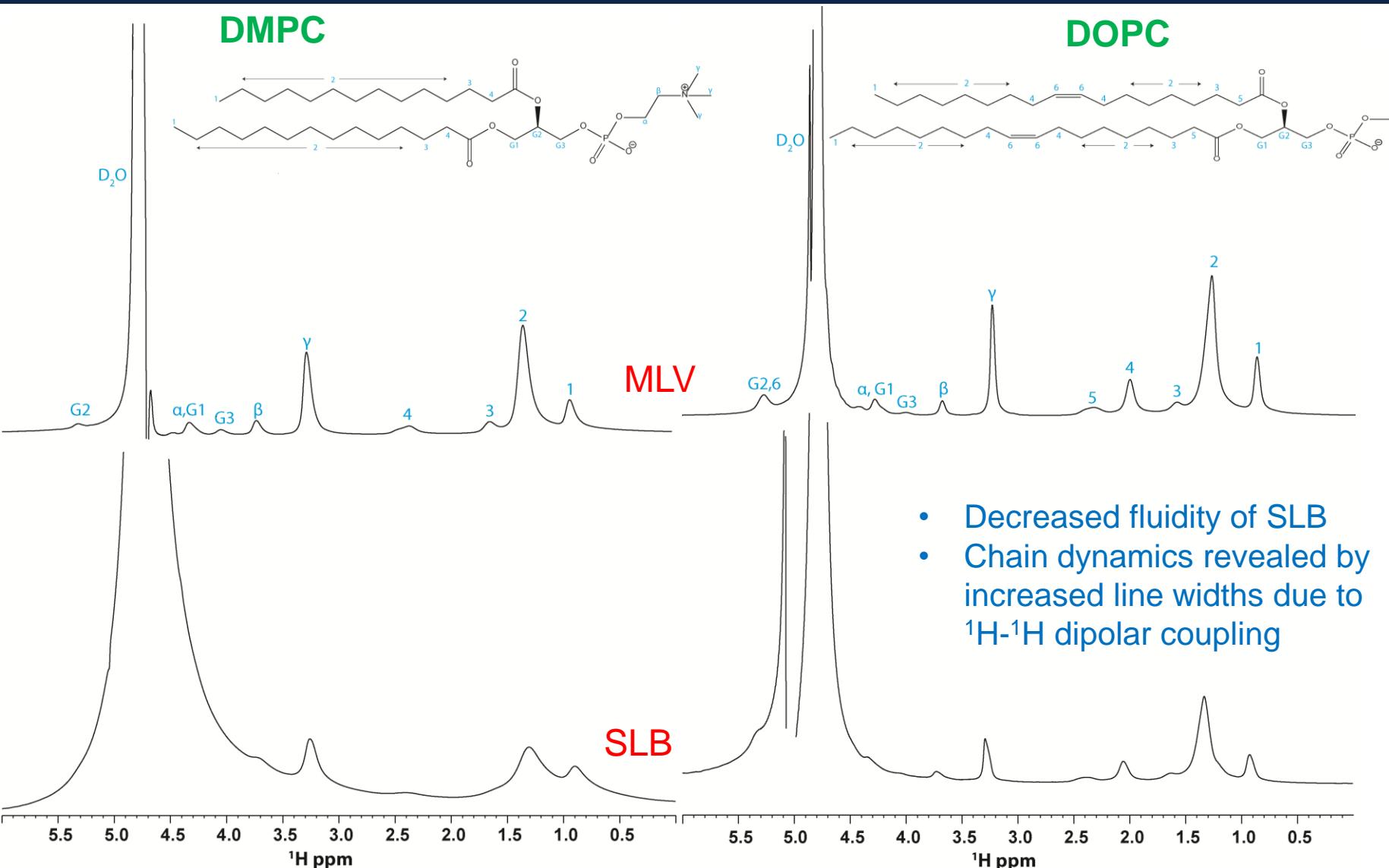
Supported Lipid Bilayer (SLB):

- Rod sonicated MLVs
- Mixed with lyophilized Bang beads ($d=320\text{nm}$, Bangs Laboratories, Inc.)
- Sample placed in water bath overnight and vortexed every hour
- Multiple washes and centrifugations to isolate SLBs

Bilayer Formation Confirmed using Static ^{31}P NMR



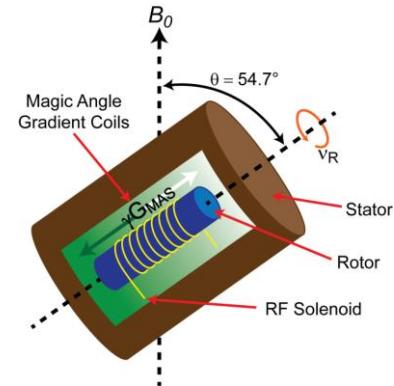
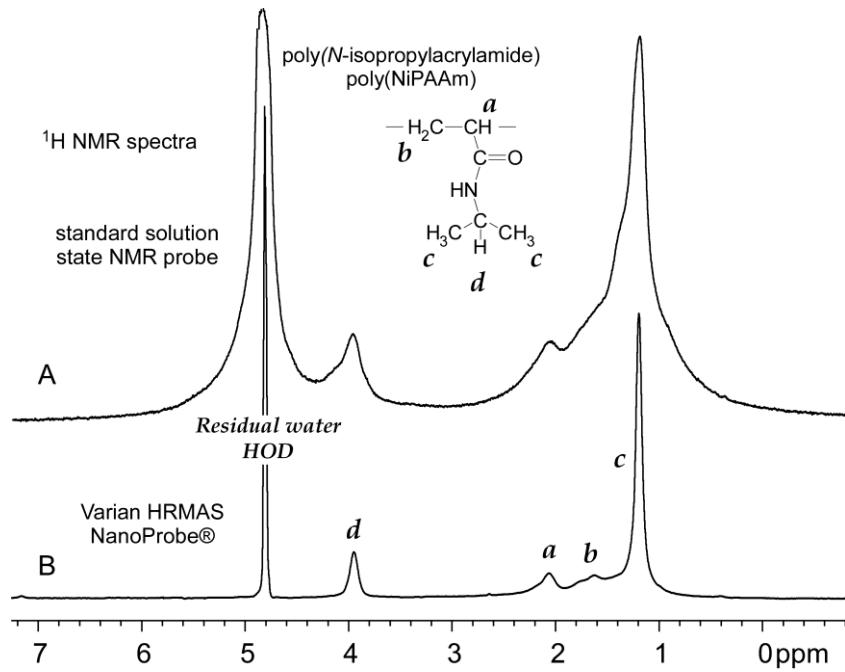
Investigation of MLV and SLB using ^1H NMR



HRMAS PFG NMR

High resolution magic angle spinning (HRMAS) pulsed field gradient (PFG) nuclear magnetic resonance (NMR)

- For liquid/solid samples (lipids) HRMAS produces improved resolution and sensitivity
- Resolution is improved because the Hamiltonians describing the dipolar, CSA and magnetic susceptibility interactions contain an orientational component ($3\cos^2\theta - 1$)
 - θ is the angle between the rotor spinning axis and the magnetic field
 - At the “magic angle” these interactions vanish



Inhomogeneity Dipolar Equations

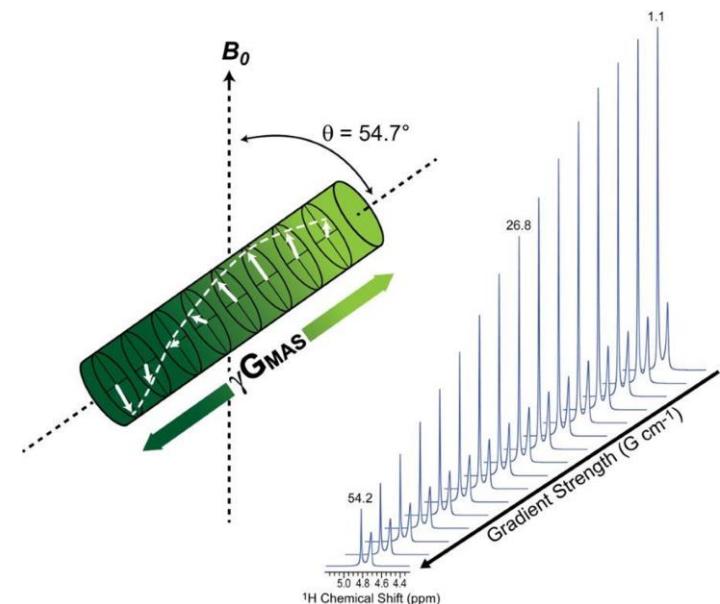
$$B(r_i, \theta_i, \phi_i) = \sum_i \frac{M_j}{r_{ij}^3} \frac{1}{2} (3\cos^2\theta_{ij} - 1) \quad 0$$

$$\begin{aligned}
 B(r_i, \theta_i, \phi_i, t) &= \sum_i \frac{M_j}{r_{ij}^3} \left[\frac{1}{4} (3\cos^2\beta - 1) (3\cos^2\beta'_{ij} - 1) \right. \\
 &\quad \left. + \frac{3}{4} \sin 2\beta \sin 2\beta'_{ij} \cos(\omega_R t + \phi_{ij}) \right. \\
 &\quad \left. + \frac{3}{4} \sin^2 \beta \sin^2 \beta'_{ij} \cos(2\omega_R t + 2\phi_{ij}) \right]
 \end{aligned}$$

$$(3\cos^2\beta - 1) = 0 \text{ when } \beta = 54.7^\circ$$

HRMAS PFG NMR

With the gradient coil along the magic angle, PFG experiments can be performed under MAS conditions



Todd M. Alam, Janelle E Jenkins, HR-MAS NMR Spectroscopy in Material Science

During PFG diffusion experiments, the application of a gradient “tags” a spin with a phase that is related to its spatial position

If the spin diffuses (changes spatial position) during Δ

- The dephasing is not refocused
- Signal intensity decreases

The loss in signal intensity with increasing gradient strength is related to the self-diffusion rate in the supported bilayer equation

$$\ln\left(\frac{I}{I_0}\right) = -\frac{2}{3}kD + \frac{2}{45}(kD)^2$$
$$k = 4\gamma^2 g^2 \delta^2 \left(\Delta - \frac{\delta}{3}\right)$$

I=integrated signal intensities

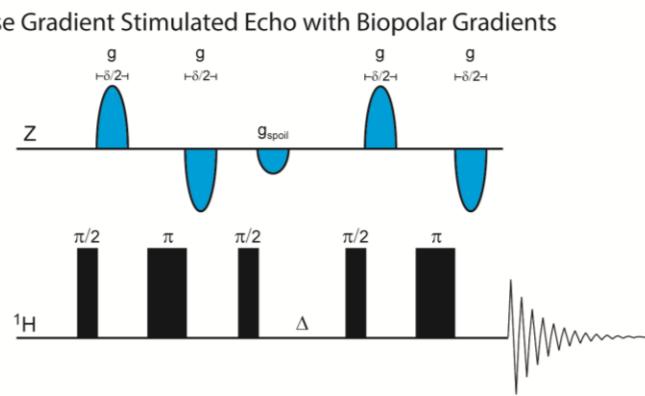
γ =gyromagnetic ratio

g=gradient strength

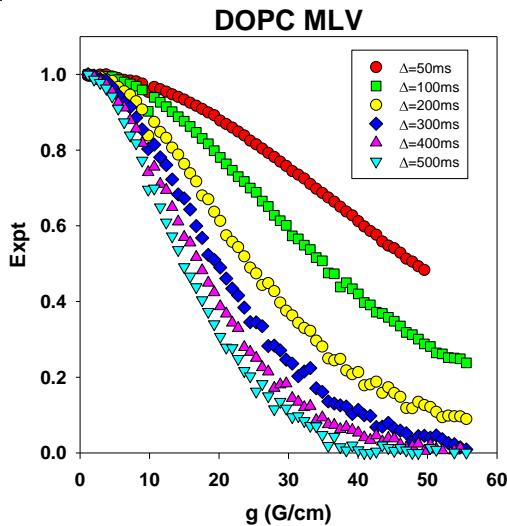
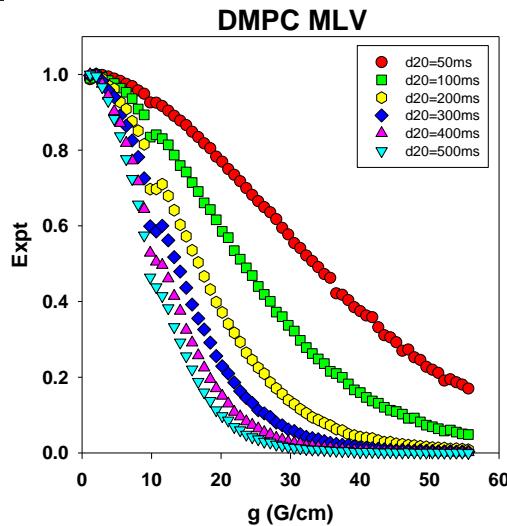
δ =gradient pulse length

Δ =diffusion time between pulses

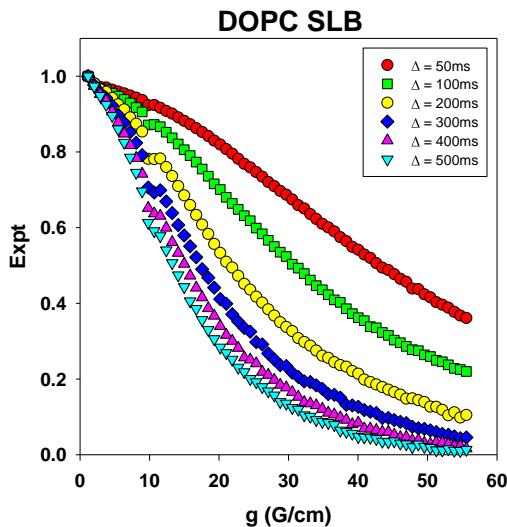
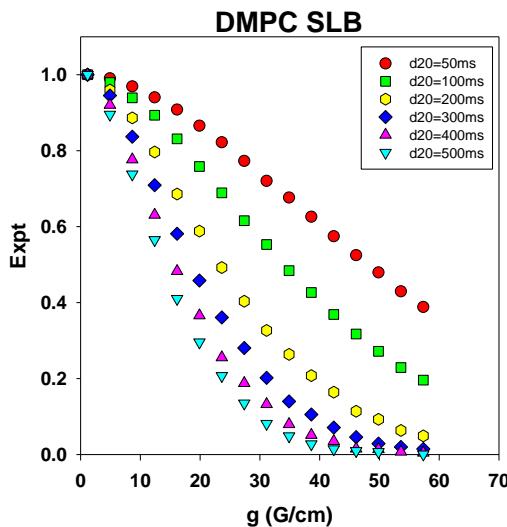
D=translational self-diffusion coefficient



PFG NMR Diffusion



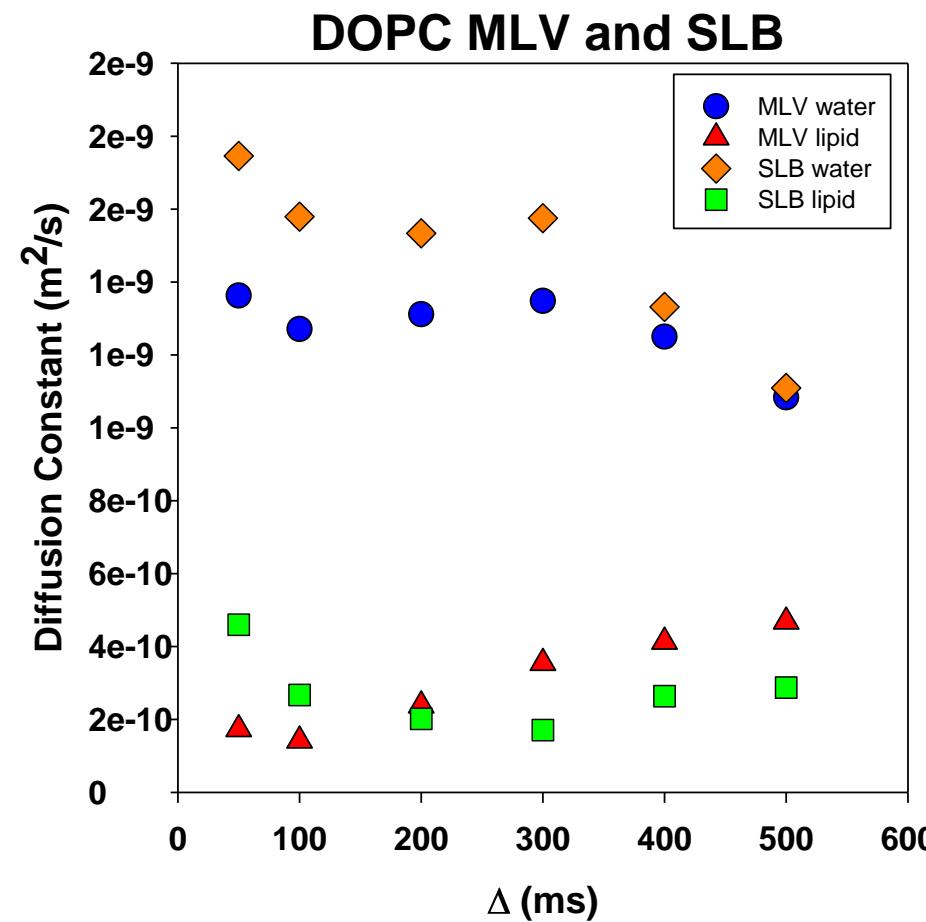
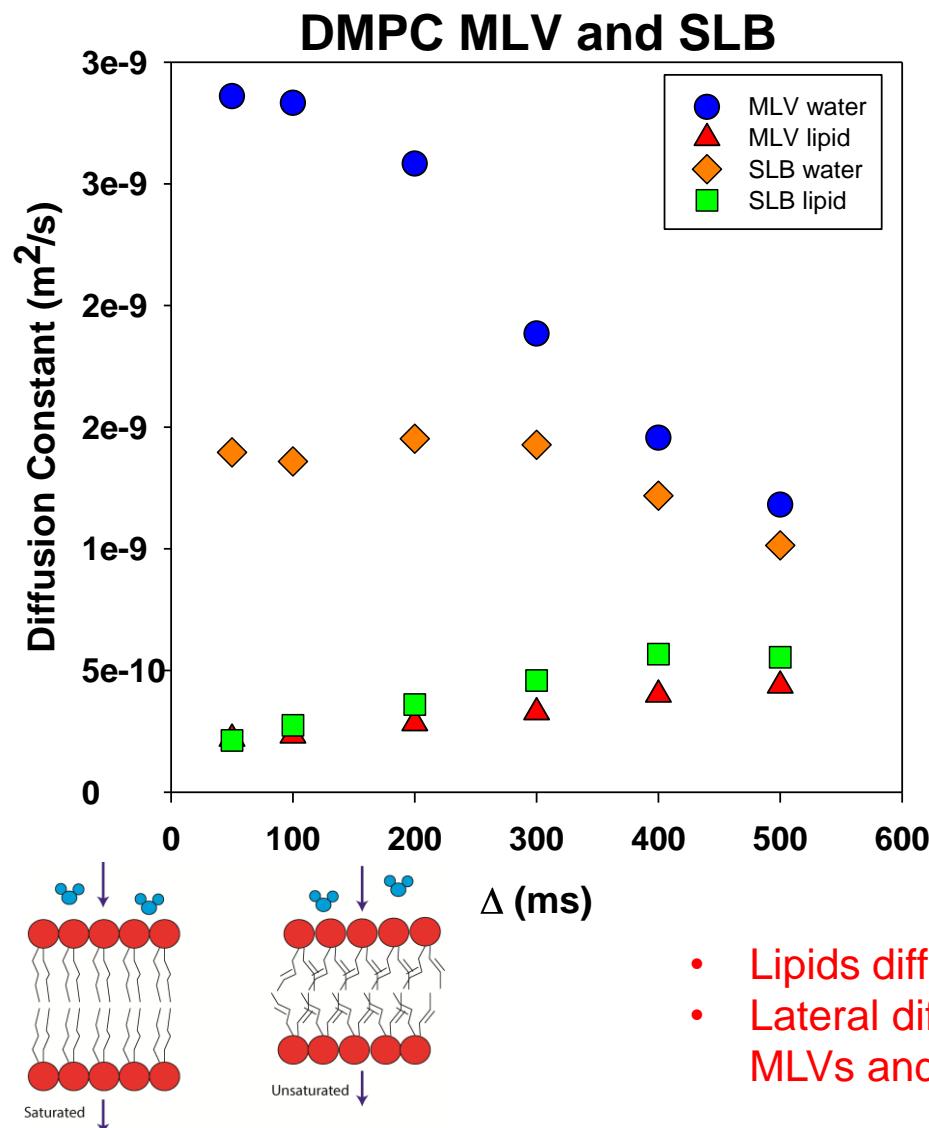
- Decay of signal intensity is a measure of diffusion rate



- Diffusion rates for MLVs are similar to SLBs

- DMPC diffuses faster than DOPC for MLV and SLB

MLV and SLB Lipid Diffusion



- Lipids diffuse an order of magnitude slower than water
- Lateral diffusion of lipids and diffusion of water are similar for MLVs and SLBs

Conclusions

- DMPC and DOPC were successfully deposited on silica beads to form SLBs
- PFG HRMAS NMR was used to determine the diffusion rates for MLVs and SLBs
- Lipids diffused an order of magnitude slower than water as predicted
- Diffusion constants for water and lipid didn't change significantly when the lipids attached to the bead
- First NMR measurement of lipid lateral diffusion on SLBs!!

Future Directions and Acknowledgements

Future Directions

- Attach POPC to Bang beads
- Collect 1D and 2D, including diffusion data, for POPC
- Form SLBs with different types of beads (nonporous vs. porous, spherical vs. globular)
- Investigate diffusion of lipid MLVs and SLBs as a function of temperature
- Study T1 spin-lattice relaxation and T2 spin-spin relaxation for lipid MLVs and SLBs
- Study the properties of the water layer between the silica and the lipid bilayer

Acknowledgements

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