

# Harnessing the power of genomics for detection, diagnosis, and mechanistic analysis of cellular responses

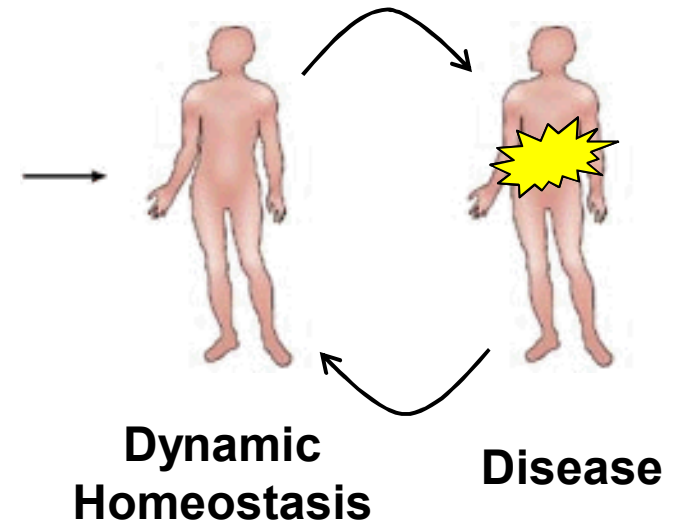
Raga Krishnakumar  
Systems Biology  
September 21, 2016

*Supported in part by the Laboratory Directed Research and Development program at Sandia National Laboratories, a multi-mission laboratory managed and operated by National Technology and Engineering Solutions of Sandia, LLC, a wholly owned subsidiary of Honeywell International, Inc., for the U.S. Department of Energy's National Nuclear Security Administration under contract DE-NA0003525.*



# *Cells constantly respond to their environment, and make decisions that govern development and disease*

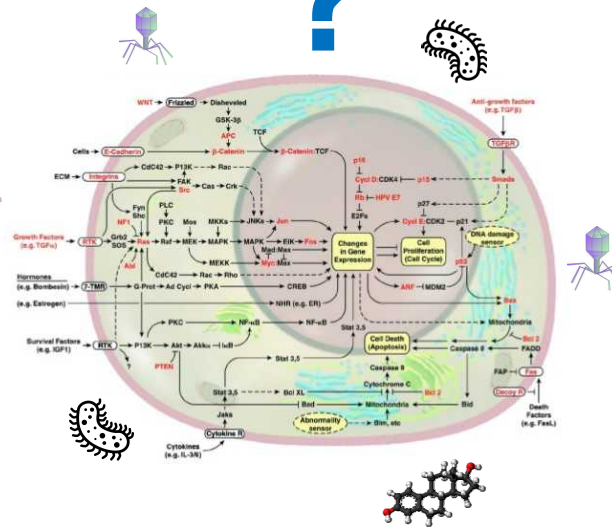
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# Cells constantly respond to their environment, and make decisions that govern development and disease

**Capture  
environmental  
changes and  
cellular responses**

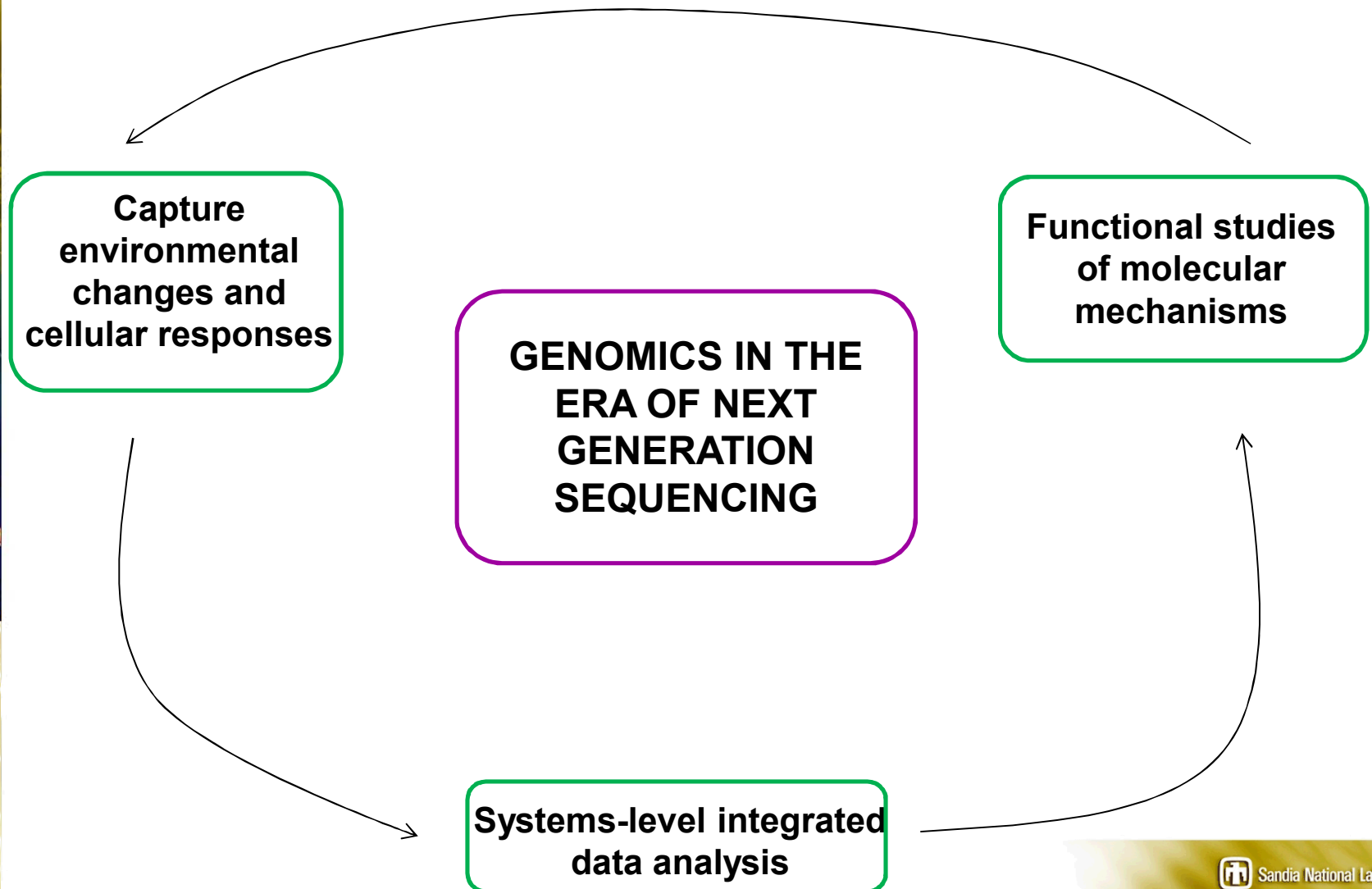
**Functional studies  
of molecular  
mechanisms**



**Systems-level integrated  
data analysis**

# ***Cells constantly respond to their environment, and make decisions that govern development and disease***

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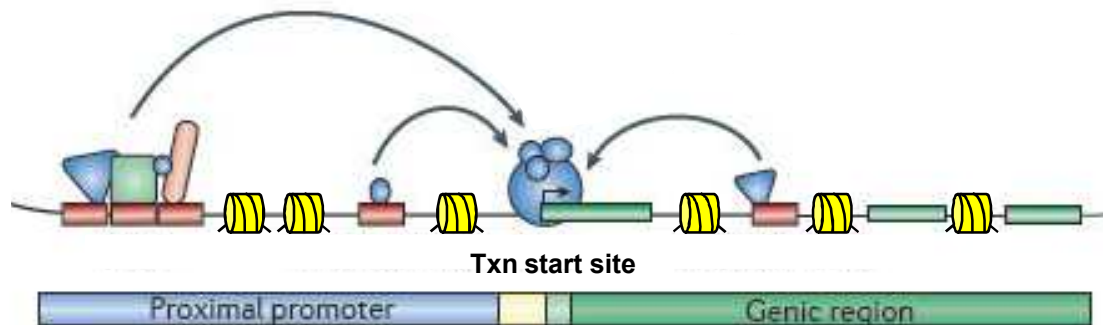
# Overview

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- Combinatorial genomics reveals novel molecular mechanisms during cell fate transitions in early development

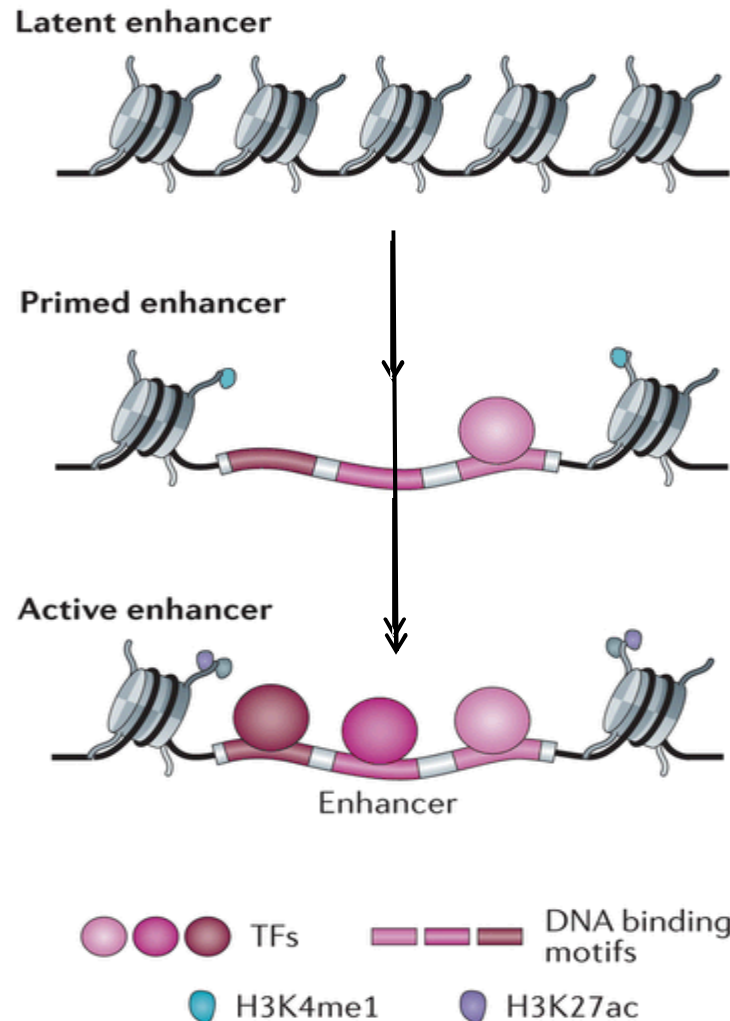
Applying lessons learnt for engineering therapeutic cells for personalized immune therapy

# *Transcriptional regulation in eukaryotes*



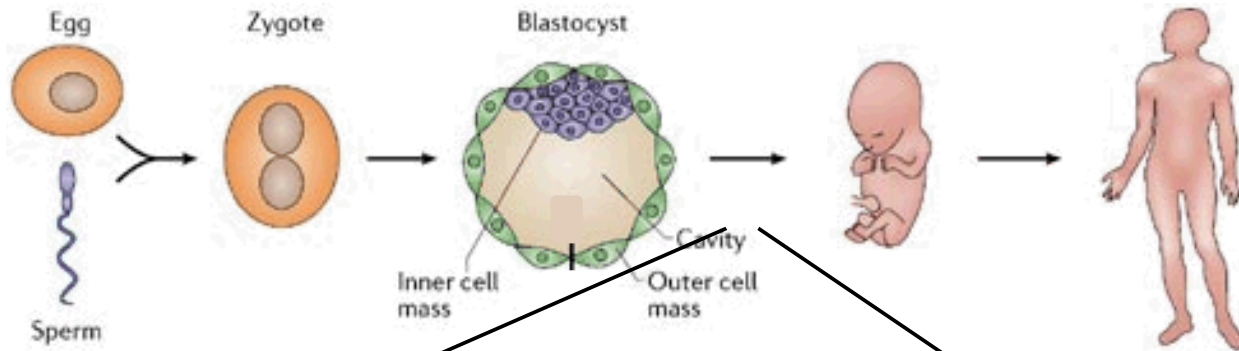
**Chromatin-bound enzymes (what) + Sequence specific TFs (where and when)**

# *Enhancers act as molecular switches for their cognate genes*

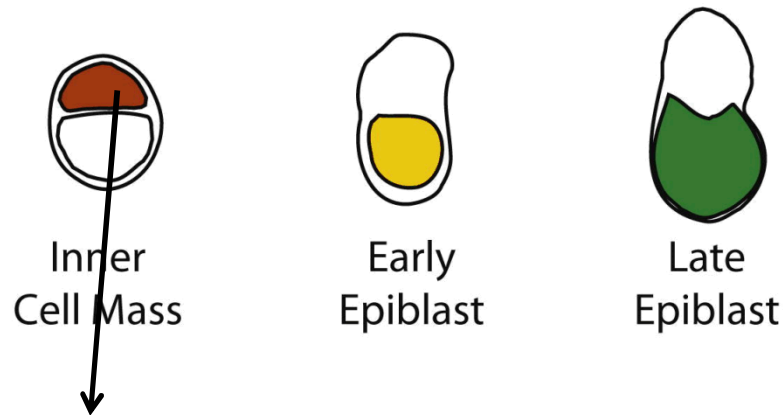


Shlyueva et al 2014 (Nature Reviews Genetics)

# Rapid cell fate choices occur in early development



## Early mouse embryogenesis



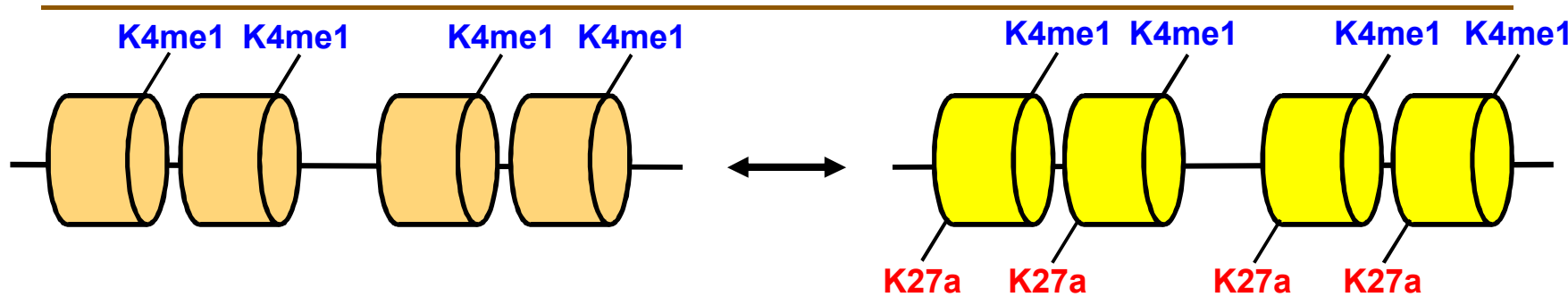
Embryonic stem cells  
(**ESCs - red**) ----> (**EpiCs - yellow**)

# ***Enhancer function in pluripotent cell fate choices***

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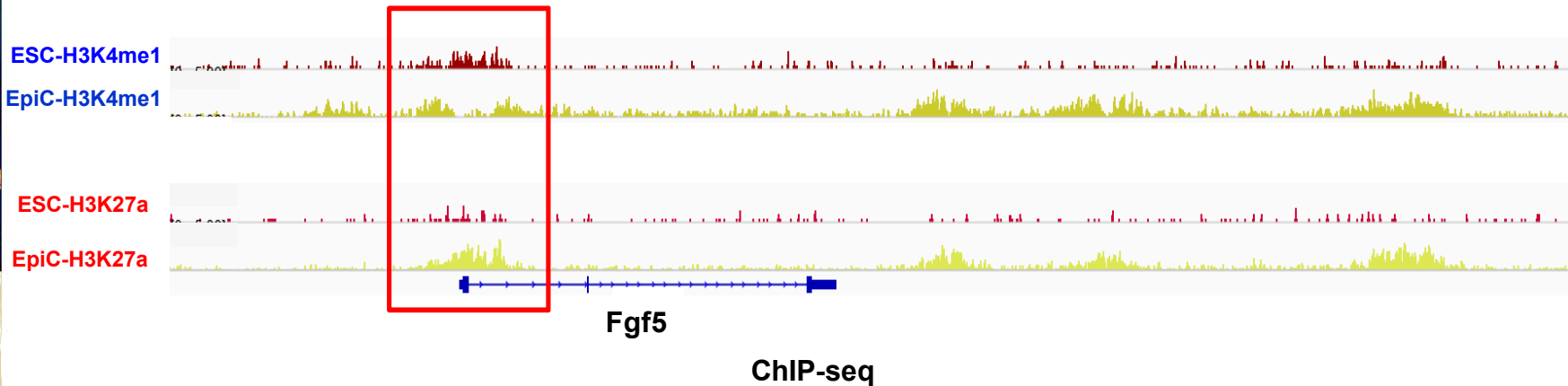
- Is gene expression regulated by enhancer priming in early mouse embryonic development?
- What factors regulate the switch between primed and active enhancers?
- What is the mechanism of this regulation?

# Profiling of enhancer chromatin in pluripotent cells



**'primed' – H3K4me1+**

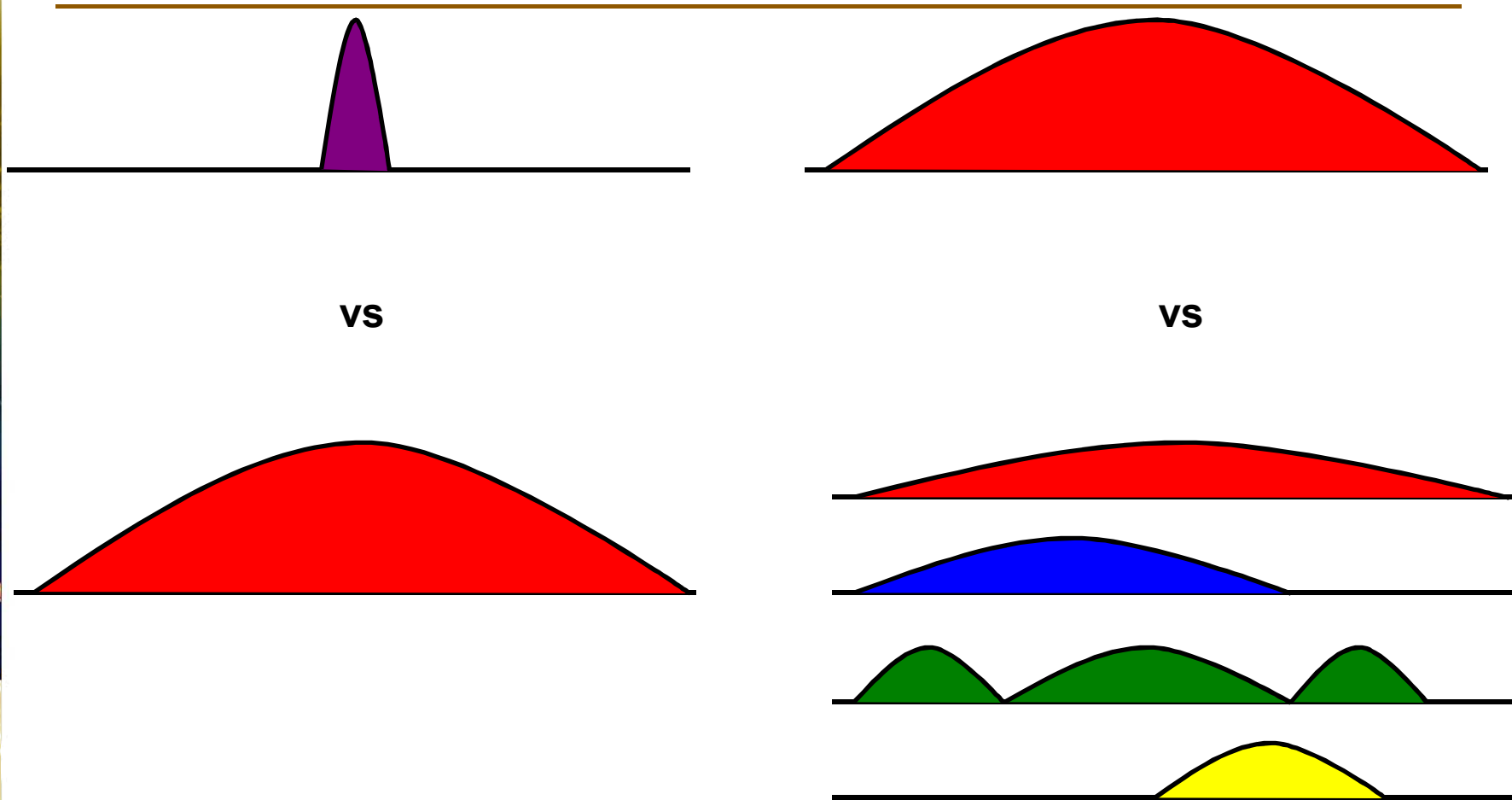
**'active' – H3K4me1+ H3K27ac+**



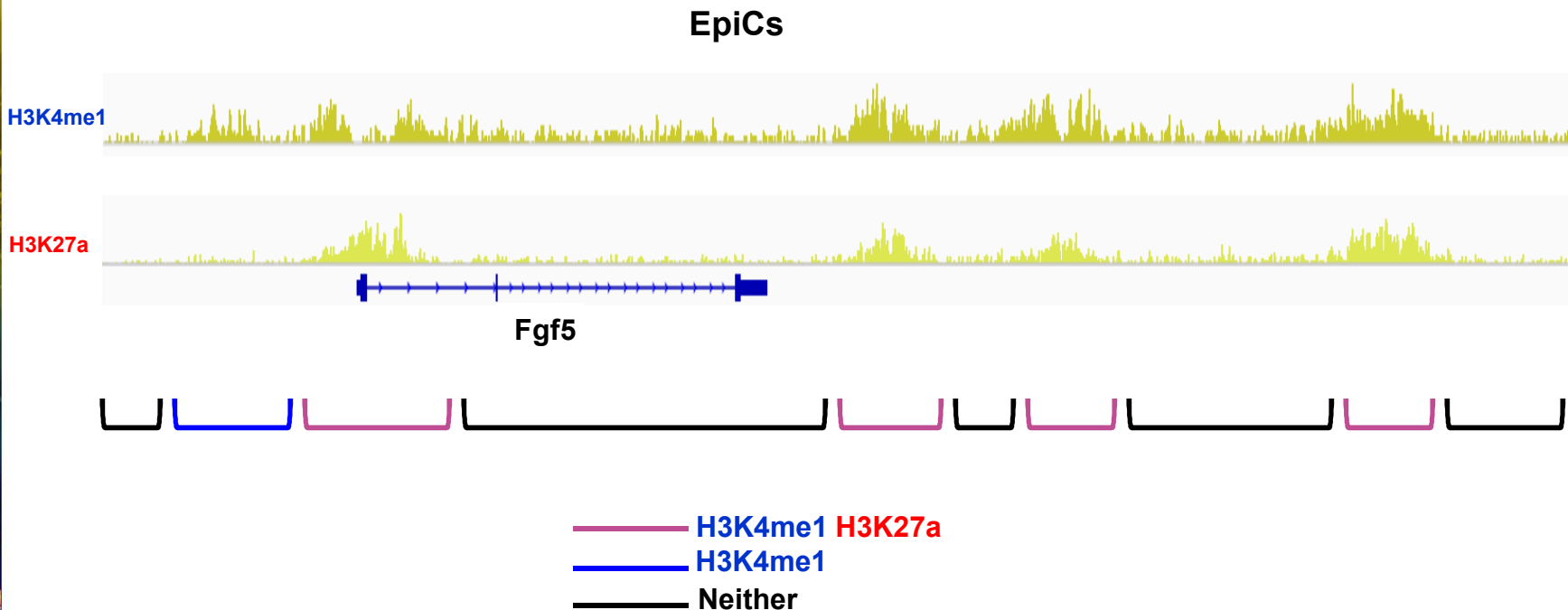
How can we consolidate the information and get something biologically meaningful out of it?



# *Peaks vs domains – understanding the chromatin landscape*

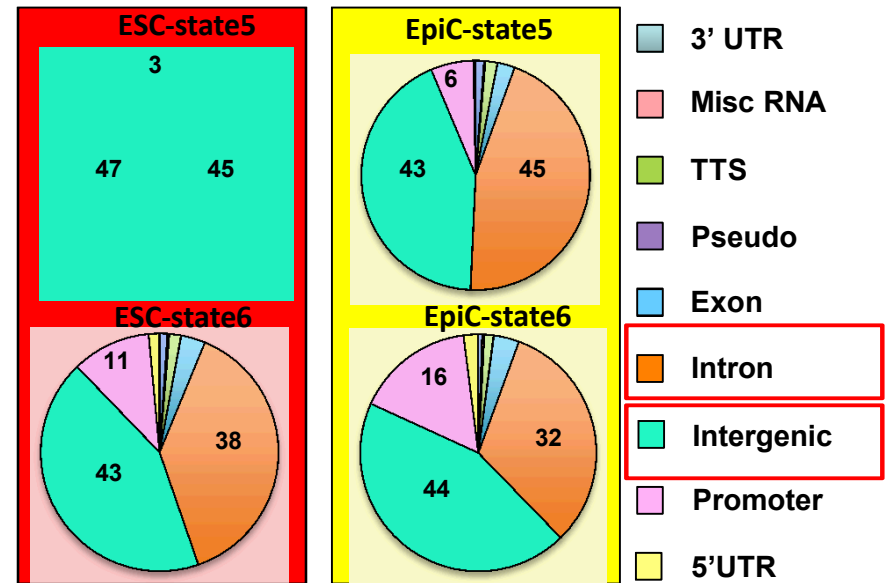
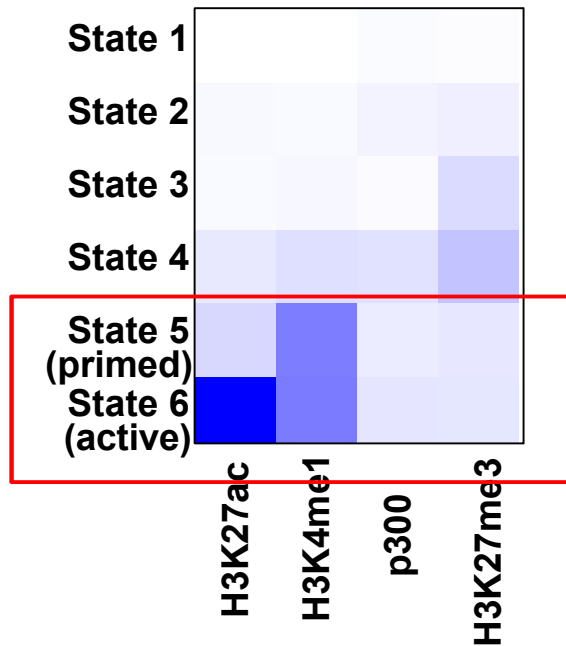


# Profiling of enhancer chromatin in pluripotent cells

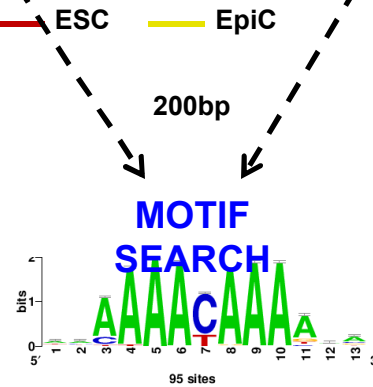
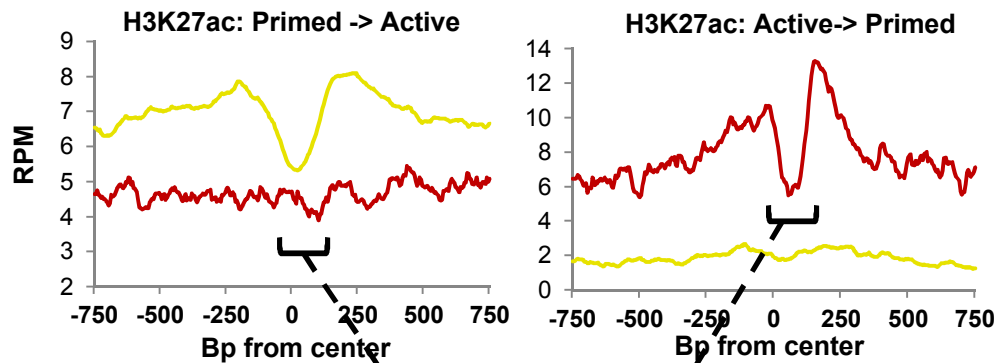
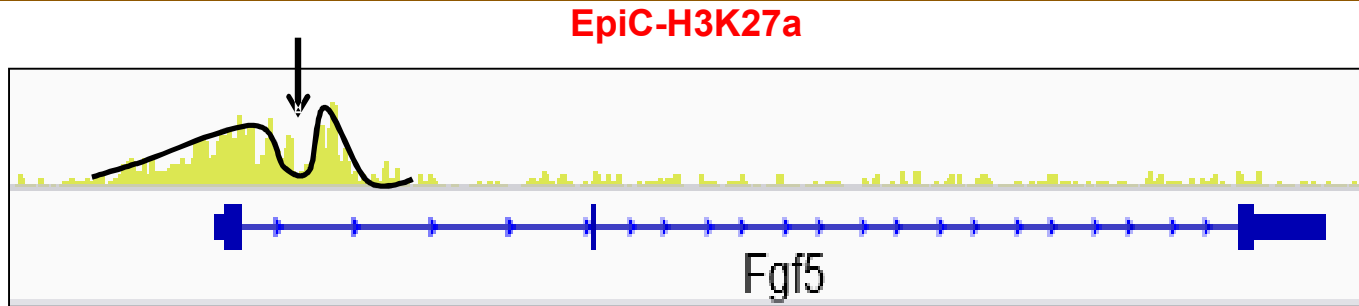


- Unknown combinations of marks
  - Unbiased discovery
- Use HMM to identify states (H3K4me1, H3K27ac, H3K27me3, p300)

# ChromHMM identified dynamic primed and active enhancer chromatin

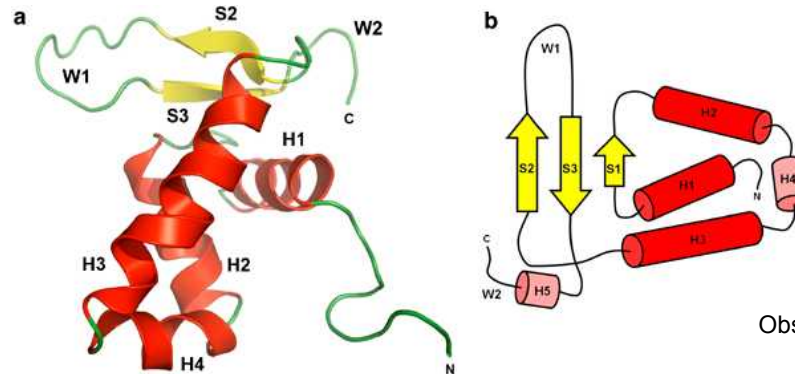


# Enhancers transitioning between primed and active are enriched for the Foxd3 motif



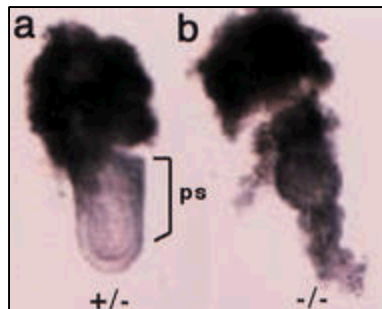
# ***Foxd3 is essential in early embryonic development***

**Forkhead family of transcription factors – involved in many early embryonic fate decisions**



Obsil and Obsilova 2008 (Oncogene)

**Foxd3 KO embryos fail to form a primitive streak and die by e7.5**

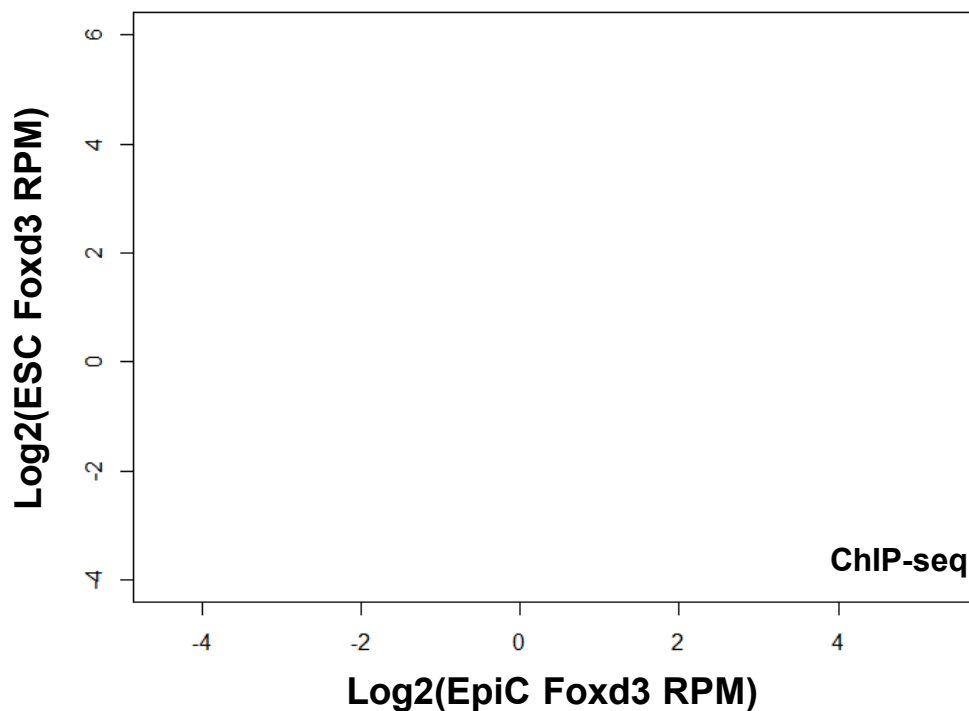


Hanna et al 2002 (Genes and Dev)

# ***Foxd3 re-localizes during the transition from naïve to primed pluripotency***

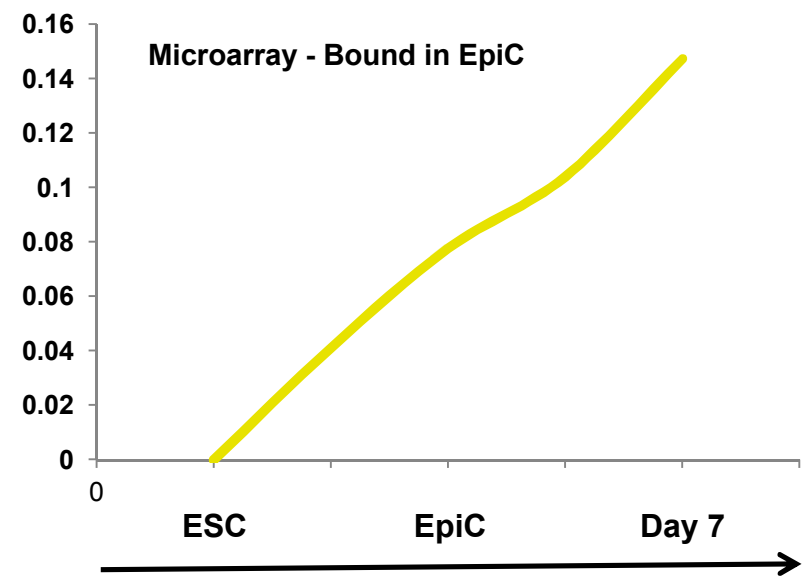
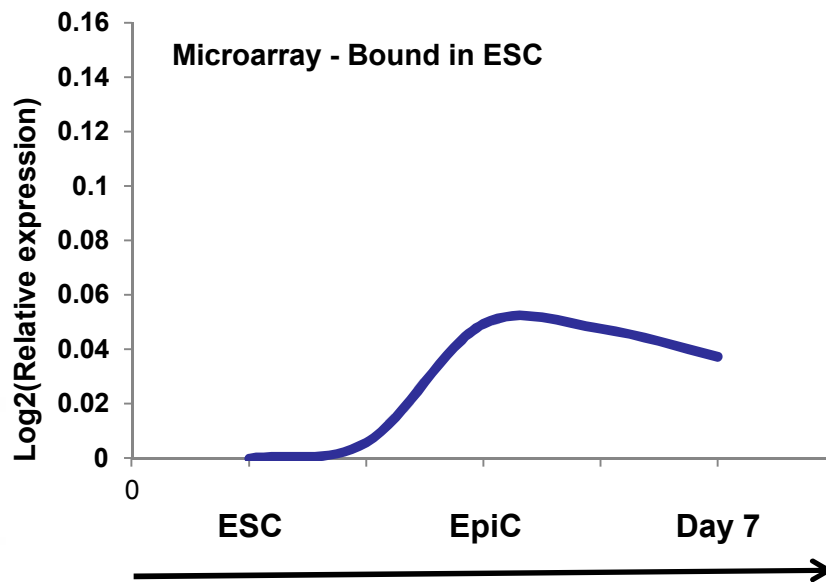
**Foxd3 ChIP-seq peak finding using MACSv1.4**

**In ESCs (y-axis) and EpiCs (x-axis)**



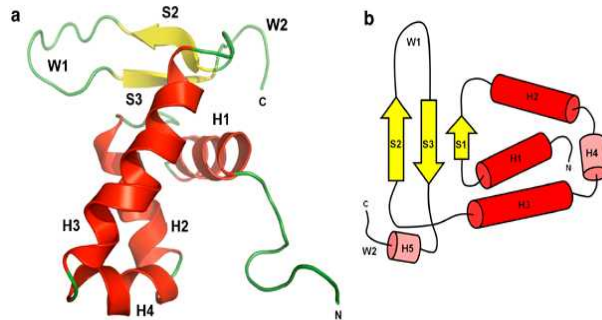


# *Foxd3 primes genes for later expression*



# Does *Foxd3* initiate enhancers or engage pre-existing ones?

**Winged helix domain can access nucleosome-occluded chromatin**



## MNase-ChIP-seq



Crosslink proteins and DNA



Sample fragmentation  
• MNase digestion



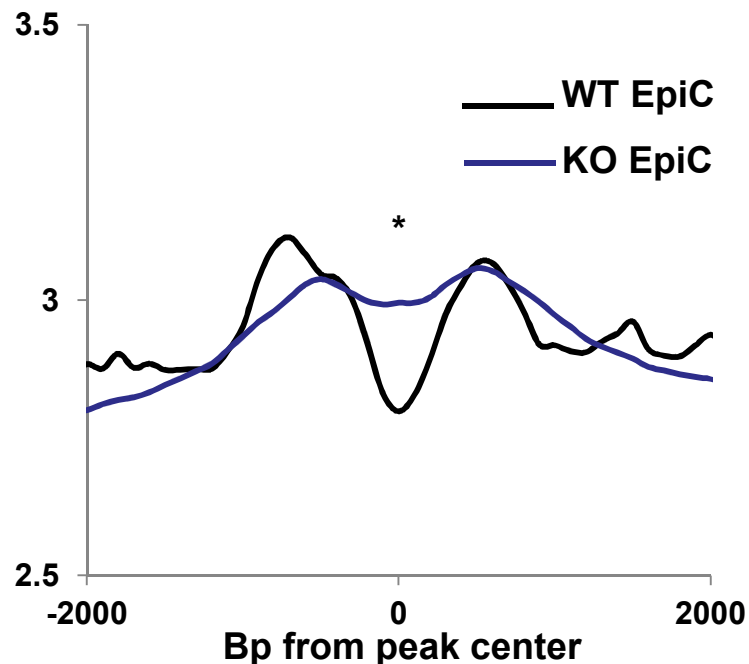
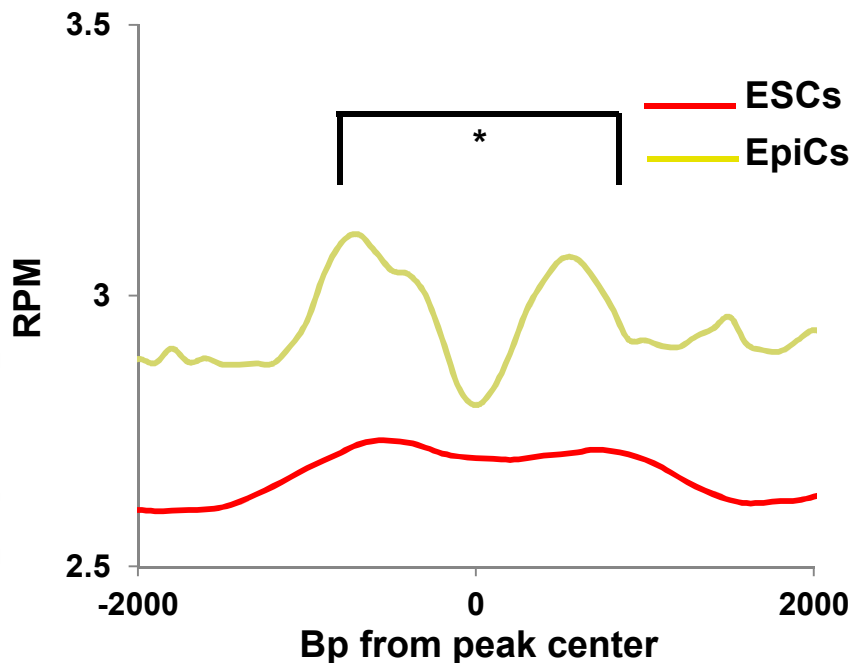
Immunoprecipitate and  
then purify DNA



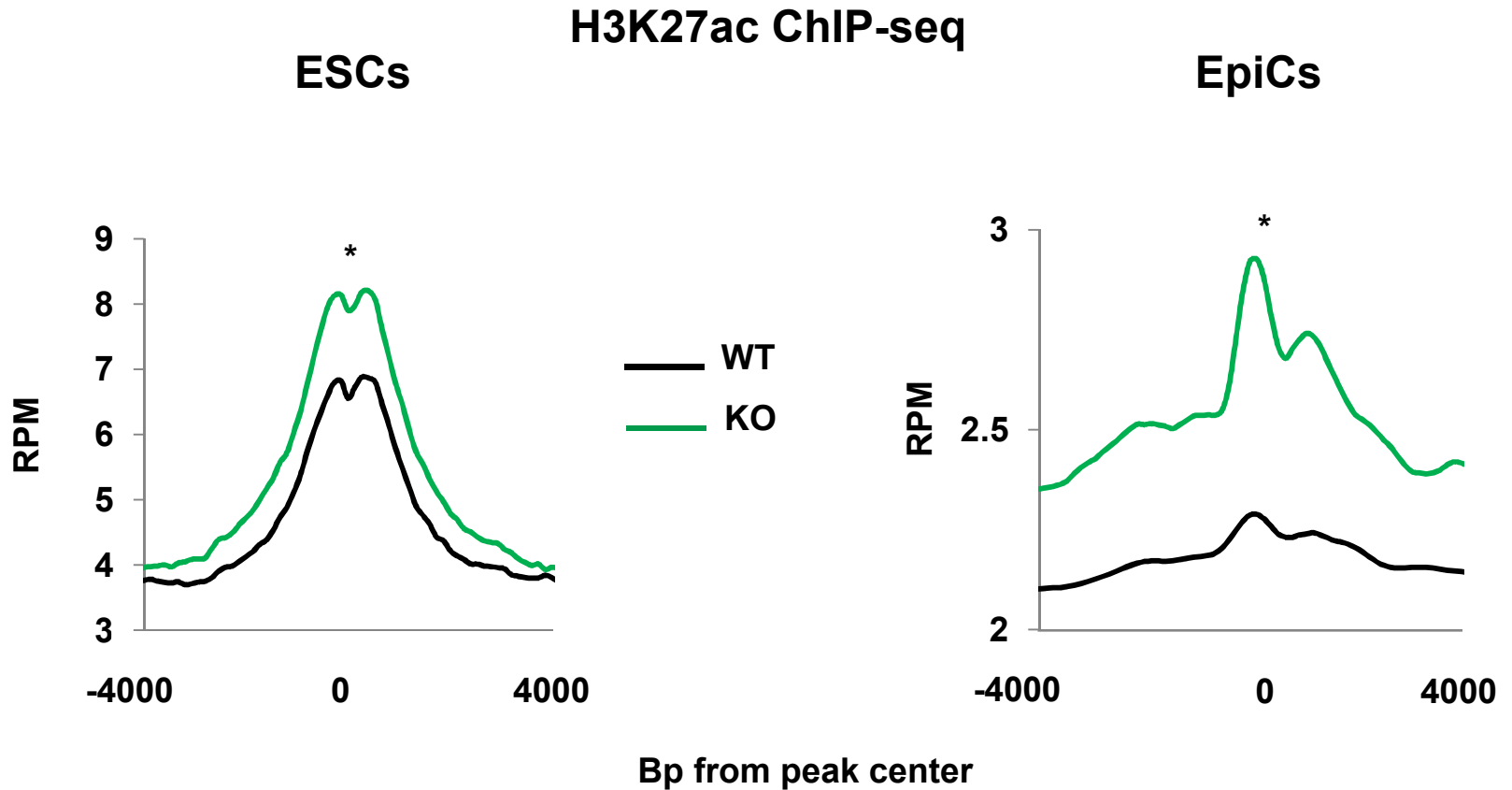
DNA library creation and sequencing

# *Foxd3 establishes nucleosome-free enhancers*

## H3K4me1 MNase-ChIP-seq EpiC sites

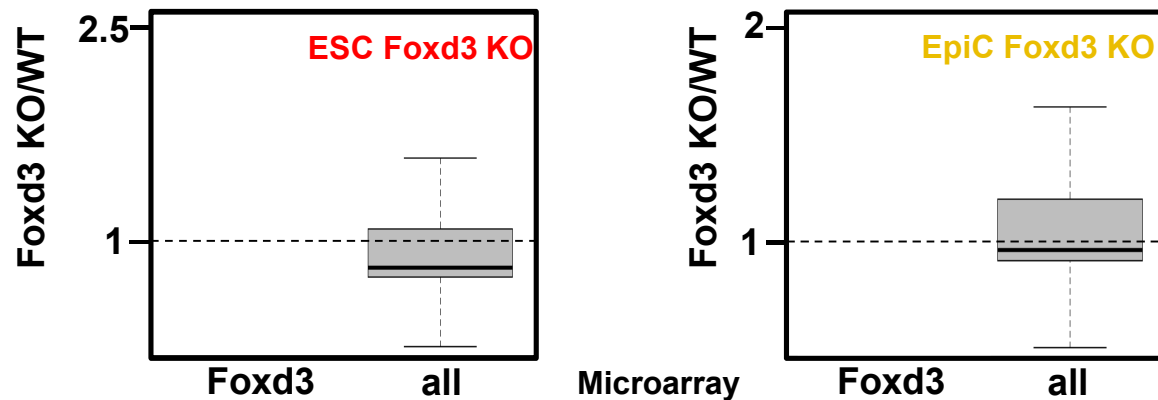


# *Foxd3 suppresses enhancer acetylation*



# ***Foxd3 knockout increases expression of neighboring genes***

## **Genes changing upon Foxd3 KO**



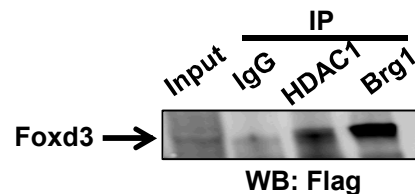
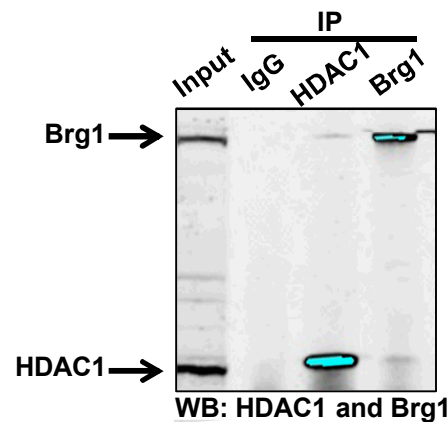
**What is the mechanism of Foxd3 action at enhancers?**

# Foxd3 is in a complex with Brg1 and HDAC1

H3K27 deacetylation – HDAC1/2 (NURD complex)

Chromatin remodeling – Brg1 (SWI/SNF complex)

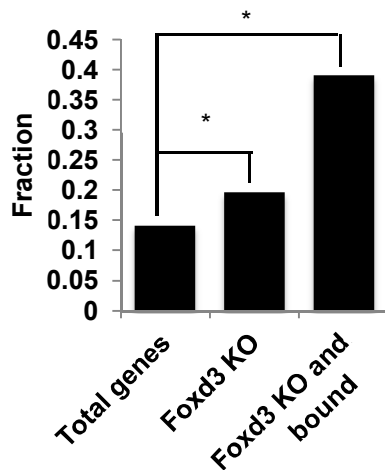
## Immunoprecipitation





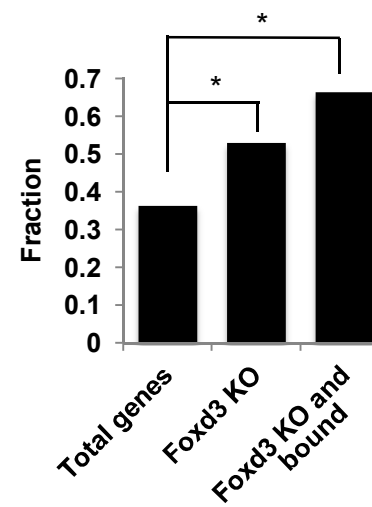
# *Foxd3 recruits BRG1 and HDACs to simultaneously establish and repress enhancers*

HDAC1/2 KO gene expression

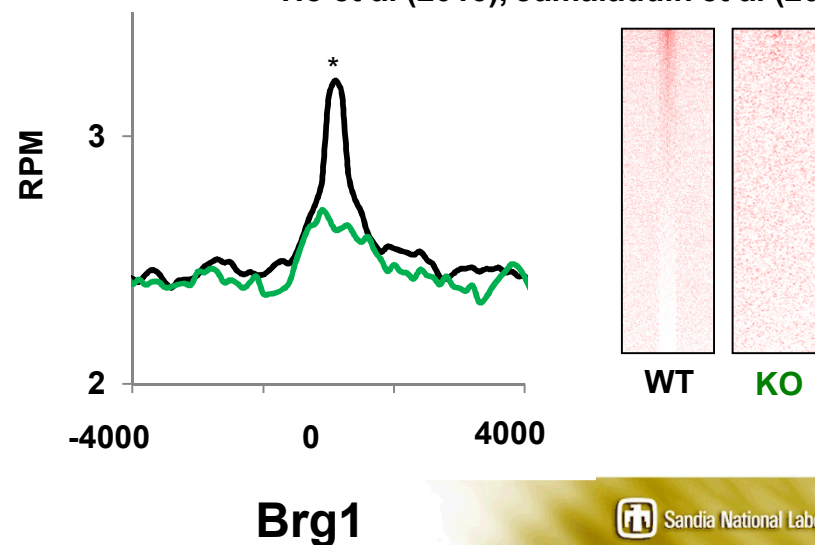
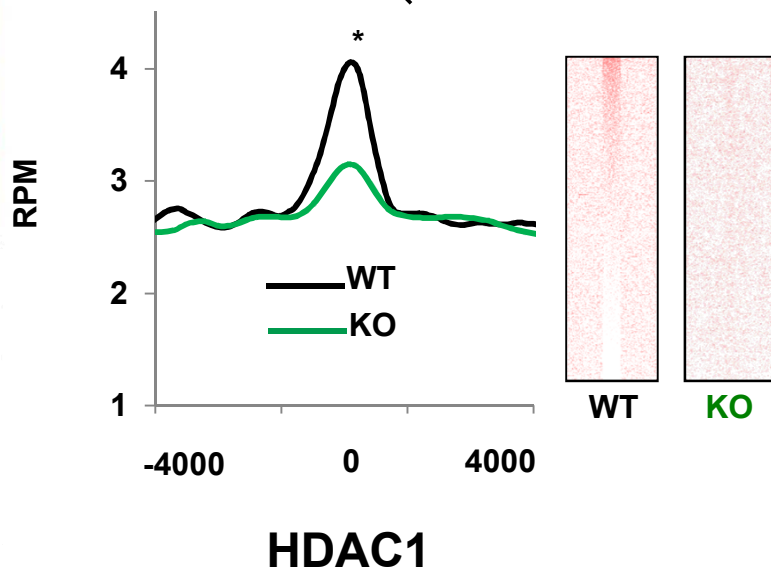


\* Chi-squared test  
p-value < 2.2e-16

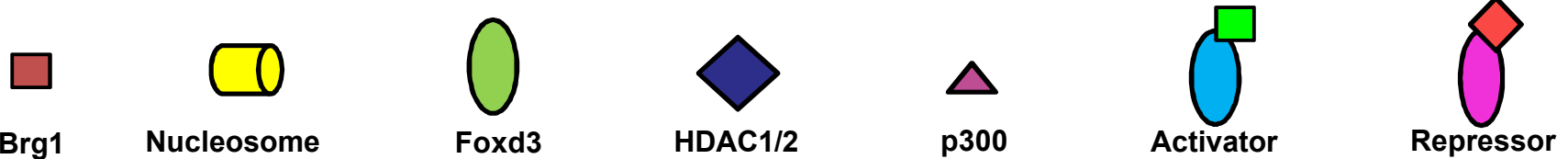
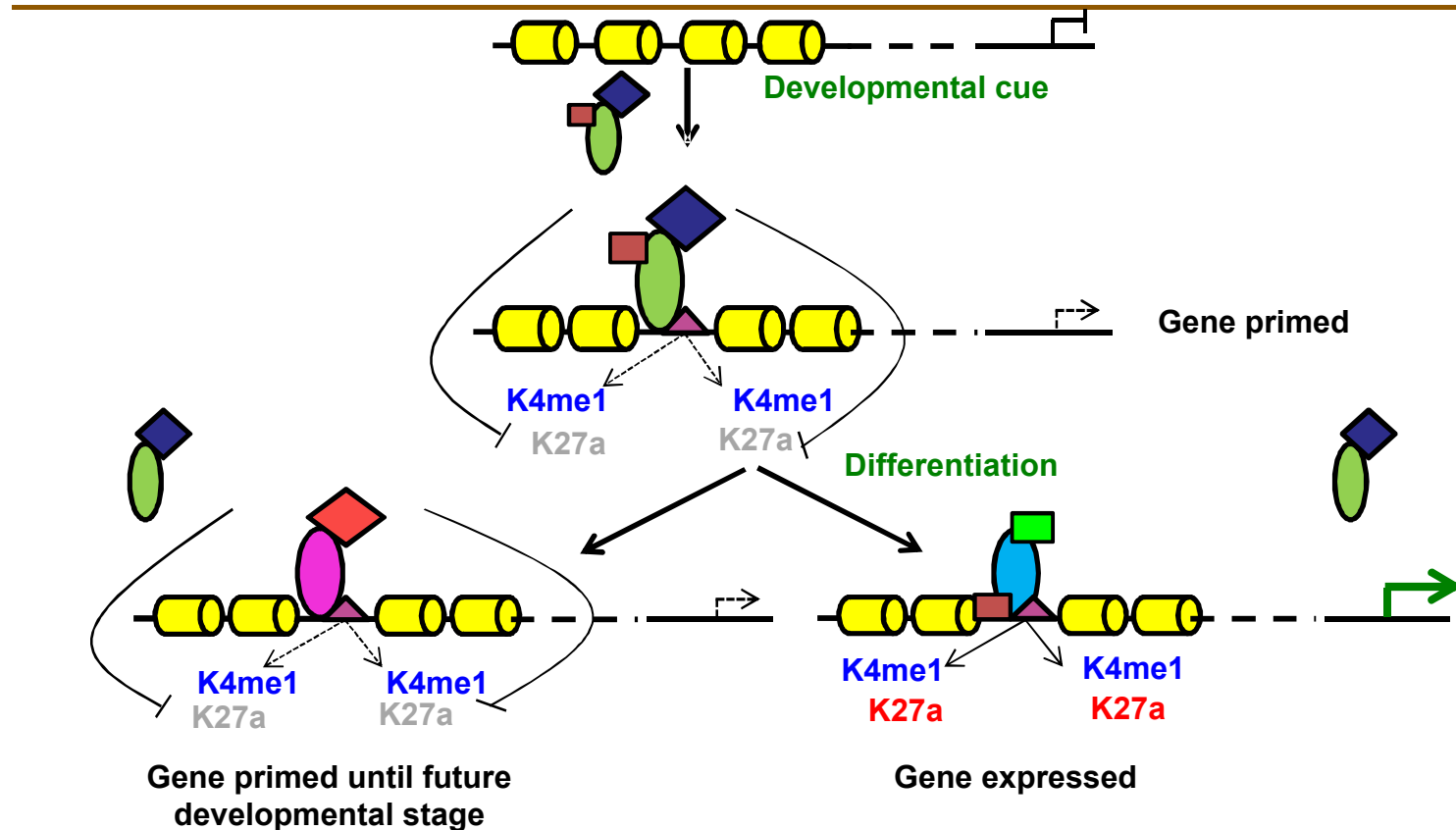
Brg1 KO gene expression



Ho et al (2013), Jamaladdin et al (2014)

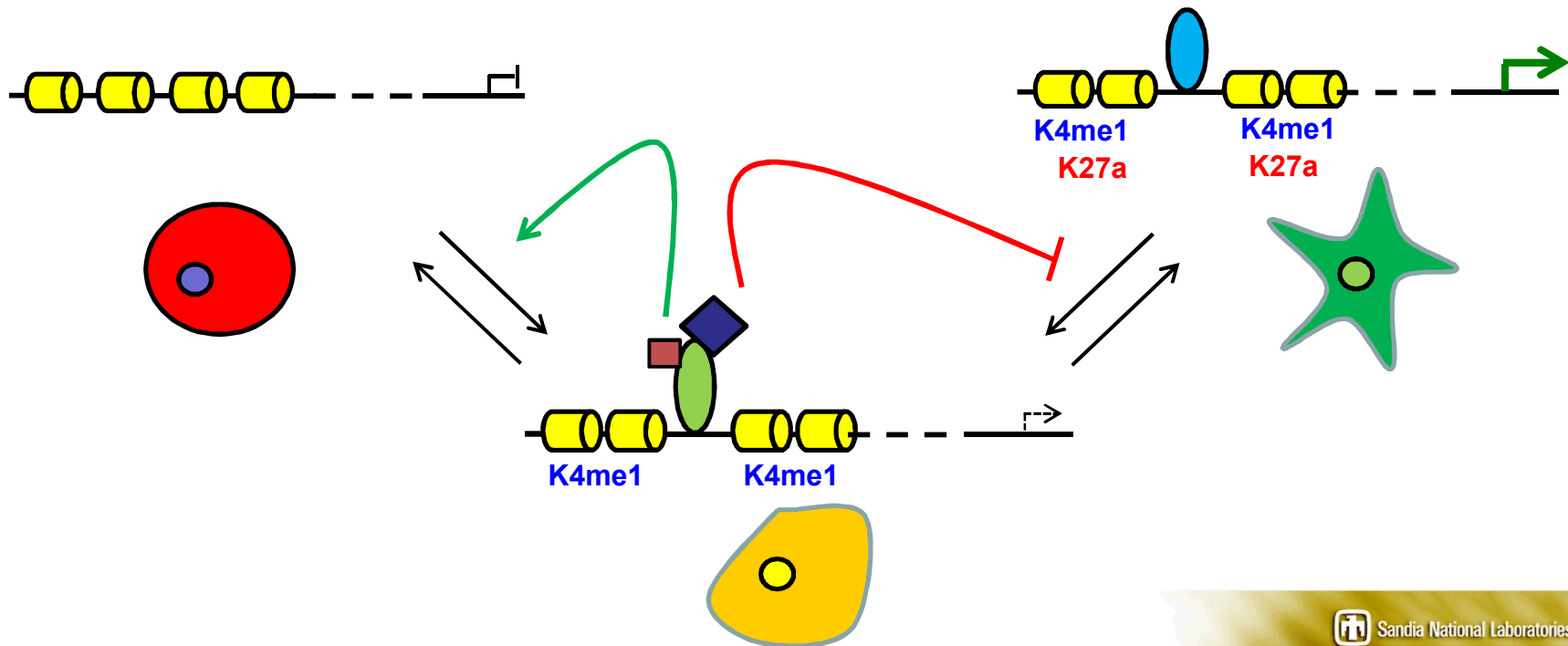


# *Foxd3 establishes and primes developmental enhancers*



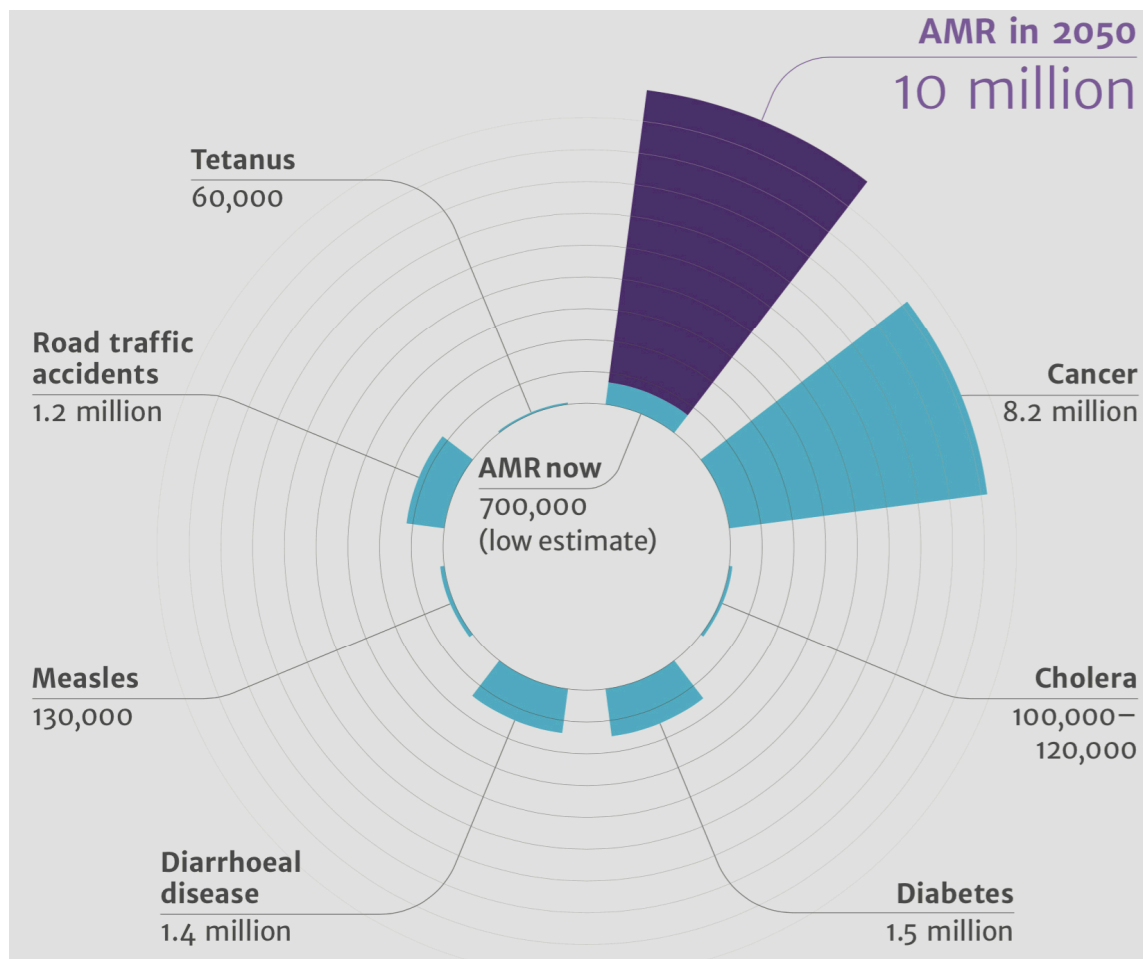
## Broad implications

- Metastable, intermediate states during cellular responses and transitions
- Identification of the regulators requires integrative genomic analysis of chromatin and gene expression

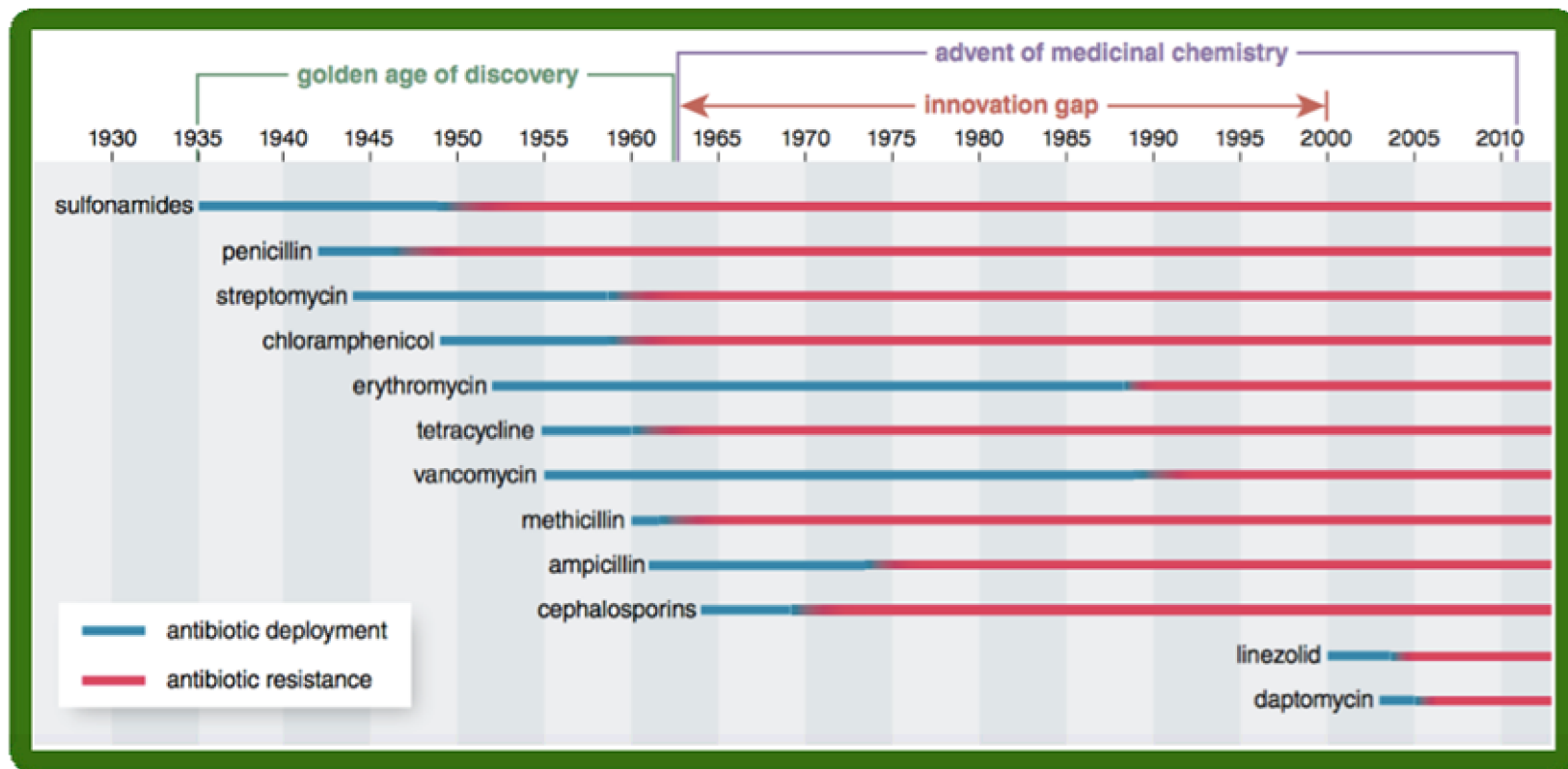


# Antimicrobial resistance – a global health crisis

AMR deaths relative to other major causes of death

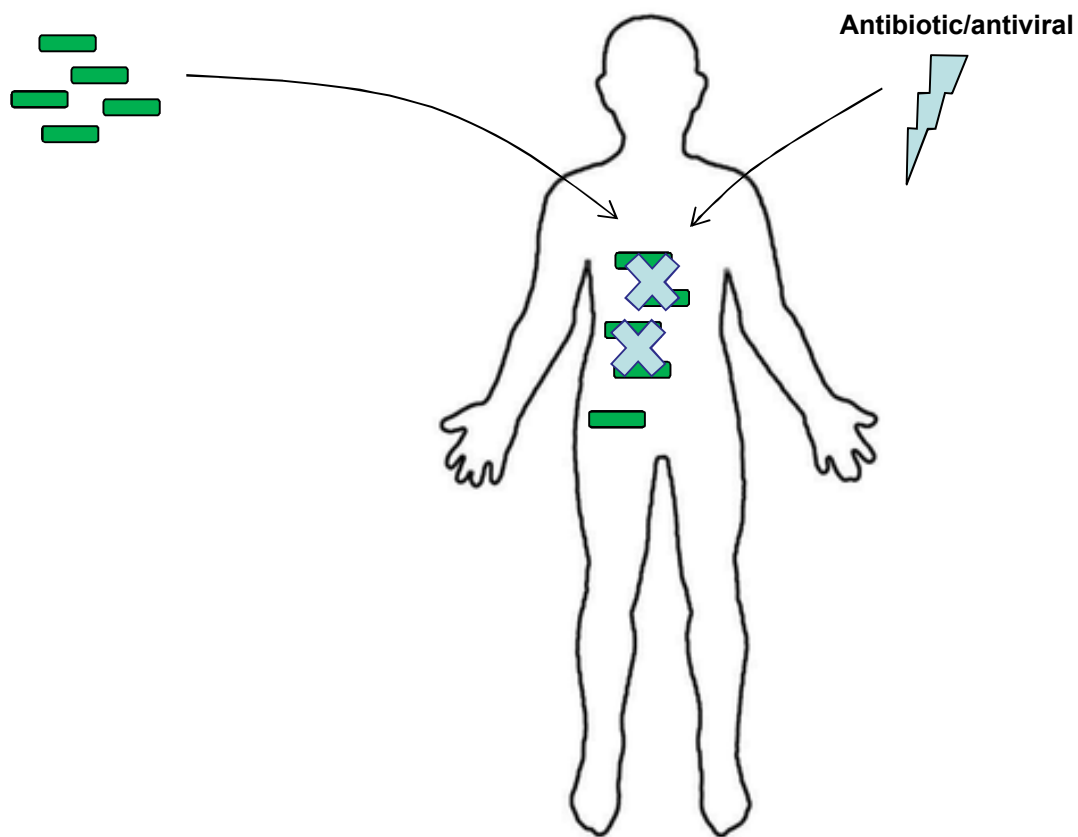


# *Antimicrobial resistance – a global health crisis*



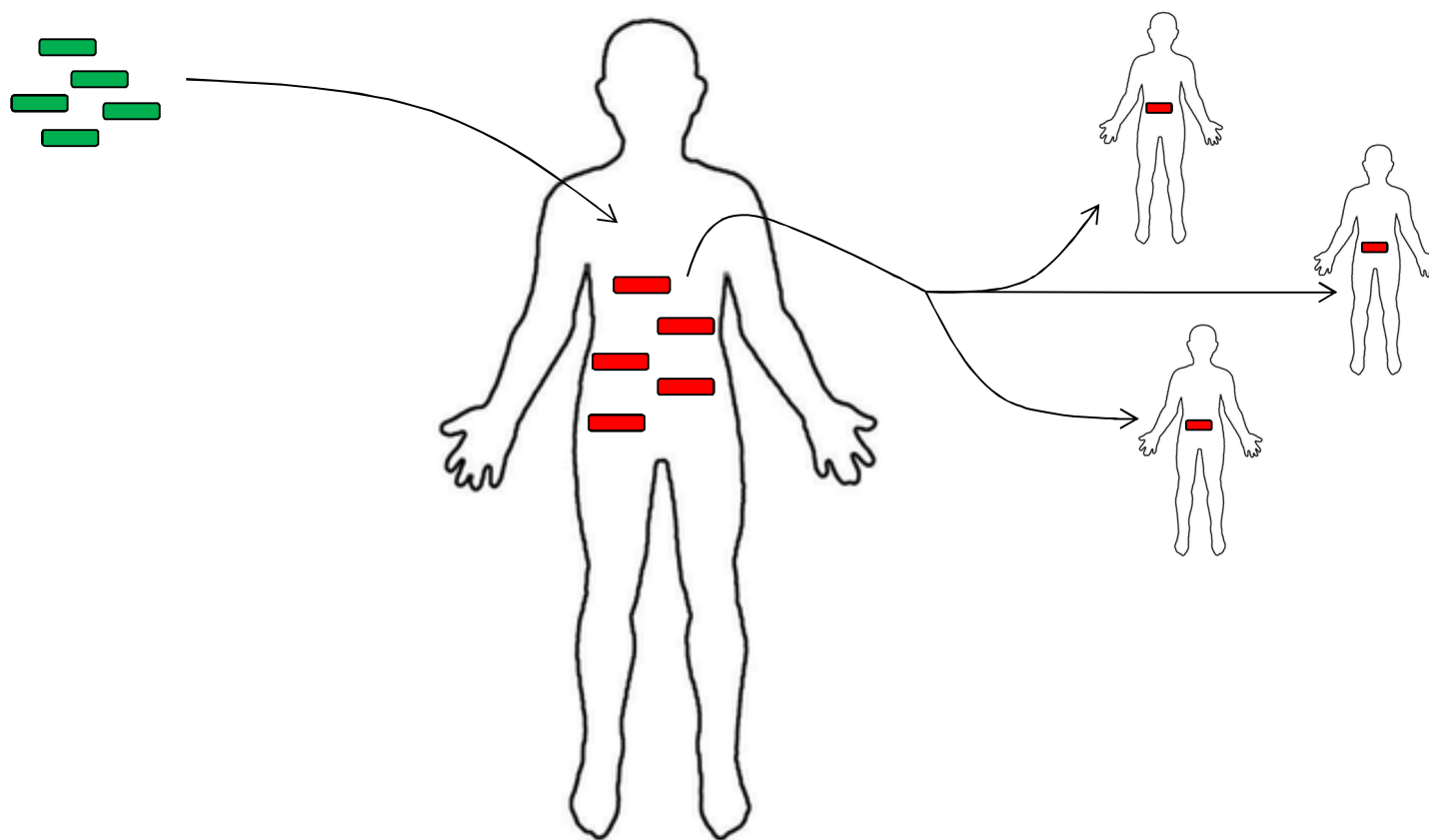
<https://antimicrobialresistance101.wordpress.com>

# *Solutions to antibiotic resistance*

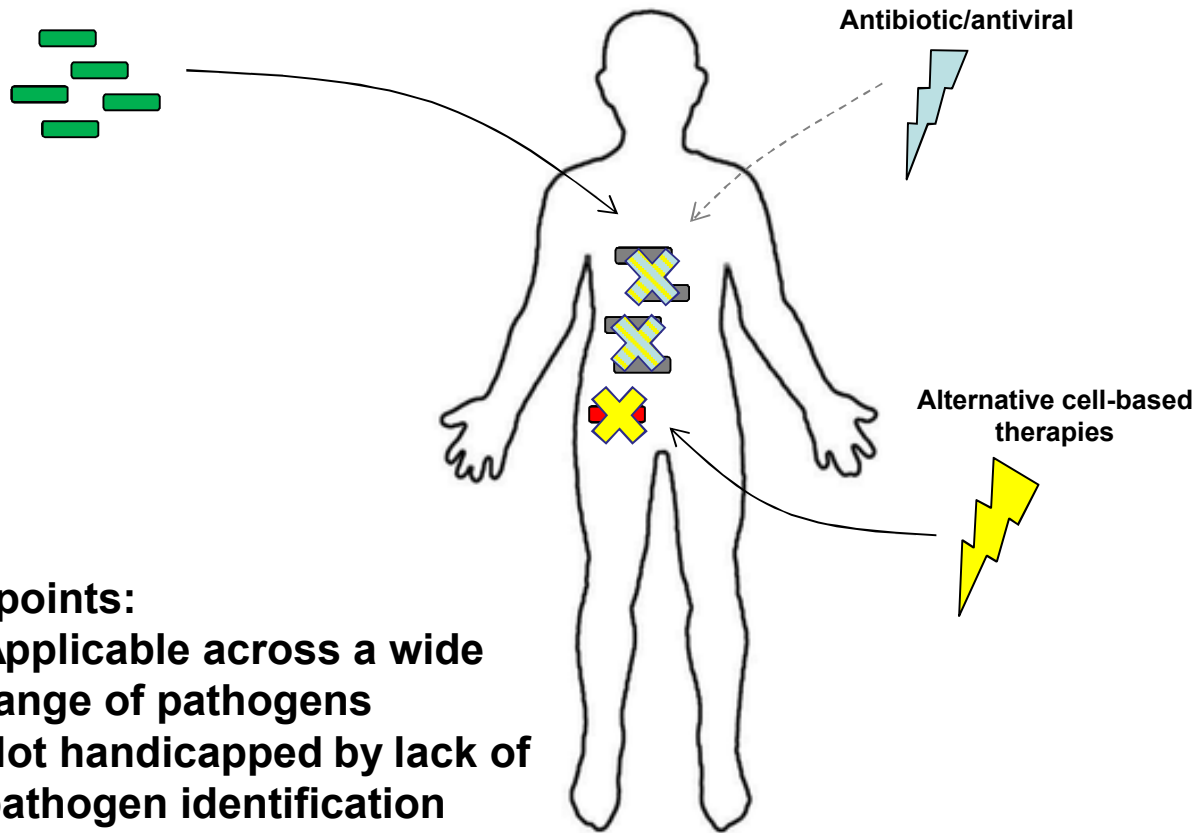




# *Solutions to antibiotic resistance*



# *Solutions to antibiotic resistance*



## **Key points:**

- **Applicable across a wide range of pathogens**
- **Not handicapped by lack of pathogen identification**

# ***New cell-based approaches to antibiotic resistance***

- Infections with no cure/AMR infections spread unchecked, and eventually can lead to death (parallels in presentation and prognosis with cancer, except with the added “bonus” of being contagious!)
- Cell-based therapies (i.e. immunotherapy) have been very successful in treating a variety of diseases including, especially cancer and autoimmune
- Challenges to tackle – make cell-based infection therapies:
  - Cheap
  - Fast
  - Effective
  - Safe

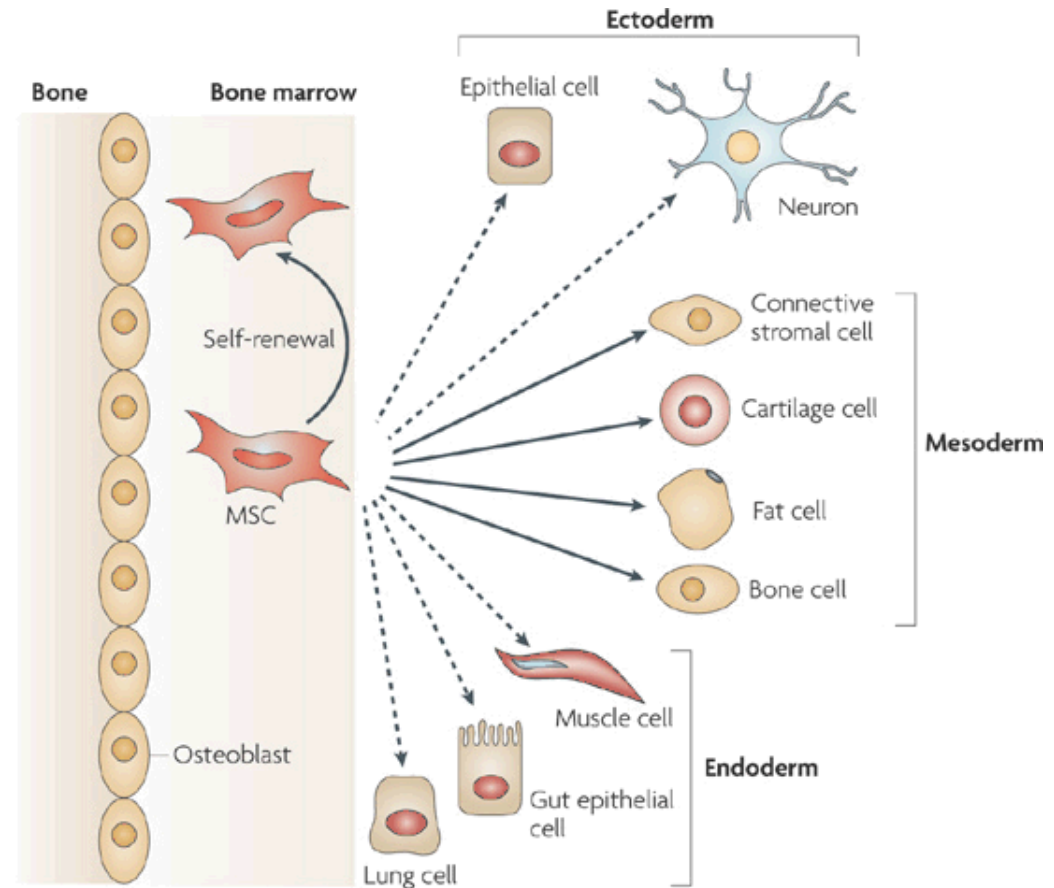
## ***New cell-based approaches to antibiotic resistance***

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- We hypothesize that by using **gene editing technology** (i.e. CRISPR/Cas9), we can rapidly convert large amounts of easily accessible cells to therapeutic cells.
- We plan to use mesenchymal stromal cells (MSCs) as our therapeutic cell type of interest
- MSCs are found in **bone marrow, adipose tissues,** and **umbilical cord tissue**, among other sources

# *MSCs can divide and regenerate tissue*

- MSCs can be passaged in culture, and differentiated down many lineages (multipotent)



Nature Reviews | Immunology

Uccelli et al, 2008

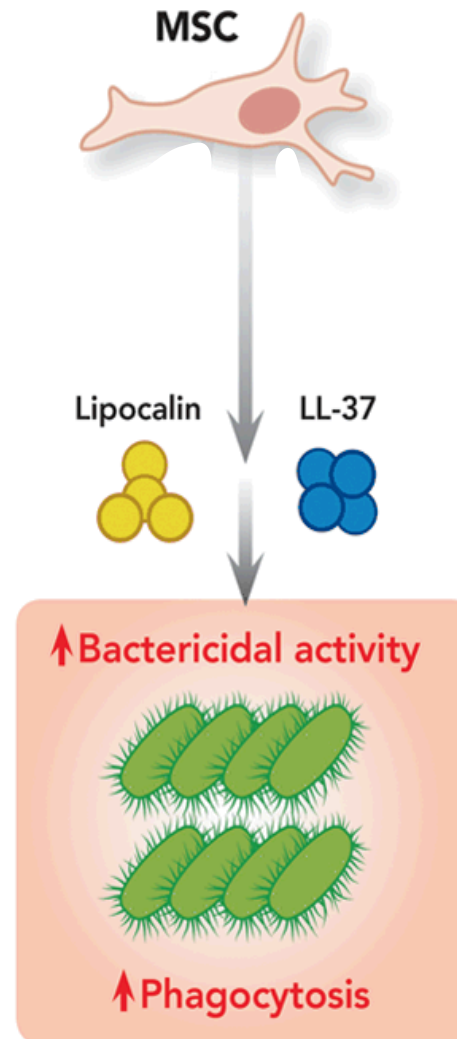




**In addition, MSCs are hypoimmunogenic, making them a good candidate for both autologous and allogeneic therapy**

# *MSCs are antimicrobial*

- MSCs secrete peptides that directly target pathogens
- Lipocalin inhibits bacterial growth by scavenging enterobactin and 'starving' them of iron
- LL-37 intercalates in and causes changes to the cell membrane of bacteria, eventually causing lysis



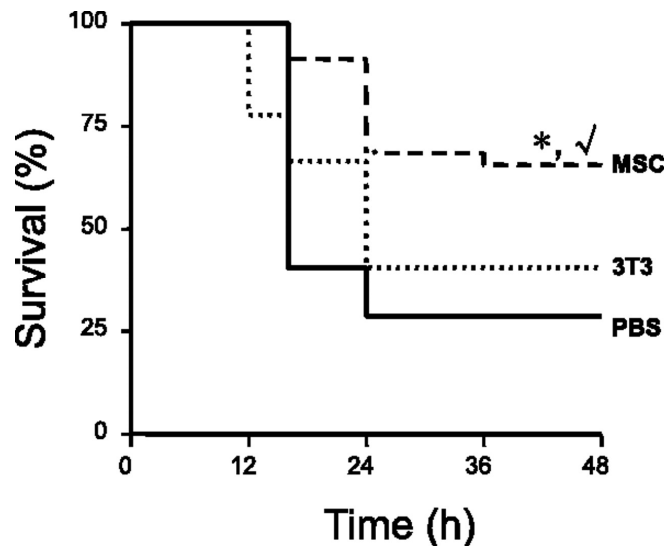
Monsel et al, 2014



# MSCs and infection in the literature

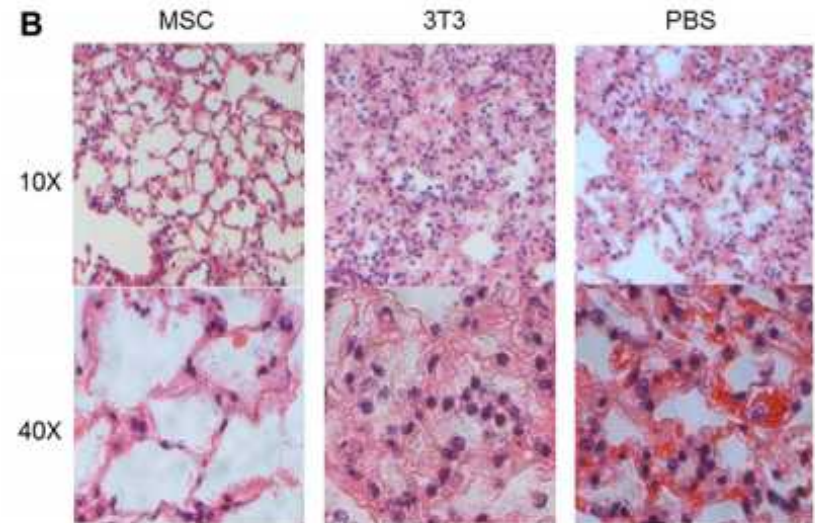
- There are numerous examples of MSCs being used to clear infection and improve survival, including in **sepsis**, **lung infection** and **wound infection**.

Survival in a *Pseudomonas aeruginosa* model of peritoneal sepsis



Krasnodembskaya et al, 2012

Histology (H&E) in a *Escherichia coli* model of lung infection



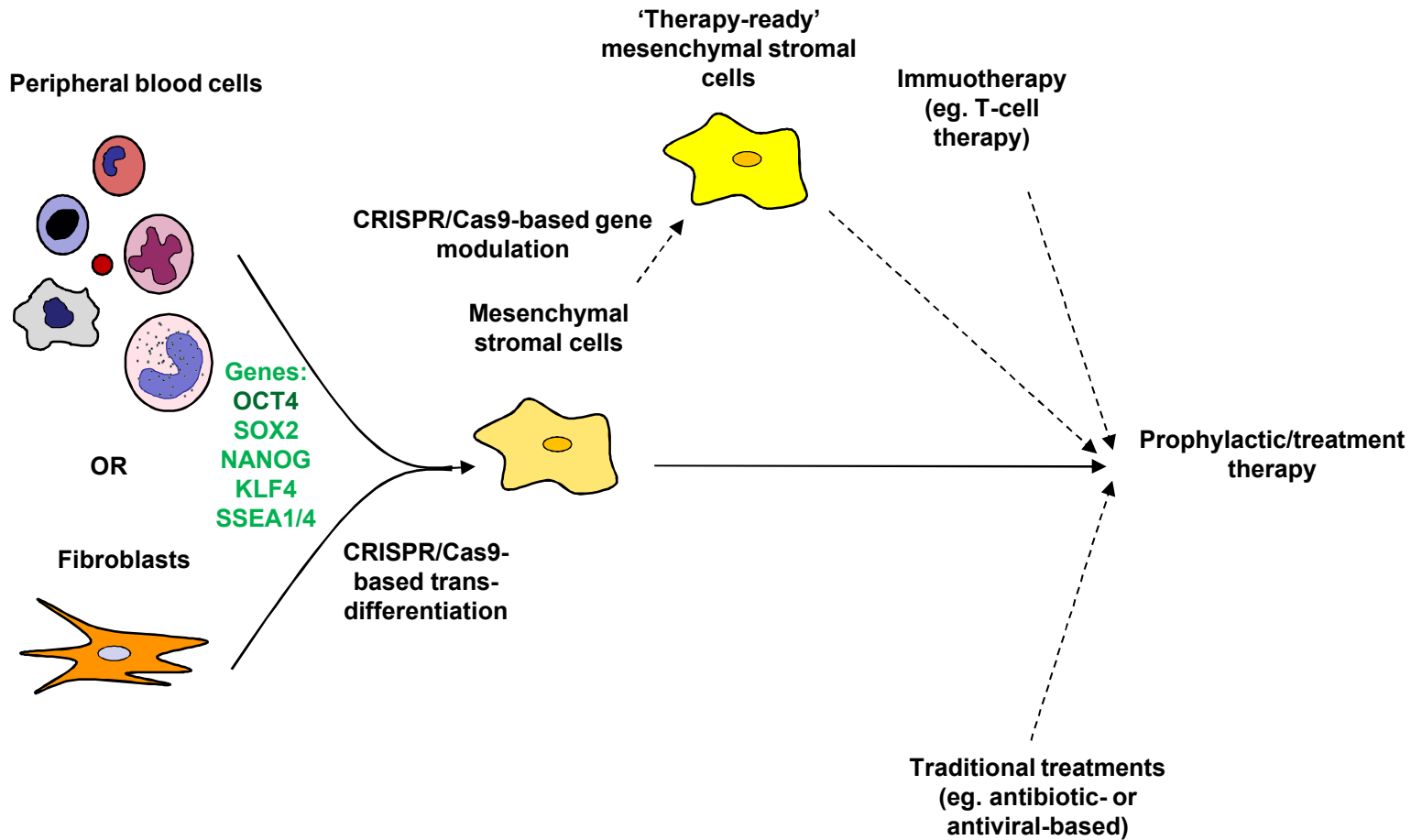
Gupta et al, 2017

## *Challenges ahead*

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- High enough quantities of high-quality, homogeneous MSCs for therapy
- Being able to fine-tune MSC function in a context-specific manner

# Engineering 'therapy-ready' MSCs



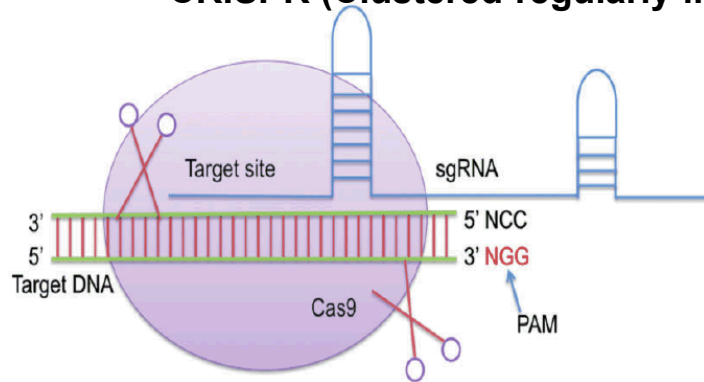
# Engineering 'therapy-ready' MSCs

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- Optimize the *production of therapy-ready mesenchymal stromal cells* (MSCs)
- Use *gene editing technology* for easy and reversible changes in gene expression
- Use both large-scale unbiased and logic-based experimental and bioinformatic approaches to optimize the cells
- Use in vitro and in vivo models of bacterial infection to test the resulting cells

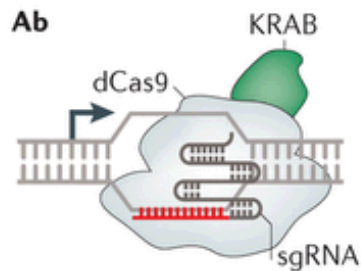
# Harnessing CRISPR/Cas9 gene editing technology

CRISPR (Clustered regularly interspaced short palindromic repeats)

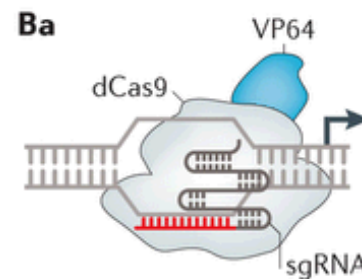


- Inactive Cas9 can be used as a targeting method for other proteins ('Break the scissors' – Jonathan Weissman)

CRISPRi - inhibition



CRISPRa - activation

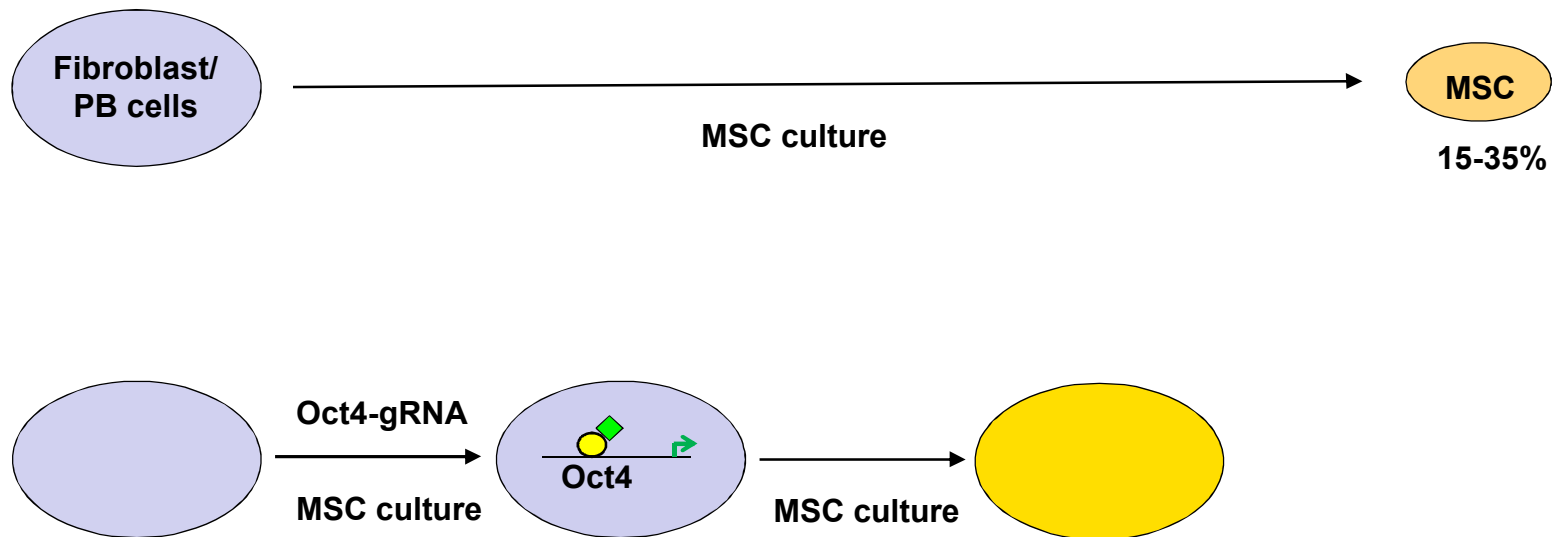


Kaur et al, 2015

Shalem et al, 2015

# *Altering endogenous gene expression profiles to modulate cell fate*

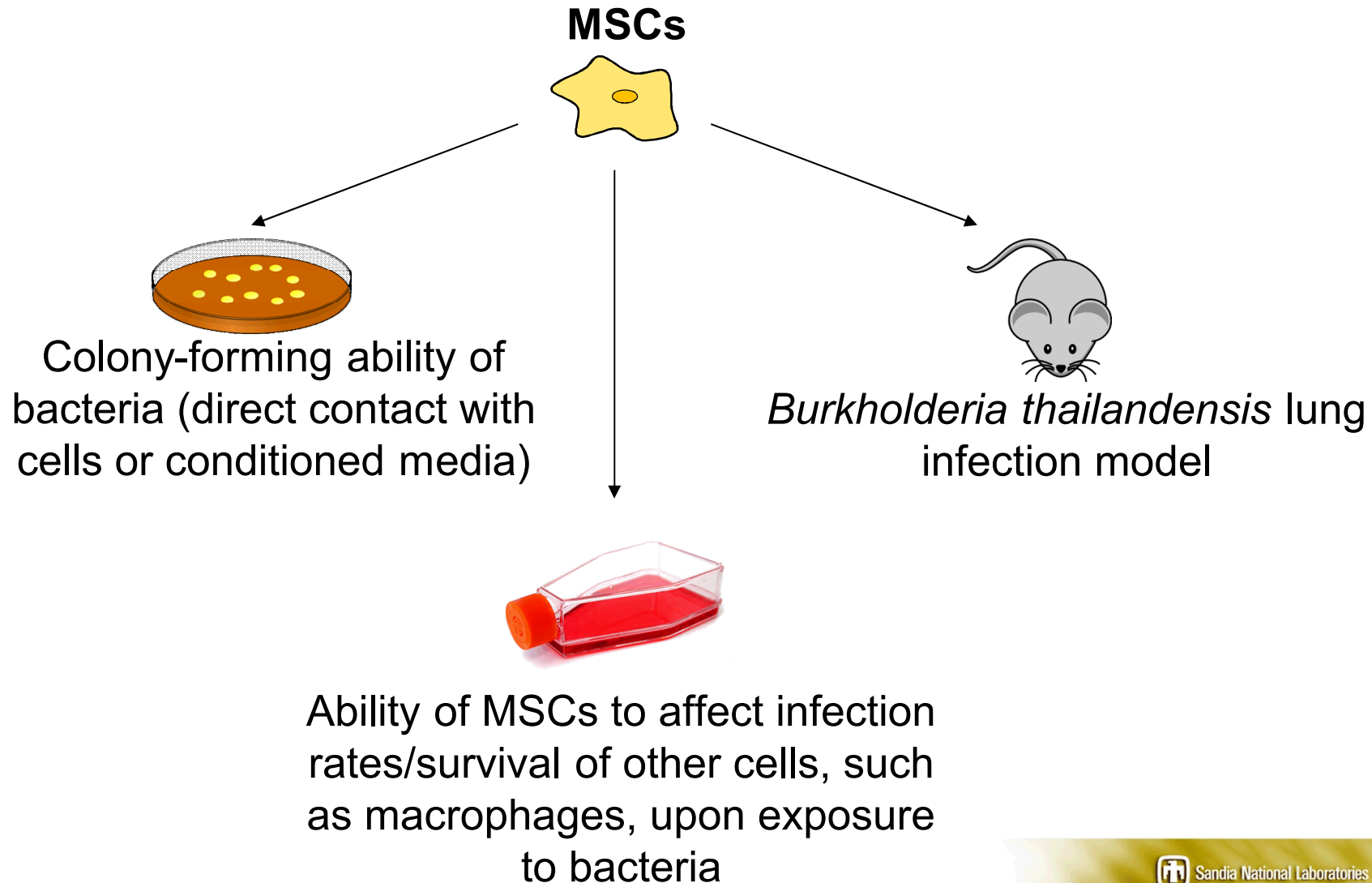
- Optimize the transition from fibroblasts to MSCs



Meng et al, 2013  
Lai et al, 2017  
Pan et al, 2017



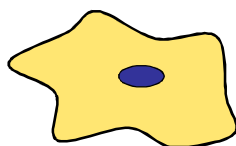
# Verifying MSC potency





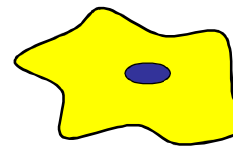
# ***Fine-tuning MSCs to enhance their potency***

Mesenchymal stem cells

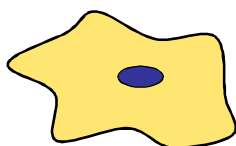


→  
Δ Culture conditions

'Therapy-ready' mesenchymal stem cells



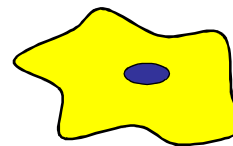
Mesenchymal stem cells



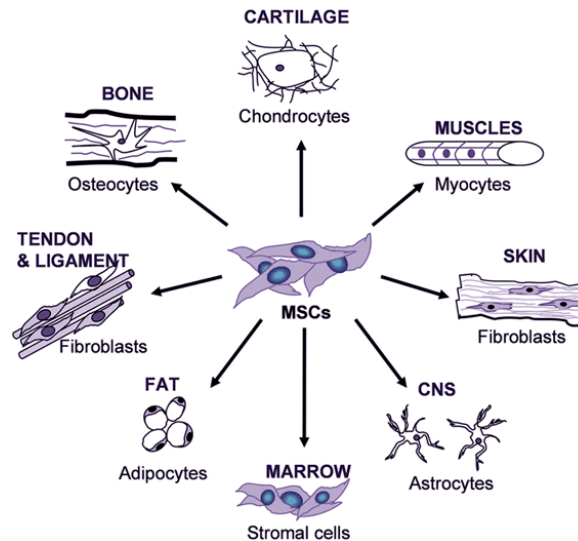
Examples:  
LL-37  
MHC1/2  
HLA-G  
PGE2  
IDO

→  
CRISPR/Cas9-based gene modulation

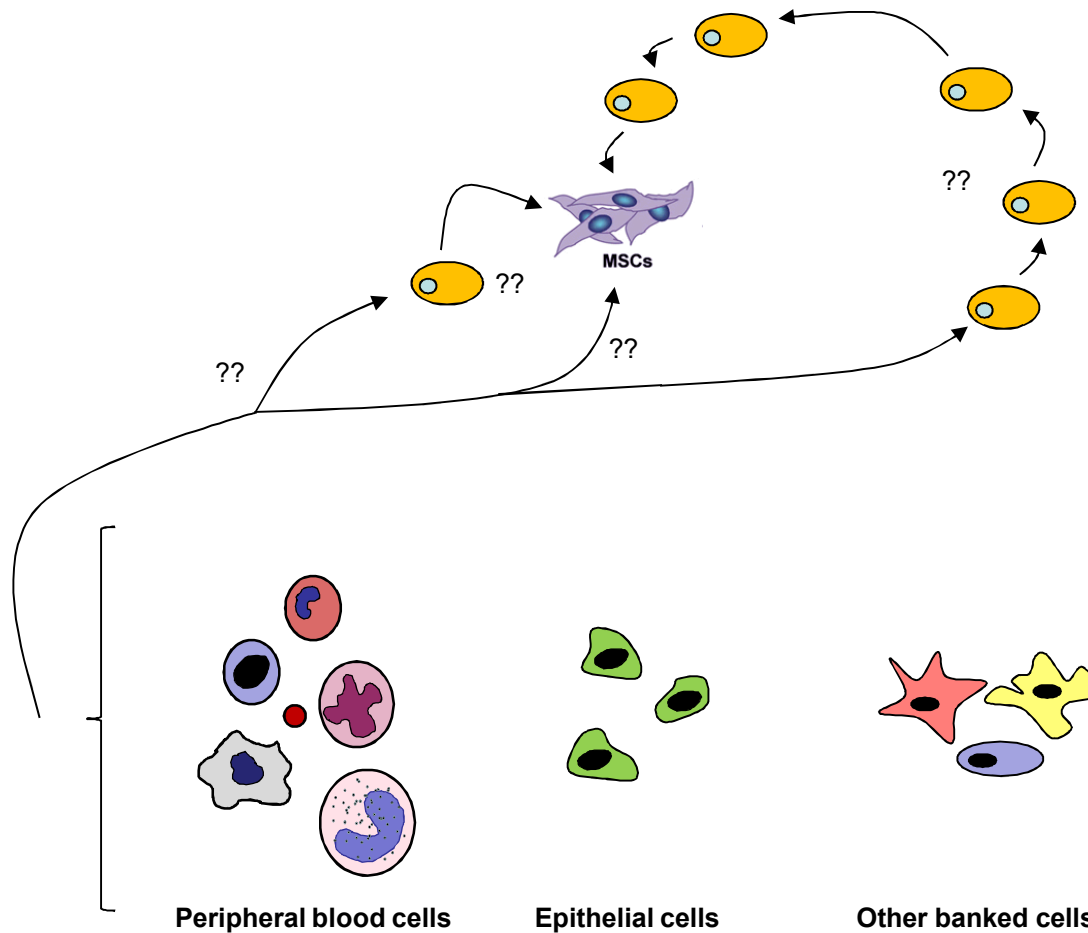
'Therapy-ready' mesenchymal stem cells



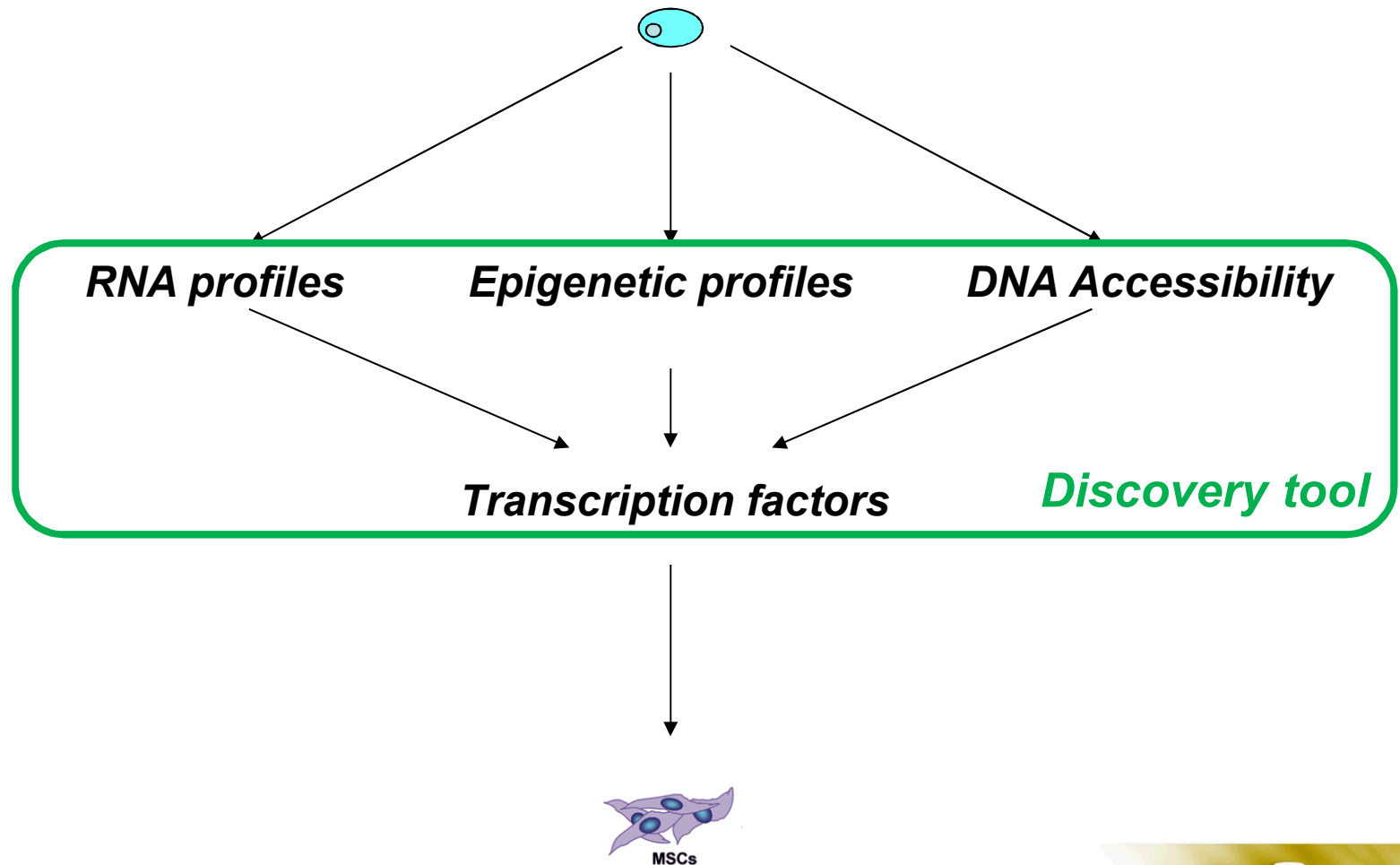
# *Expanding the types of cells that can be converted to MSCs*



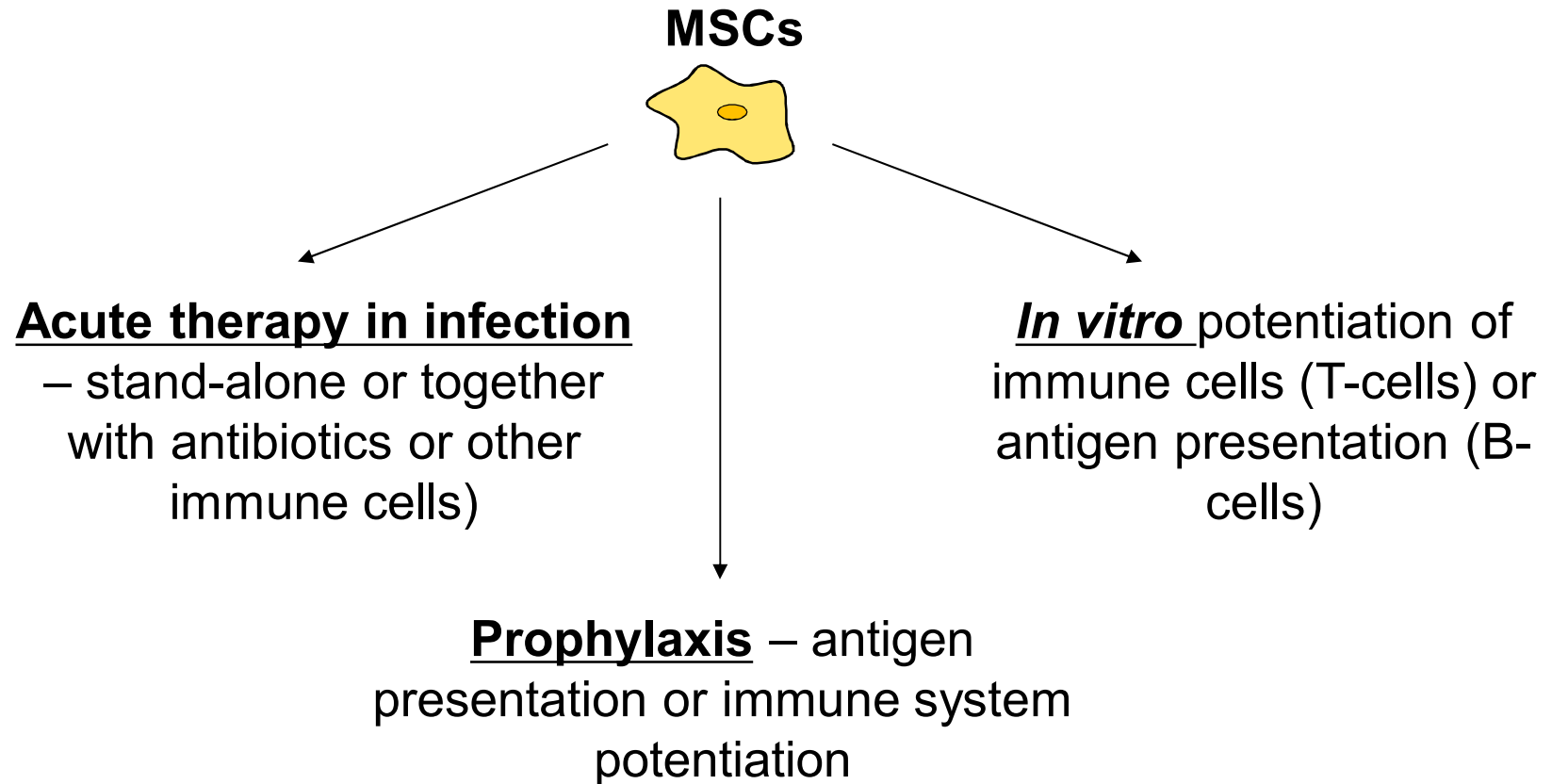
# Expanding the types of cells that can be converted to MSCs



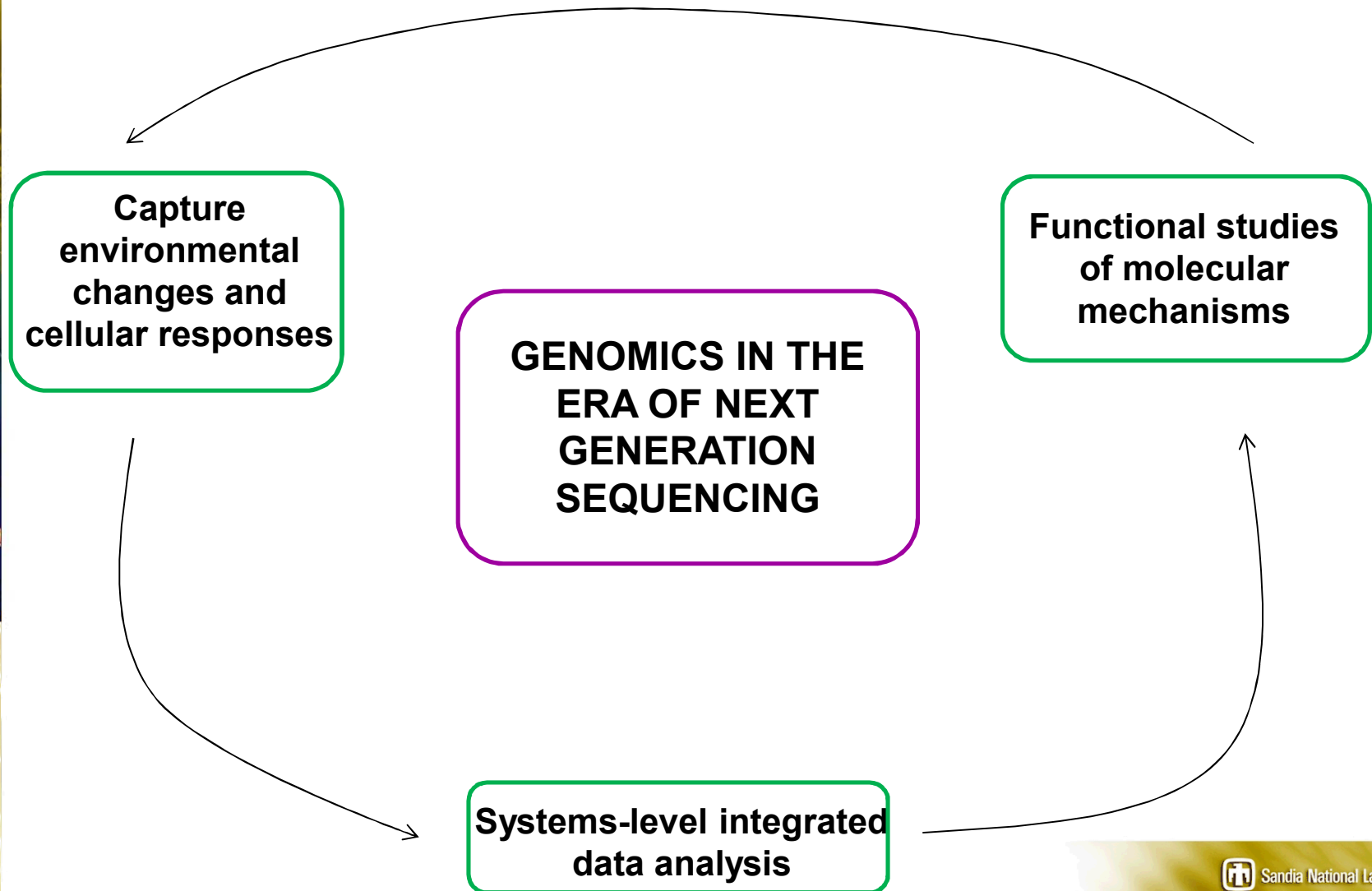
# ***Bioinformatic analysis to identify key factors for converting cells***



# Clinical uses



# *Collecting, synthesizing and deciphering genomic information for engineering cell fates*



# Acknowledgements

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Robert Blelloch and Blelloch lab

8623, 8625 and 8621

Amy Chen

Jacob Freimer

Patricia Labosky (NIH)

Danial Muhammad

Alex Pankov (UCSF, Costello lab)

Barbara Panning (UCSF)

Marisol Pantovich

Ronald Parchem

Jennifer Plank (NIH)

Mike Bartsch

Sara Bird

Cathy Branda

Steve Branda

Harrison Edwards

Hari Jayamohan

Ken Patel

Joe Schoeniger

Anupama Sinha

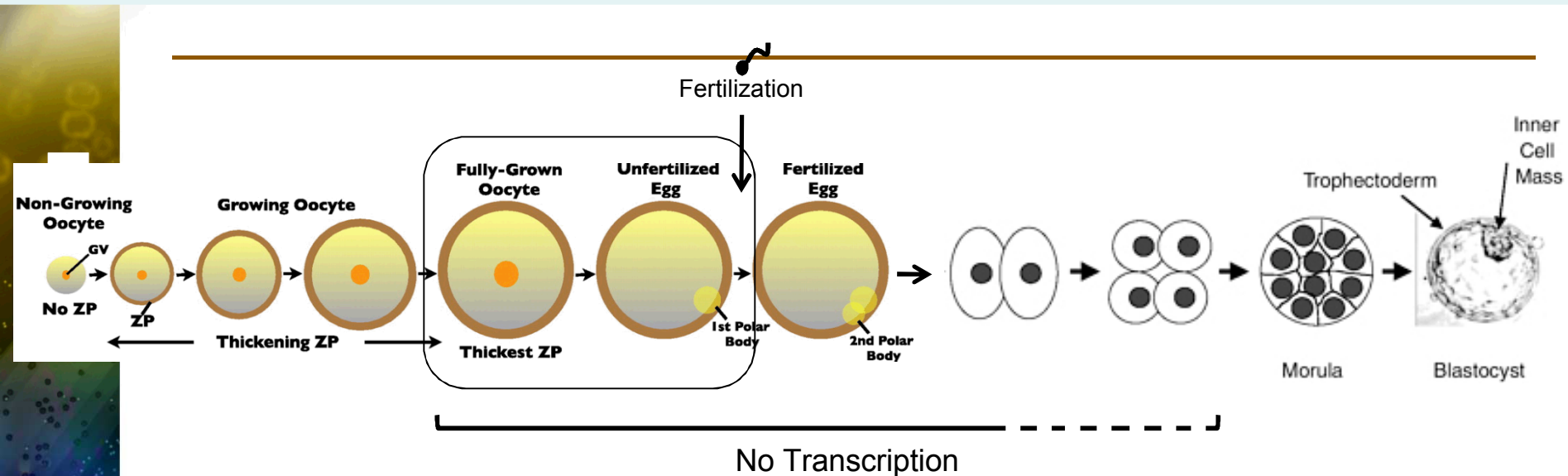
Kelly Williams

Funding – NIH, AP Giannini foundation

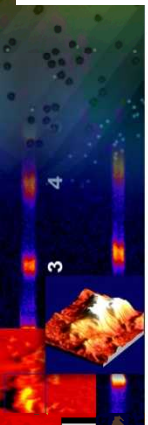
Funding – Sandia LDRD



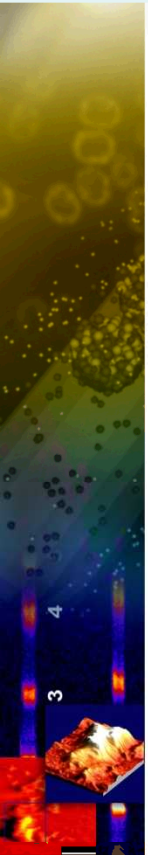
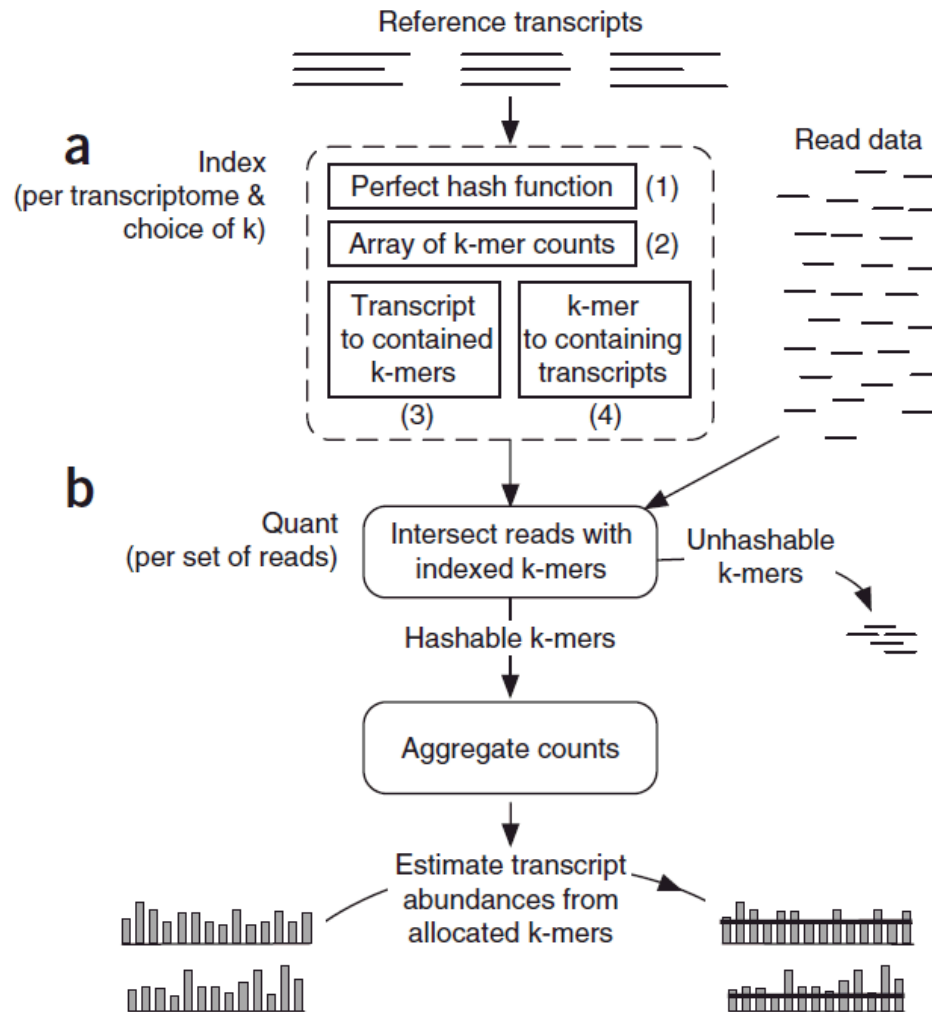
# *Oogenesis happens in the absence of transcription*



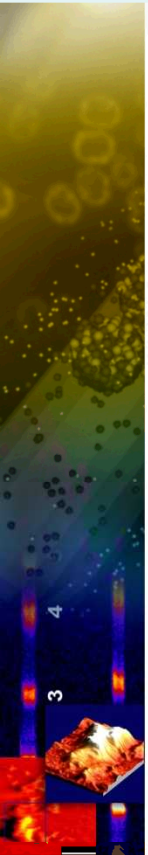
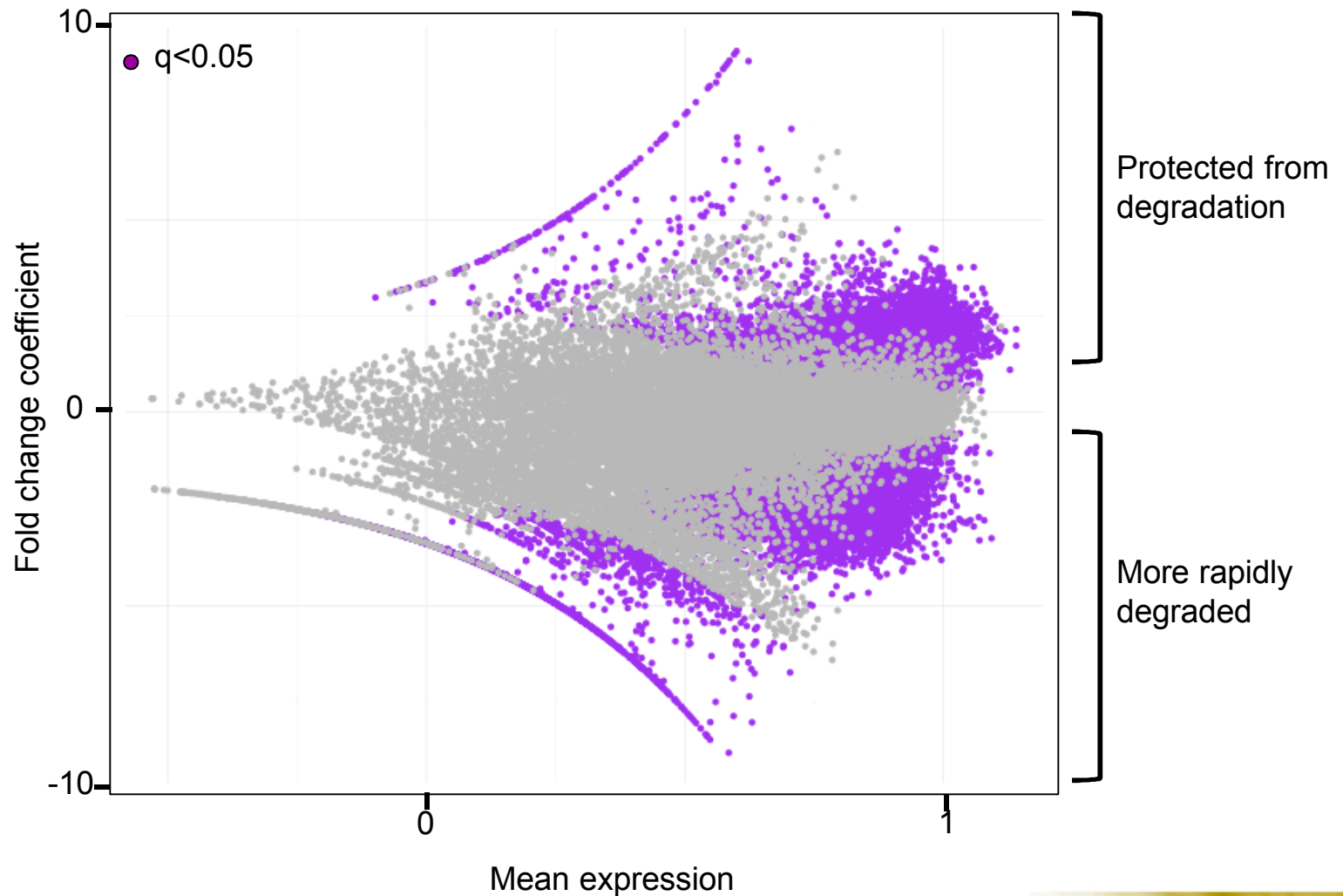
Regulation of mRNA stability and  
RNA seq of GV (fully grown) and MII  
(unfertilized) oocytes



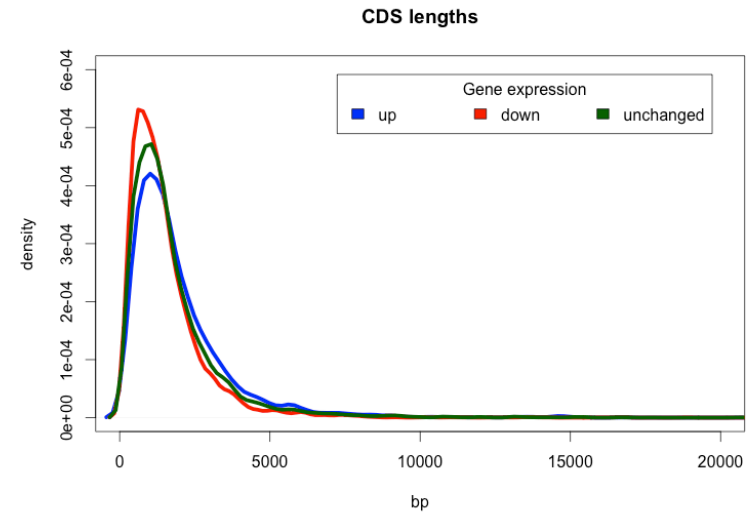
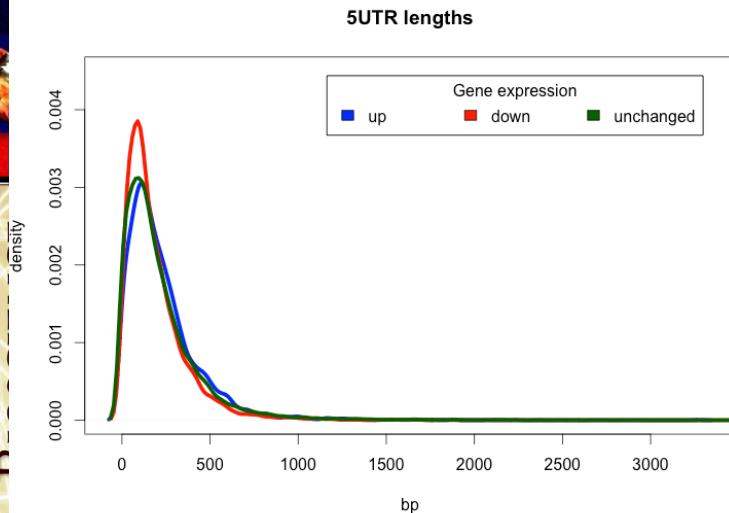
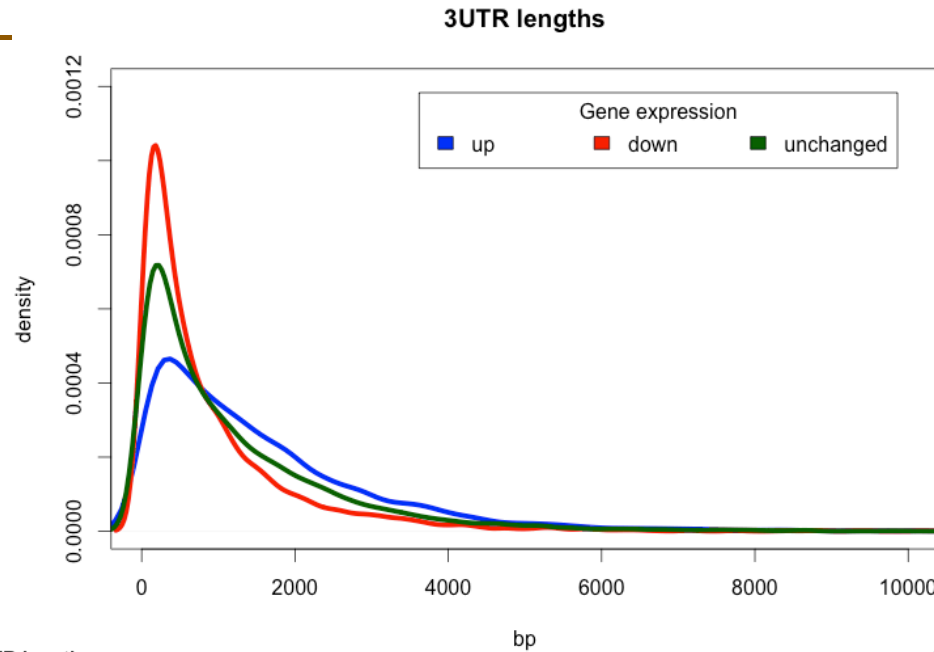
# Transcriptome mappers aka “alignment-free mappers”



# *Dramatic changes in RNA stability during oocyte maturation*

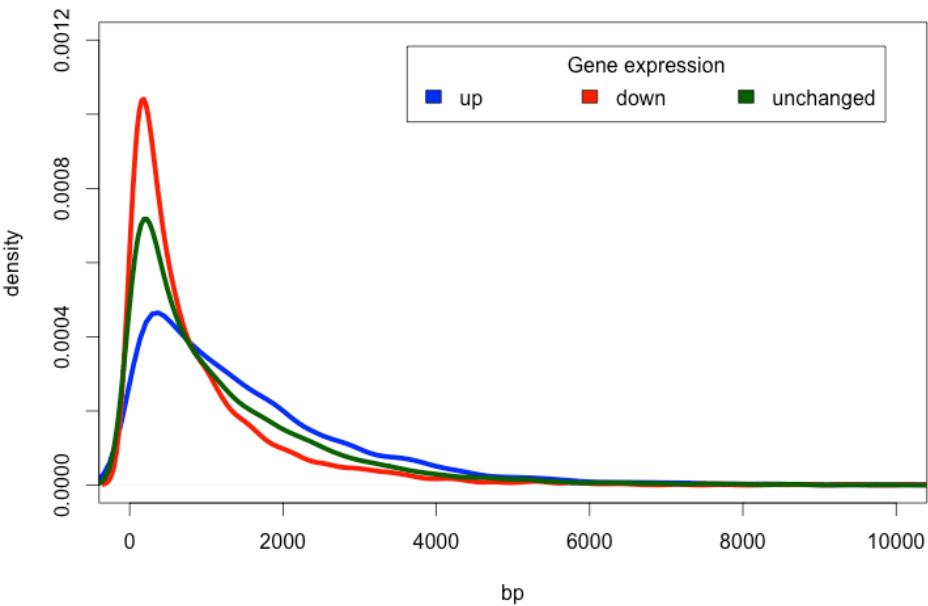


# Changing genes from GV to M2 have different 3'UTR lengths

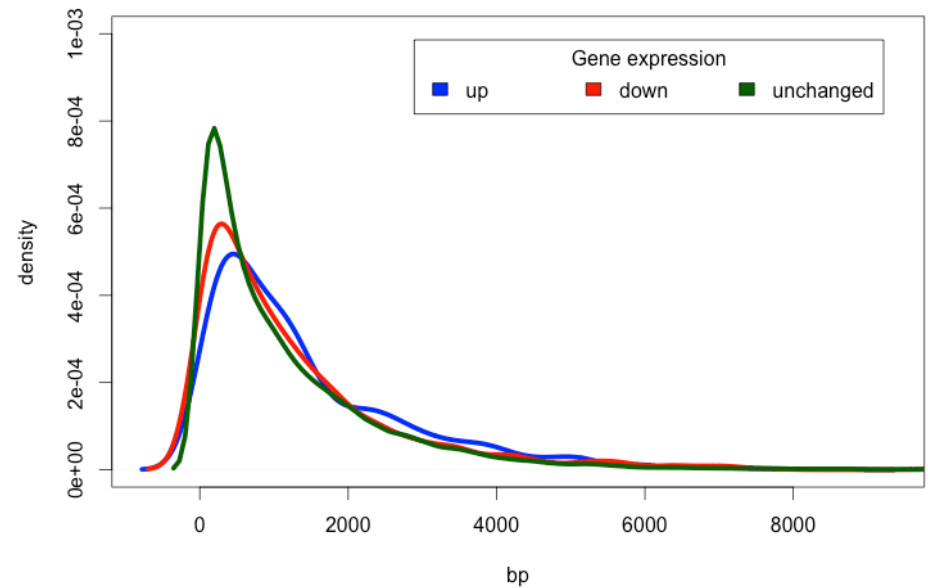


# Changing genes from GV to M2 have different 3'UTR lengths

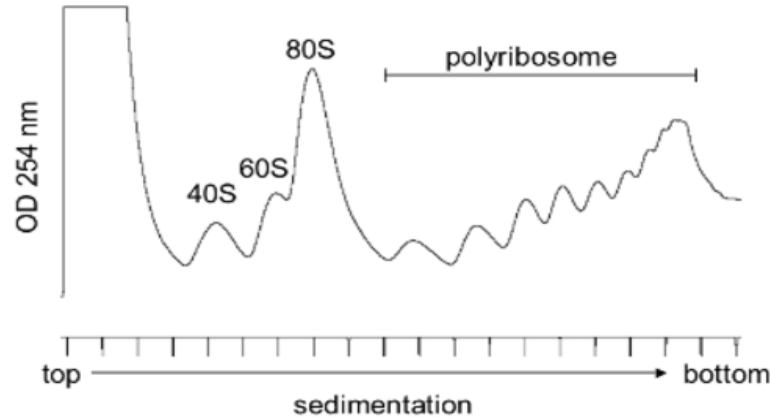
GV to M2



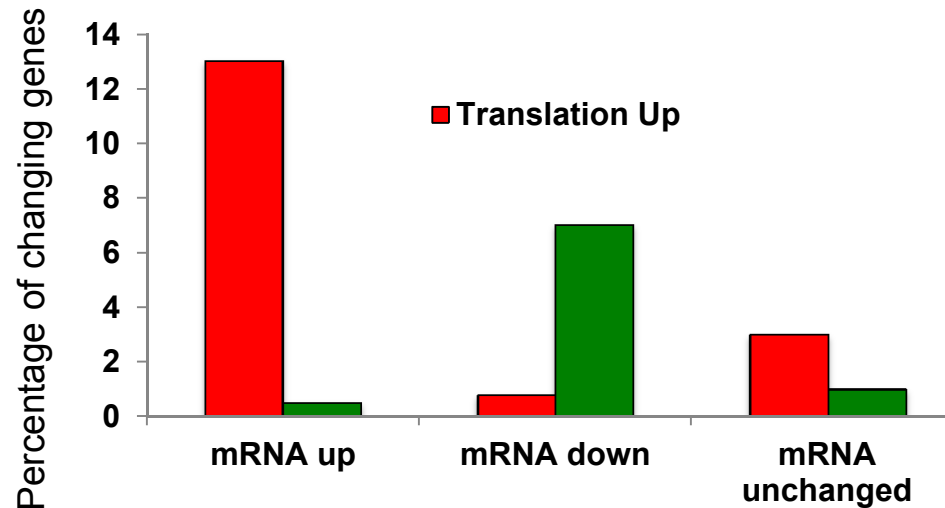
ESC to EpiC



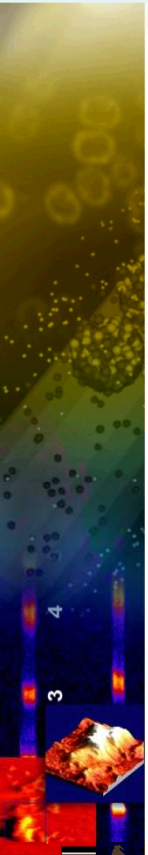
# Relationship of stability with translational efficiency



From Bor et al 2010 (Nature Protocol Exchange)



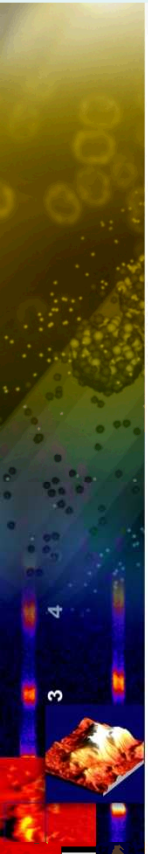
Polysome data from Chen et al 2010 (Genes and



## *Ongoing analyses/Future directions*

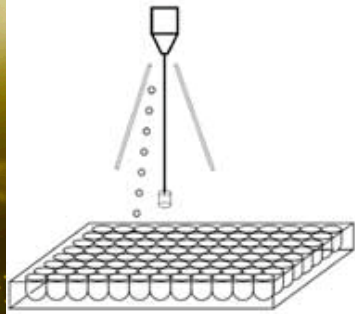
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- Are there any proteins/structural features in longer UTRs that protect them from degradation?
- Can we stabilize transcripts by manipulating the degradation machinery?
- How do these changes relate to oocyte function?

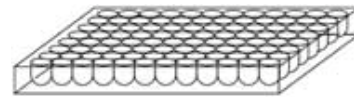




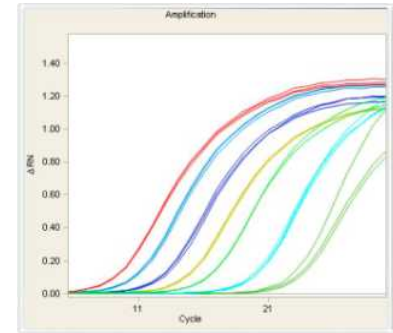
# Single-cell miRNA qPCR shows homogeneity in populations



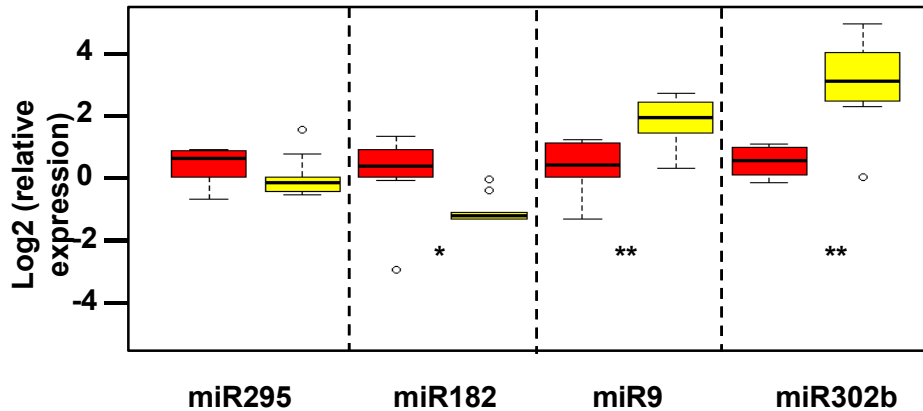
In plate:  
Lysis and RT



Taqman  
probe-based  
qPCR

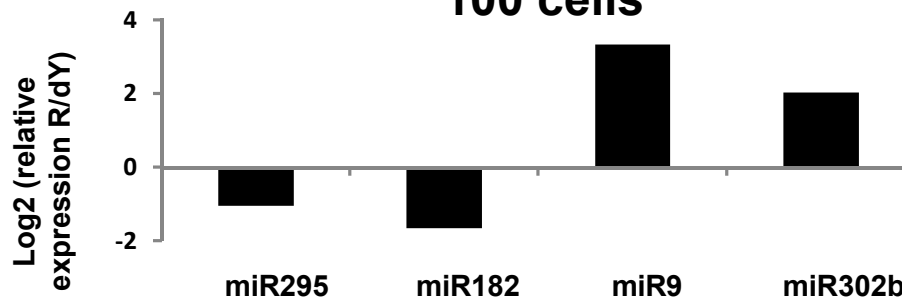


Single cell

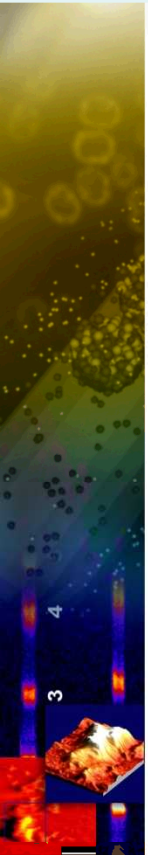
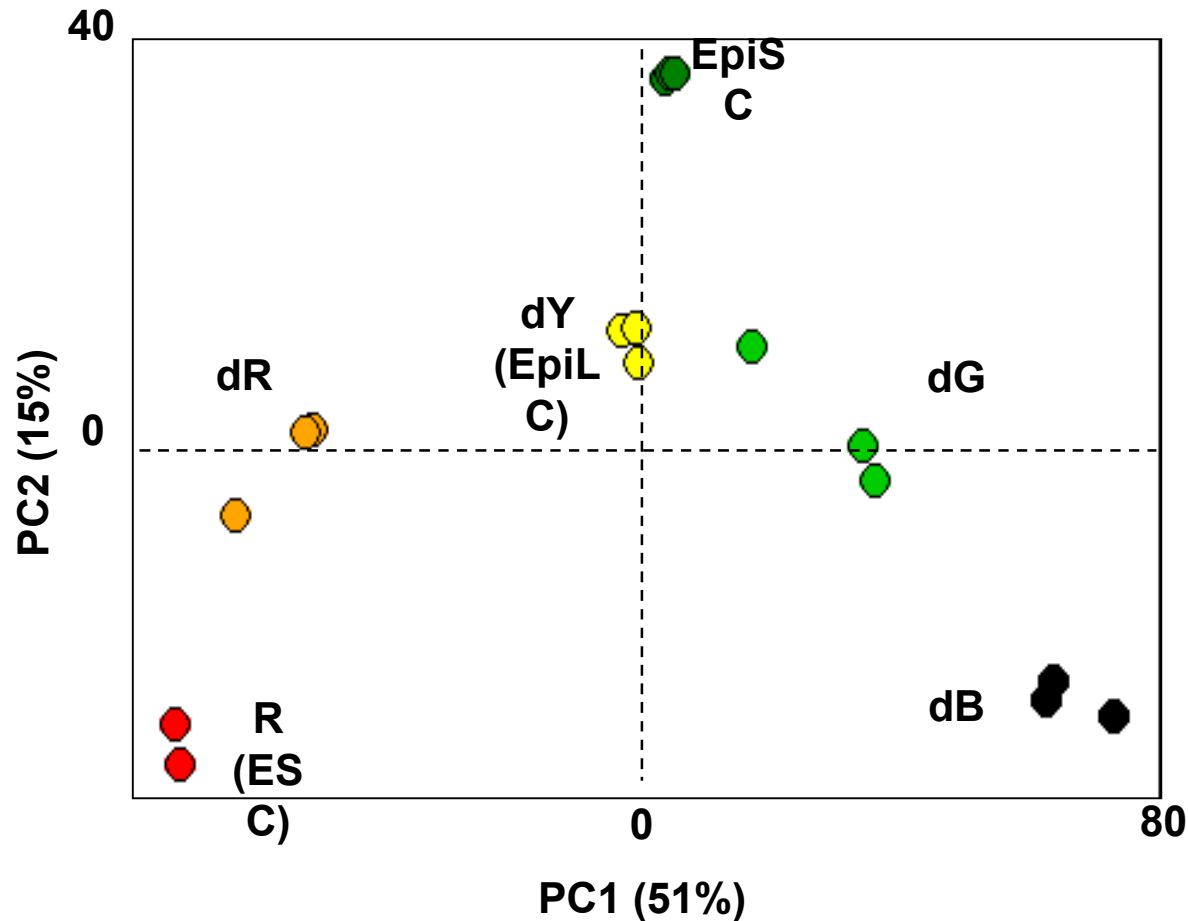


■ ESCs  
■ EpiLCs

100 cells

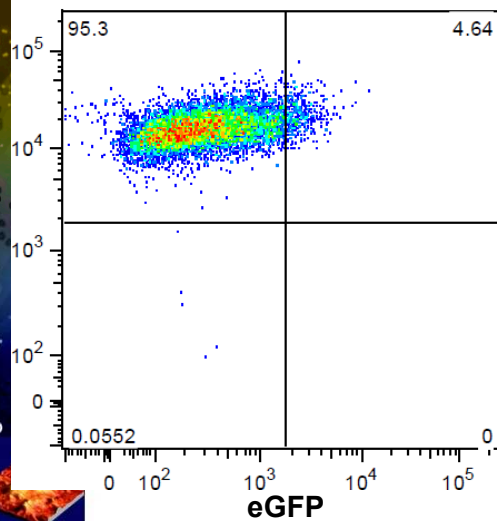


# Principal component analysis of populations demonstrates relationships during differentiation

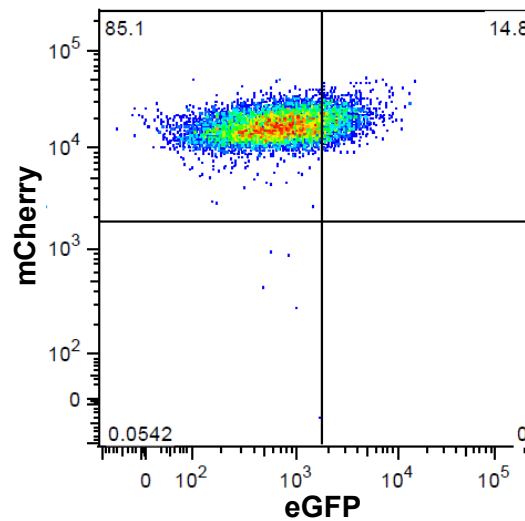


# Homogeneous population of ESCs differentiating to EpiLCs

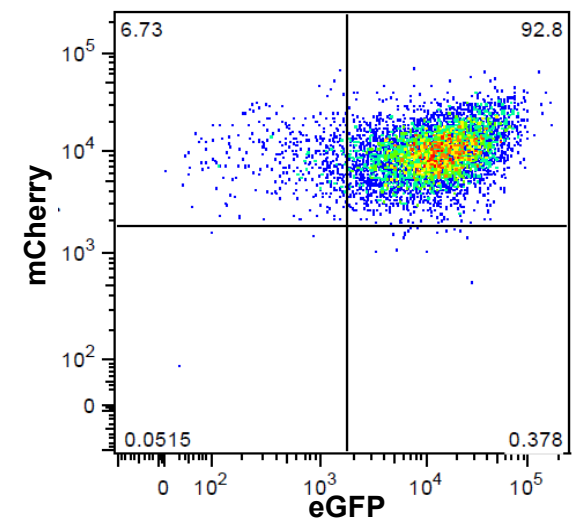
R - ESC



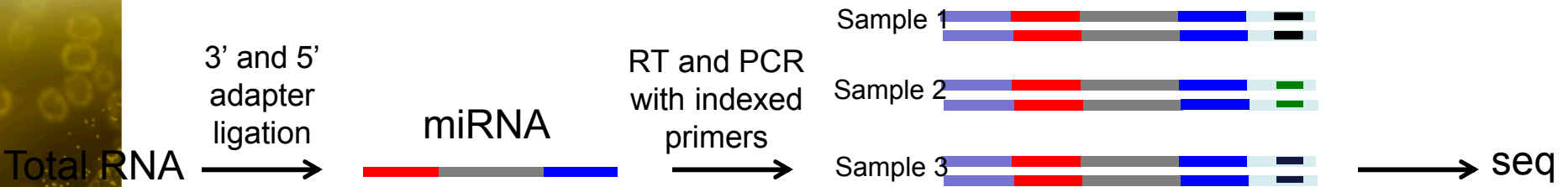
dR



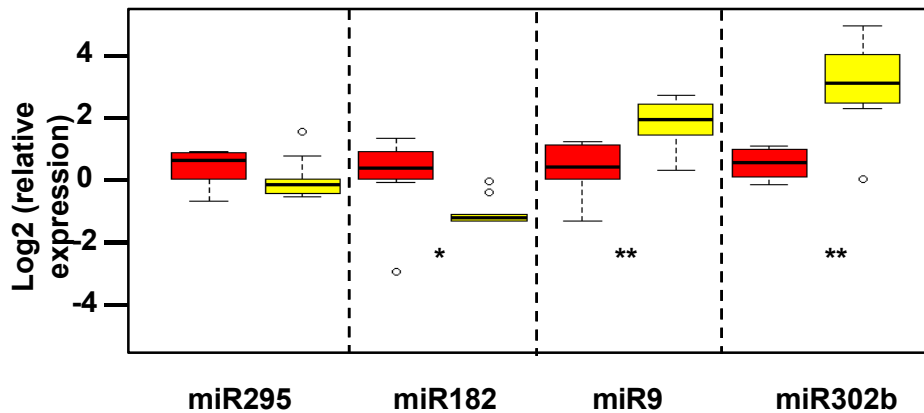
dY - EpiLC



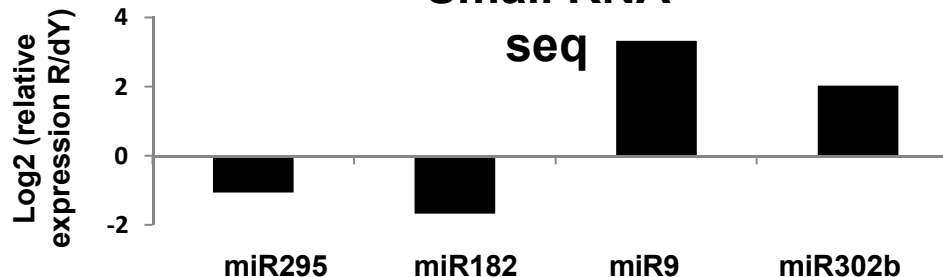
# Single-cell miRNA qPCR shows homogeneity in populations



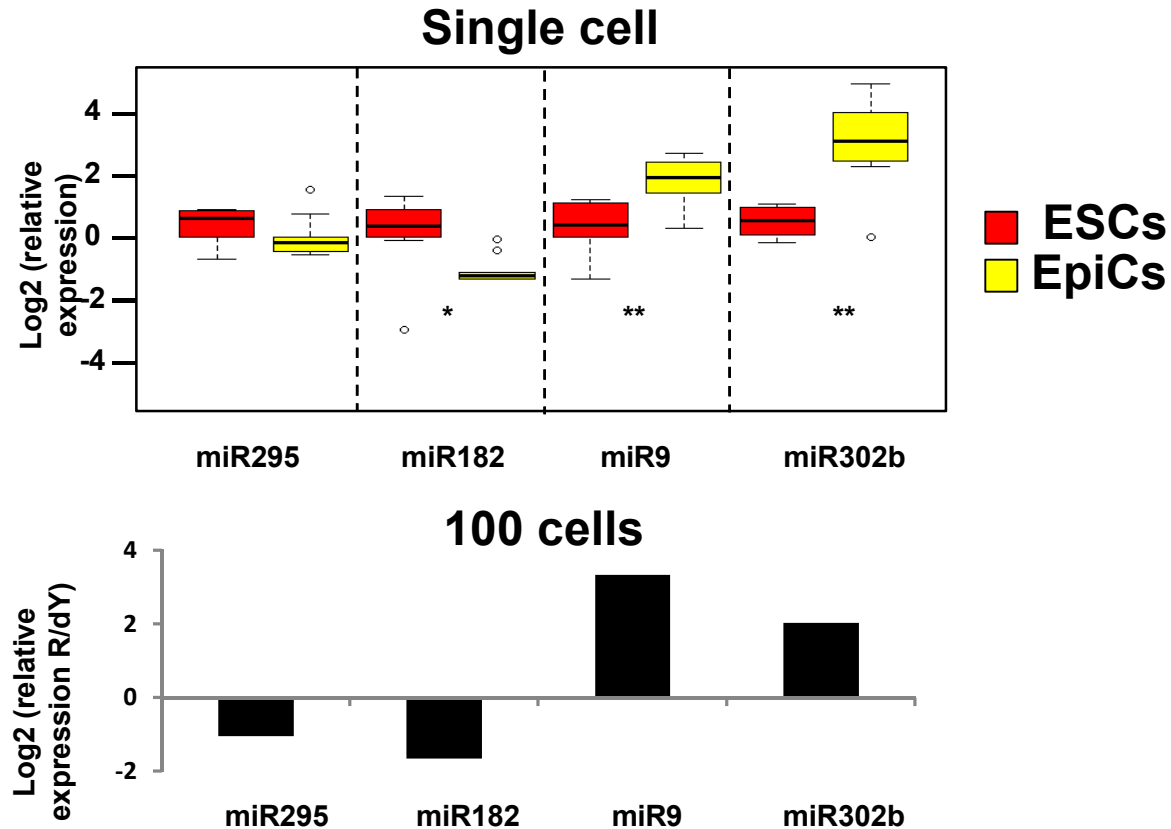
## Single cell



## Small RNA

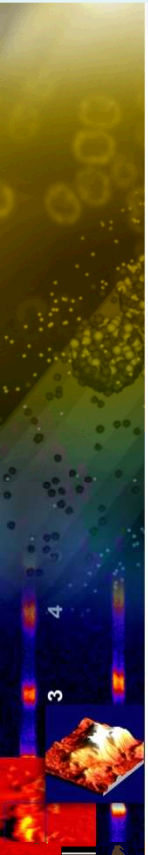


# Single-cell miRNA qPCR shows homogeneity in populations



# Using ChromHMM to Identify Chromatin Domains in ChIP-seq Data

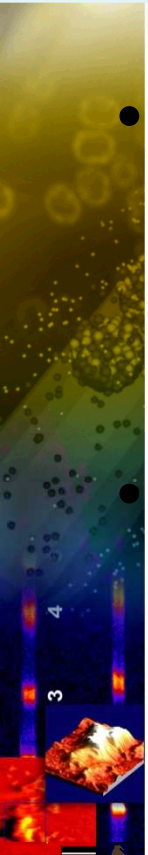
- Uses hidden markov models to determine combinatorial patterns of chromatin marks
  - H3K4me1
  - H3K27ac
  - H3K27me3
  - p300
- Input: data + number of “states” you want to identify
  - upper limit depending on number of conditions and chromatin marks
- Model re-tested on independent replicates





# *Which genes does Foxd3 regulate?*

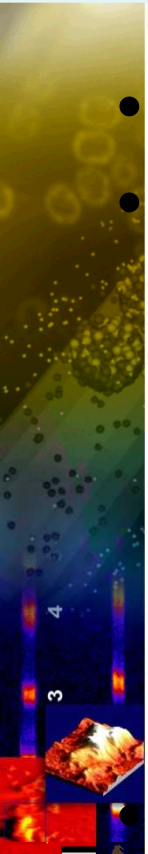
- Neighboring gene:
  - Didn't focus just on expressed genes, since Foxd3 is known to be a repressor
- In the future need to combine information from a number of data sets to determine cognate genes of regulatory regions:
  - Chromatin landscape
  - Gene expression
  - 3D interaction data
  - Functional experiments



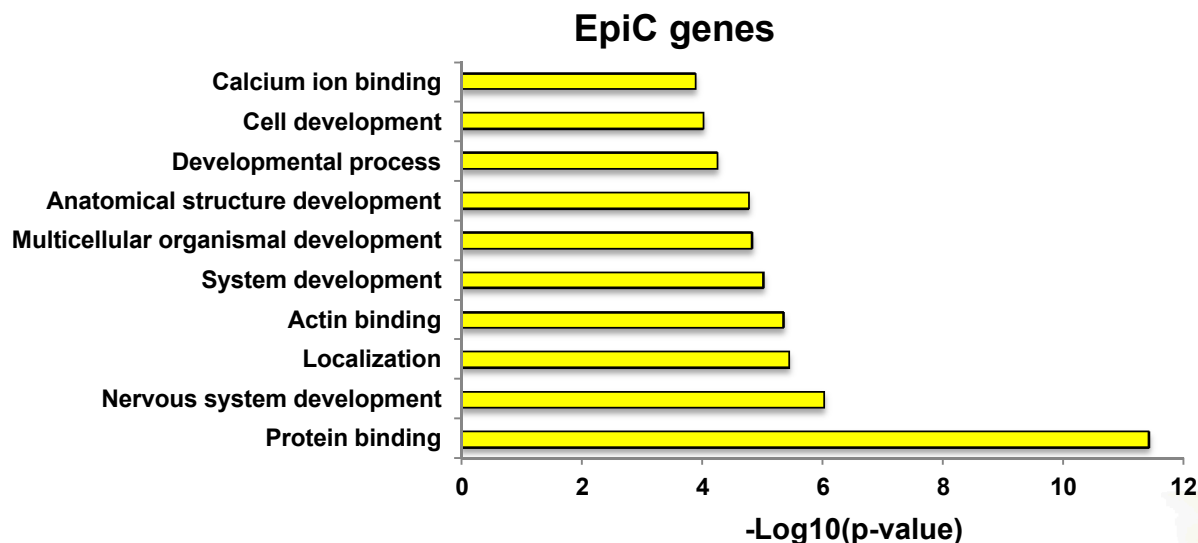
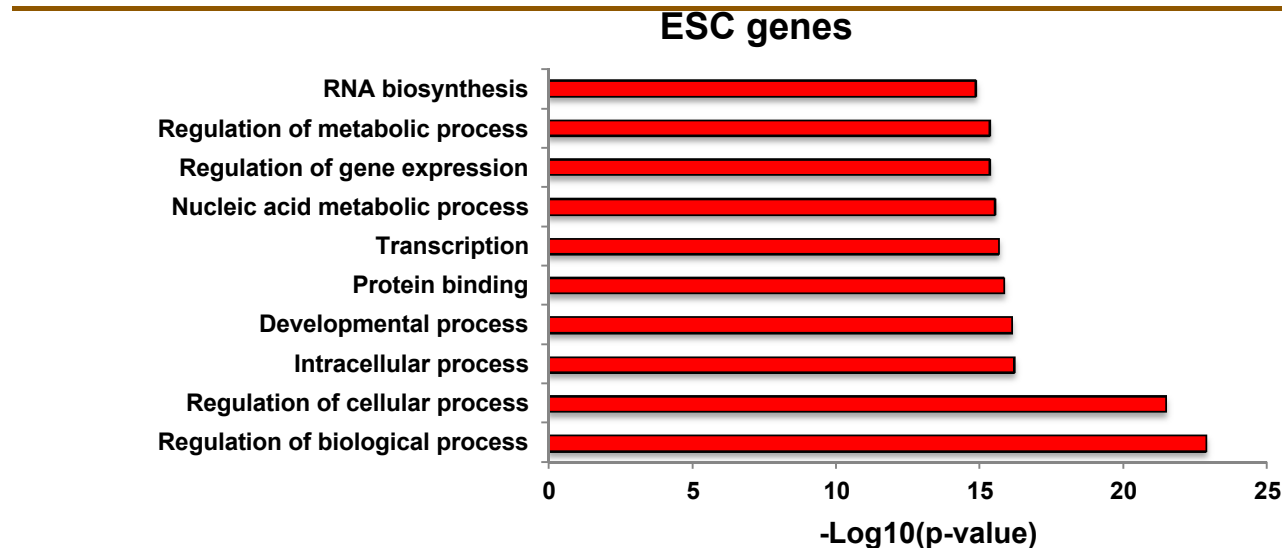
# Gene Ontology – linking genomic locations to biological function

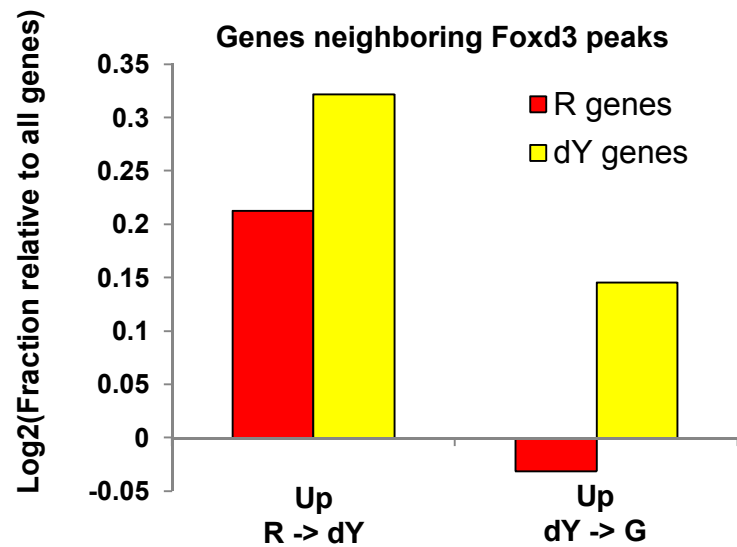
- Assigning function to binding – big challenge!
- Gene ontology:
  - Choosing the right background gene set
    - All annotated genes? Expressed genes? If array, genes on array?
  - Choosing the correct statistical test
    - Hypergeometric? Binomial? Chi-squared?
  - Correcting for multiple-hypothesis testing
    - Bonferroni, sidak, FDR?

Most methods yield similar but not identical results – showing trends vs follow-up on specific conclusions – **GoSTAT (Beissbarth and Speed 2004)**



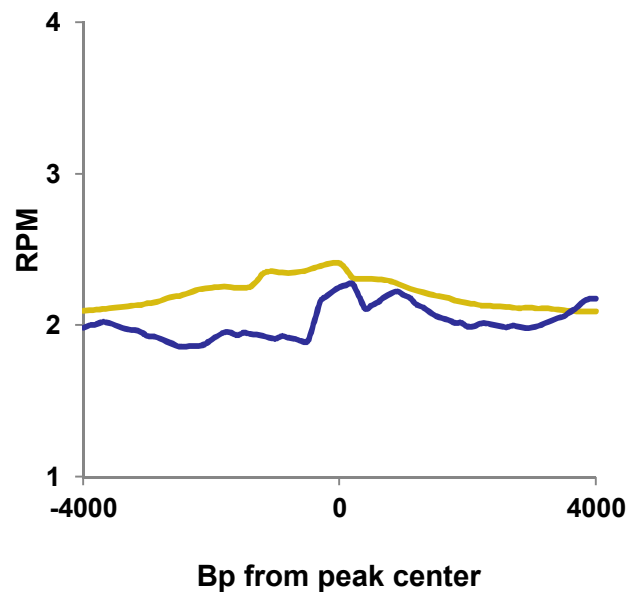
# Genes near *Foxd3* binding sites are involved in distinct pathways in ESCs and EpiCs



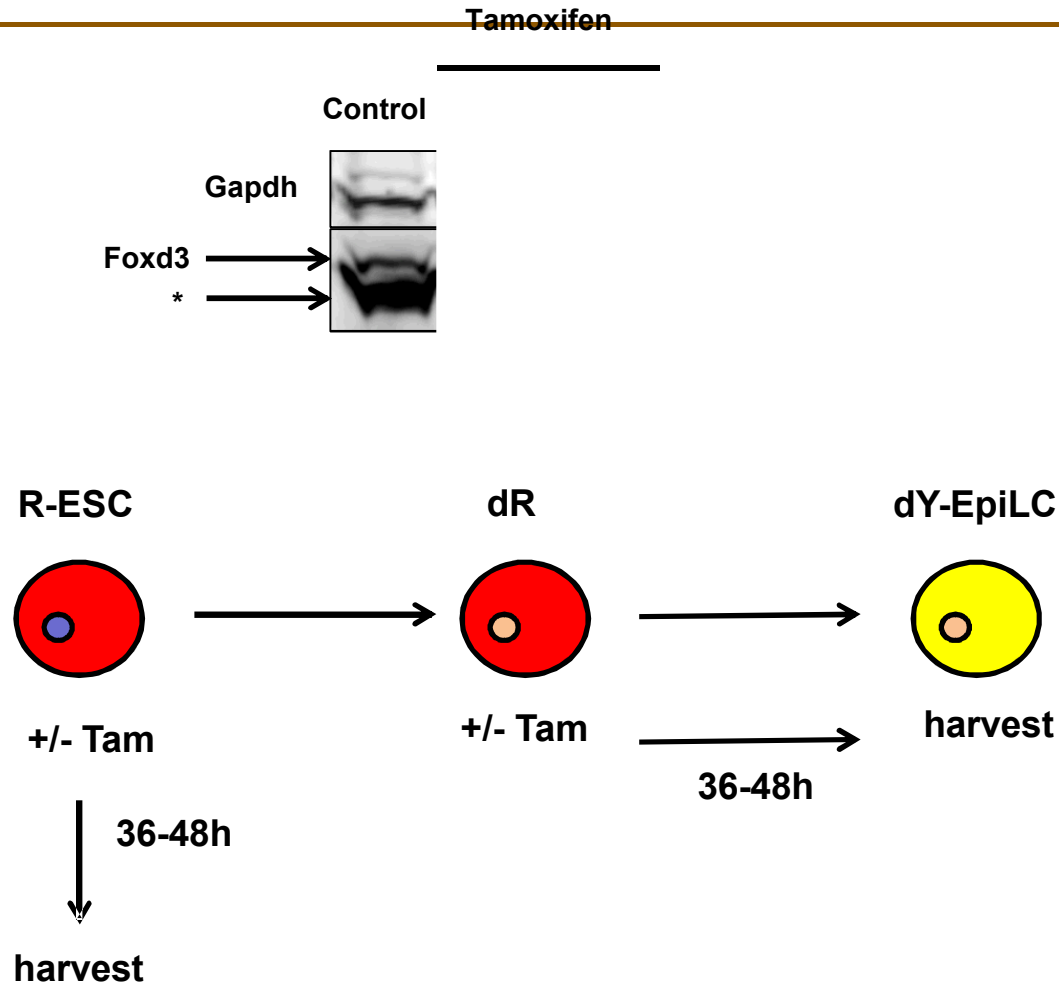




### ChIP-seq: H3K27ac

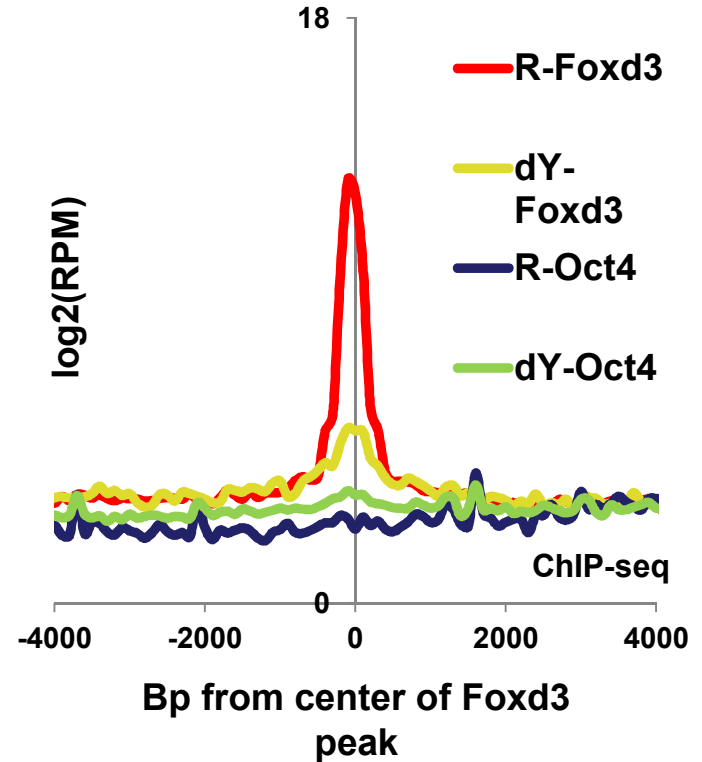
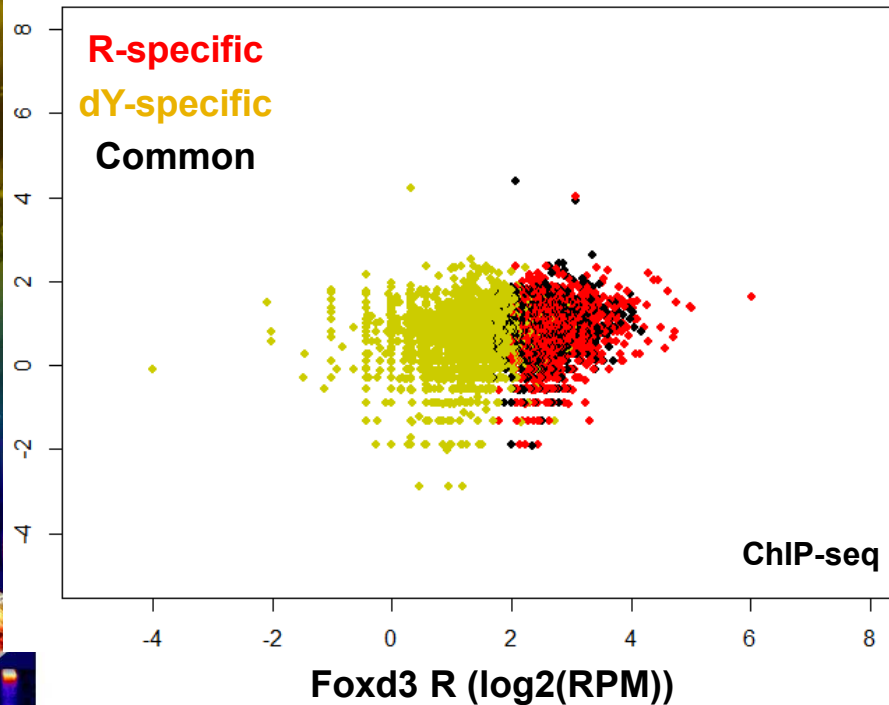


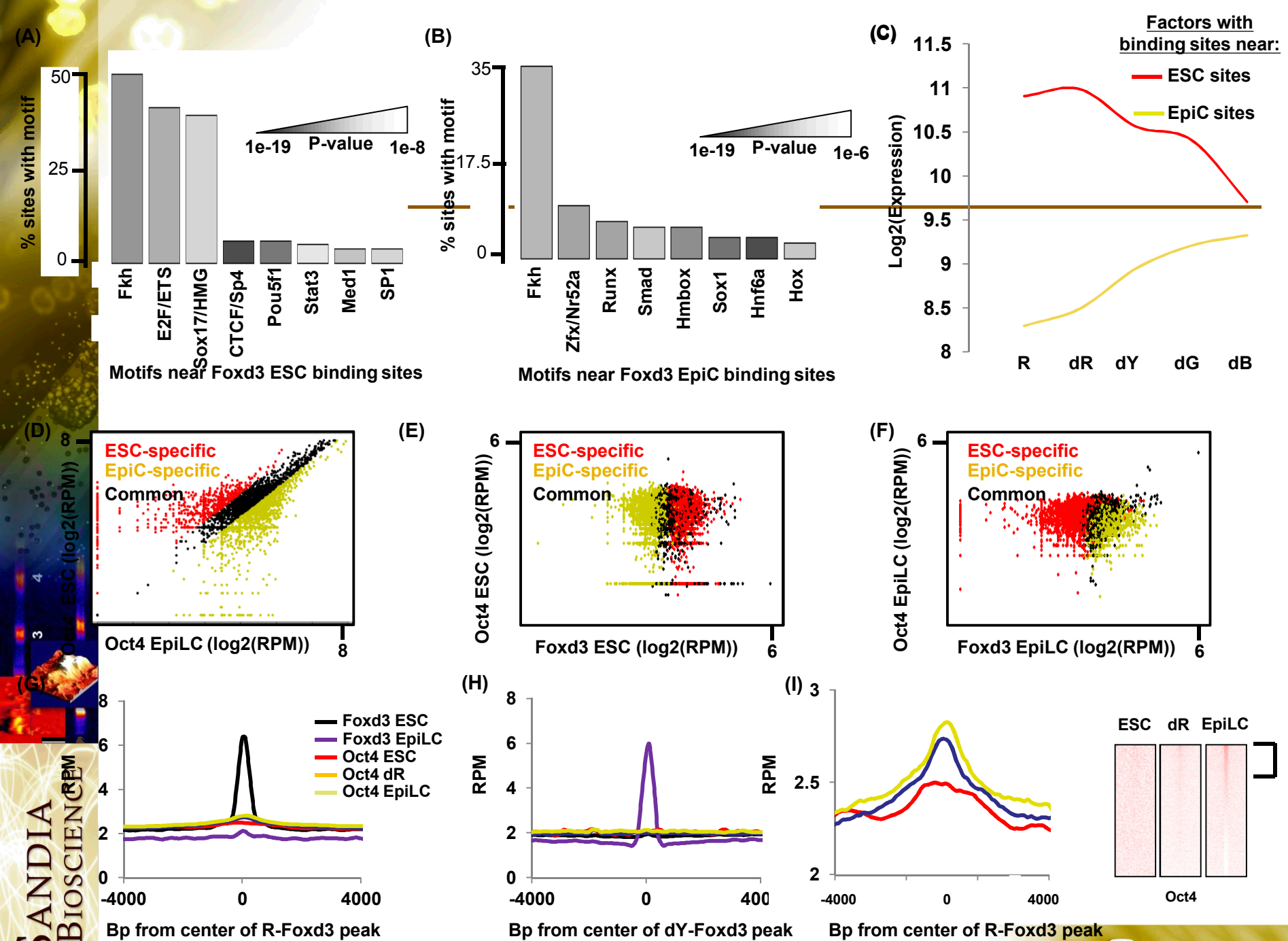
# Foxd3 knockout ESCs and EpiLCs





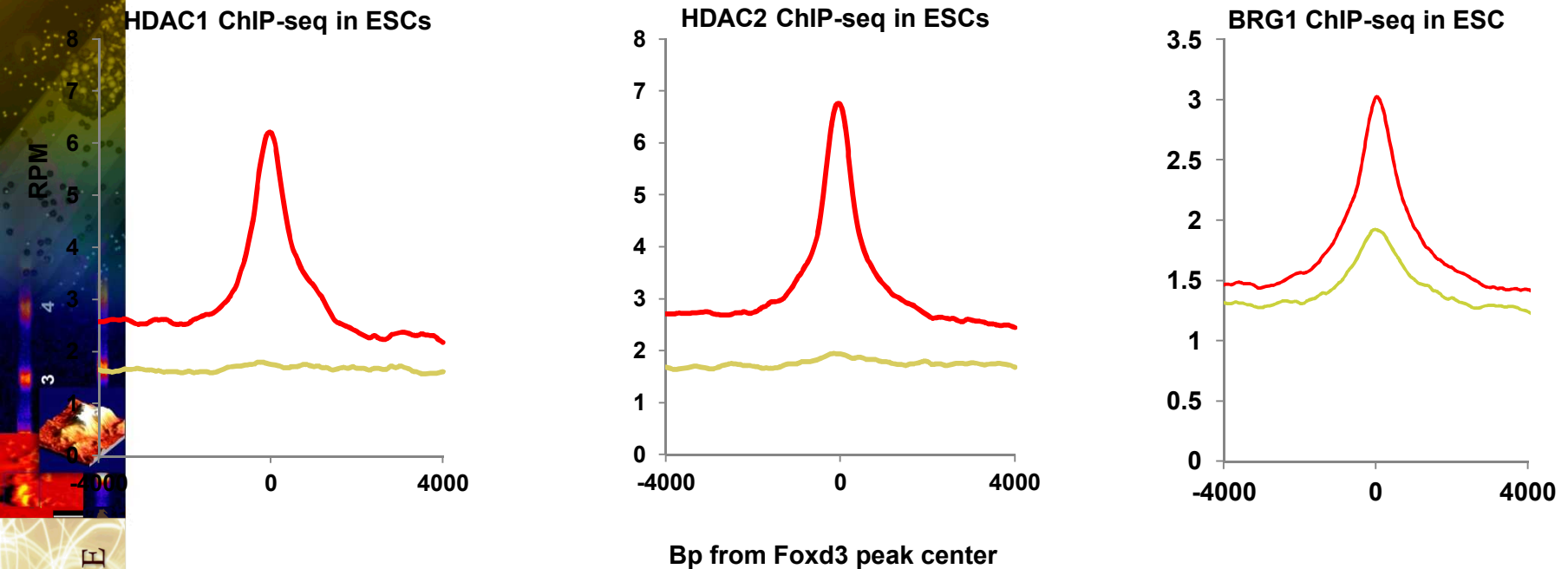
# Foxd3 does not co-localize with Oct4





# Brg1 and HDAC1/2 globally co-localize with Foxd3 in ESCs

— ESC Foxd3 peaks — EpiLC Foxd3 peaks

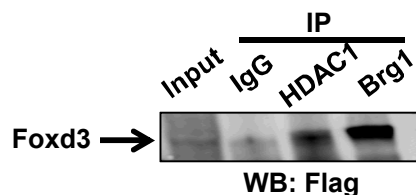
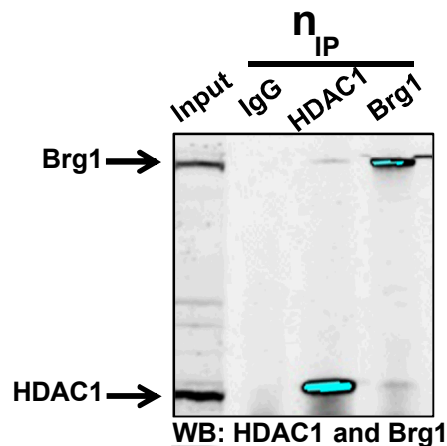


# *Foxd3 is in a complex with Brg1 and HDAC1*

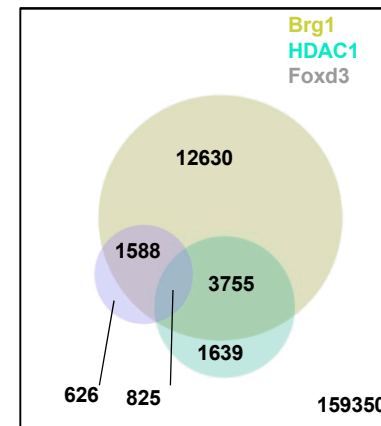
H3K27 deacetylation – HDAC1/2

Chromatin remodeling – Brg1 (SWI/SNF complex)

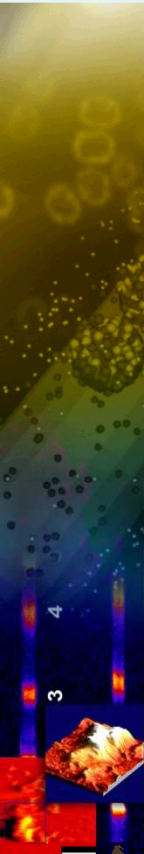
Immunoprecipitation



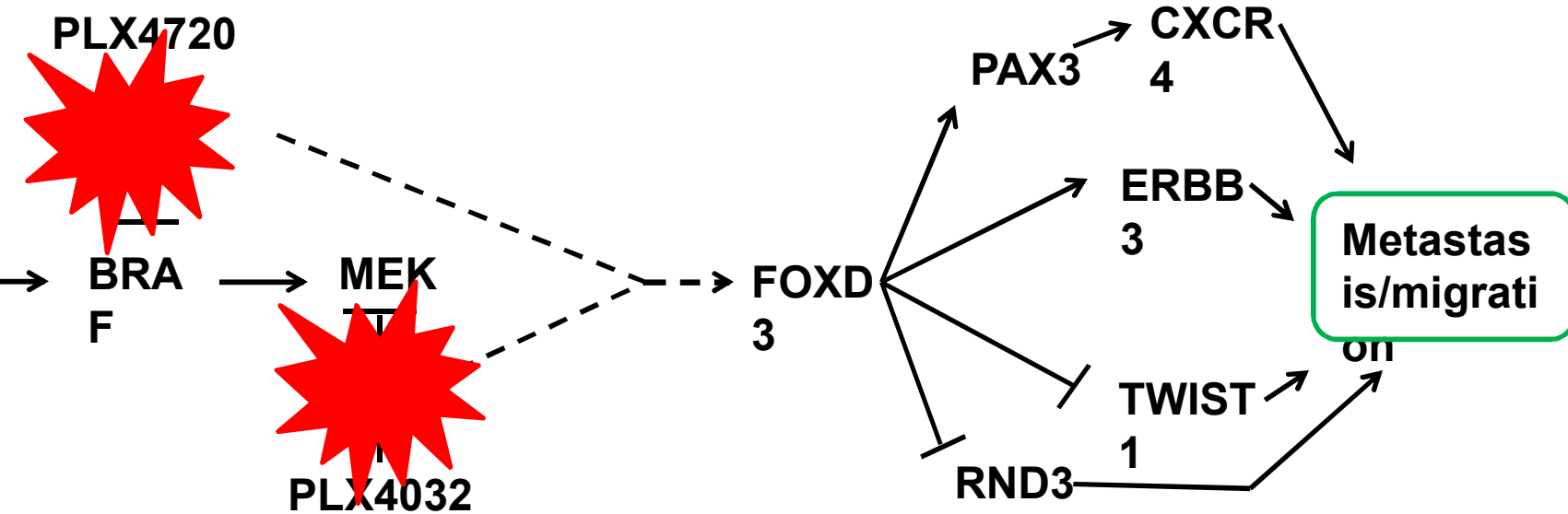
Overlap of ChIP-seq peaks



Chi-squared test p-value < 2.2e-16



# Example - Foxd3 expression regulates cell fate in melanoma



Katiyar and Aplin 2011 (Mol Cancer Res)

Kubik et al 2015a, b (JBC)

Weiss et al 2014 (Mol Cancer Res)