

# Understanding Receptor Regulation for Designing, Targeting and Delivering Novel Therapeutics

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Adrienne C. Greene, Ph.D.

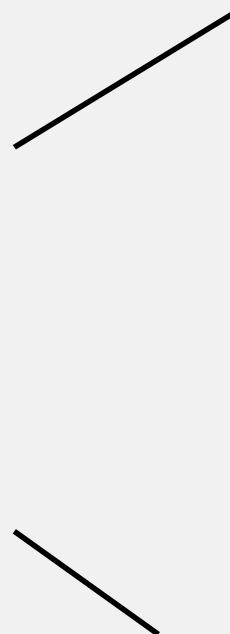
Sandia National Laboratories

Case Western Reserve University

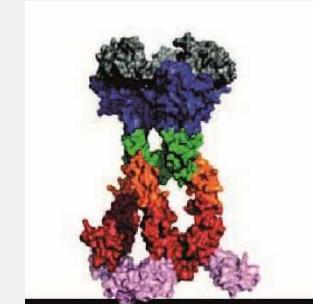
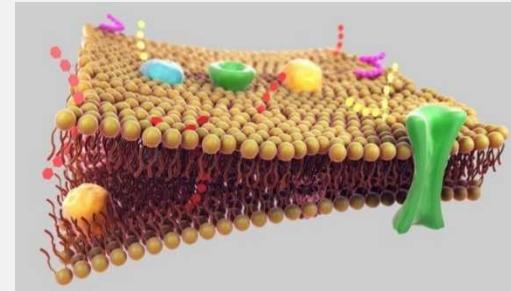
12/05/2016

# Understanding Regulatory Mechanisms of Biological Systems

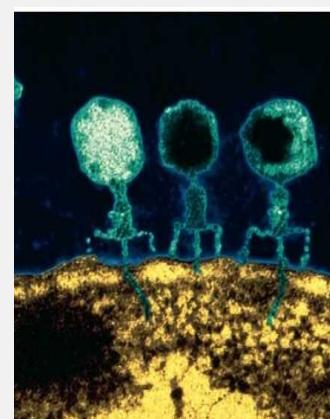
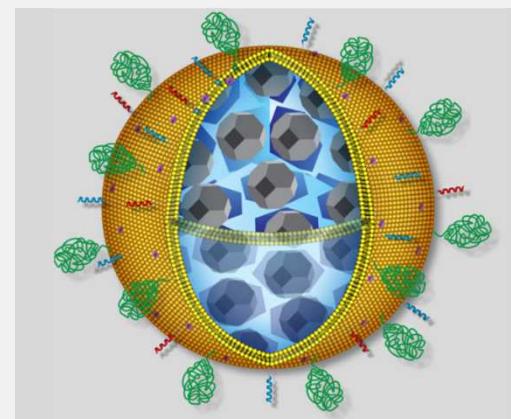
Goal: Understand biological systems for applications-based technologies (therapeutic target design, diagnostic tools, biosensing and/or bioengineering technologies)



1. Receptor Signal Regulation

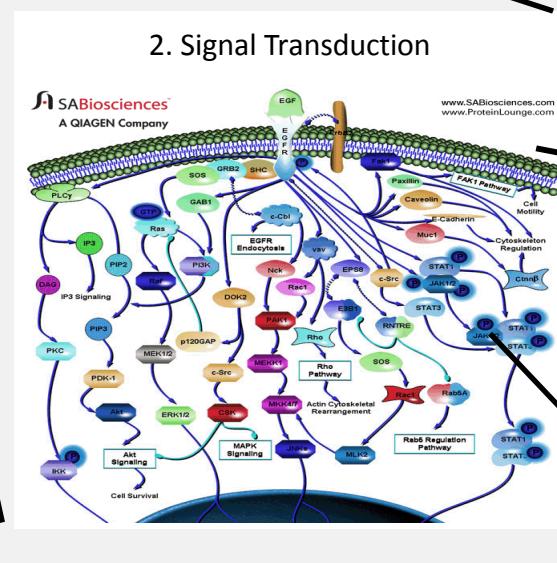
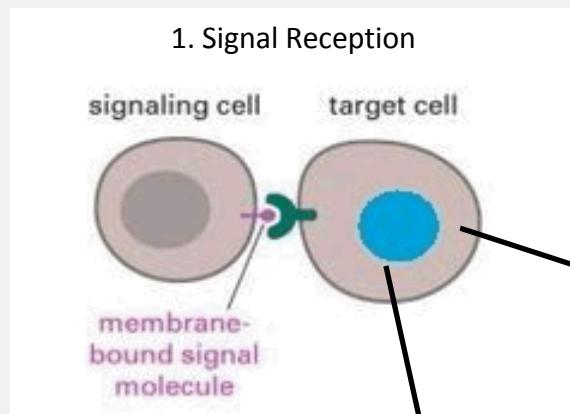


2. Targeted Therapeutic Delivery

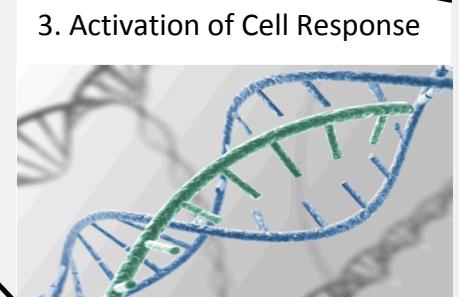


# Complex Regulation of Cell Signaling

One mechanism by which cells communicate with each other is through direct physical contact

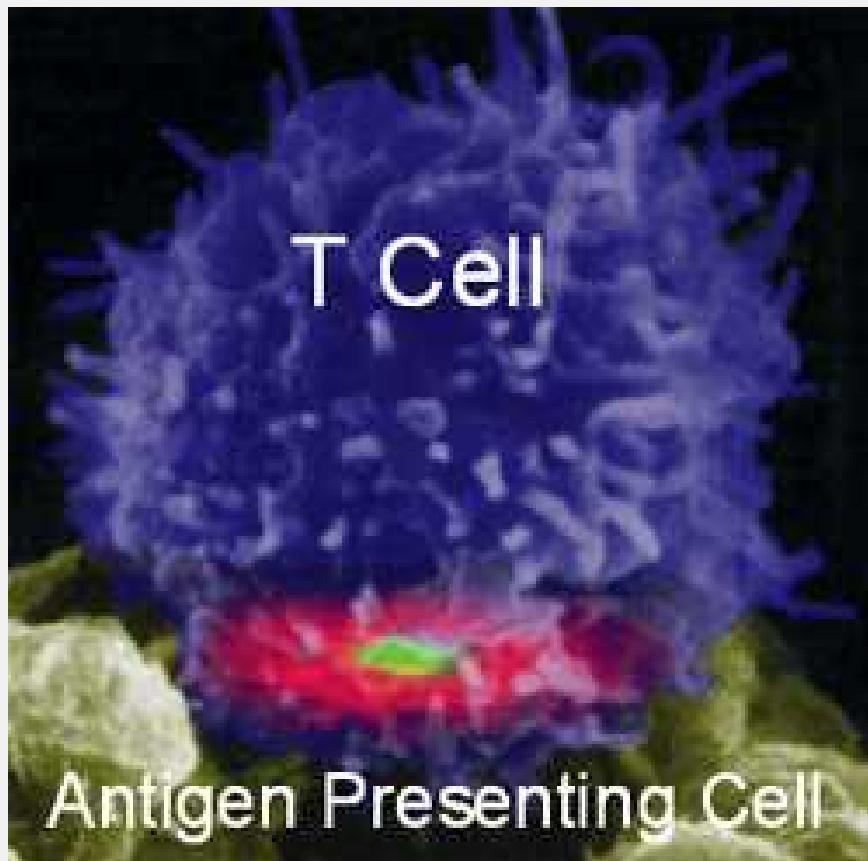


- Cell signaling is “noisy”
  - A single cell input results in a complex cascade of intracellular responses
  - Typically, there are many simultaneous signal inputs, resulting in highly stochastic cell response

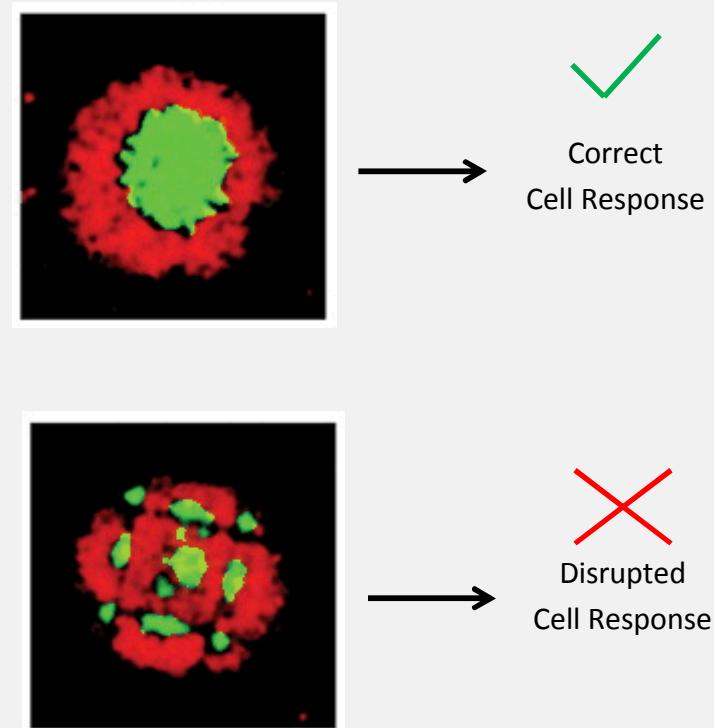


# Clustering at a Cell-Cell Interface Regulates Signaling

Receptor-ligand organization (clustering) at a membrane is a mechanism for cells to overcome biological noisy signaling environments and impart a downstream signal appropriately



<http://www.bme.columbia.edu/~kam/research/research.htm>

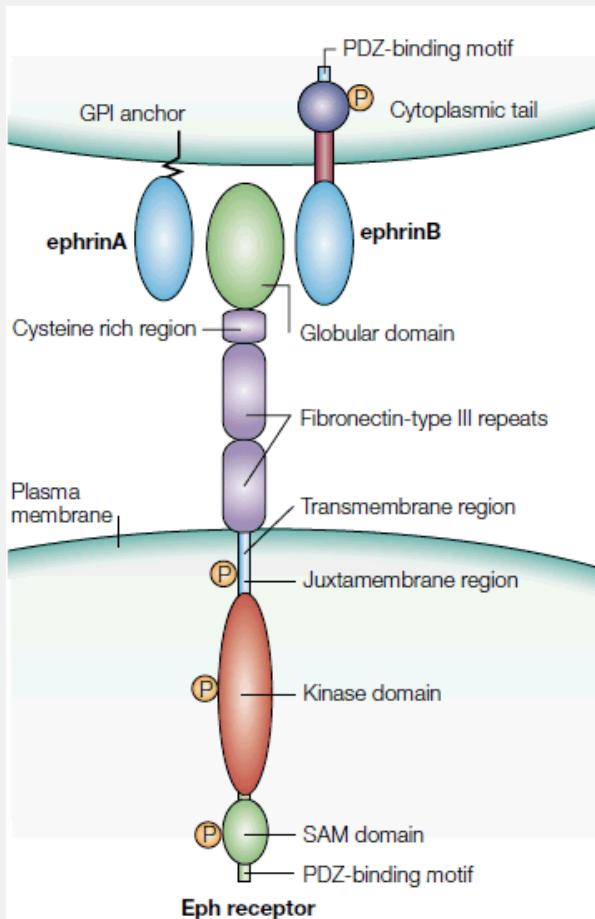


Same # of Signal Inputs!

# Eph Receptor Tyrosine Kinase Signaling

Receptor: EphA2

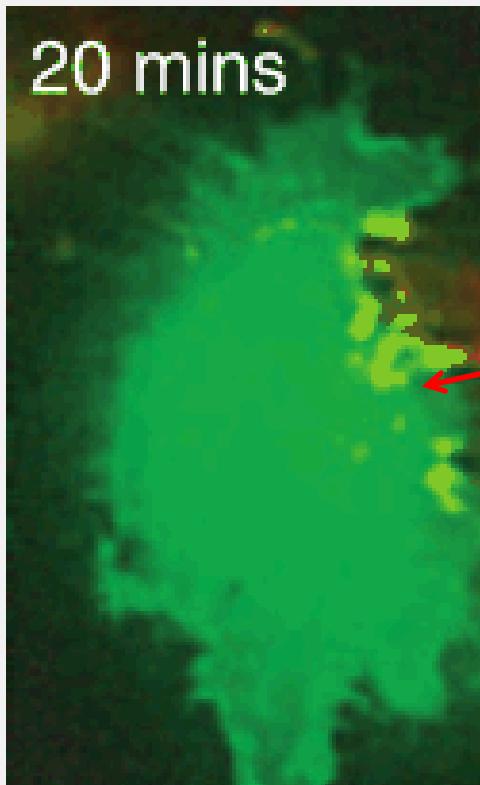
Ligand: ephrinA1



- Bidirectional signaling
  - Bidirectional signaling
- Overexpressed in aggressive cancers
- Highly overexpressed in MDAMB231 cells: invasive and metastatic breast cancer cells
- *Often expressed in triple negative breast cancers (no ER, PR or Her2)*
  - Key therapeutic target

# Activation of Eph Receptors by ephrin Ligands

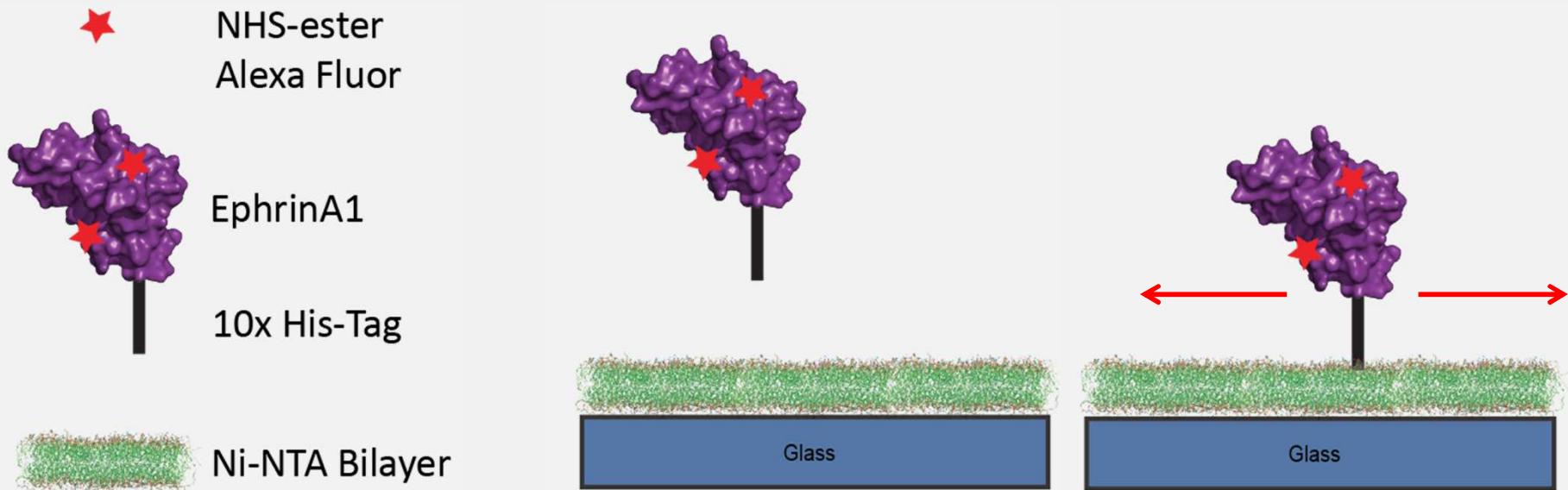
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- Typical Eph receptor activation assays use:
  - co-cultures
  - soluble ephrin
- Forward EphA2 signaling (and likely clustering) must be important in the context of a membrane
- Need to use membrane-bound ephrin ligand to more accurately understand Eph receptor forward signaling

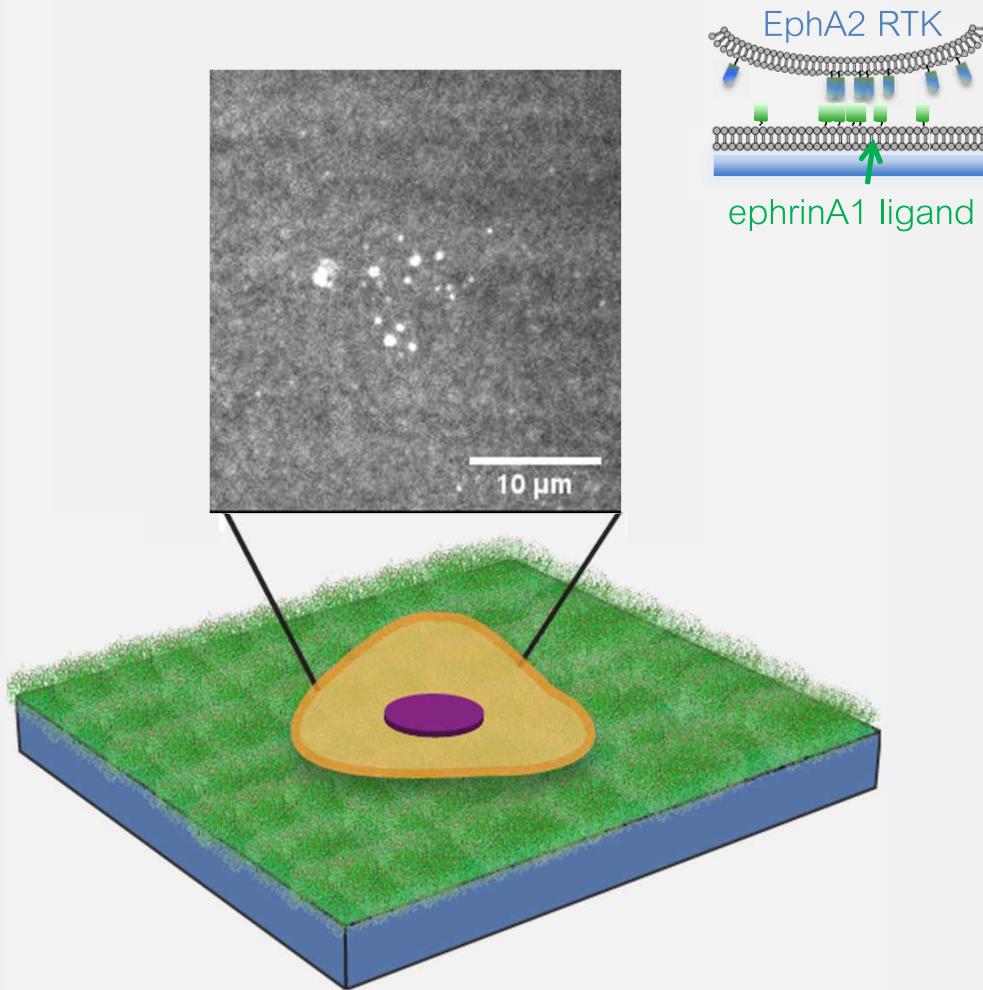
# Linking ephrin Ligands to a Supported Lipid Membrane

Ephrin ligands can be purified and linked to a supported lipid membrane, creating a synthetic mimic of the ephrin-expressing cell



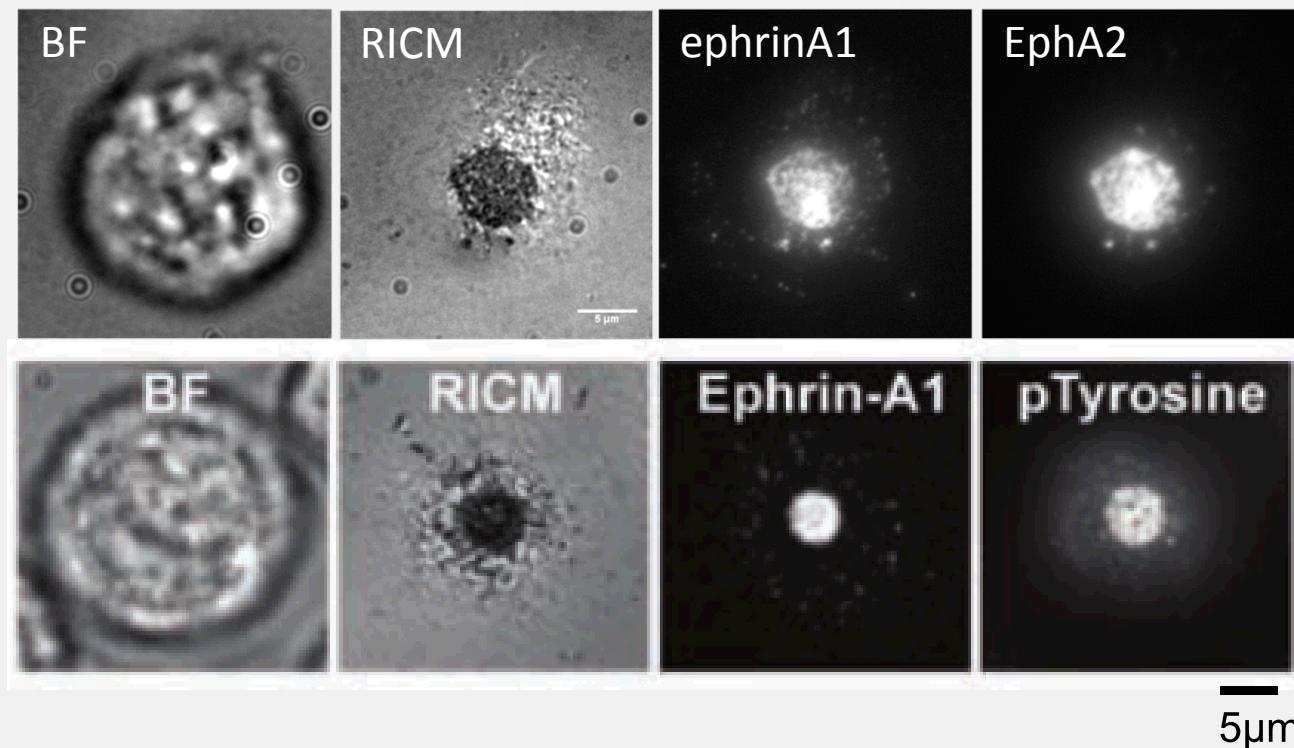
# Receptors and Ligands Cluster at the Cell-Membrane Interface

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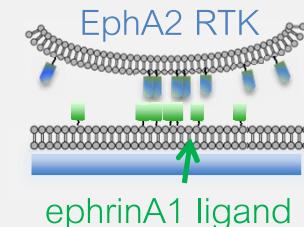


- MDAMB231 cells are seeded onto an ephrinA1 bilayer
- Receptor-ligands undergo higher ordering reorganization at the cell-membrane interface
- High resolution microscopy can be used to probe the importance of spatial organization

# Membrane-Bound Ligands Activate Cell Receptors



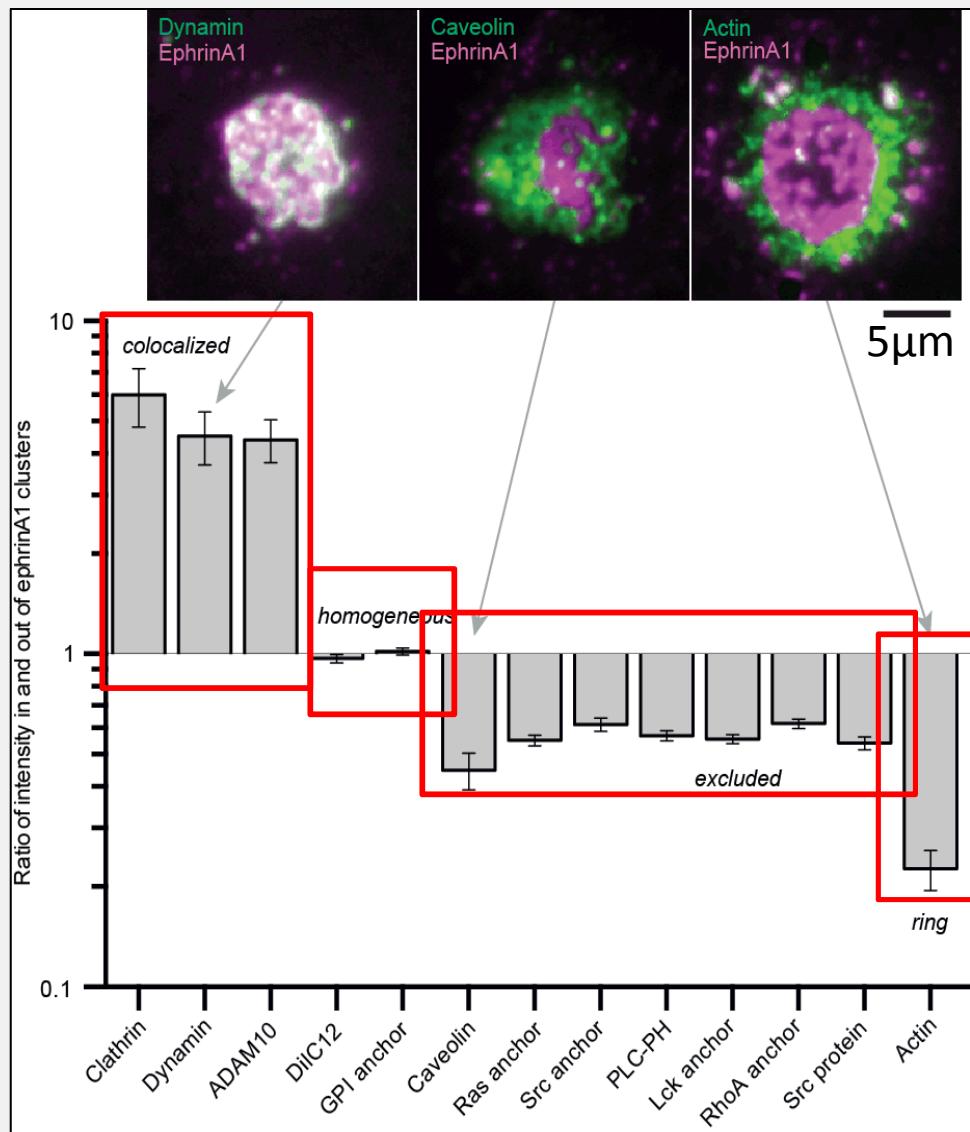
- Ephrin ligands on a supported membrane activate Eph receptors
  - Causes Eph receptors to be phosphorylated
  - Activates an actomyosin-driven contractility



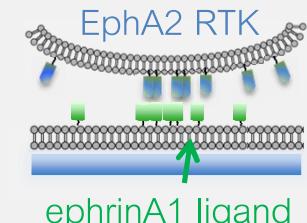
Salaita, K., et al. 2010. *Science*. 327(5971): 1380-5.

Xu, Q., et al. 2011. *Biophys J*. 101(11): 2731-9.

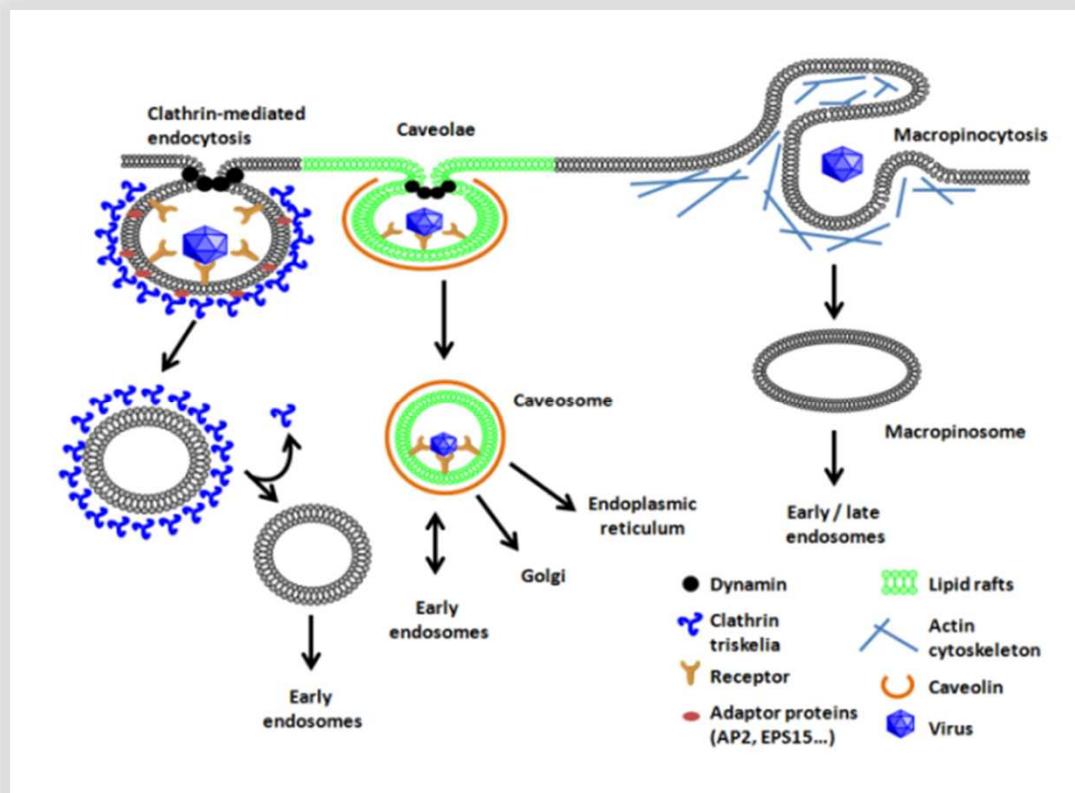
# EphA2-ephrinA1 Clusters Contain Endocytosis Molecules



- Localization of signaling molecules to ephrinA1 was measured
- Four types of localization were characterized
  - Colocalization
  - Homogenous distribution
  - Anti-localization
  - Ring formation
- Maybe these clusters are important sites of endocytosis?

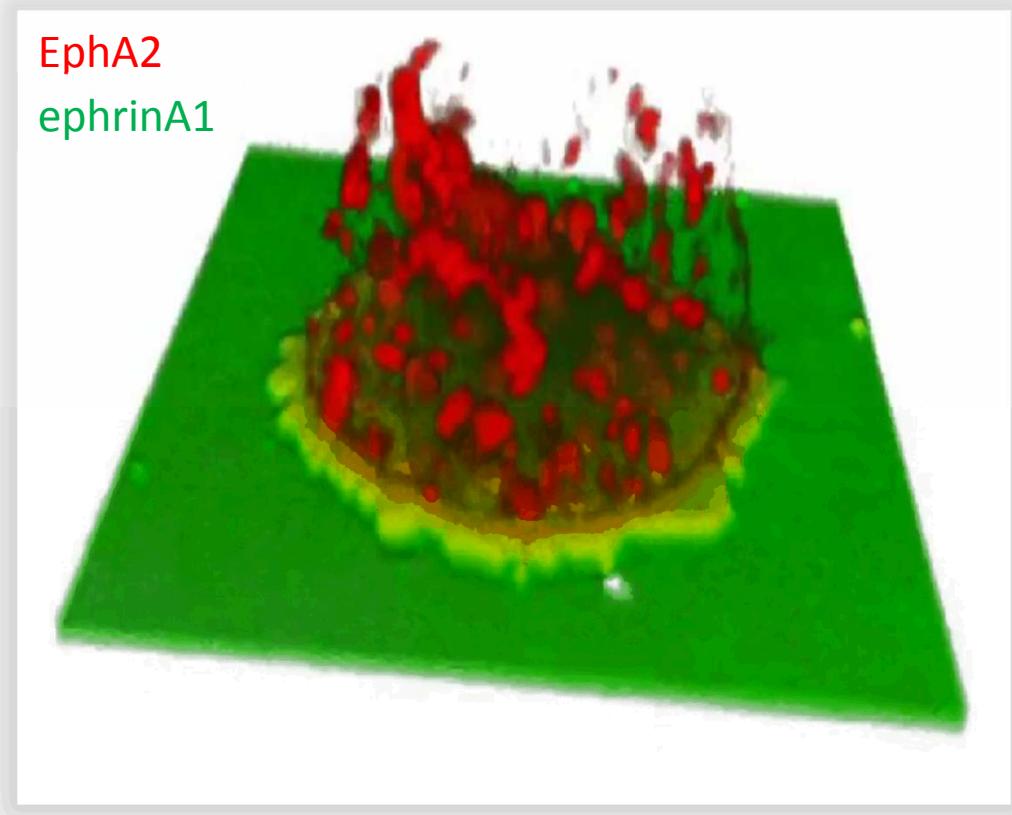


# Clusters are Likely Sites of Clathrin-Mediated Endocytosis



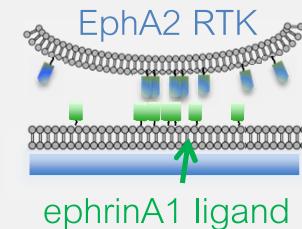
- Endocytosis is a mechanism to downregulate cell signaling, resensitize cells to signaling, recycle receptors and/or degrade signaling molecules
- Perhaps EphA2-ephrinA1 clusters are large sites of clathrin-mediated endocytosis (CME)

# Detecting Changes in EphA2 Endocytosis is Challenging



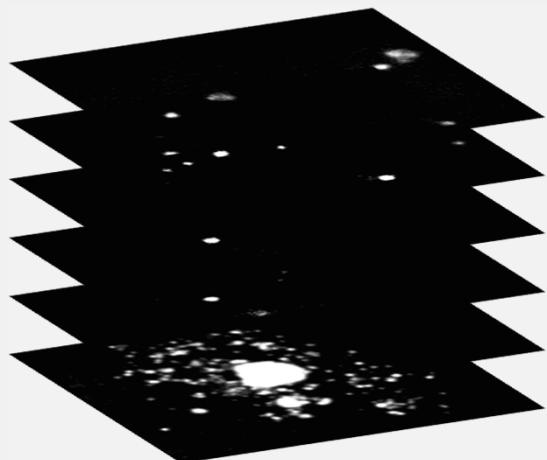
## Problems:

- Non-specific labeling
- Lots of background fluorescence
- Consistency with permeabilization and/or labeling
- Always a high background level of internal EphA2

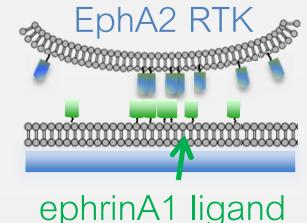


# Developing a Live-Cell Endocytosis Assay

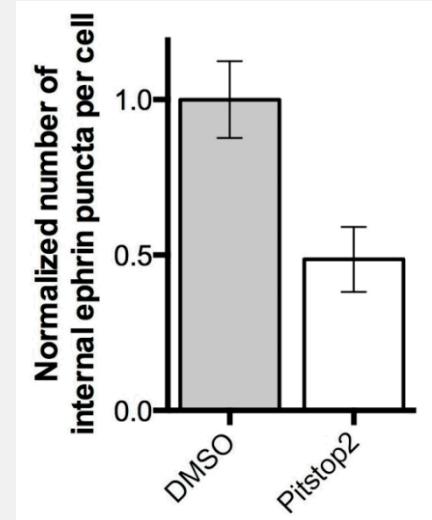
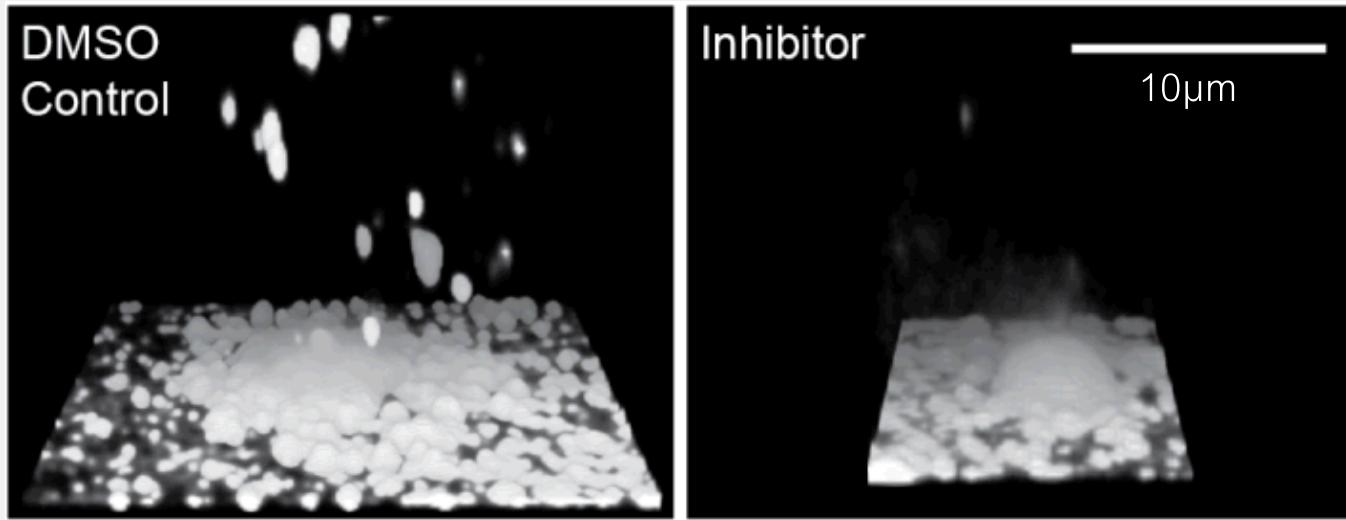
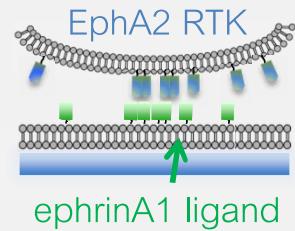
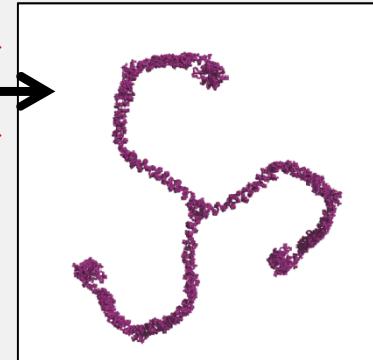
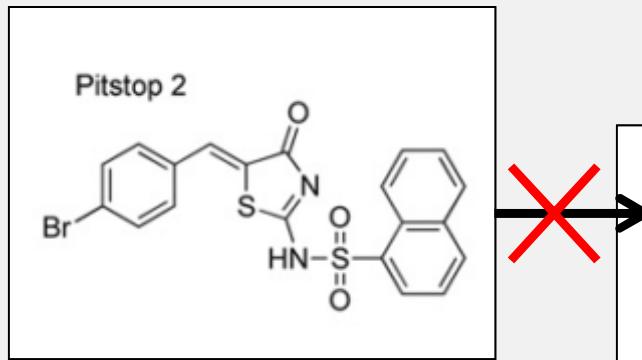
0.5μm Z-slices



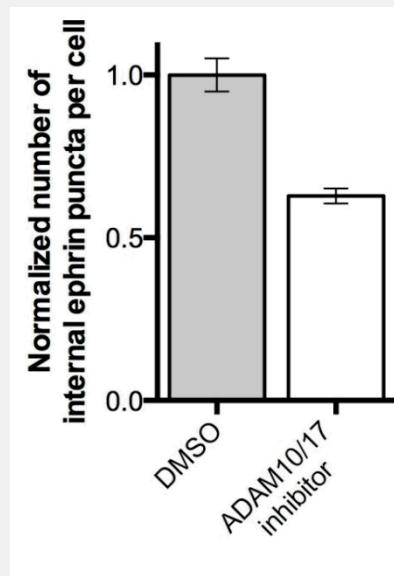
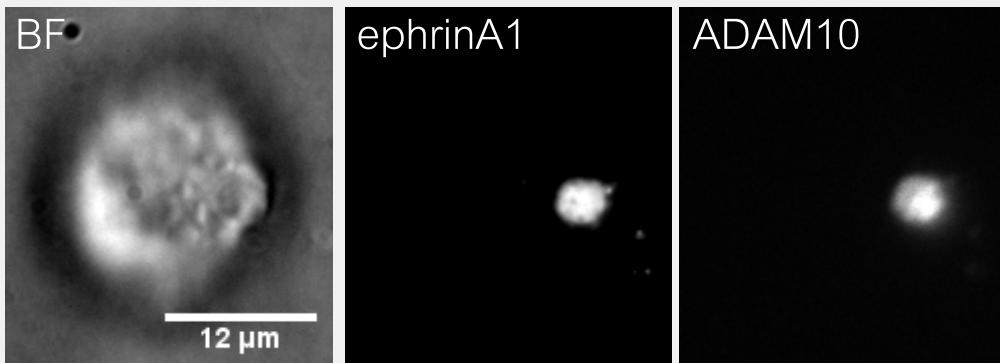
- Detect internal ephrinA1 using spinning disc confocal microscopy
- All signal must have come from the bilayer, so high signal to noise!
- Detection of small changes, very quantitative, cleaner/more reproducible assay than antibody staining
- Easy background threshold
- Selects for specific punctate spots using a size threshold



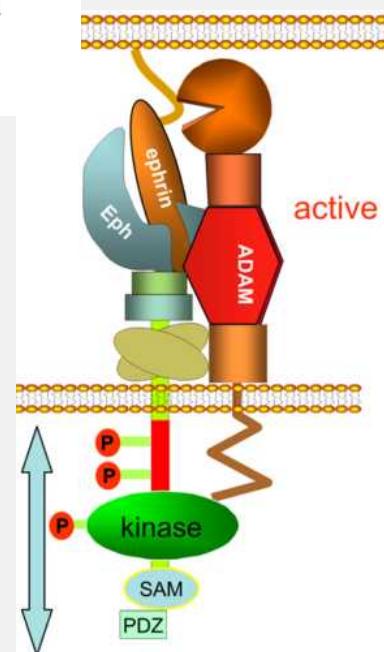
# Trans-Endocytosis of ephrinA1 Requires Clathrin



# Trans-Endocytosis of ephrinA1 Requires ADAM10

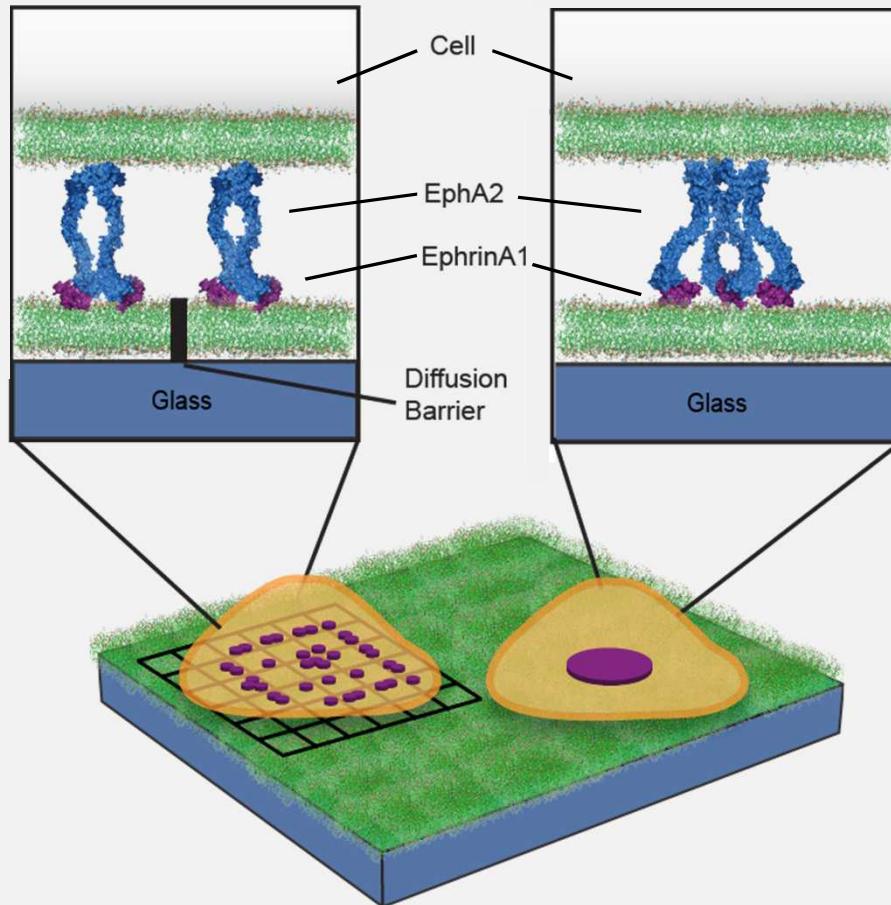


- ADAM10 is a disintegrin and metalloprotease thought to be responsible for the trans-cleavage of ephrinA1
- ADAM10 is necessary for efficient ephrinA1 endocytosis
  - ADAM10/17 was inhibited with a small molecule

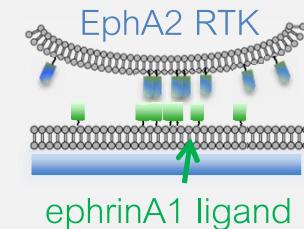


# Using Diffusion Barriers to Alter Clustering: Creating a “Spatial Mutation”

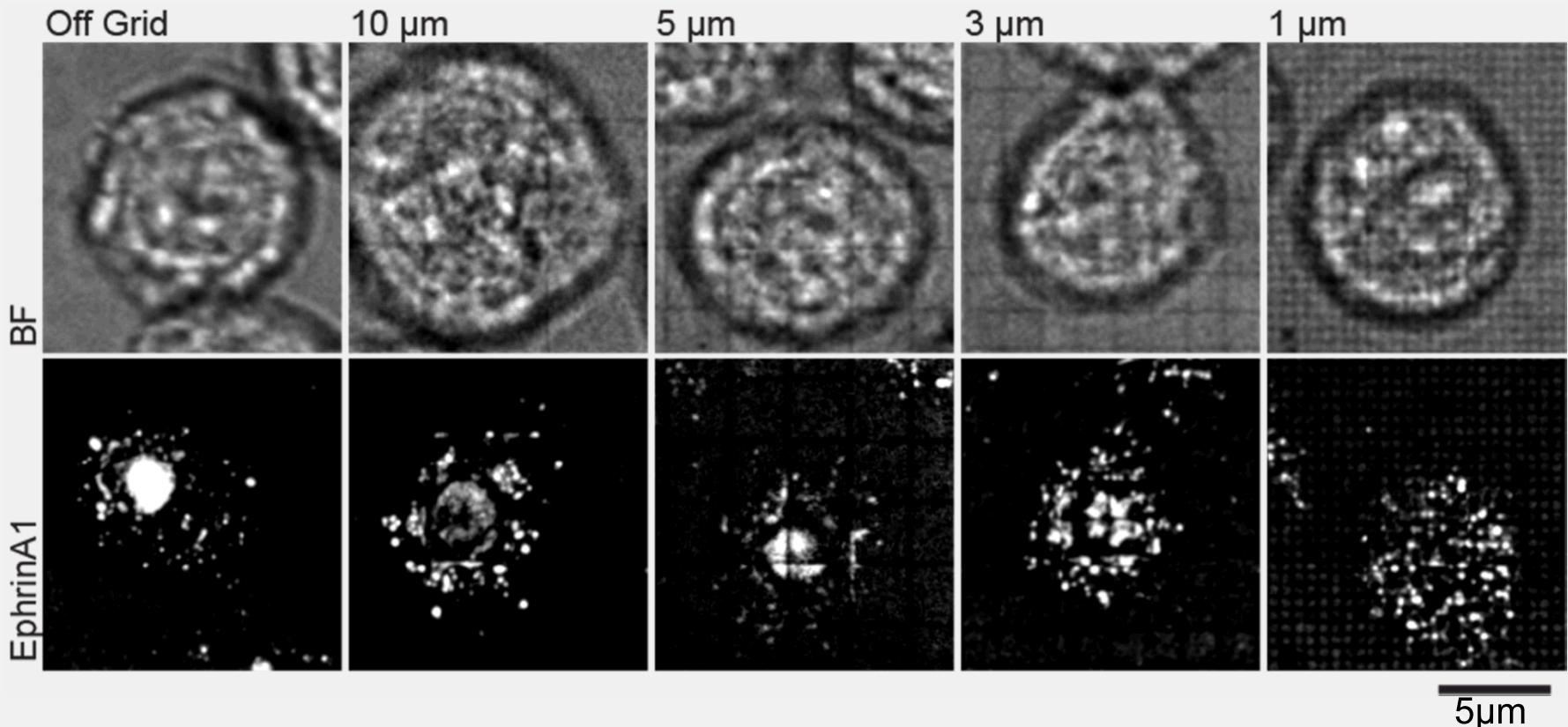
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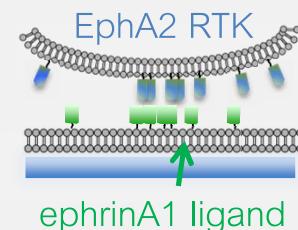
- Using electron beam lithography, diffusion barriers can be created to restrict receptor-ligand mobility
- This assay allows us to probe the importance of EphA2-ephrinA1 reorganization in the context of downstream signaling



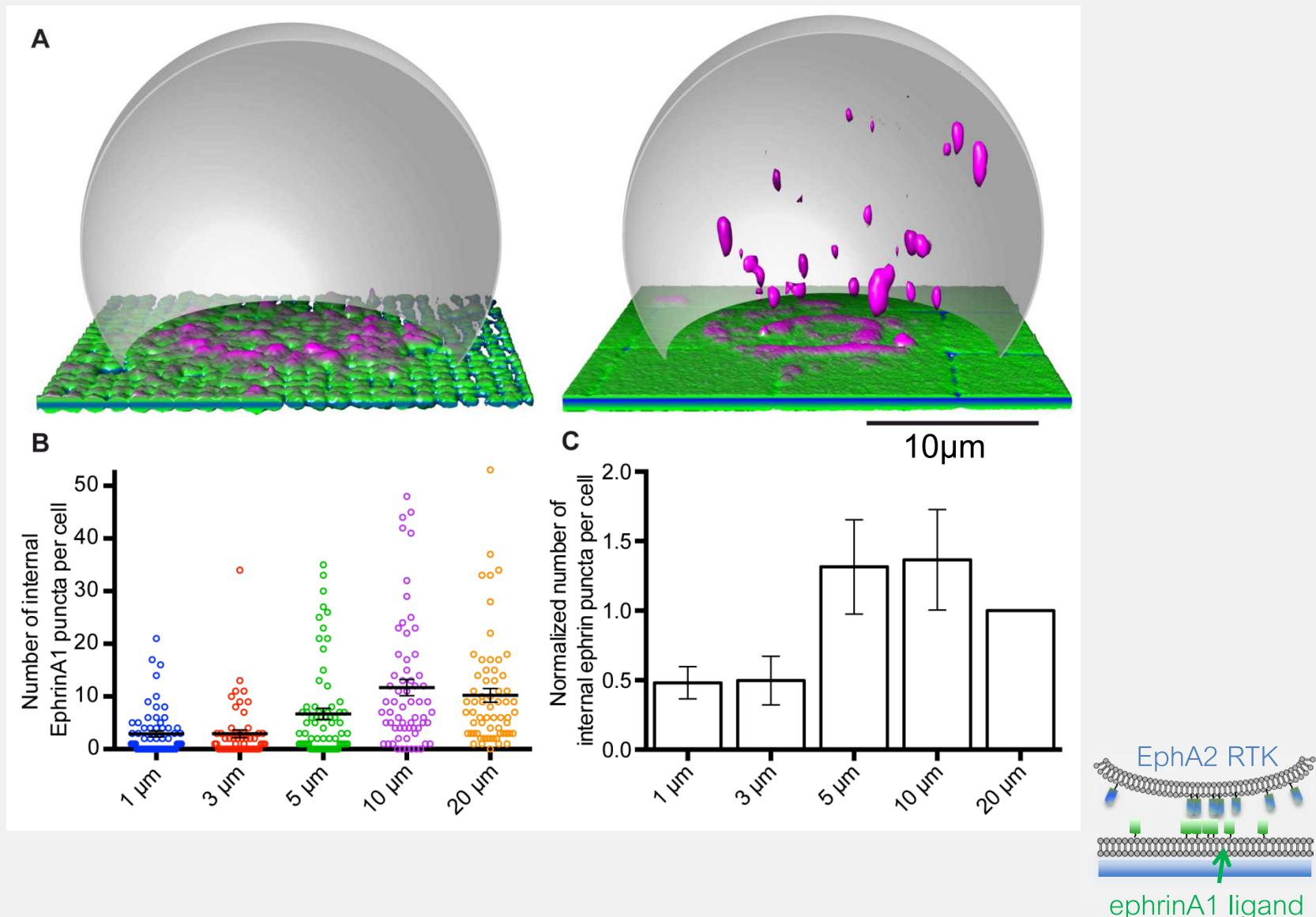
# Diffusion Barriers Alter EphA2-ephrinA1 Endocytosis



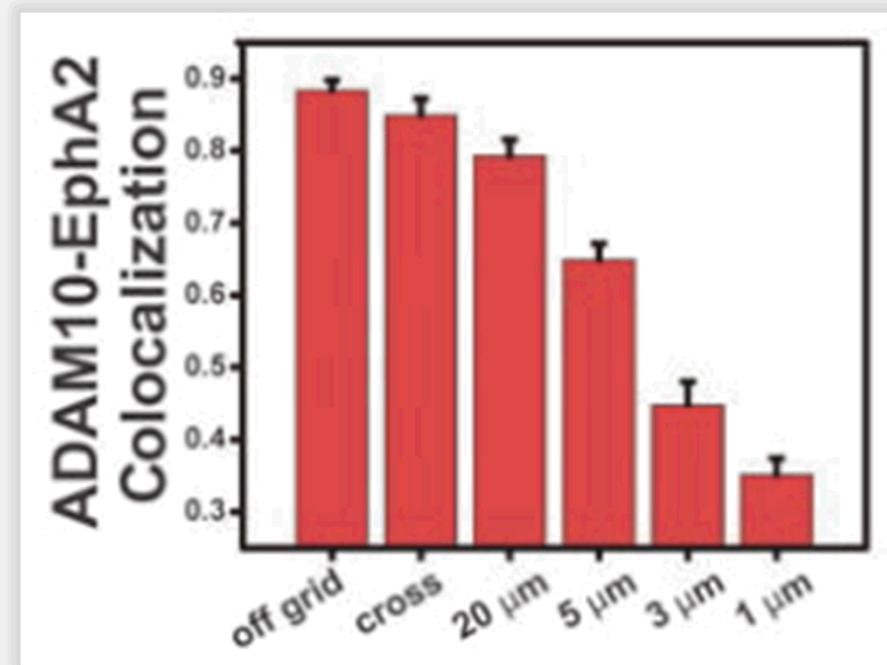
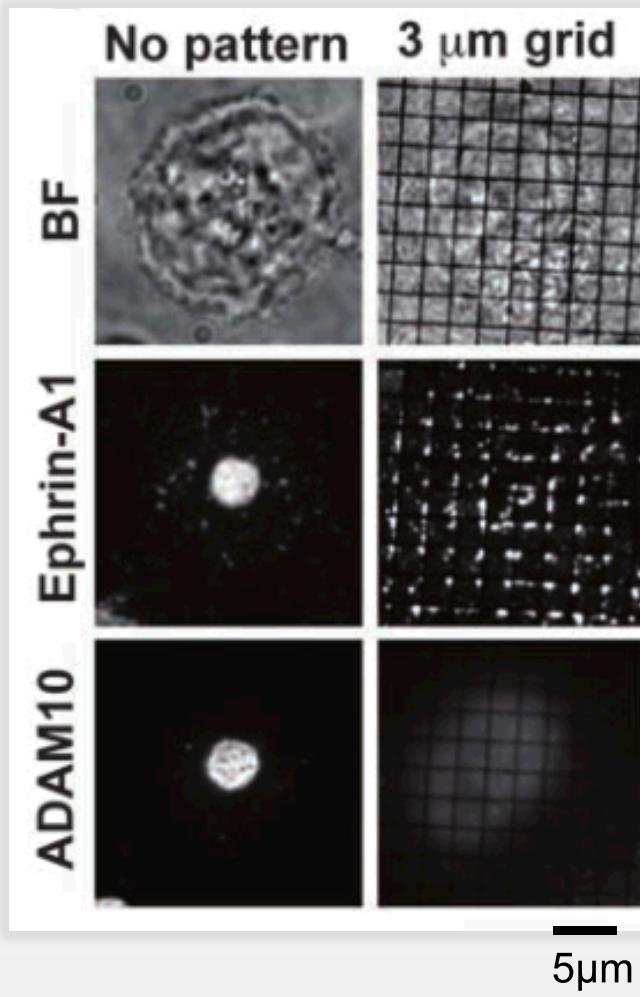
- Imaged using TIRF microscopy
- Only clustering and reorganization is altered
- Chemical composition remains the same



# Trans-Endocytosis of ephrinA1 is Altered on Grids

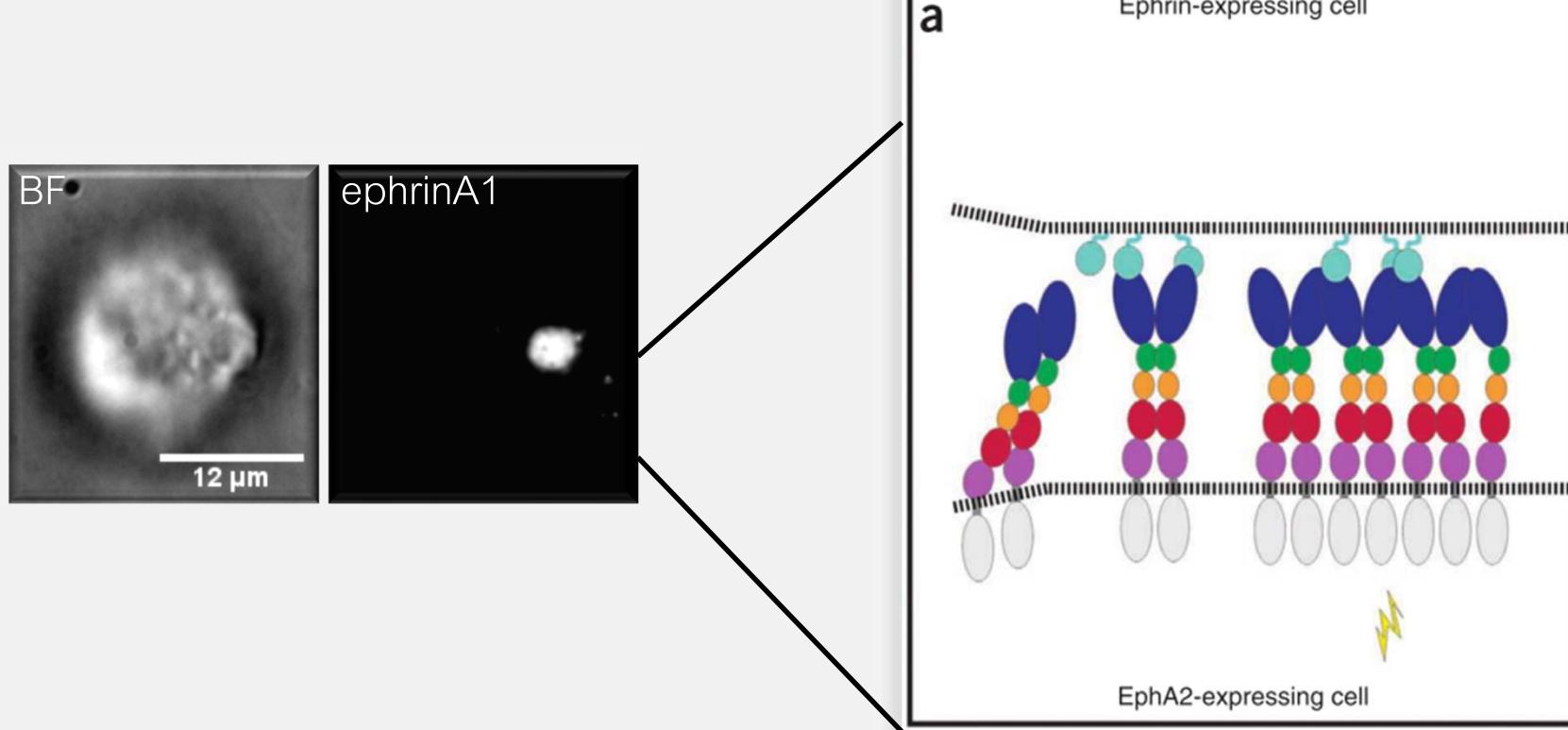


# Recruitment of ADAM10 is Decreased on Grids



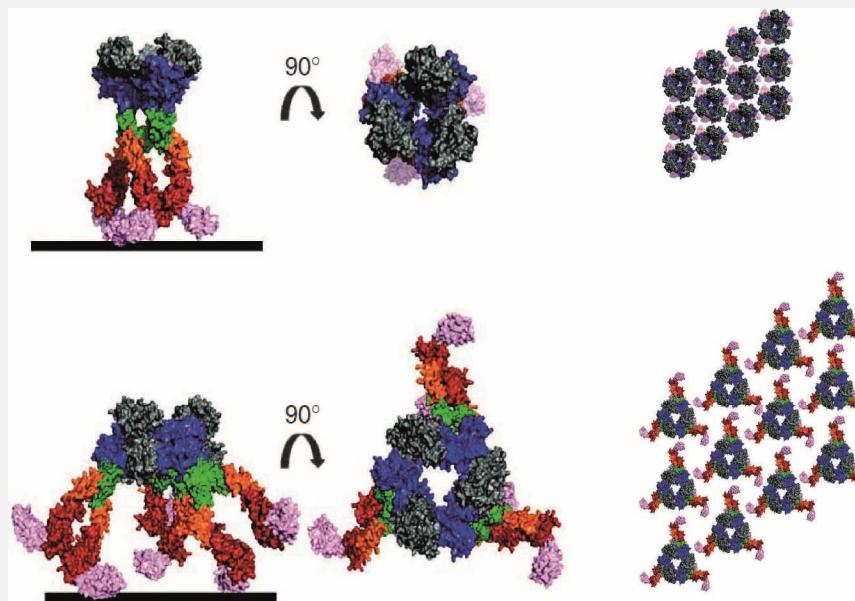
# Structure and Function of EphA2 Clustering

*How does clustering from the micron down to the nanoscale regulate signaling?*



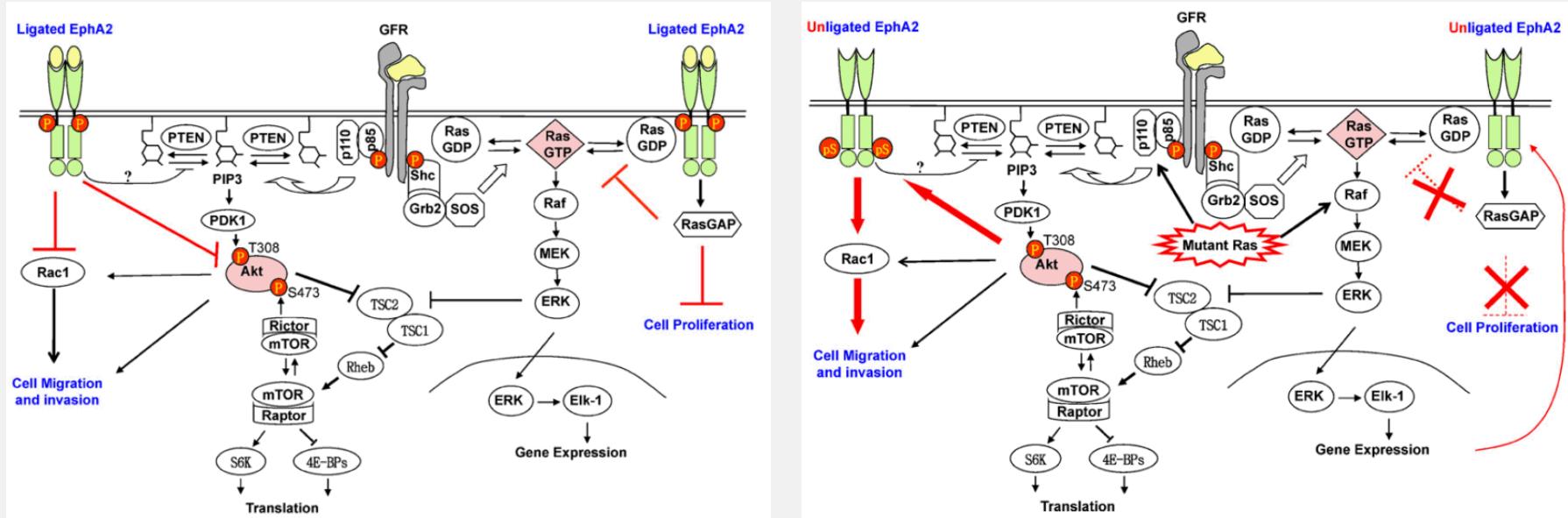
# Molecular Architecture of EphA2 Clustering

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- EphA2 forms large-scale oligomers
  - Both in *cis* with other EphA2 receptors and in *trans* with ephrinA1 ligands
- We need to understand more about Eph molecular structure and clustering and how that alters signaling

# EphA2 Signals in a Ligand-Dependent and Independent Manner



- Requires fine-tuning the balance of signaling based upon ligand-dependent and ligand-independent signaling; also context and cell type specific
- How Eph clusters (i.e. the stoichiometry of receptor/ligand binding) also changes this signaling map

# Why Do We Care About EphA2 Clustering?

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Invest New Drugs (2013) 31:77–84  
DOI 10.1007/s10637-012-9801-2

## PHASE I STUDIES

### Phase 1, open-label study of MEDI-547 in patients with relapsed or refractory solid tumors

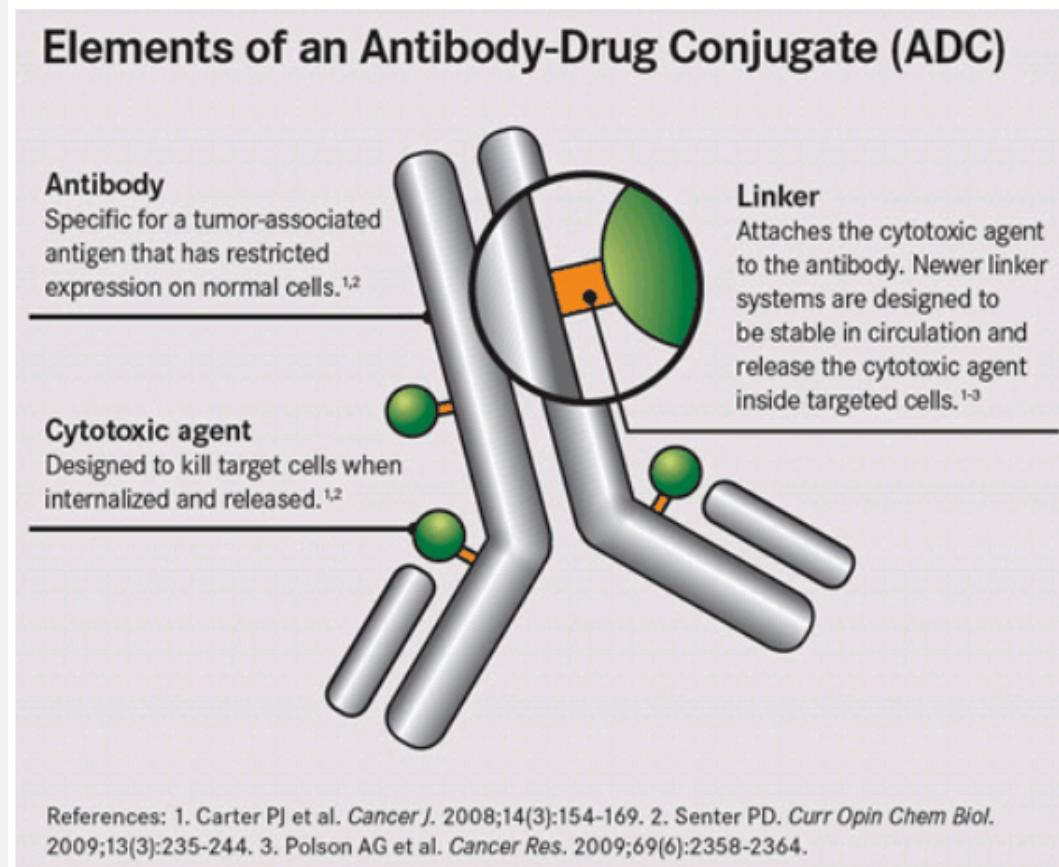
Christina M. Annunziata · Elise C. Kohn ·  
Patricia LoRusso · Nicole D. Houston ·  
Robert L. Coleman · Manuela Buzoianu ·  
Gabriel Robbie · Robert Lechleider



- Drug trial for targeting EphA2 expressing cells (e.g. triple negative breast cancers) failed in the Phase I Trial
- 6 women entered (breast, ovarian, endometrial and colon cancer patients)
  - Trial had disastrous effects; all women withdrew due to adverse affects (hemorrhage, liver disorder, etc.)

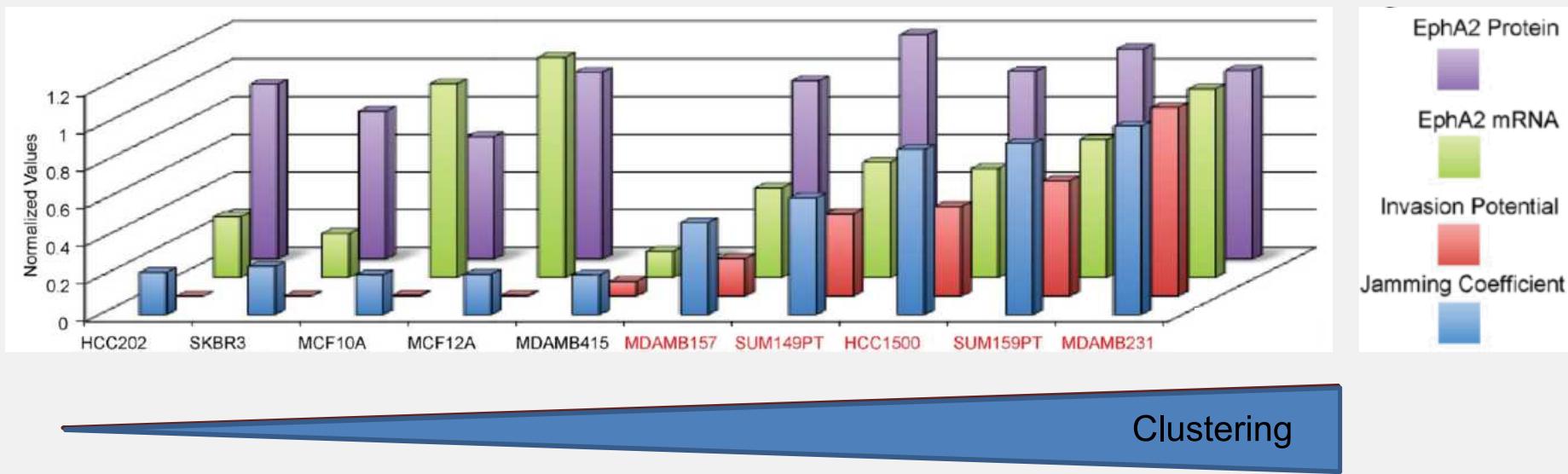
# What Went Wrong with the Trial?

- Monoclonal anti-EphA2 antibody covalently attached to auristatin (microtubule inhibitor)
  - Likely NOT due to non-specific toxicity of auristatin
- Likely due to the antibody components of the antibody-drug conjugate
- In one patient, it caused pulmonary metastases
- **Maybe the artificial clustering induced by the antibody caused toxicity?**



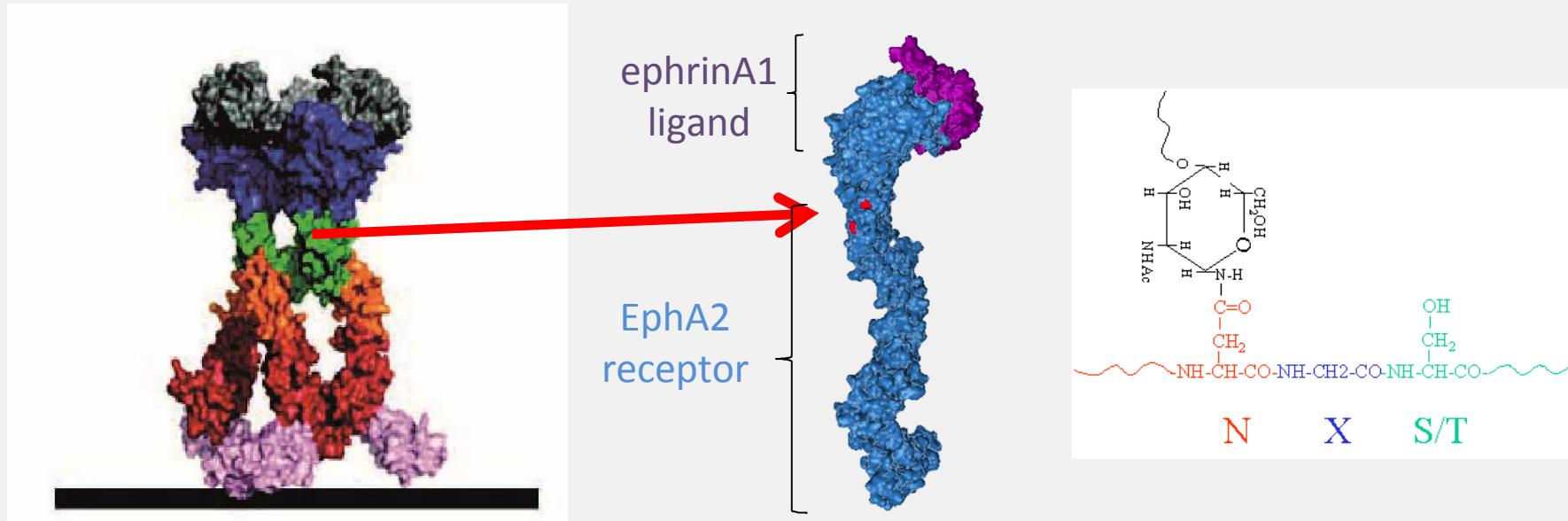
# Clustering Correlates to Disease State

- The most aggressive cancers have the most tightly clustered EphA2 receptors*



- Invasion potential of these cells only correlates to the EphA2 clustering phenotype
- Tightly clustered EphA2 indicative of more dangerous cancers?
  - Could the antibody-drug target be inducing more EphA2 clustering and causing an increased disease state of the cell?

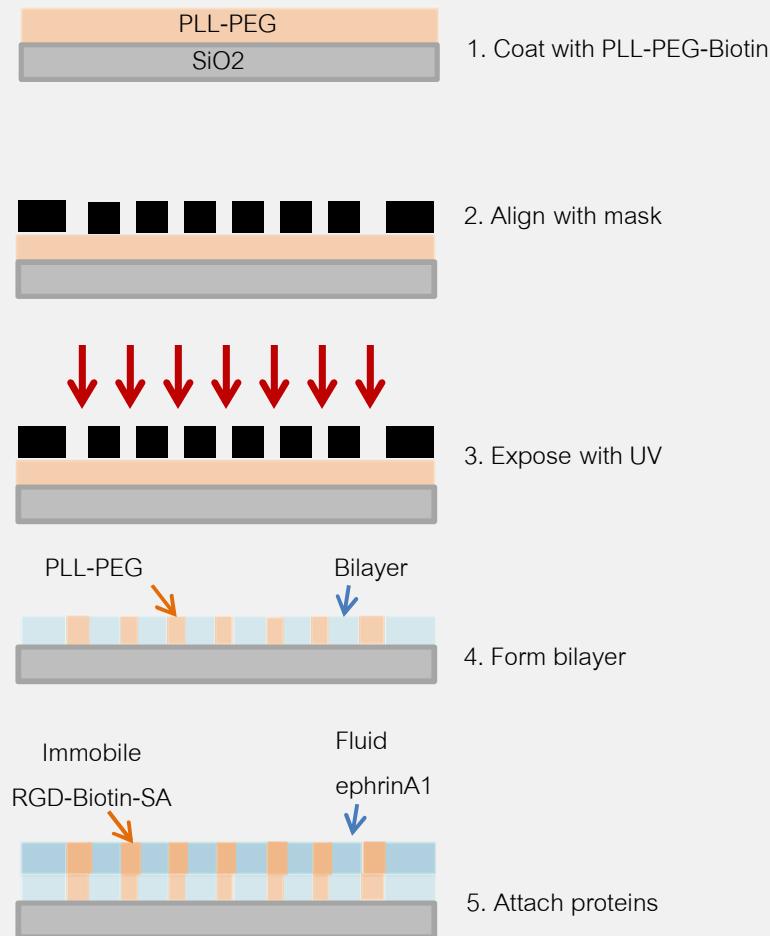
# Genetic Mutation of EphA2 *cis* Clustering



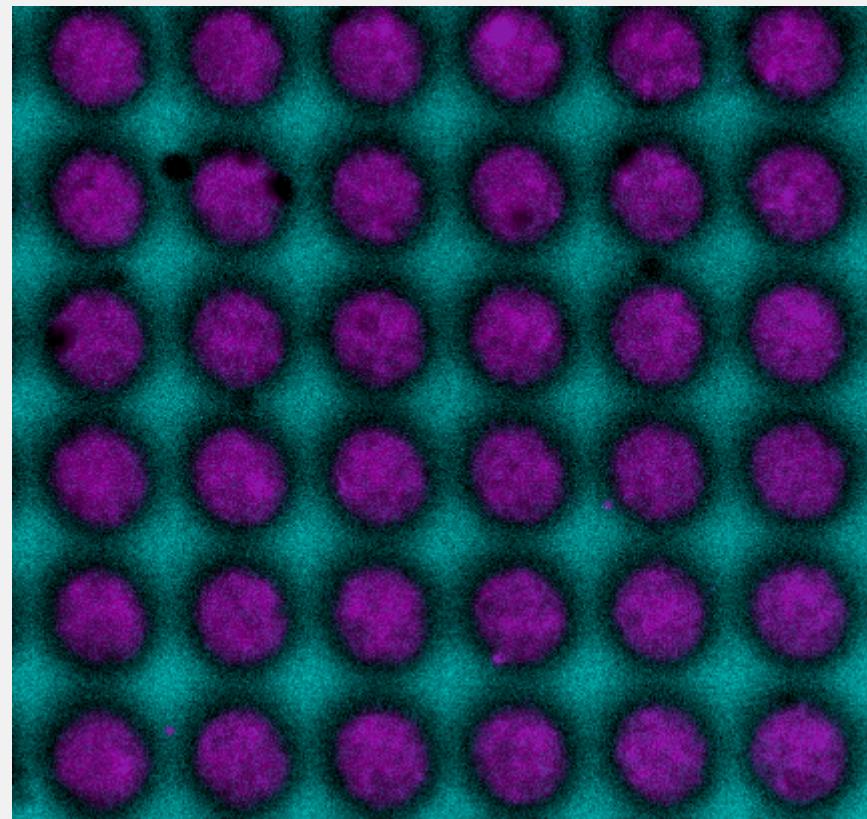
- Disrupting Eph clustering at the nanometer/angstrom scale requires genetic mutations
  - Target the sushi domain of the receptor to disrupt EphA2 *cis* clustering
- To do this in an endogenous context requires genome-editing
- We used **CRISPR/Cas9** to permanently introduce two point mutations into the sushi domain resulting in an N-linked glycan in the domain

# Confining Clustering to Determine Cluster Structure

- Confine clusters to defined regions and limit ligand density
- Flatten cells using integrin adhesions

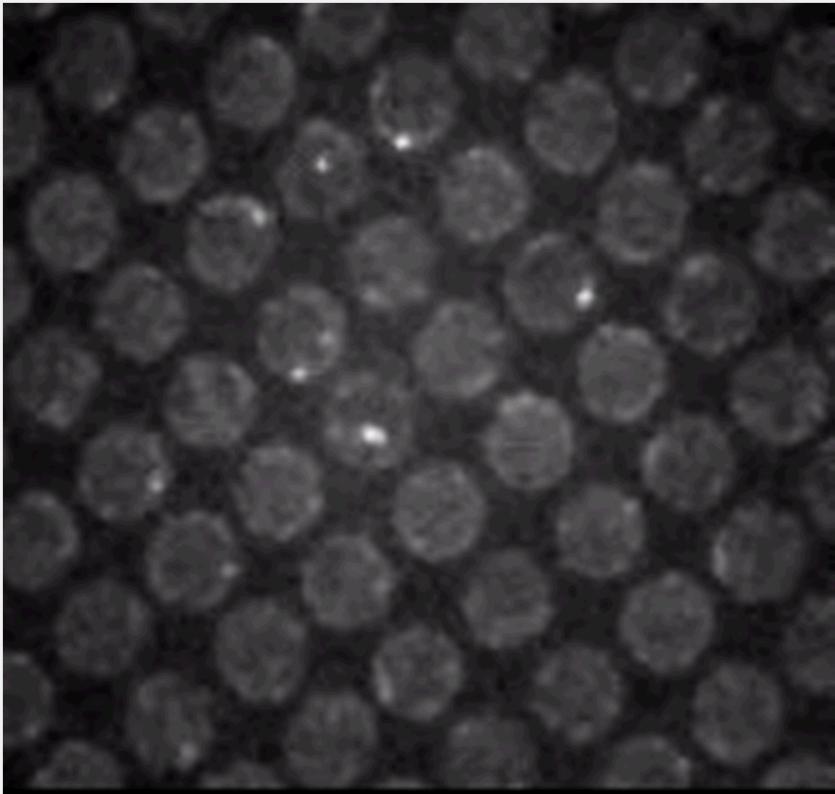


Immobile RGD, fluid ephrinA1

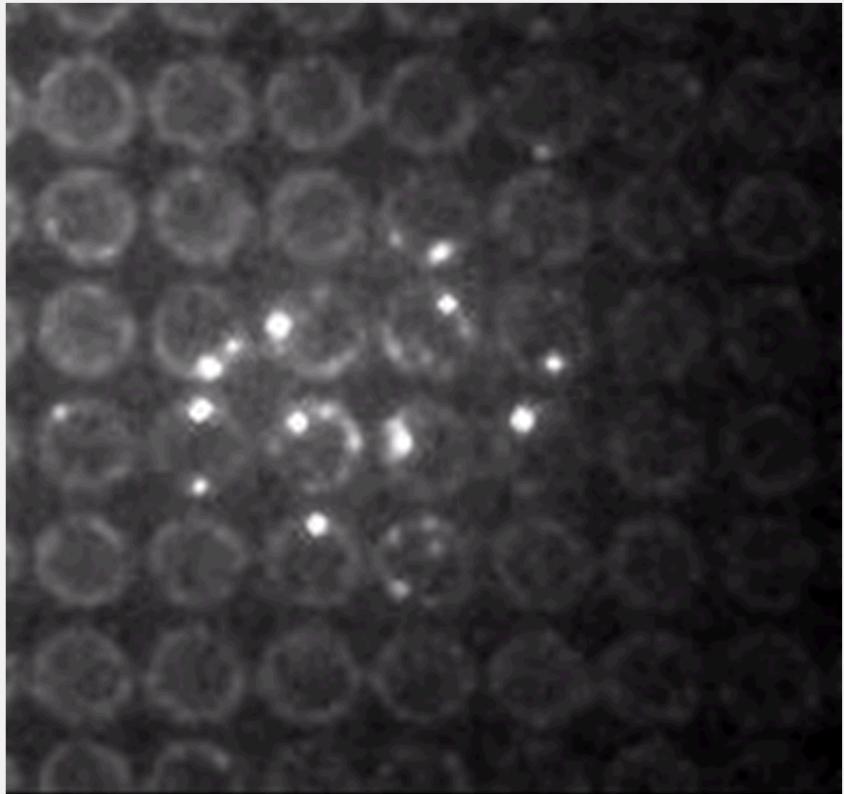


# Mutant Clusters are More Dynamic and Transient

Wildtype



Mutant



5 $\mu$ m

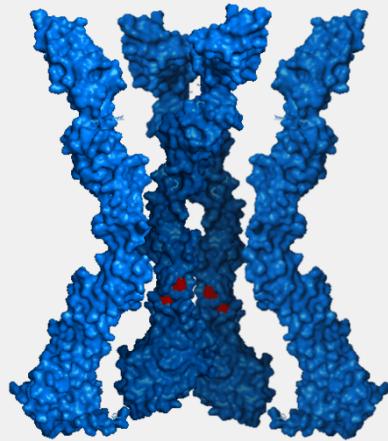
- Ligand density can be confined while adhering the cells using photolithography
- Mutants cluster faster and less definitively

Imaged every 10 seconds for 6 minutes

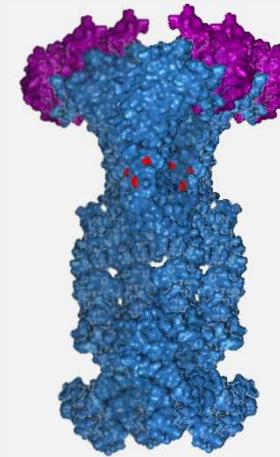
Movies are 5 frames per second

# Disrupting EphA2 *cis* Interactions is Necessary for Clustering

Side view of unligated EphA2



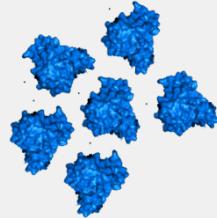
Side view of ligated EphA2



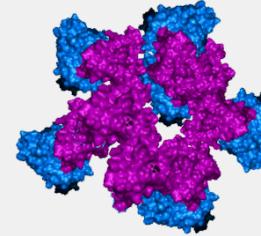
Rearrangement  
of Eph-Eph  
interactions



Top-down view of unligated EphA2



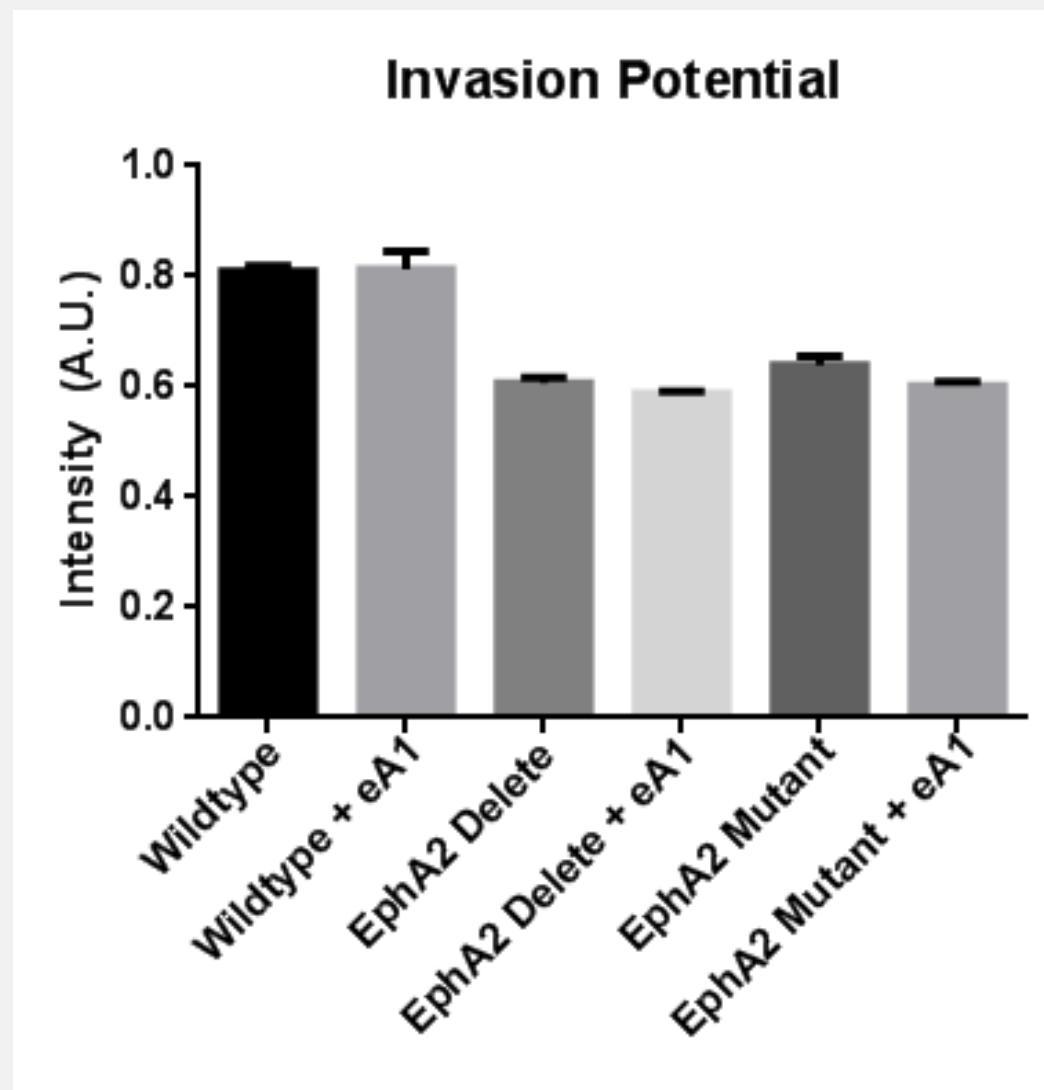
Top-down view of ligated EphA2



- Sushi domain mediates stable Eph-Eph interactions within the cell
- Micron-scale clustering requires binding to ephrinA1 and a disruption of Eph-Eph interactions

# Changing Molecular Clustering Changes Invasion Potential

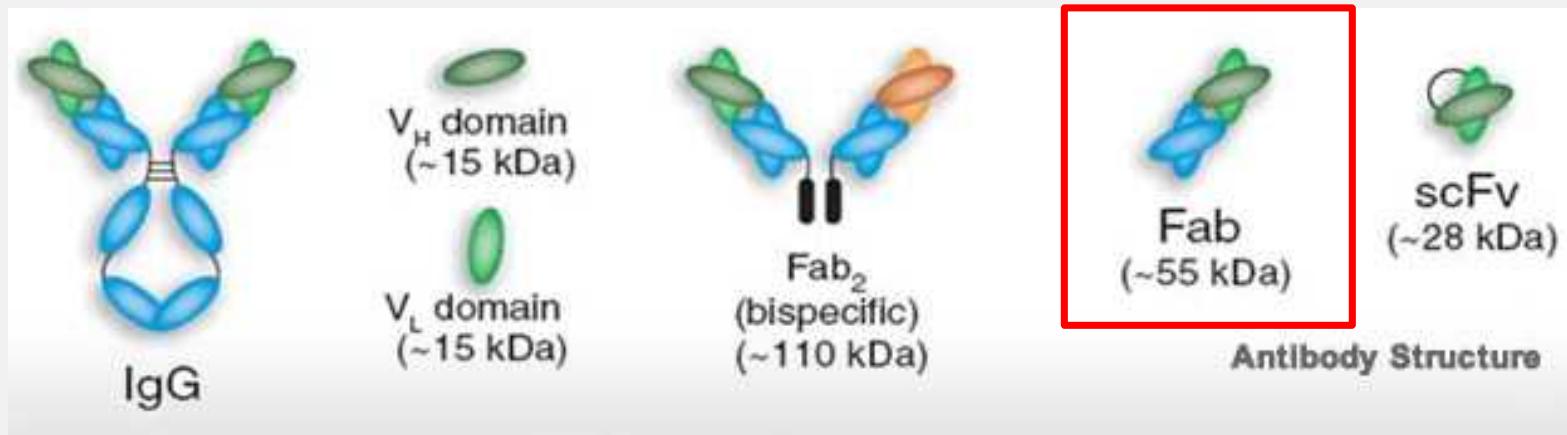
- Invasion potential measures the ability of a cell to leave a colony [tumor] and migrate through certain barriers; it is the hallmark of metastasis
- EphA2 with the sushi domain mutation is ~25% less invasive than the wildtype



# Part 1 Conclusions

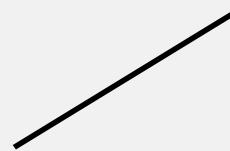
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- The importance of spatio-mechanical regulation of signaling systems across length scales is becoming increasingly evident
- Understanding spatio-mechanical regulation of Eph receptors will provide insights into the misregulation of EphA2 in disease, particularly cancer
- Alternate strategies for targeting these kinds of receptors can be developed

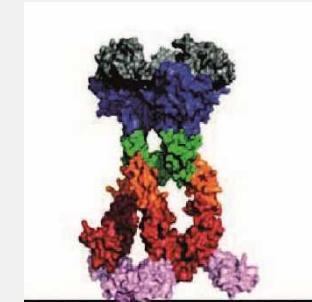
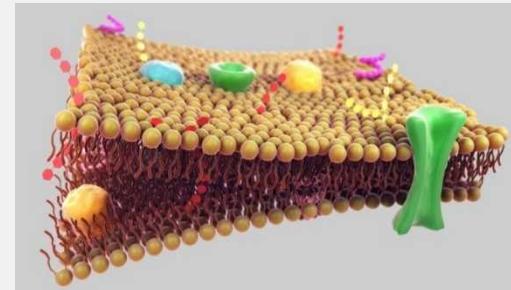


# Understanding Regulatory Mechanisms of Biological Systems

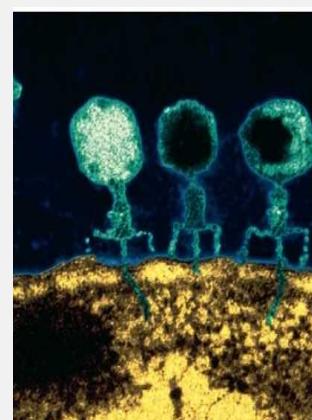
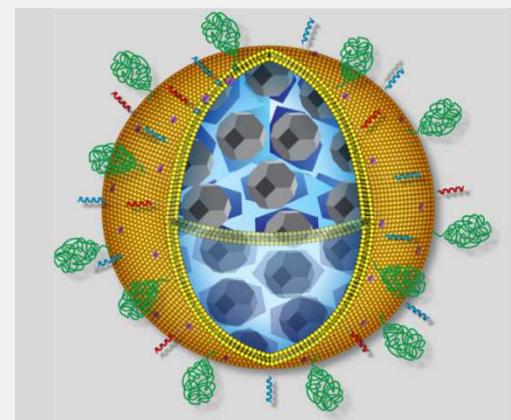
Goal: Understand biological systems for applications-based technologies (therapeutic target design, diagnostic tools, biosensing and/or bioengineering technologies)



1. Receptor Signal Regulation



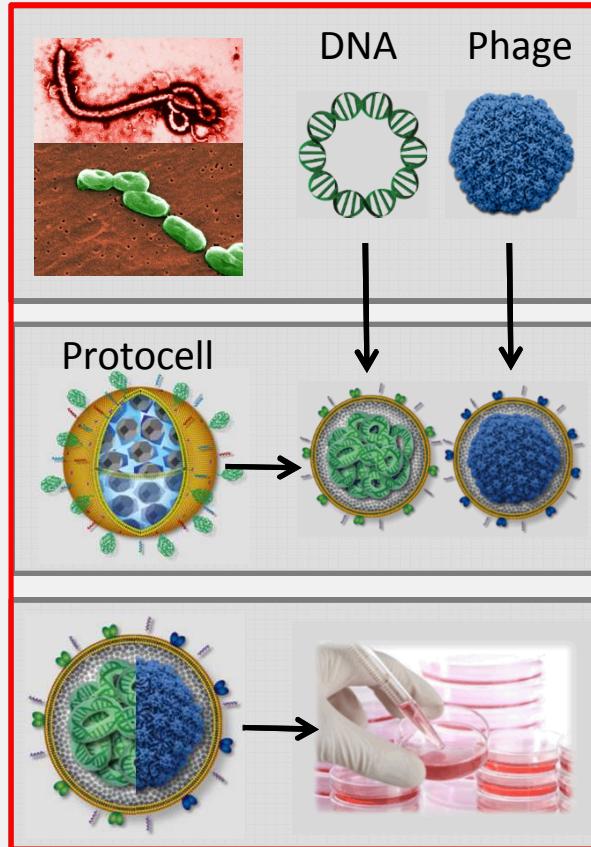
2. Targeted Therapeutic Delivery



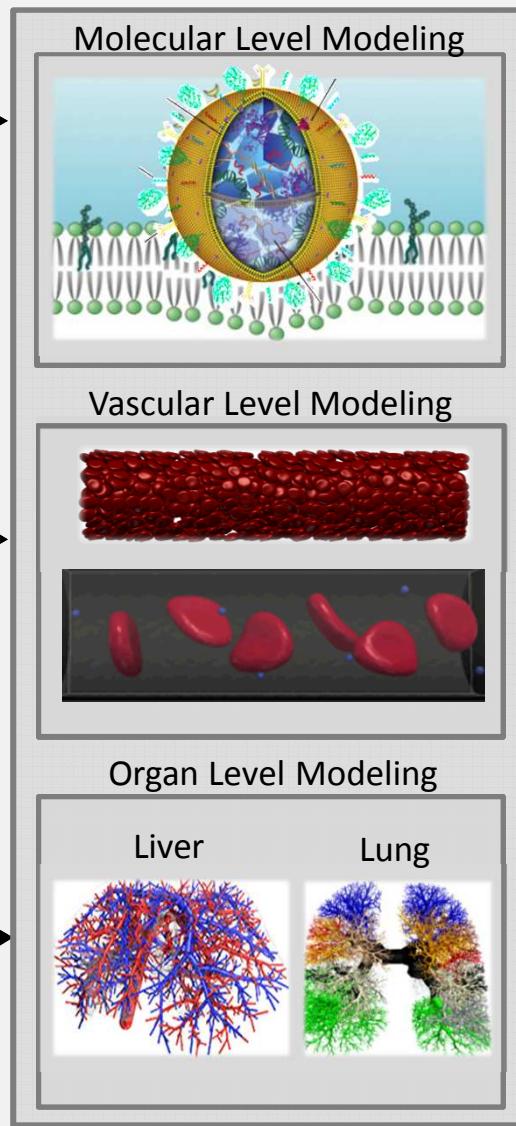
# Targeted Delivery of CRISPR/Cas9 Therapeutics

CRISPR/Cas9 Toolbox  
Packaging Therapeutics  
Testing *in vitro*  
Model Organisms

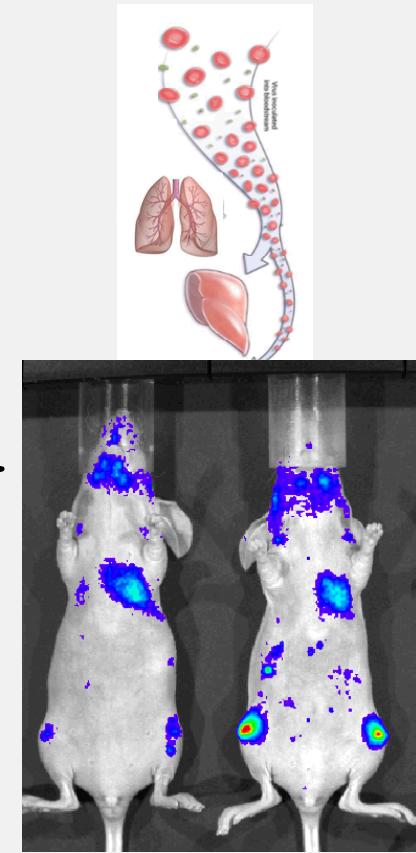
## Experimental Approaches



## Modeling Approaches



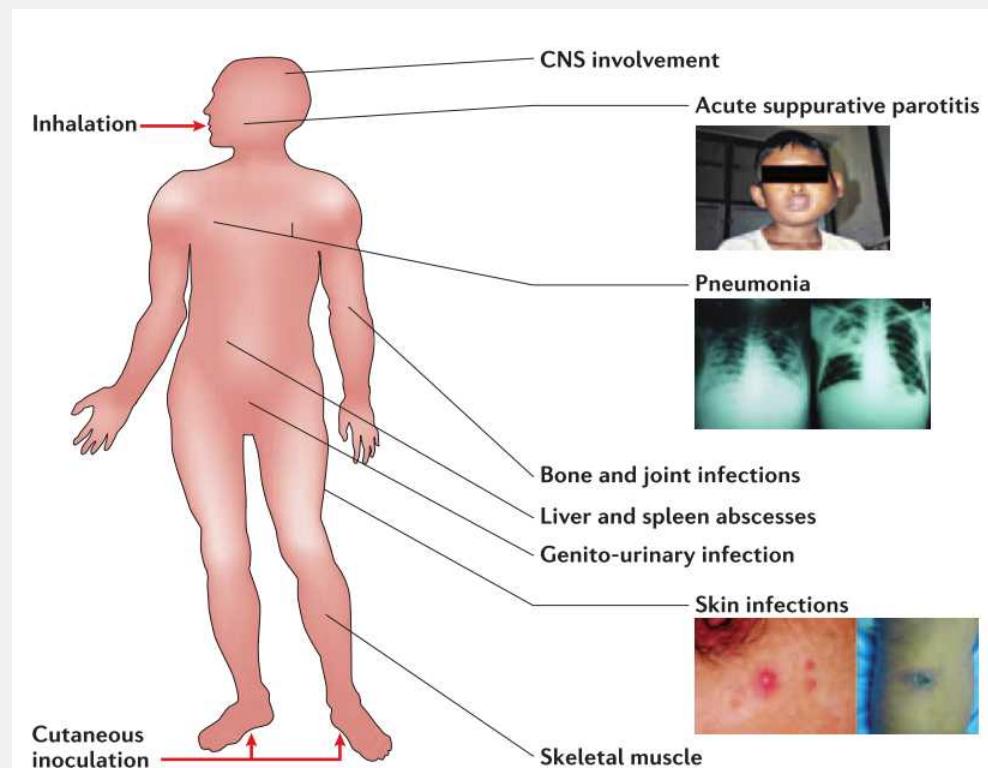
## Optimization



**Goal: Targeted Therapeutic Delivery**

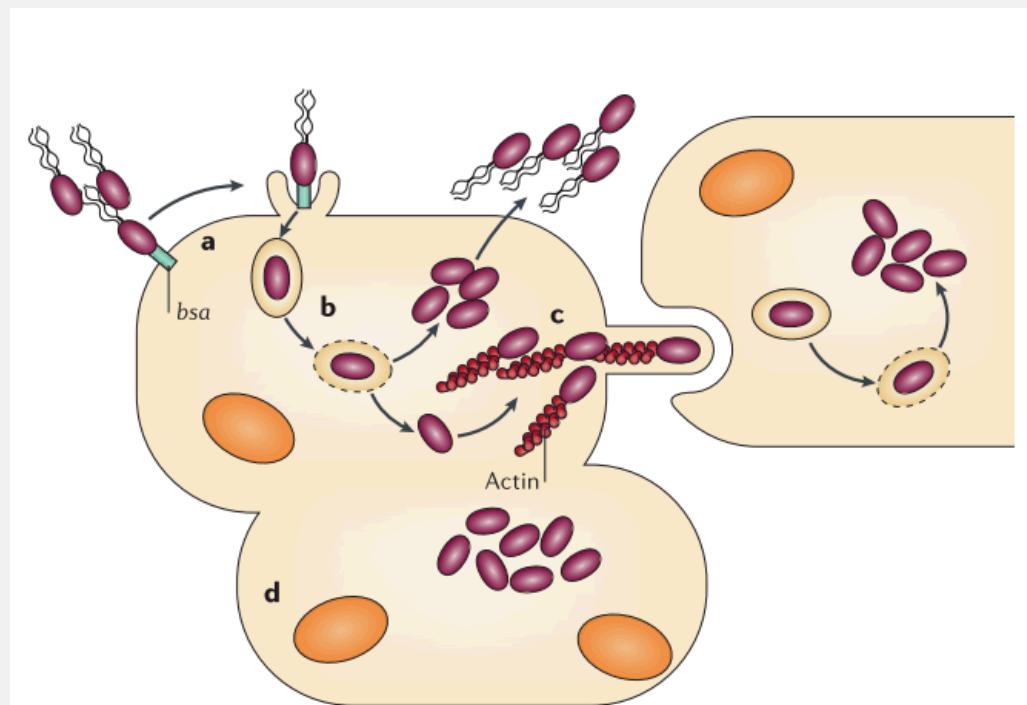
# *Burkholderia pseudomallei*

- *Burkholderia pseudomallei* is a highly drug resistant, intracellular gram-negative bacterium
- Common in Southeast Asia and Northern Australia
  - 20-50% mortality rate
- Infection is acquired by inoculation, inhalation, and aspiration
- Causes melioidosis
  - Pneumonia
  - Bone pain
  - Abscesses
  - Brainstem encephalitis
- **Goal: Develop novel treatment strategies**



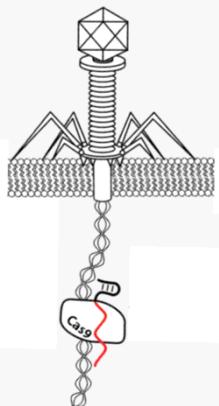
# The Life Cycle of *Burkholderia pseudomallei*

- Life cycle involves:
  - Adherence and entry into host cells
  - Phagosome escape
  - Cytosolic replication
  - Actin propelling within the cell
  - Spreading to neighboring cells

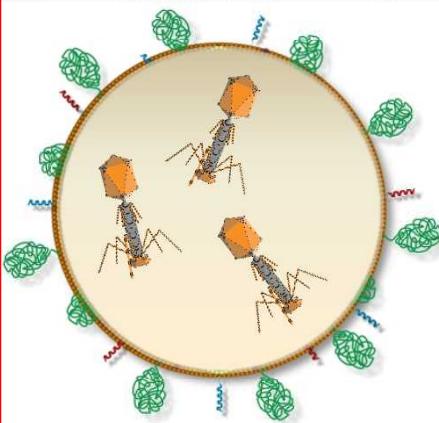


# How to Target Resistant Bacteria

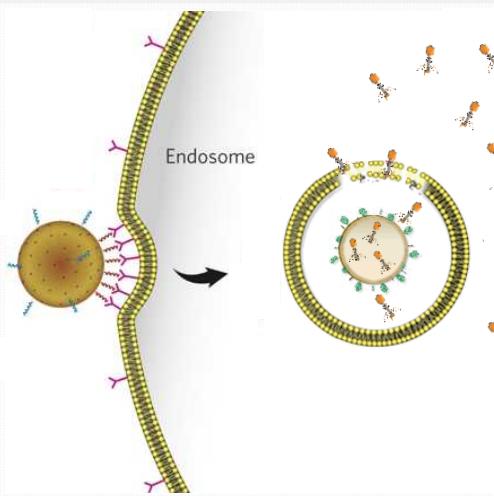
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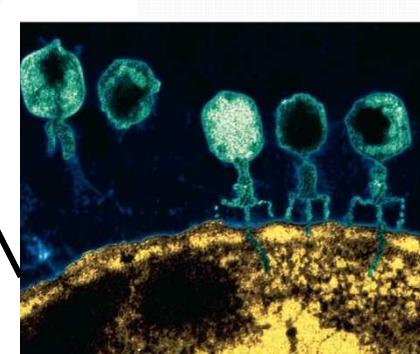
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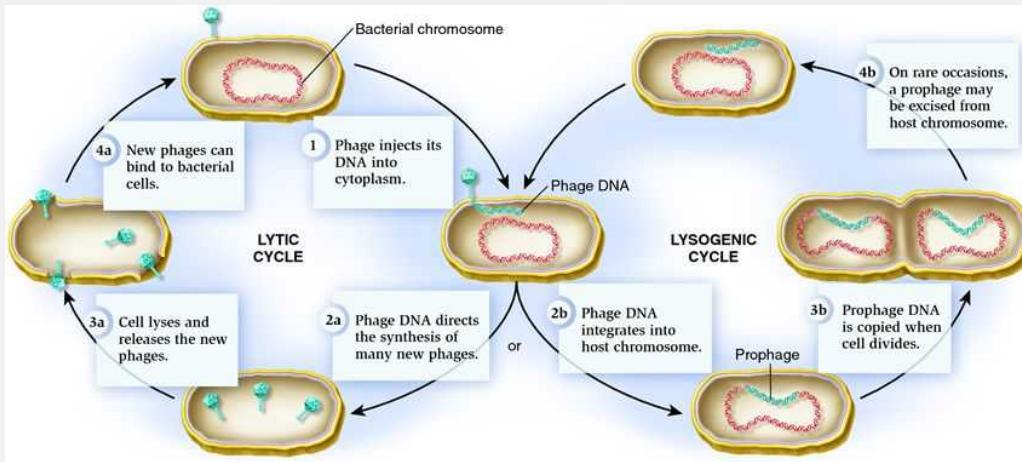
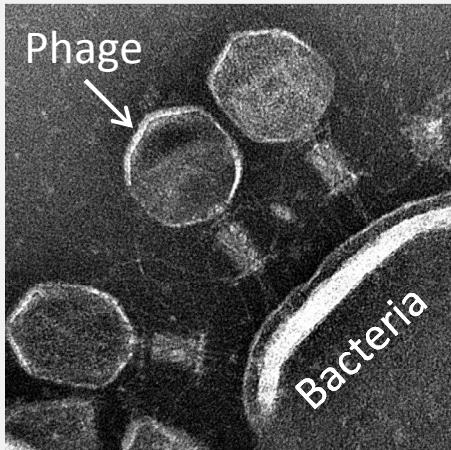
4. Bacteriophage Deliver CRISPR/Cas9 to Intracellular Bacteria



Protocell masks bacteriophage from initiating an immune response

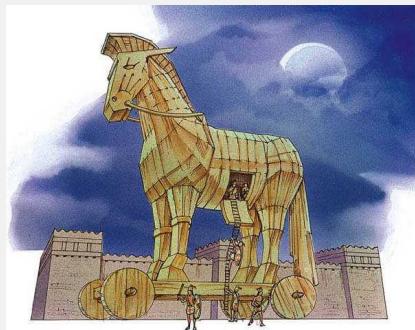
# Inhibit Antibiotic Resistant Bacteria Using Bacteriophages

## Bacteriophages: Bacteria-Specific Viruses



### Lytic Phage

- Fast bacterial killing on own
- Challenging to genetically modify



### Lysogenic Phage

- Can exist without killing the bacteria
- Easier to genetically modify to contain CRISPR/Cas9; these phage will carry CRISPR as a “Trojan horse” for resistant pathogens

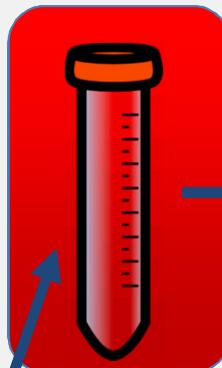
Goal: *Use Bacteriophage as a Trojan Horse for Delivering CRISPR/Cas9 Therapeutics*

# Environmental Isolation of Bacteriophage

Identify regions of interest to isolate phage based on epidemiology



Incubation



Filtration



Concentration



Sample types:

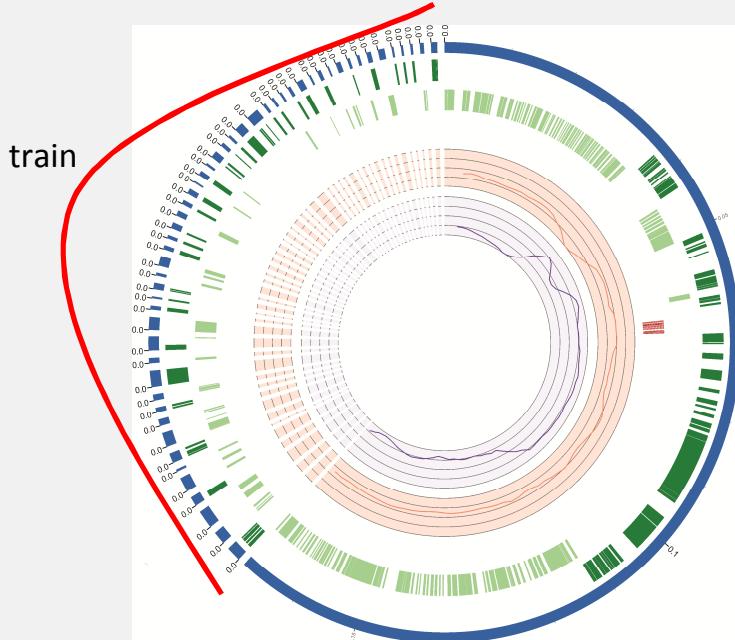
- Water
- Soil
- Sediment



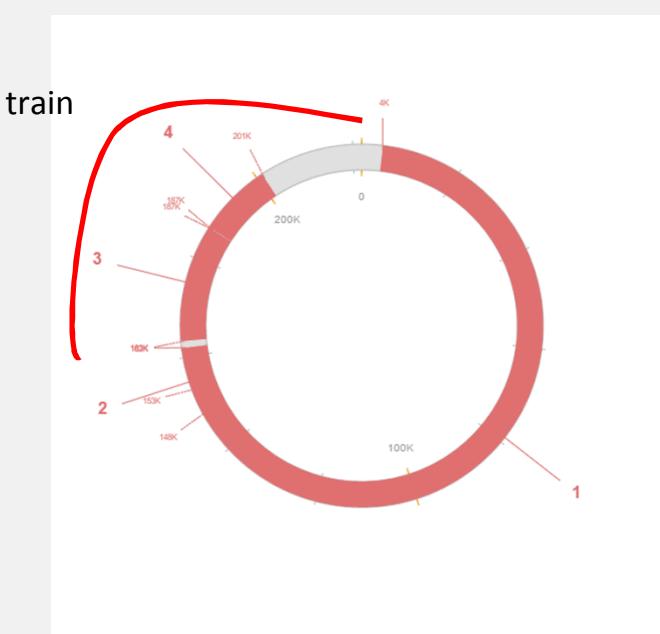
# Environmental Isolation of Bacteriophage

- Isolated 2 new bacteriophage against *Burkholderia*
  - Isolated from samples collected in Louisiana (ME) and Arizona (AE)
  - Tested stability; currently purifying high titer stocks
  - Sequencing in progress
  - One is **lytic** and one appears to be **lysogenic**

CE-7 + train PATRIC annotation

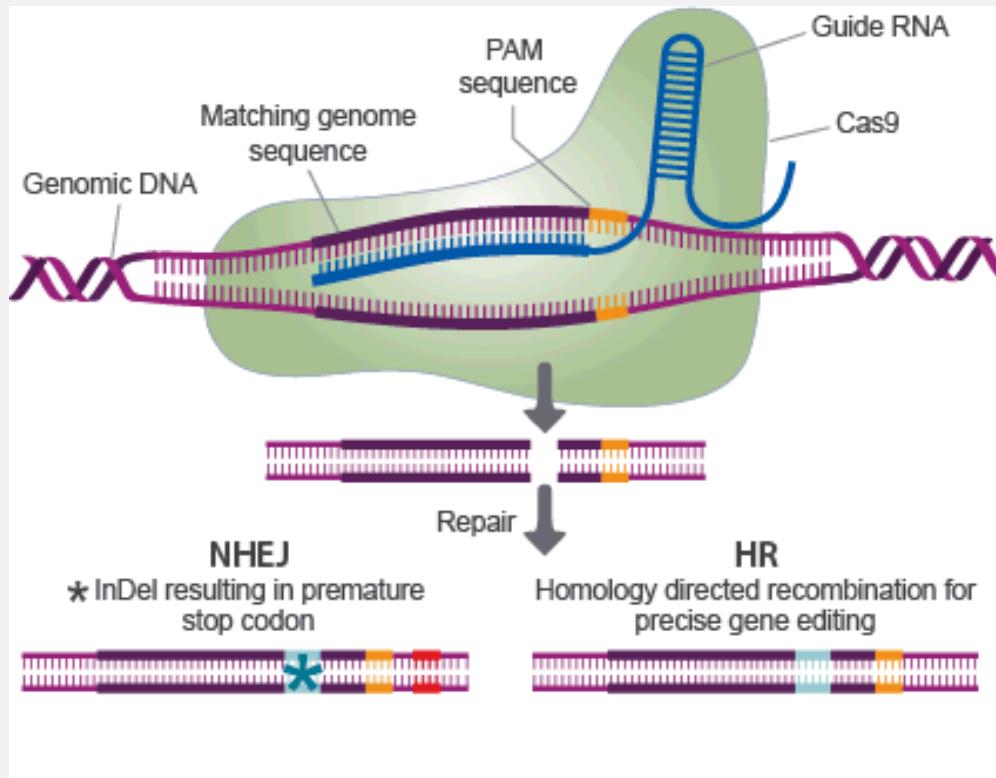


CE-6 + train PHASTER annotation



# CRISPR/Cas9 Gene Editing Technology

- CRISPR is a novel targeted gene-editing technology
  - Can introduce point mutations, **delete** genes, and add genes in specific genomic loci
- The technology uses an RNA-guided endonuclease to introduce a DNA break and relies on cellular DNA repair mechanisms to selectively edit genes



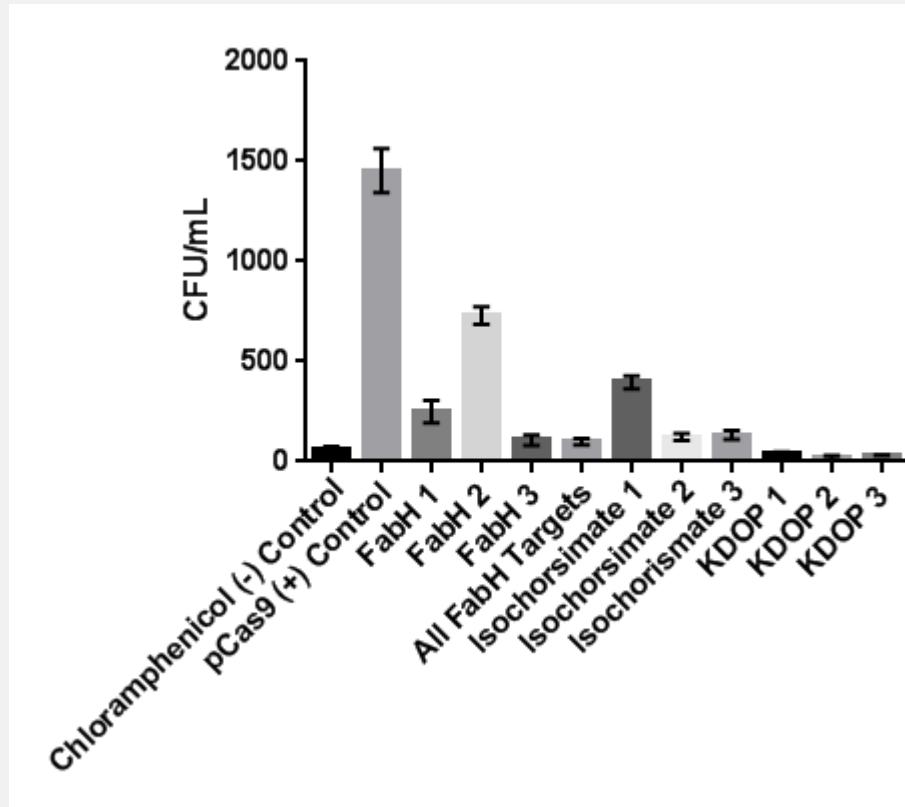
# Building a CRISPR/Cas9 Antimicrobial Library

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- Overall Goal: Use CRISPR/Cas as an antimicrobial to target intracellular bacteria using bacteriophage delivery
  - *Disrupt gene expression permanently*
- Target essential (genes required for viability) genes in *Burkholderia*, with no off target effects in humans or other organisms
  - FabH (fatty acid and phospholipid metabolism)
  - Isochorismate (iron uptake)
  - KDOP Synthase (synthesis/degradation of lipo- and polysaccharides)
- *Hypothesis: Using CRISPR/Cas9 to induce a double strand break in essential genes would result in cell death, thus acting as an effective “antibiotic”*

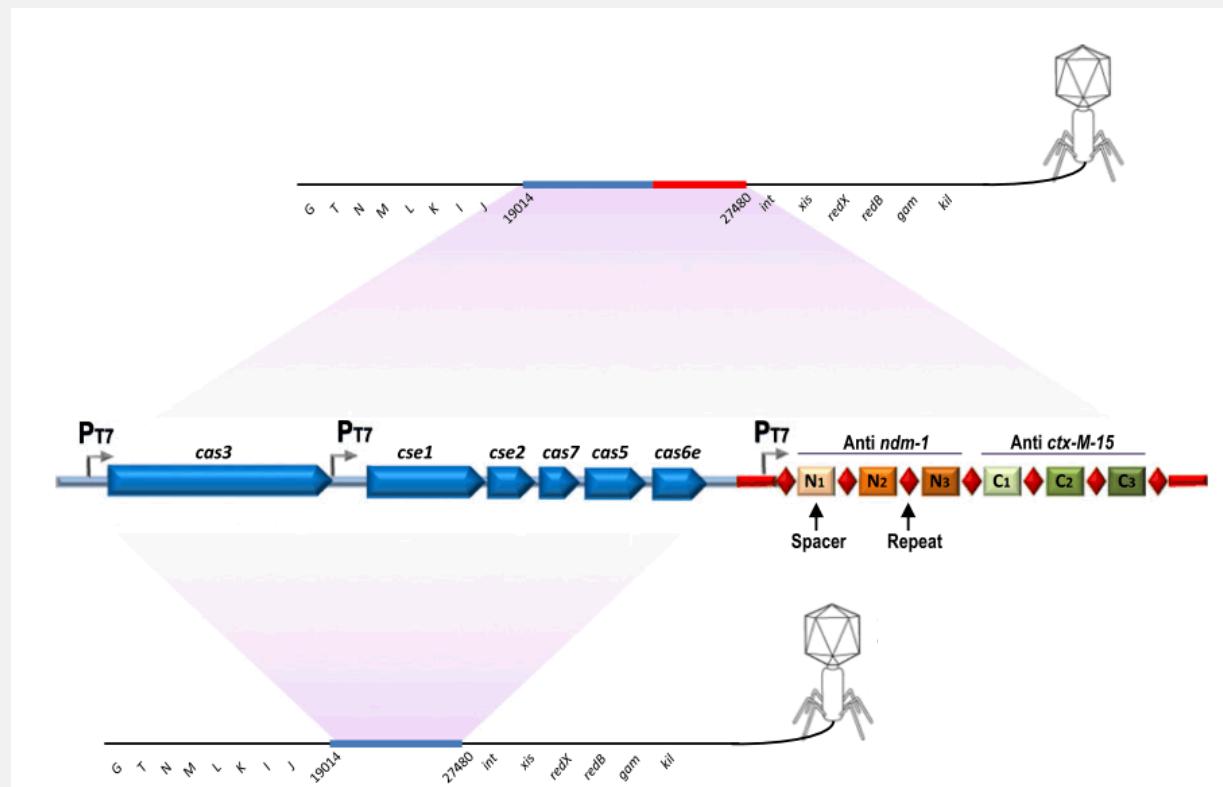
# CRISPR Targets Kill *Burkholderia*

- Tested gene targets by directly delivering CRISPR/Cas9 components
  - Cloned sgRNA targets into vector expressing both Cas9 and sgRNA
  - Delivered plasmid encoding CRISPR and Cas9 to bacteria and monitored changes in viability
  - All CRISPR/Cas9 targets result in significant cell death



# Putting it All Together: Encoding CRISPR/Cas9 Targets into Bacteriophage

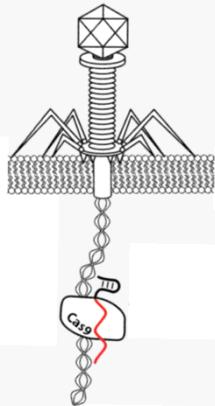
- Next steps:
  - Genetically incorporate CRISPR/Cas system into bacteriophage genomes using molecular biology techniques



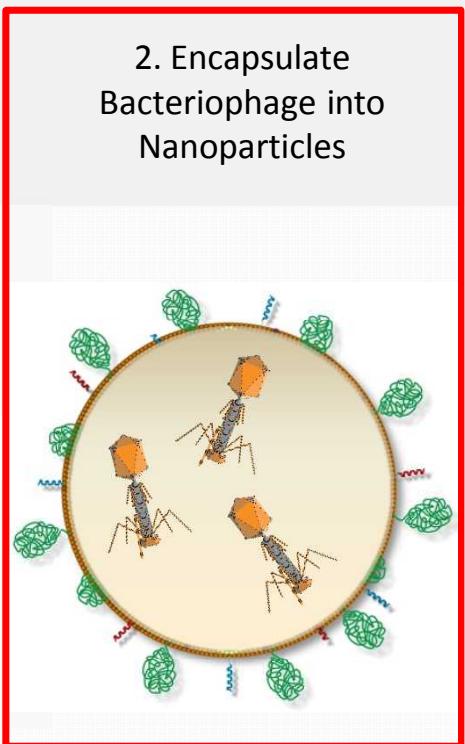
# How to Target Resistant Bacteria

- Using protocells, we can package and deliver a variety of therapeutic cargos, including bacteriophage, to specific cell types

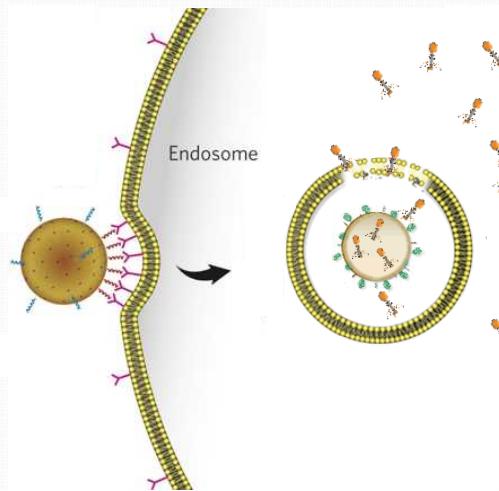
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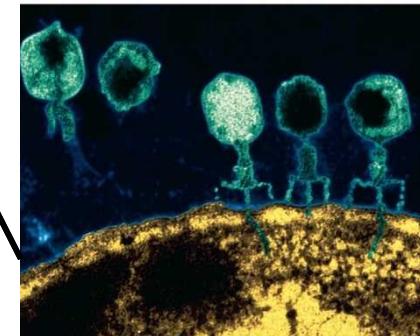
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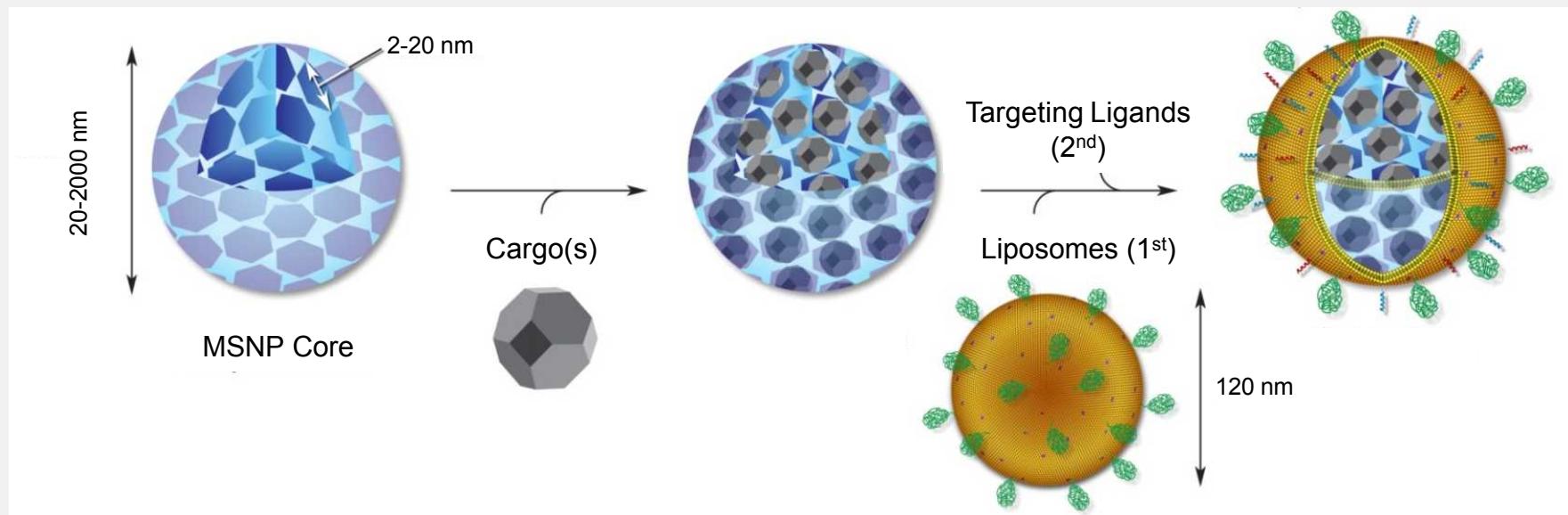
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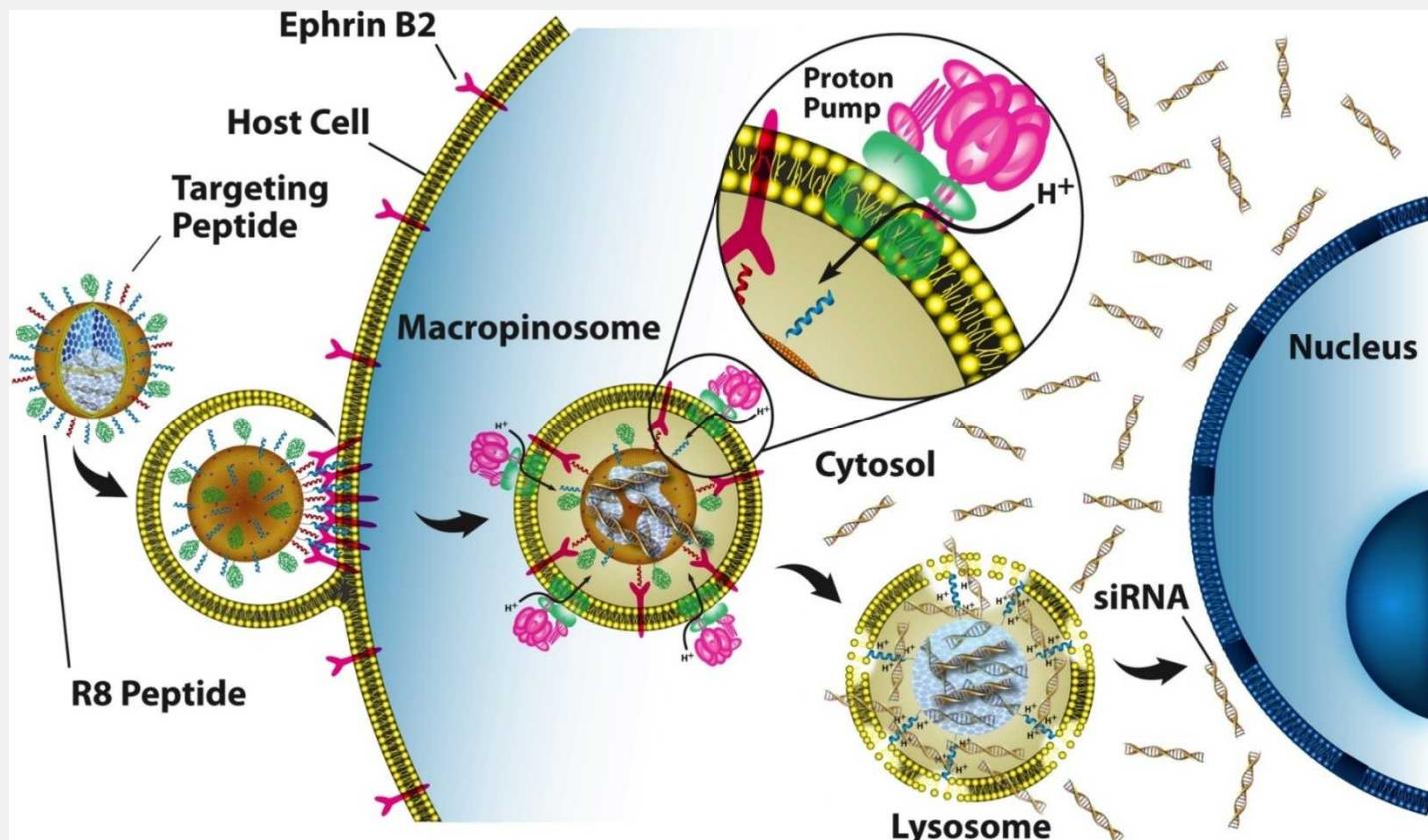
# Using Protocells to Deliver Therapeutic Cargoes

- Protocells are mesoporous silica nanoparticles (MSNP) that can be *easily* loaded with complex mixtures of cargo molecules (DNA, RNA, small molecules, proteins)
- Protocells have a 100-1000 fold higher capacity than other nanoparticles
- Properties of both the MSNP core and the bilayer shell can be precisely modulated to effect behavior



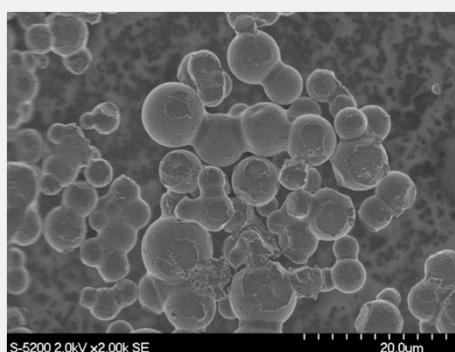
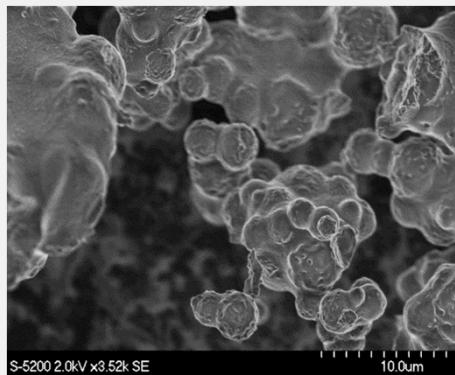
# Using Protocells to Deliver Therapeutic Cargoes

- Functionalizing the supported lipid membrane with targeting, self and endolysmotic peptides, the protocells can:
  - Be selectively delivered to specific cell types
  - Escape the lysosome
  - Deliver the therapeutic cargo



# Encapsulation of Bacteriophage into Protocells

- Tested encapsulation of bacteriophage
  - Bacteriophage were successfully encapsulated
  - Activity was maintained after release from the nanoparticle!

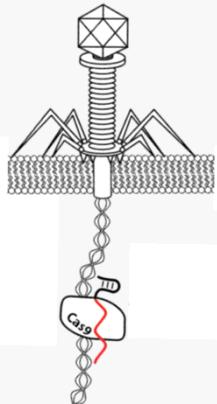


Phage	<u>Initial Activity</u> (pfu/mL)	<u>Encapsulated Activity</u> (pfu/mg)	<u>Loss of Activity (Log</u> pfu)
<b>PL-4A</b>	5.50E+10	1.05E+08	0.5
<b>PL-4B</b>	1.50E+10	4.00E+07	0.6
<b>CE-6</b>	2.38E+10	1.93E+06	2.0
<b>CE-7</b>	1.50E+10	6.50E+06	1.4

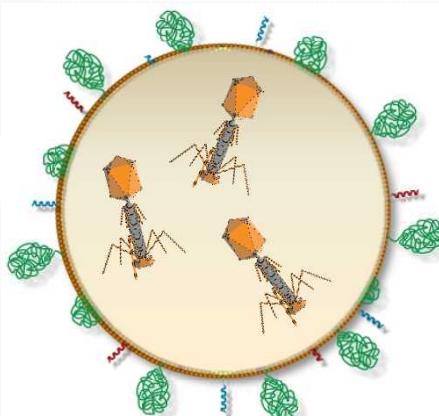
# Next Steps

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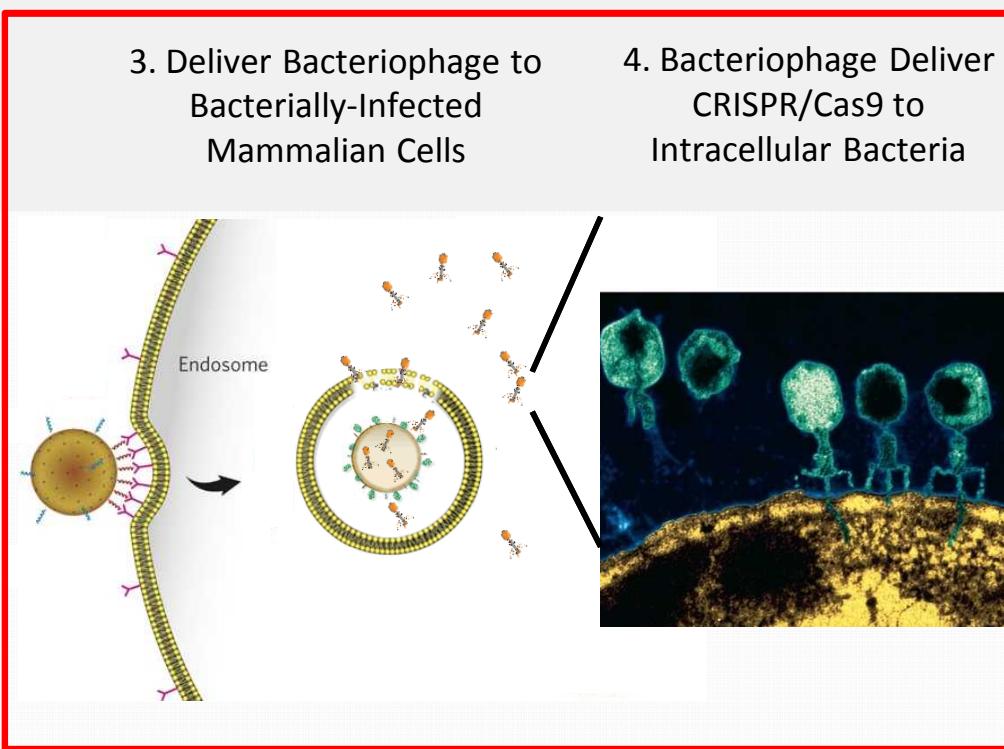
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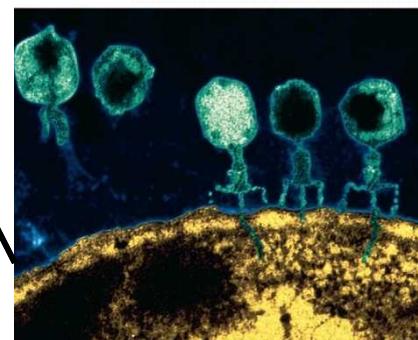
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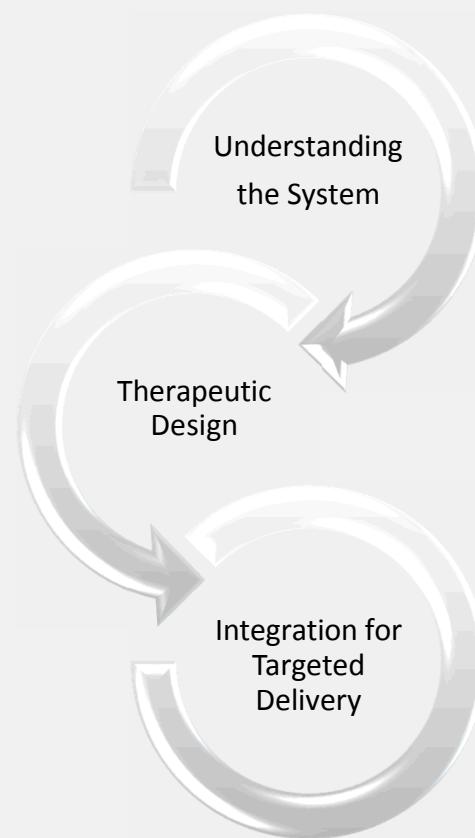
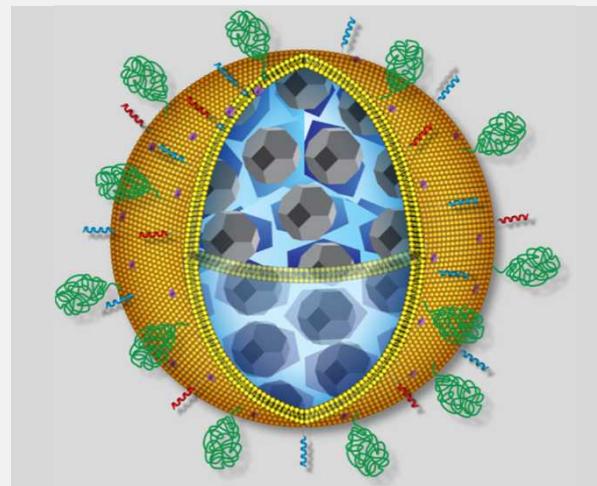
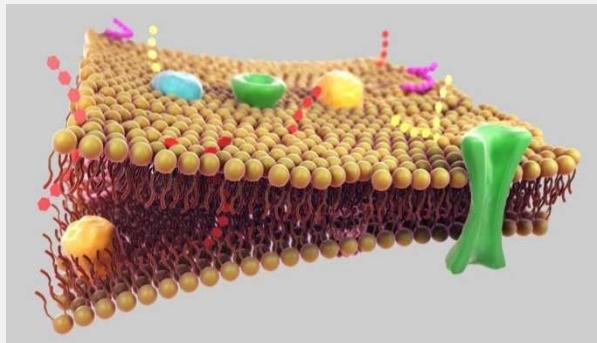
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Protocell masks bacteriophage from initiating an immune response

# Understanding Biological Systems for Integrated Science Applications

Goal: Understand biological systems for designing targeted therapeutics. We can couple our expertise with protocells and cancer cell biology to design and selectively target/deliver novel, more effective chemotherapies



# Acknowledgements

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## Sandia National Laboratories

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Dr. Carlee Ashley	Patrick Fleig
Dr. Darryl Sasaki	Kevin Crown
Dr. Steve Branda	Marissa Anderson
Dr. George Bachand	

Nano-arrays  
Membranes Nanoharvesters

Gels Kinesin Endocytosis  
Assay Motors Metastasis

Signaling Biology Cells  
Genome-Editing Neurons  
EphA2 Microtubules  
Clustering

Cell Growth

Spatio-Mechanical DNA  
CRISPR Polymersomes  
Proteins

