



Genetic Evaluation of Pond Crashes during ATP³ Unified Field Study

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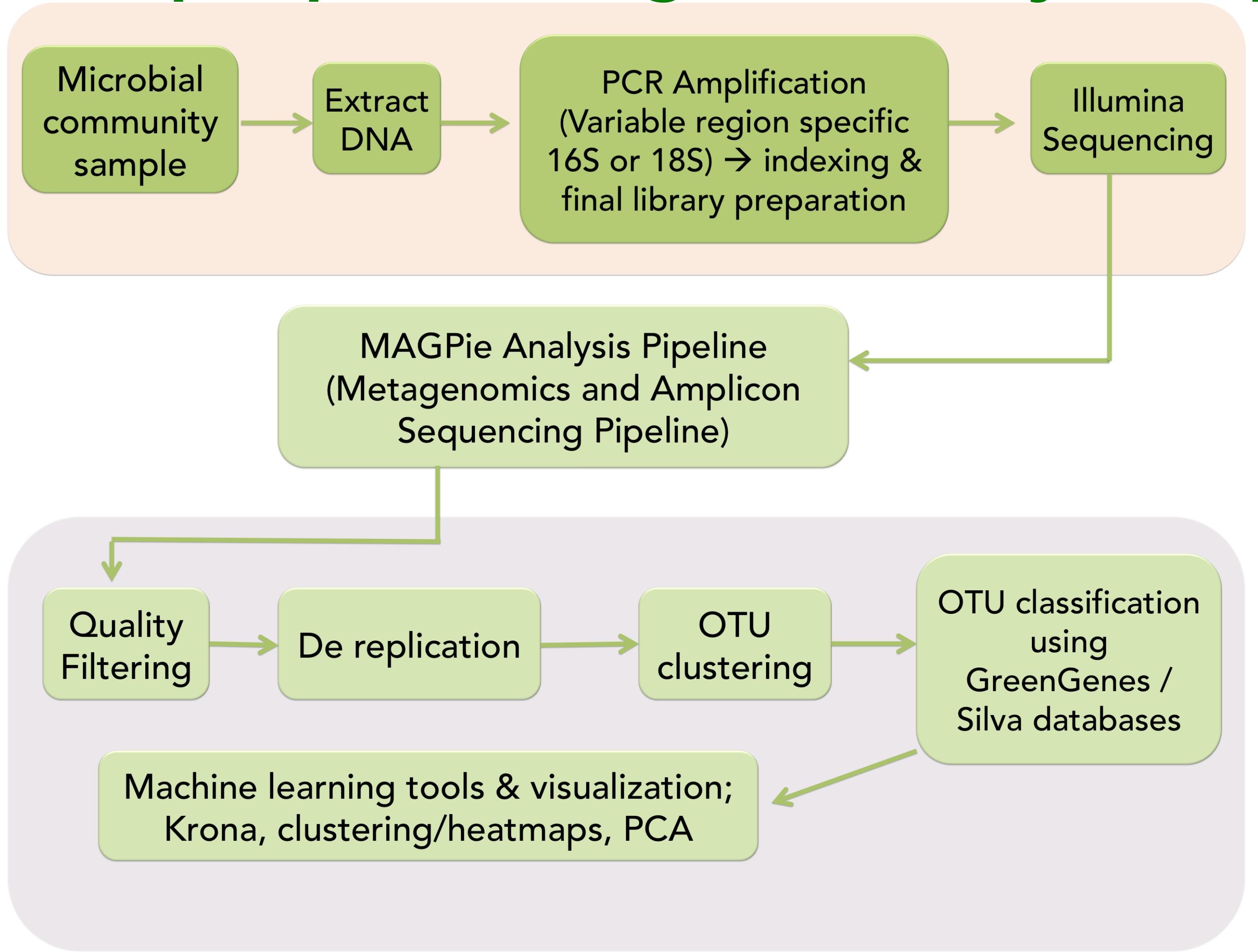
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Abstract

The Algae Testbed Public Private Partnership, ATP³, is a Department of Energy supported consortium funded to generate high impact data in order to quantify the seasonal and geographic variations on the production of microalgae biomass for use in production of biofuels and bio-products. We use metagenomic amplicon sequencing to identify deleterious species that contribute to pond instability and consequent reduction in annualized production. Such diagnosis of the root causes of pond crashes is critical to for development of inexpensive screening and monitoring tools for early crash detection, as well as engineering countermeasures. For this Unified Field Studies, cultivation trials, two strains, *Nannochloropsis oceanica* and *Chlorella vulgaris*, were grown over four seasons: Spring, Summer, Fall and Winter at geographically distinct consortium member sites. Routine samples were preserved and archived onsite for genetic analysis in event of pond crash. At Sandia National Labs microbiome analysis by next generation DNA sequencing was carried out to provide a presumptive identification of the biological agent(s) responsible for the pond crash. Fragments of the small subunit ribosomal RNA gene were amplified by PCR and sequenced. Though analysis of the sequencing data with novel Metagenomics Pipeline (MAGPie) combined with data-mining tools we identified bacterial and eukaryotic communities responsible for pond crashes.

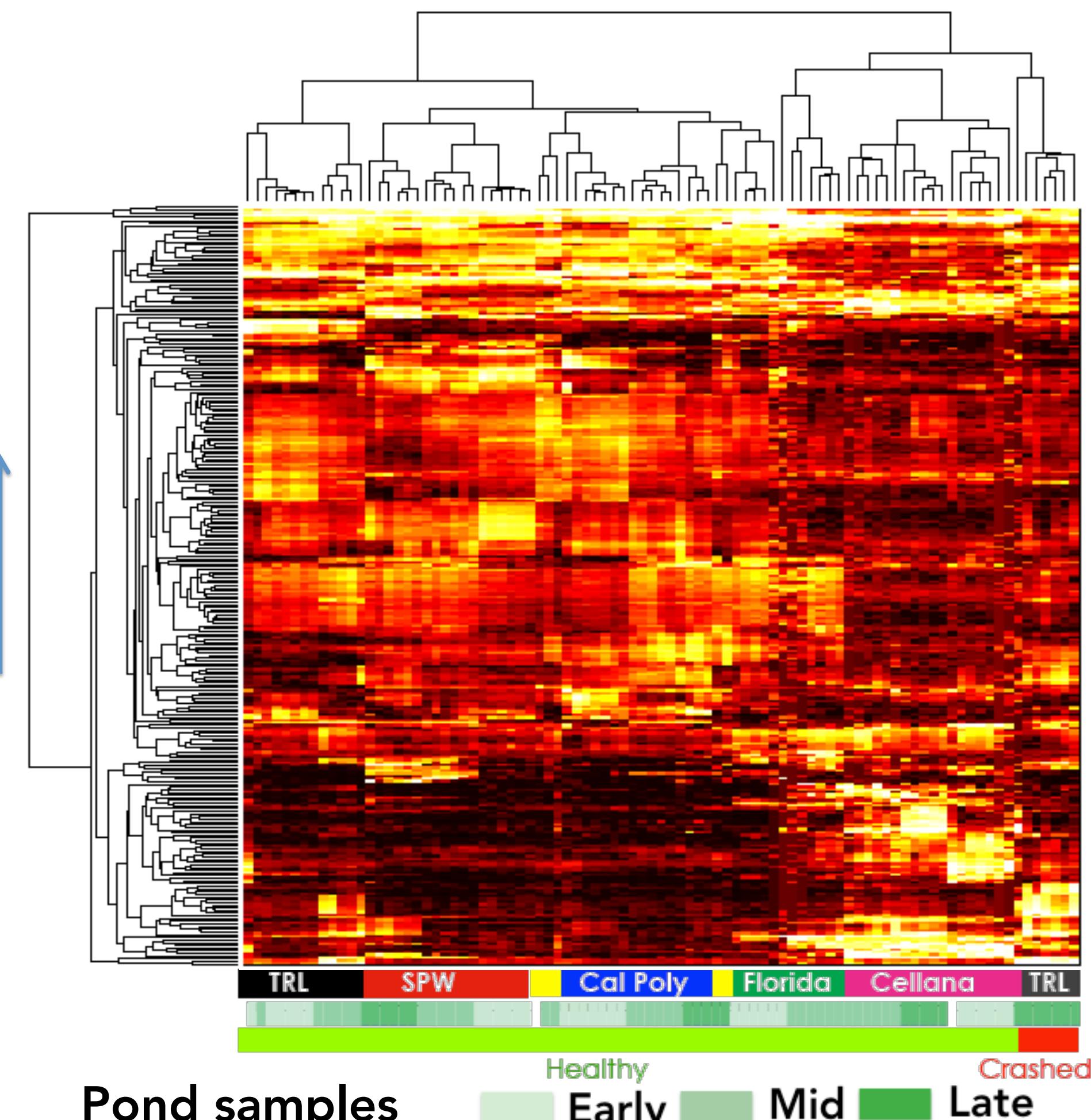
Sample processing and Analysis Pipeline



The biomass samples from the sites were sent to Sandia from the six ATP³ sites is processed to extract DNA followed by 2 steps of PCR amplification for a specific variable region in rRNA to amplify and then sequence. For these samples we amplify V3-V4 region of 16s rRNA to target prokaryotes and V4 region of 18s rRNA to target Eukaryotes.

The sequence reads were analyzed by new software pipeline **MAGPie (MetAGenomics and Amplicon Sequencing Pipeline)** for OTU clustering and classification. We have integrated Usearch program for analysis and has significantly faster than other traditional softwares (Qiime). Further we use machine learning tools to study and identify contaminants or deleterious species on our datasets

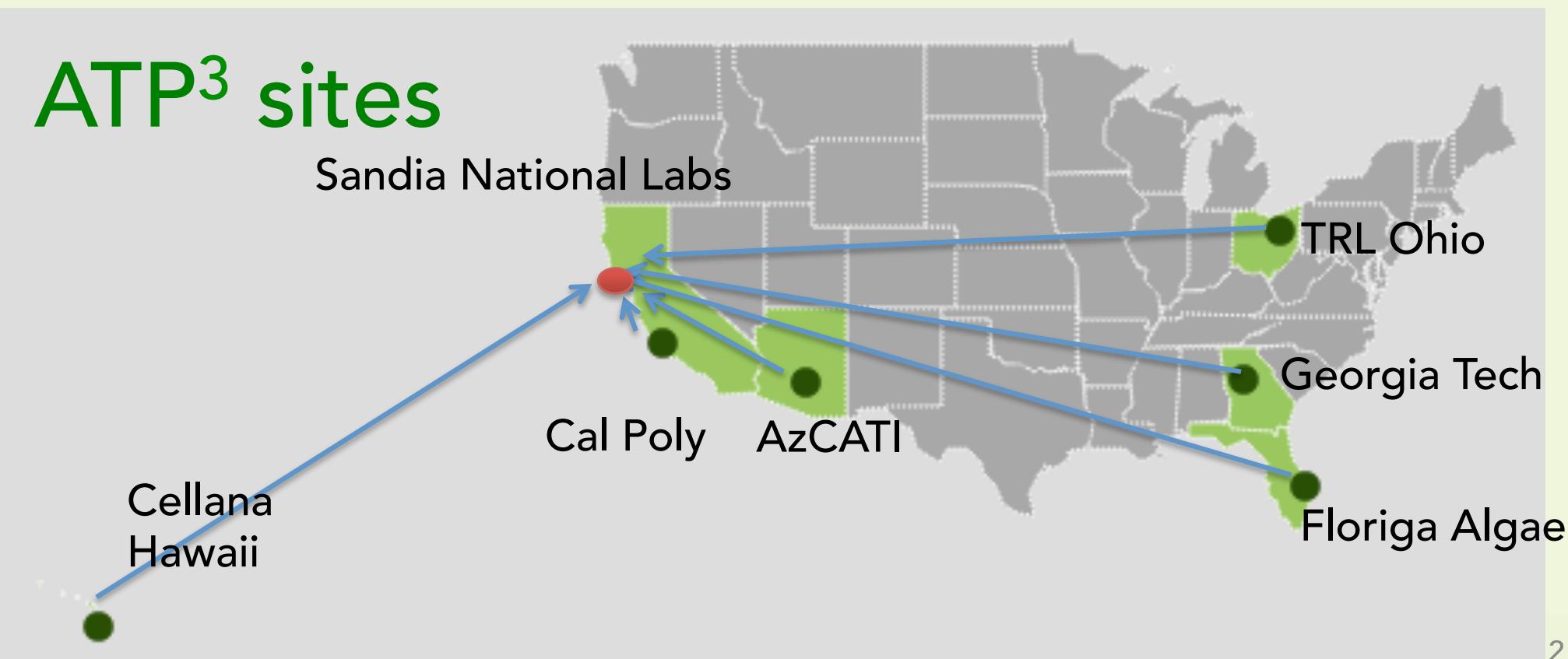
Results: 16s analysis for spring season



The figure on the left shows a heat map of bacterial community structure for spring season for all the 6 sites. The data was clustered both with respect to column and row using a correlation coefficient (below) and also by rows

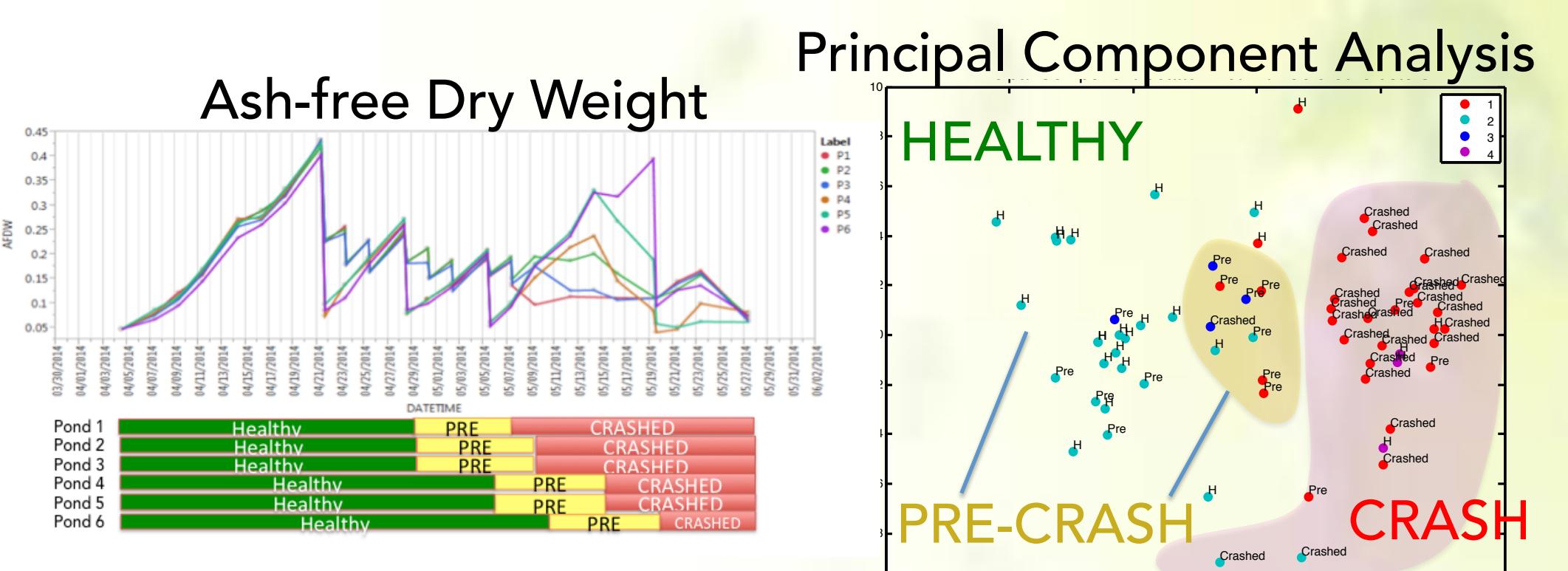
$$d_{st} = 1 - \frac{(x_s - \bar{x}_s)(x_t - \bar{x}_t)'}{\sqrt{(x_s - \bar{x}_s)(x_s - \bar{x}_s)'} \sqrt{(x_t - \bar{x}_t)(x_t - \bar{x}_t)'}}$$

The first set of rectangular bands at the bottom of the heat map shows pond location. It is clear that each ATP³ site has its own signature microbial community. One other key observation can be drawn from the heat map and the column dendrogram that the microbial community structure of site in Cellana is quite distant to all the other sites. The second set of bands are colored with respect to the date the samples was taken. It was observed that the microbial community structure can be clustered according to early, mid and late cultivation days. This was replicated for all 6 ponds in the site. And last the pond sample corresponding to a crash had a unique and significantly different community structure than normals and we were able to detect the change

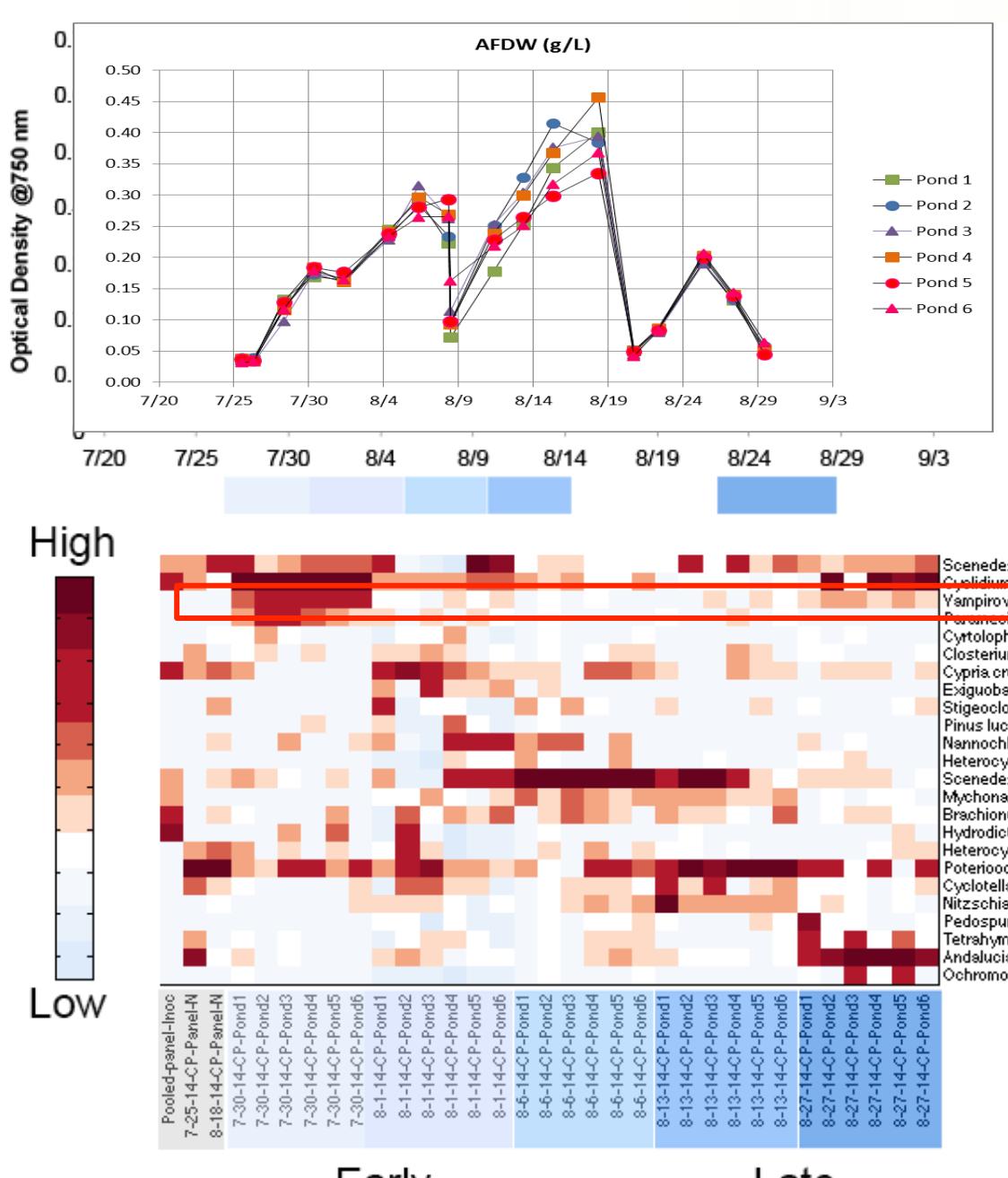


TRL Spring 16s analysis

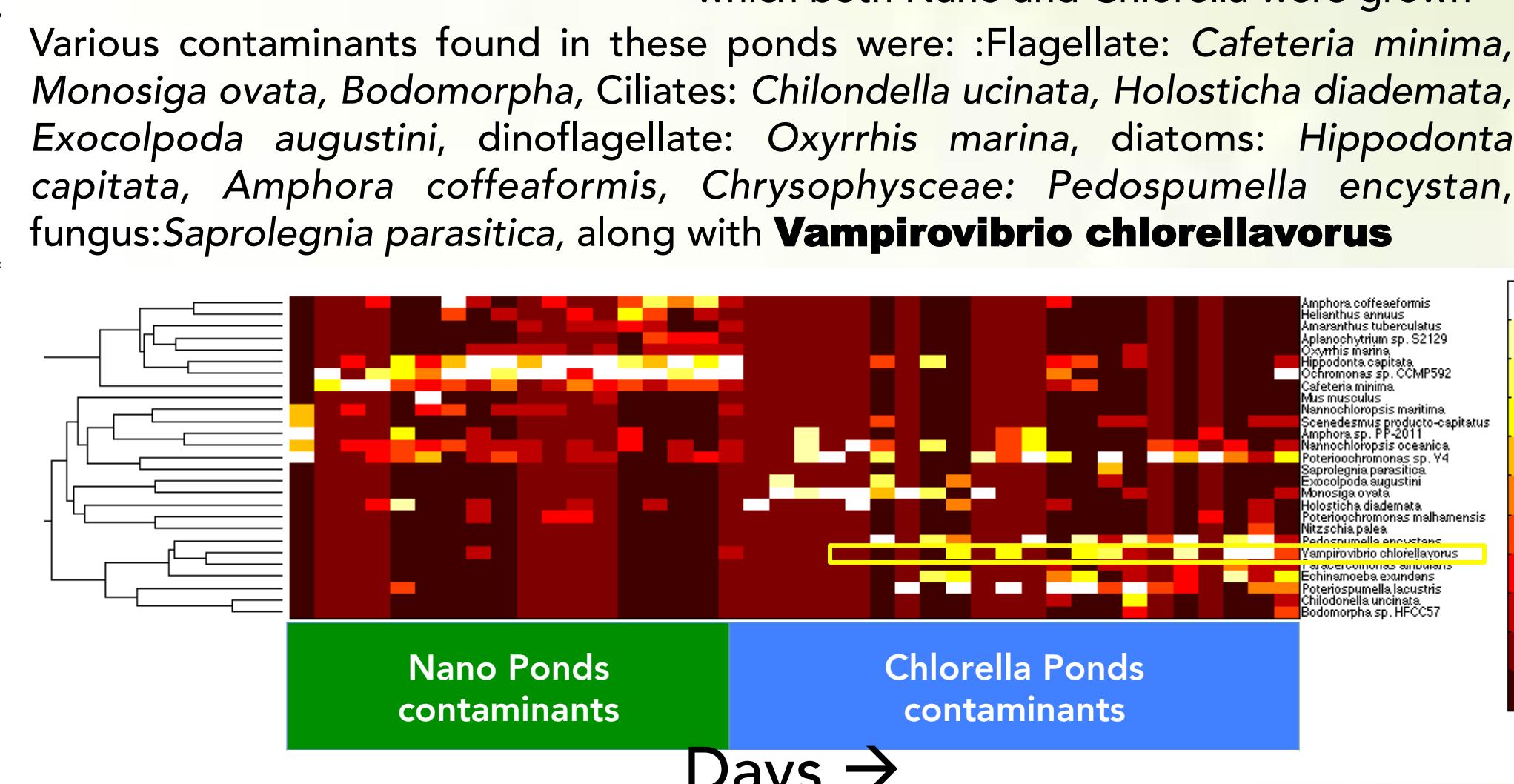
For site at TRL we did an in-depth analysis for its spring run to study a unique crash event. We focused on 16s prokaryotic community structures to see the smoking gun of the crash by clustering MAGPie output both w.r.t the samples and bacterial taxa. 4 major clusters were observed for healthy, pre-crash, crash, late crash samples. And similar results were obtained by principal component analysis



CalPoly Summer



The figure on the left shows OD measures in the samples. The bands below going from white to blue in gradient shows the time course of the sample collection. The heat map shows eukaryotic species clustering on the y axis to show the emergence of deleterious species in the ponds.



AzCATI Fall

Similar to previous examples we show a heat map of contaminants for the Fall season in which both Nano and Chlorella were grown. Various contaminants found in these ponds were: Flagellate: *Cafeteria minima*, *Monosiga ovata*, *Bodomorpha*, Ciliates: *Chilodella ucinata*, *Holosticha diademata*, *Excolopoda augustini*, dinoflagellate: *Oxyrrhis marina*, diatoms: *Hippodonta capitata*, *Amphora coffeaforni*, *Chrysophyceae*: *Pedospumella encystan*, fungus: *Saprolegnia parasitica*, along with **Vampirovibrio chlorellavorus**

Microscopy observations at the sites confirmed/identified the contamination as: ciliates (*Cyclidium glaucoma*), chrysophyte (*Poterioochromonas sp.*), flagellate (*Andalucia godoyi*). *Scenedesmus*, a few diatoms and **Vampirovibrio chlorellavorus**

Summary

On the right we show all the eukaryotic contaminants identified in the pond crashes by sequencing and computational pipelines. They range from diatoms. Ciliates, bacterias, rotifers, amoebas, and competing algae species. These results from sequencing analysis did match the microscopy observations at sites.

The advantages of using NGS methods is that we get a rough estimate of species abundance and making observation is not dependent on any a priori knowledge. Although several potential sources of bias can distort abundances and presence does not imply causality.

