



LAWRENCE  
LIVERMORE  
NATIONAL  
LABORATORY

LLNL-TR-704348

# Enzyme-Embedded, Microstructural Reactors for Industrial Biocatalysis

S. E. Baker, J. M. Knipe, J. Oakdale, J. Stolaroff

October 4, 2016

## Disclaimer

---

This document was prepared as an account of work sponsored by an agency of the United States government. Neither the United States government nor Lawrence Livermore National Security, LLC, nor any of their employees makes any warranty, expressed or implied, or assumes any legal liability or responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product, or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commercial product, process, or service by trade name, trademark, manufacturer, or otherwise does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States government or Lawrence Livermore National Security, LLC. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States government or Lawrence Livermore National Security, LLC, and shall not be used for advertising or product endorsement purposes.

This work performed under the auspices of the U.S. Department of Energy by Lawrence Livermore National Laboratory under Contract DE-AC52-07NA27344.

## Abstract

In this project we explored enzyme-catalyzed methane conversion to methanol. Industrial biological approaches to methane conversion using whole organisms are predicted to be more energy efficient than chemical approaches, but are limited by mass transfer of the gas phase reactants, methane and oxygen, to the organisms. We demonstrated that 3D printing the enzyme particulate Methane Mono Oxygenase (pMMO) embedded in a polymer can improve the kinetics of methane to methanol conversion. This improvement was likely due to the ability to increase the surface area of the catalytic material using 3D printing. We also demonstrated the first continuous use of pMMO in a flow-through reactor. In order to understand the fundamental kinetic properties of pMMO, we conducted an in-depth study of pMMO kinetics using analytical tools developed in our lab. Finally, we developed a new copolymer system that allowed tuning of the gas permeability of the biocatalytic material.

## Background and Research Objectives

Natural gas flaring results in 13 million tons of CO<sub>2</sub> released annually. A technology to efficiently convert methane to other hydrocarbons is highly sought-after as a profitable way to convert “stranded” sources of methane and natural gas (sources that are small, temporary, or not close to a pipeline) to liquids for further processing. Such a technology would also provide an incentive to reduce methane emissions from landfills and wastewater treatment plants.

Methane is a potent greenhouse gas, with about 70 fold higher

global warming potential than CO<sub>2</sub>, and emissions must be reduced to achieve Paris Agreement climate goals. The only known true catalysts (industrial or biological) to convert methane to methanol under ambient conditions with 100% selectivity are mono-oxygenase enzymes (MMOs), which convert methane to methanol in methanotrophic bacteria. Therefore, industrial biological methane conversion is projected to have significantly lower energy and capital costs than chemical conversion.

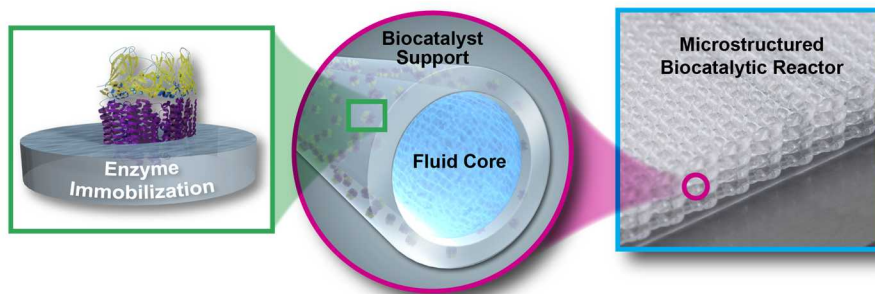


Figure 1. A biocatalytic reactor is fabricated by printing enzymes embedded in a polymer. The reactor consists of a lattice of tubes for continuous product removal and has a high gas/biocatalytic contact area to improve mass transfer and throughput.

However, the current reactors for biological methane conversion have low throughput and suffer from mass transfer limitations.

In this project we sought to explore **1)** the properties and kinetics of isolated pMMO for conversion of methane to methanol, which enables a higher carbon conversion efficiency than whole cell conversions, and **2)** how new polymeric materials and **3)** manufacturing methods impact pMMO activity and enable new reactor design to overcome throughput and mass transfer limitations. (An example reactor design is shown in Figure 1). We have achieved all of the goals of the project; we have filed one patent and published one high impact paper describing results from objective 3 (Blanchette et al 2016). We plan to submit two additional papers describing results from objectives 1 and 2.

## **Scientific Approach and Accomplishments**

### **Objective 1. Kinetics of pMMO**

Despite decades of research into determining the location and nature of the active site in pMMO, an in-depth study of the enzyme kinetics has not yet been published. However, in order to design materials and reactors around methane oxidation by pMMO, the enzyme kinetics must be understood. For example, a reactor might be designed to improve mass transfer, or to reduce product inhibition, if these kinetic and mechanistic parameters were known. Therefore, one of the core objectives of this project was to uncover the kinetics, and therefore the overall mechanism and rate limiting steps involved in methane oxidation catalyzed by this remarkable enzyme.

We developed two new methods for measuring the kinetics of pMMO: 1) a new stopped-flow spectrophotometric assay that measures the rate of reducing agent (NADH) consumption as a proxy for methanol production, and 2) a measurement of the rate as a drop in pressure in the headspace over a reaction vial containing pMMO and the gas phase reactants over time. Importantly, we have verified that the kinetic parameters determined from both of these methods were similar. We have tested several potential mechanisms against the data, and found a mechanism that corresponded well. This mechanism suggests that activation of oxygen is likely the slow step in methane oxidation, and that this oxidation step is irreversible. Furthermore, the magnitude of the kinetic constants show when the concentrations of reactants become limiting, which will be informative for reactor design. Finally, the data indicate that product inhibition is an important factor which influences pMMO activity. These findings will have implications for design of both methane oxidation catalysts and bioreactors.

## Objective 2. Polymer design: tuning the properties of the biocatalytic material

The nearly limitless range of properties accessible using synthetic polymers and hydrogels can be coupled with the extraordinary catalytic properties of enzymes to create biocatalytic materials with enhanced function. We sought to create such a biocatalytic material by embedding (crosslinking) pMMO in polymers with tunable properties. Because methane conversion requires gas phase reactants, we focused on tuning the gas permeability. After finding that pMMO retained up to 100% physiological activity in a polymer composed of polyethylene glycol (PEG), we focused on synthesizing polymers that contained amphiphilic mixtures of PEG, to retain enzyme activity, and polydimethyl siloxane (PDMS), to provide gas permeability.

A critical challenge in the synthesis of PDMS/PEG block copolymers suitable for crosslinking enzymes is that the material must be water soluble in order to mix it with aqueous enzymes, and must be curable under mild conditions in order to preserve the enzyme

activity. In fact, to our knowledge there are no reports of crosslinking enzymes in PEG-PDMS block copolymers in the literature. We discovered that certain random brush

copolymers satisfied both of these criteria. The PDMS-PEG copolymer materials had 10 fold higher gas permeability than PEG alone. Our mesoscale model of one of the polymer variants, corroborated by x-ray scattering data, indicates that the polymer forms a network of PDMS and PEG domains at high volume percent of polymer (>50%) in water (Figure 2, inset). We successfully incorporated pMMO into this polymer and measured the activity as a function of polymer volume percent. Our major finding was that the activity, while lower than that of pMMO in PEG at lower polymer concentrations, increased as the polymer concentration increased, the opposite of the trend seen in PEG (Figure 2). Combined, these results indicate that tuning the network

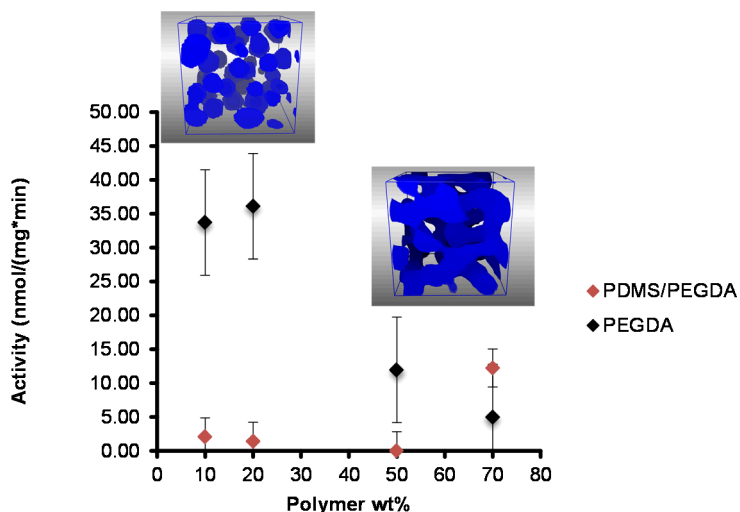


Figure 2. Simulations and x-ray data indicate that as the polymer concentration increases, the PDMS-PEG copolymer changes from a micellar structure with isolated PDMS domains (inset, left) to a continuous network structure, providing a pathway for gas permeability (inset, right). Remarkably, the activity of pMMO increased when the copolymer was in the network structure, indicating that this structure was necessary for transport of reactants and products.

structure of polymers at the nanoscale may allow tuning of the biological activity of enzymes embedded in them.

### Objective 3. Tuning the polymer/reactor architecture using 3D printing

In an effort to develop a biocatalytic material that can be molded into controlled, predetermined structures with tunable surface areas, we created a polymeric material based on PEG, which could be embedded with pMMO and used as a feedstock for 3D printing (Figure 3). We

discovered that at low polymer concentrations the pMMO-PEG material had the same catalytic activity as pMMO in the cell. We explored printing this material into geometries with varying surface area, and found that the pMMO activity increased as the surface area of the material increased. (Figure 4) Furthermore, we found that this printed material allowed high loadings of pMMO, enabling high volumetric productivity. Our results with printed pMMO show that the strategy of printing catalysts into reactor geometries that are designed for maximally efficient use of reactants, energy, and reactor volume has

promise both for methane conversion and other biocatalytic processes. The synthesis of the catalytic polymer material allowed pMMO to be used in a continuous reactor for the first time rather than in a batch process.

The ability to 3D print the pMMO membrane allowed a comparison of different membrane thickness suspended at the gas-liquid interface. We found that thinner printed membranes produced more methanol over time, consistent with the previous results that suggested the pMMO is mass-transfer limited, and increasing the surface area is a promising approach to utilizing pMMO most effectively.

### Impact on Mission

This project is directly relevant to the Laboratory's focus on Materials for Energy. Specifically, it has increased Laboratory expertise in the strategic focus areas of climate



Figure 3. Printed pMMO structures with varying surface area to volume ratios.

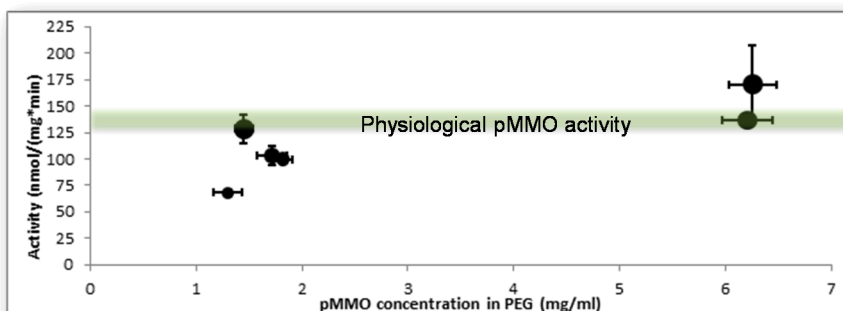


Figure 4. pMMO activity as a function of pMMO concentration in PEG. Larger bubbles correspond to higher surface area to volume ratios.

and energy security and engineered materials. In addition, the project demonstrated a new application for the Laboratory's capability in engineered/manufactured materials: advanced manufactured reactors. We will continue to explore this area for other energy-relevant applications. We have hired/supported two new postdoctoral staff members on this project.

## **Conclusion**

We have demonstrated that polymer synthesis and advanced manufacturing enable the properties of a biocatalytic material to be tuned to the application. Specifically, we showed that pMMO can be embedded in a polymer while retaining its native activity, and that printing this biocatalytic polymer into high surface area structures with high enzyme loading improves this activity. The kinetic model that we developed for pMMO will be instructive both for a fundamental understanding of the enzyme, and the design of bioreactors that use pMMO. The advanced manufactured bioreactor concept can be applied to many applications, including microbial electrosynthesis and syngas fermentation. Co-development of the materials, the manufacturing, and the biocatalysts is needed for commercial realization of this technology and to expand the use of enzymes for energy applications.

## **References**

Blanchette, C. (2016), "Printable Enzyme-Embedded Materials for Methane to Methanol Conversion," *Nature Communications*, **7**, 11900.

This work was performed under the auspices of the U.S. Department of Energy by Lawrence Livermore National Laboratory under Contract DE-AC52-07NA27344