



Tethered Supported Lipid Bilayers Understanding Viral Entry

Nicole Zawada, University of California, San Diego, B.S. Human Biology, est. June 2014

Darryl Sasaki, Ph.D., Org: 8621, Biotechnology and Bioengineering Department

Carl Hayden, Ph.D., Org: 8353, Combustion Chemistry Department

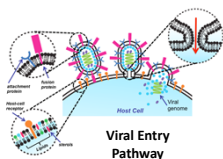
Sandia National Laboratories, U.S. Department of Energy

August 2, 2012



Background

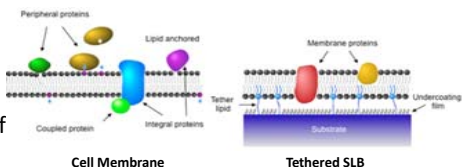
that makes up living organisms. It acts as a selectively permeable barrier responsible for regulating all substances that pass between the interior of the cell and the external environment. A major role of the cell membrane is to protect the cell from outside forces. However, the membrane is susceptible to viral entry.



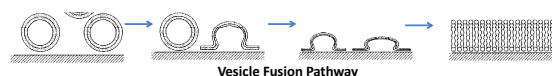
Pathogenic viruses are a serious threat to human health and national security. Our goal is to develop platforms for detecting and identifying viral particles, and understand pathways for infection.

Many biodefense priority viruses enter host cells by lipid membrane fusion. We are working on the development of cell membrane mimetics that can be used to detect and evaluate viral particle binding and fusion.

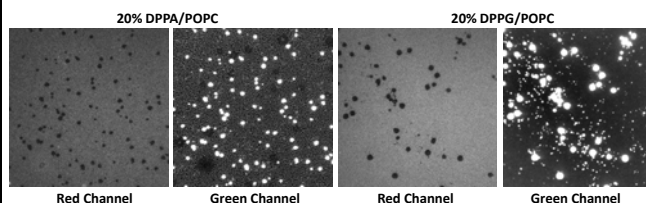
In order to study the binding and fusion process, simplified models of the cell membrane called supported lipid bilayers (SLBs) can be used. SLBs are phospholipid membranes adsorbed to a planar substrate. The presence of the substrate can cause unwanted interactions with the bilayer, so we are developing tethered SLBs (t-SLBs) to separate the SLB from the substrate.



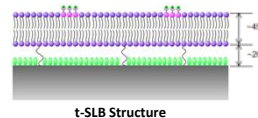
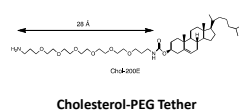
Results



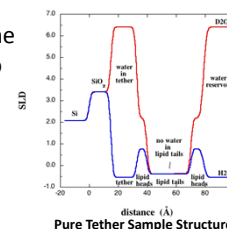
We formed SLBs on glass through vesicle fusion and adhesion, and demonstrated the formation of functionalized domains by selectively binding cationic polymers.



We are now working on constructing t-SLBs in order to improve membrane fluidity, protein function, and stability. A typical SLB on glass has a 10 Å water cushion, compared to a 20 Å cushion in our t-SLBs.

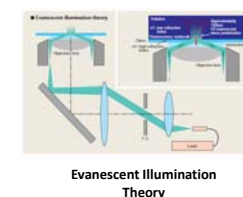
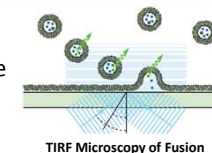


In order to study and characterize the t-SLBs, we used neutron reflectivity to observe their structure. Pure tether samples have been studied; we are in the process of studying t-SLBs with a cushioned substrate.

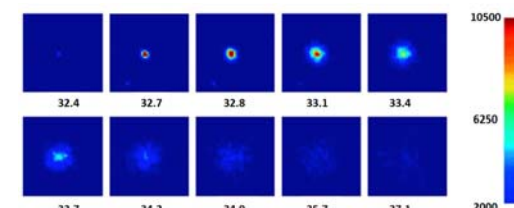


Conclusion

We will be using total internal reflection fluorescence (TIRF) microscopy to observe and study the fusion of viral particles to the membrane.



Previous work (below) has been done studying viral particle binding and fusion using typical SLBs. Our research will be taking this a step further and observing this binding and fusion on t-SLBs, which would more accurately mimic the cell membrane.



Single Particle (Sindbis) Hemifusion/Fusion Event on Supported Membrane
Wessels L., Elting M.W., Scimeca D., Weninger K., *Biophys. J.* (2007) 93: 526-538.

Acknowledgements: Research for this work was supported by the U.S. Department of Energy, Division of Materials Sciences and Engineering and the Laboratory Directed Research and Development program at Sandia.