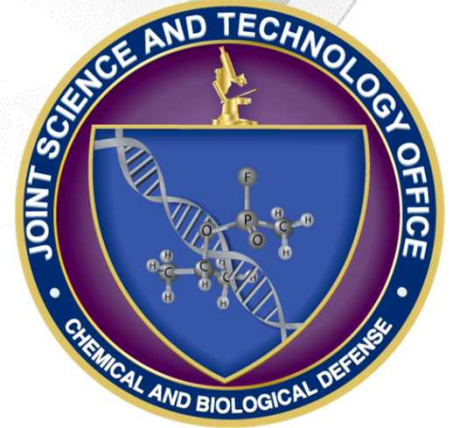


Real-Time, Autonomous Biosurveillance for Vector-Borne Viral Pathogens

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Project Overview

- Develop and field-test an *autonomous sensor* to detect presence of mosquito-borne viruses (West Nile, etc) with daily reporting
- Integrate sensor data into BSVE along with mapping & visualization software and predictive models.
- 3-year effort began 2014 (~16 months in)
- Partnership between Sandia National Laboratories (Livermore)
 - Systems engineering, microfluidic assays, statistical modeling
- ...and UC Davis Center for Vectorborne Diseases (CVEC)
 - Virology, entomology, and ecology of vectorborne disease
 - Integrated with public health and vector control districts in CA



Sandia National Laboratories

UC DAVIS
UNIVERSITY OF CALIFORNIA



Why arboviruses?

- “Arbovirus” = Arthropod-borne virus
 - Mostly RNA viruses; carried by mosquitoes, ticks, flies, *etc*
- West Nile swept the globe in a few years – what’s the next epidemic to emerge?
- Threat to military personnel overseas.



Dengue
Yellow fever
Chikungunya



West Nile virus*
St. Louis encephalitis virus*
Japanese encephalitis virus
Rift Valley fever virus
Equine encephalitis viruses
(WEE*, VEE, EEE)



Transmits malaria
(not a virus!)



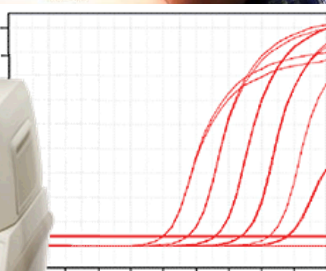
Ground truth for arboviruses

Vector surveillance



~2 weeks

Adulticide spraying



Molecular assays

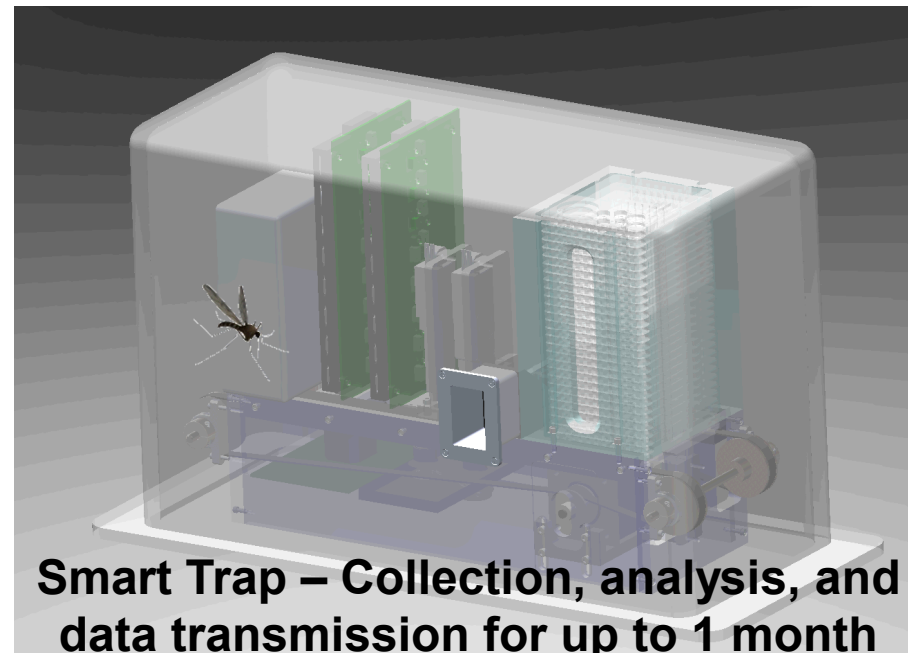


Sugar-based surveillance

- Besides blood, mosquitoes feed on sugar for energy.
- Infected mosquitoes deposit virus while sugar-feeding.
- We propose to exploit sugar-feeding in an *autonomous field-based detector*: the **Smart Trap**
- Needs to do a complex job, but be cheap enough to deploy in a network for spatial surveillance



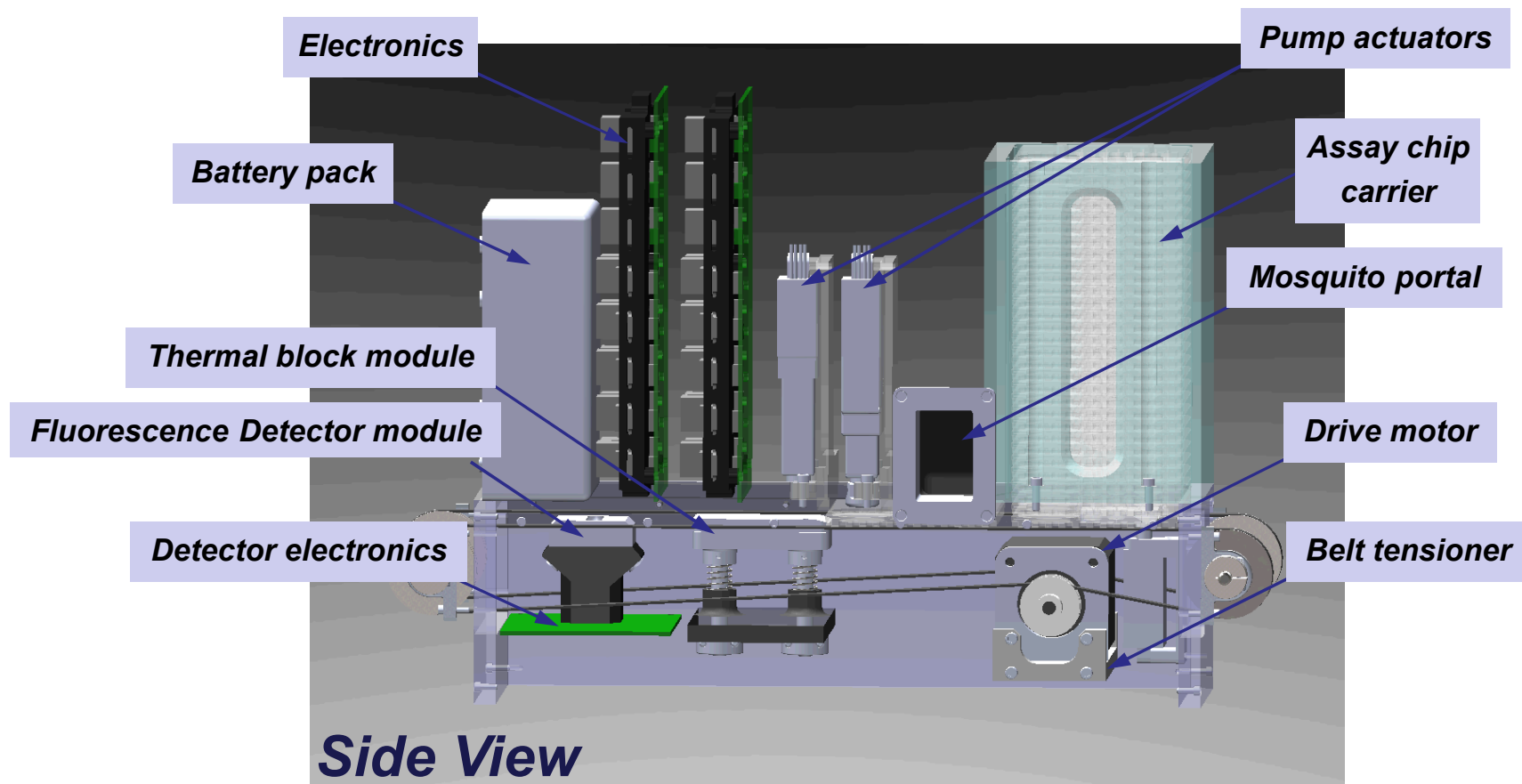
Sugar baits used for field collection followed by laboratory analysis



Smart Trap – Collection, analysis, and data transmission for up to 1 month



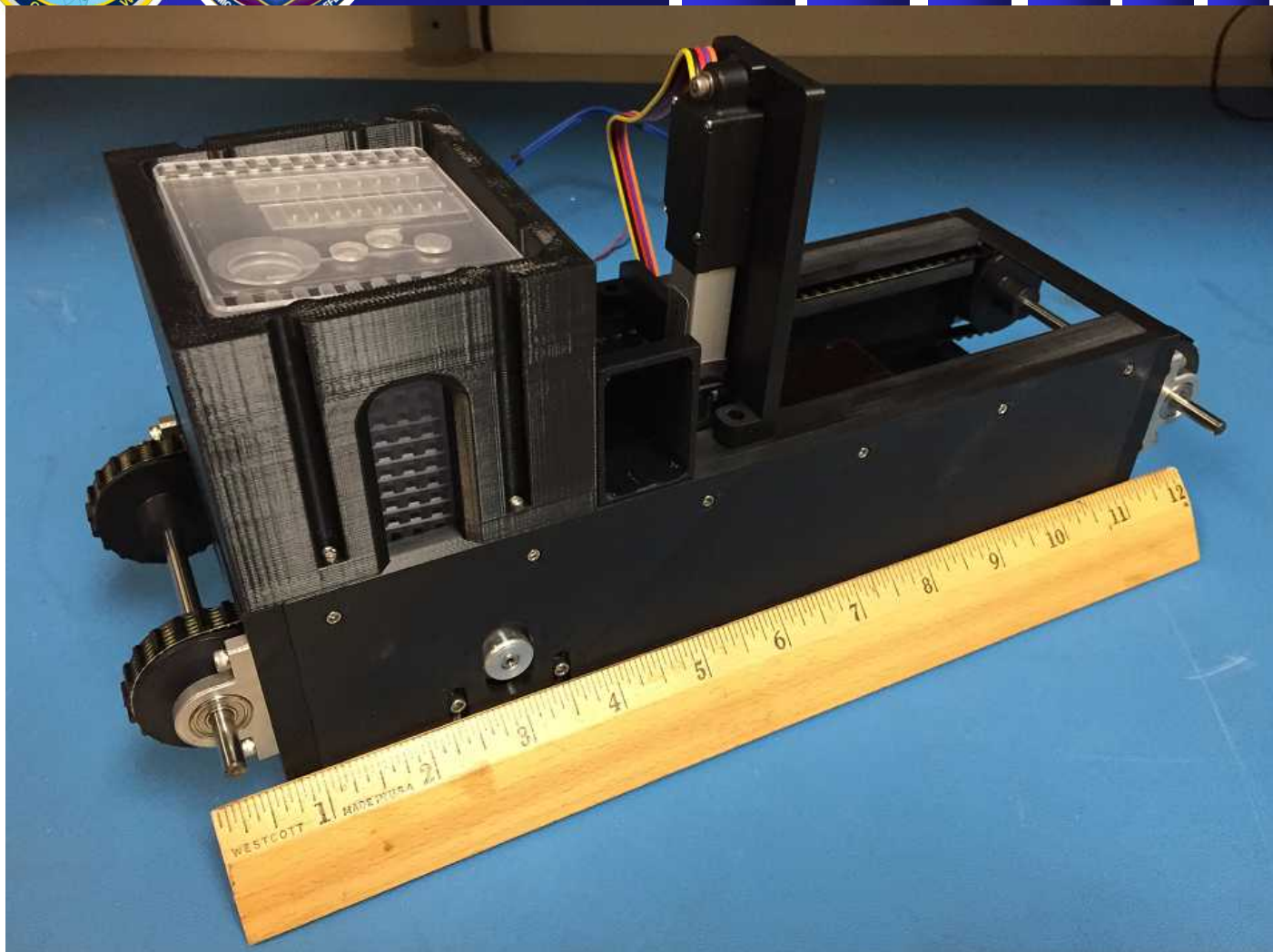
Smart Trap Concept



System cover and side covers removed for illustration purposes



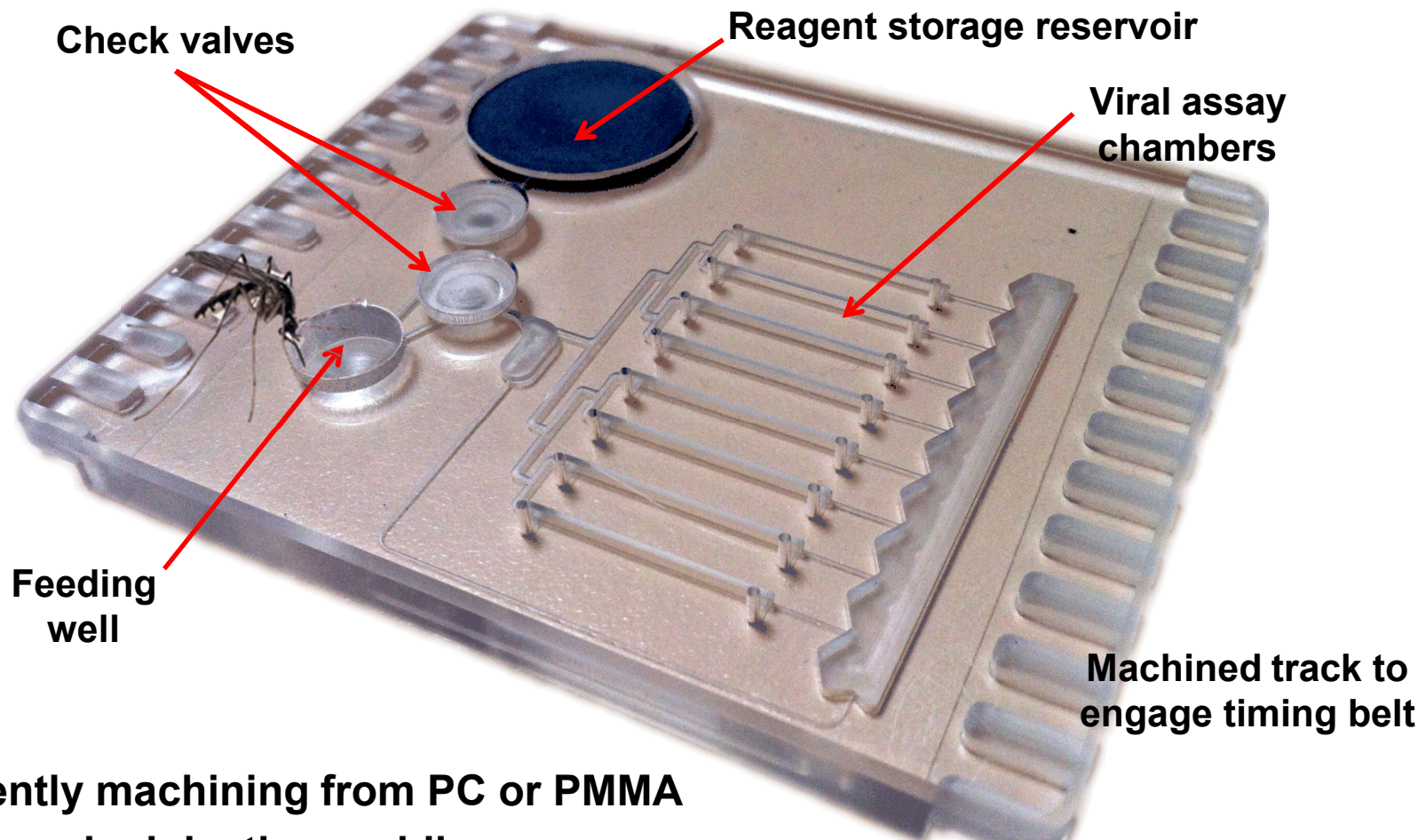
Smart Trap Prototype



Approximate size : 14 x 6 x 8 inches



Daily assay cartridge



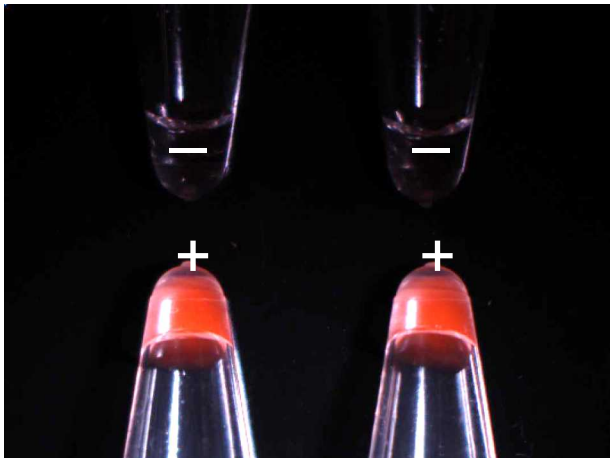
- Currently machining from PC or PMMA
- Scale-up by injection molding



Viral assays by RT-LAMP

- Isothermal amplification of viral RNA (63 °C)
- Requires minimal sample prep / RNA extraction not necessary
- Detects <0.1 PFU virus in ~30 minutes
- Created novel primer sets for WEEV and SLEV
- Novel Sandia-developed detection chemistry gives bright and distinctive fluorescence endpoint signal

WNV RT-LAMP endpoint



Implemented in Smart trap with a simple LED and photodiode detector

Also works with an LED flashlight and a color camera (e.g. smart phone) with a red plastic film as an emission filter.

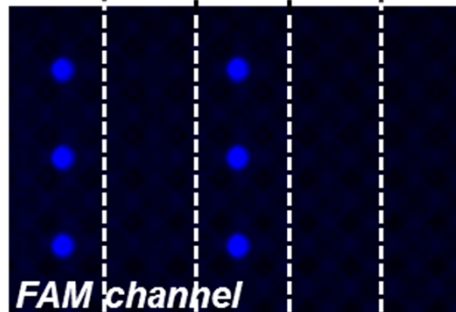


Potential for clinical diagnostic

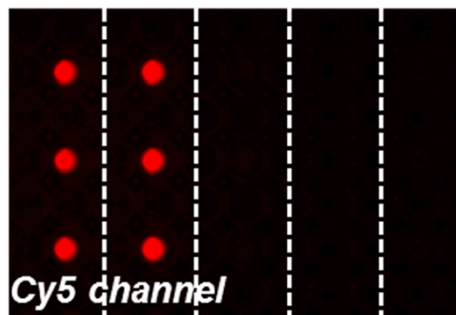
Single-tube multiplexing Ebola and *Plasmodium*

P. falc. + - + - -
EBOV + + - - -

Malaria
result



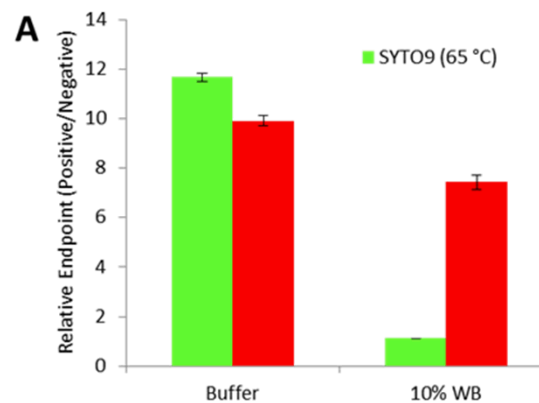
Ebola
result



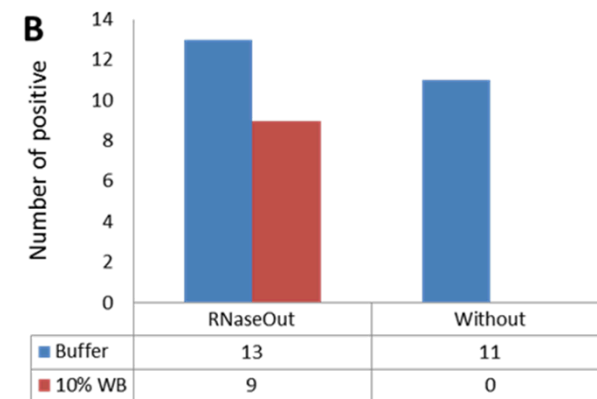
Adaptable to bright
visual readout

Newly designed RT-LAMP primers for Ebola GP gene target both historic (1976) and recent (2014) isolates; detect 200 copies in about 20 minutes

RT-LAMP with new detection technique enables detection of Ebola RNA directly in whole blood (10% of total reaction volume, no RNA extraction)



1000 copies EBOV RNA
(100% detection rate)

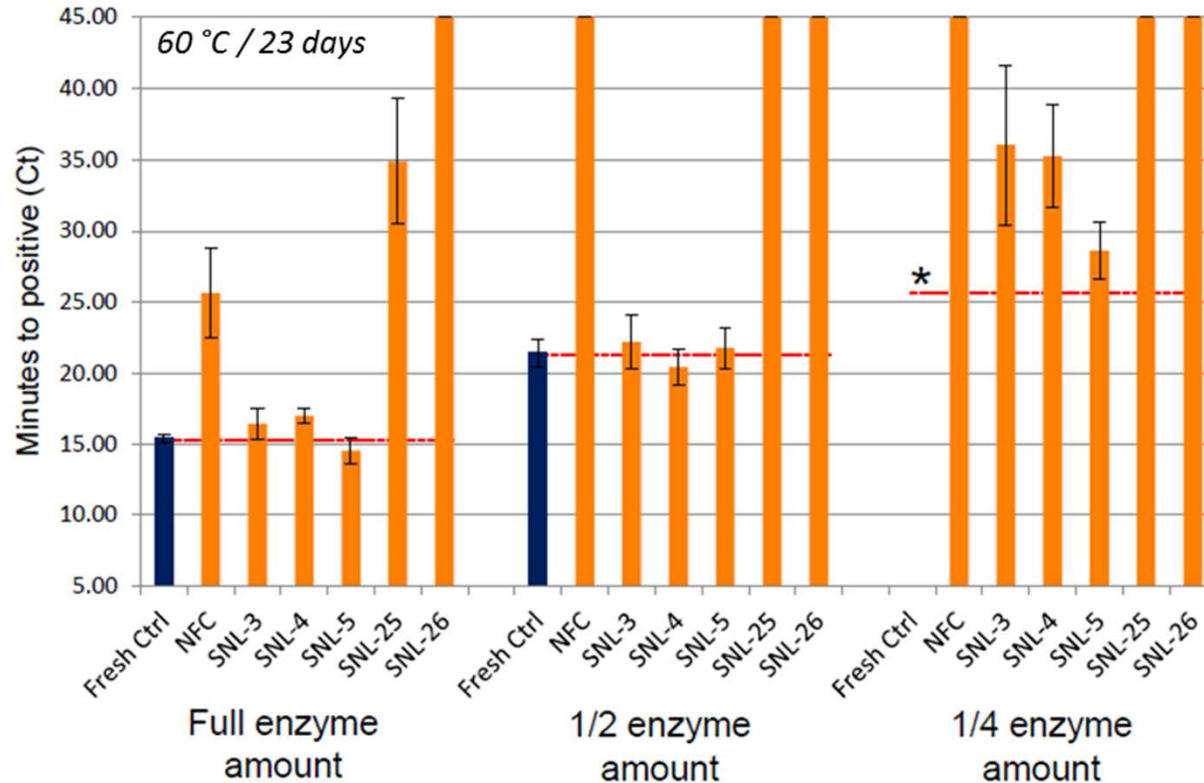


200 copies EBOV RNA ×
20 replicates per condition
~50% detection rate



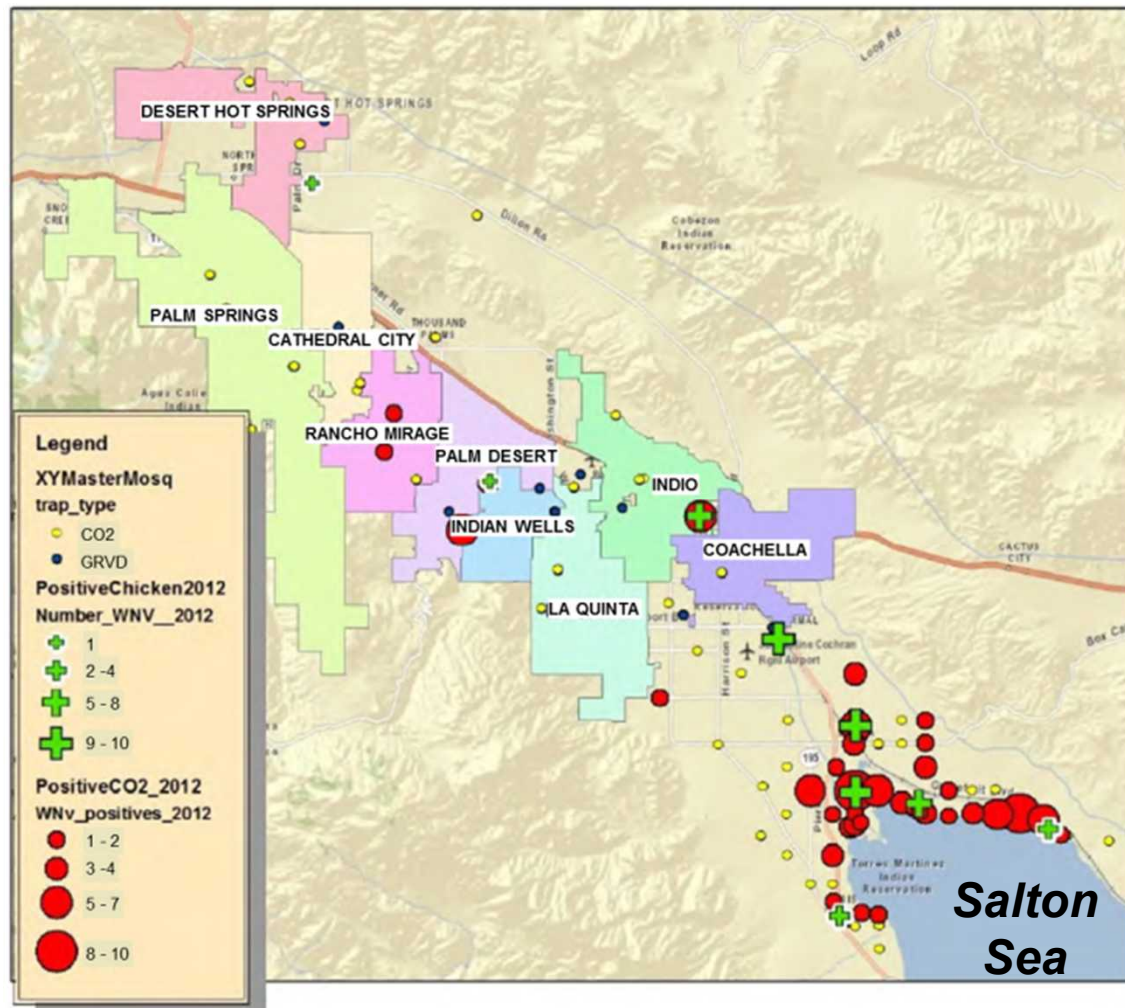
Field stabilized reagents

- Partnered with Biomatrix to formulate RT-LAMP assay reagents for long-term stability in dry form in assay chip; rehydrate with “bait fluid” to run assay.
- Minimal loss in activity after >3 weeks at 60 C, even with reduced enzyme





Field test planned 2016

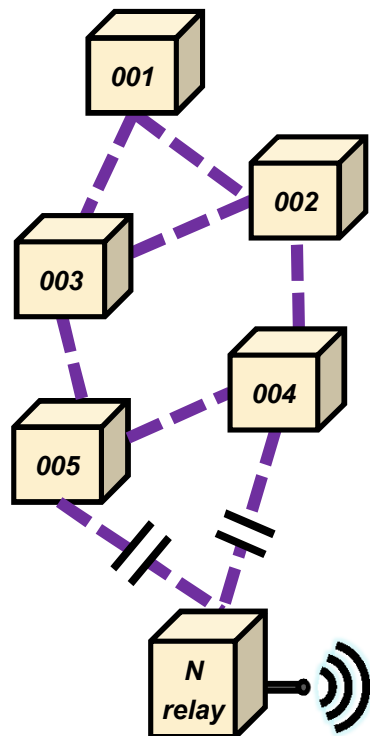


- Will deploy Smart Trap network near Salton Sea
- Perform field test concurrently with conventional vector surveillance for WNV (traps & sentinel chickens)



Trap wireless networking

Private wireless trap network



Cellular-enabled relay node

SMS-coded "result" messages

001A%k...

002z4m...

003B*r...

004ybA...

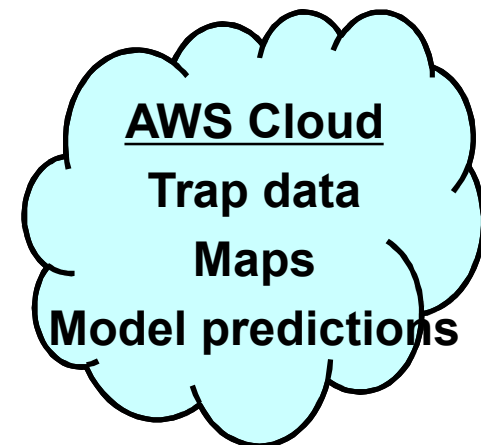
005xcD...

⋮

NNNj#z...



**Translate results using BSVE
PON diagnostic XML schema**

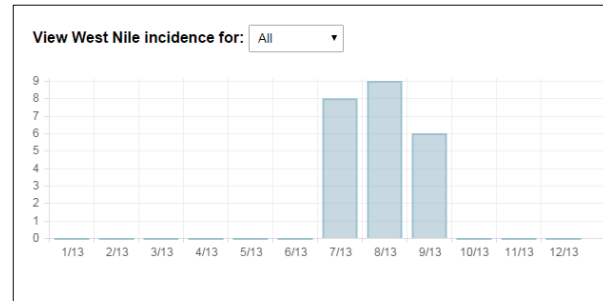


BSVE



AWS screenshot

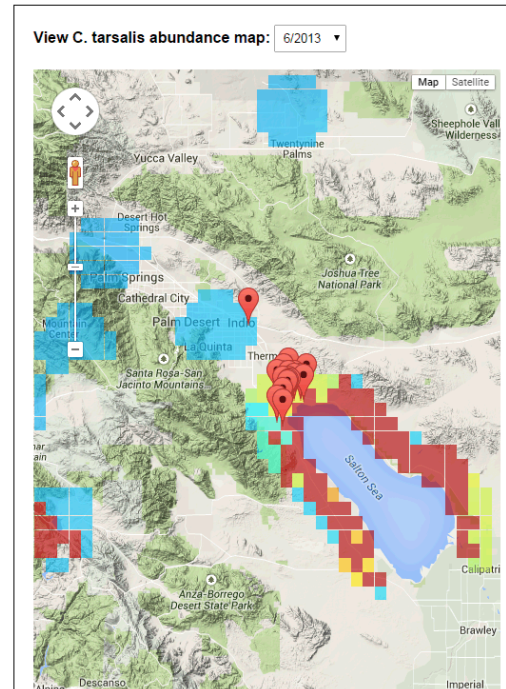
- Trap data stored using PON diagnostic XML schema
- Trap data viewable as time series, or search by location & date
- Behind the scenes: CA vector control data, environmental data, trap data, and statistical model (R script) produce vector abundance map and infection rate map
- Model is anchored at trap locations; interpolates through space



Trap Data

Search:

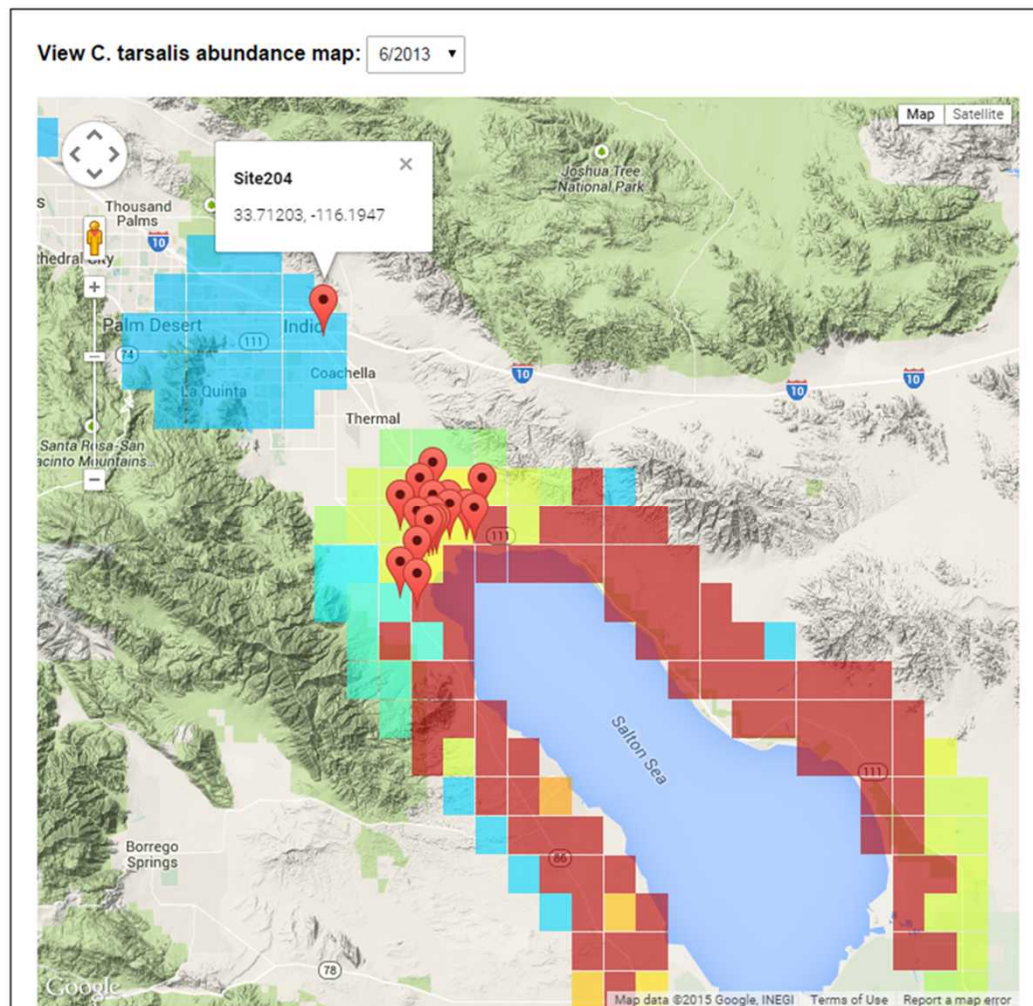
Time	Site ID	West Nile
1/2013	Site121	Negative
1/2013	Site13	Negative
1/2013	Site17	Negative
1/2013	Site204	Negative
1/2013	Site30	Negative
1/2013	Site33	Negative
1/2013	Site34	Negative
1/2013	Site35	Negative





AWS site visualizations

- Pushpins at trap locations
- Model predictions overlaid on map
- Map results selectable by date





Future prospects

- Just beginning year 2 of 3-year Smart Trap effort
- Maturing hardware & field testing components
- How to attract the most mosquitoes?
- Correlating sugar-feeding to conventional trapping & surveillance
- Adapting to other targets
 - Other viruses (Dengue, chikungunya, JEV, etc)
 - Plasmodium parasites
 - Other arthropod vectors: flies, ticks, etc?
- RT-LAMP / LAMP assay useful for simple yet sensitive PON test
 - human/veterinary use
 - virus, bacteria, parasite, drug resistance/virulence genes
 - Compatible with blood, saliva, urine, stool, etc.
 - Stabilization without cold chain