

# *Real-Time, Autonomous Biosurveillance for Vector-Borne Viral Pathogens*

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# Project Overview

- Develop and field-test an *autonomous sensor* to detect presence of mosquito-borne viruses (West Nile, etc) with daily reporting
- Integrate sensor data into BSVE along with mapping & visualization software and predictive models.
- 3-year effort began 2014 (~16 months in)
- Partnership between Sandia National Laboratories (Livermore)
  - Systems engineering, microfluidic assays, statistical modeling
- ...and UC Davis Center for Vectorborne Diseases (CVEC)
  - Virology, entomology, and ecology of vectorborne disease
  - Integrated with public health and vector control districts in CA



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# Why arboviruses?

- “Arbovirus” = Arthropod-borne virus
  - Mostly RNA viruses; carried by mosquitoes, ticks, flies, etc
- West Nile swept the globe in a few years – what’s the next epidemic to emerge?
- Threat to military personnel overseas.



*Aedes aegypti*  
Dengue  
Yellow fever  
Chikungunya



*Culex* spp.  
West Nile virus\*  
St. Louis encephalitis virus\*  
Japanese encephalitis virus  
Rift Valley fever virus  
Equine encephalitis viruses  
(WEE\*, VEE, EEE)



*Anopheles* spp.  
Transmits malaria  
(not a virus!)



# Ground truth for arboviruses

## Vector surveillance



»» **~2 weeks**



## Molecular assays

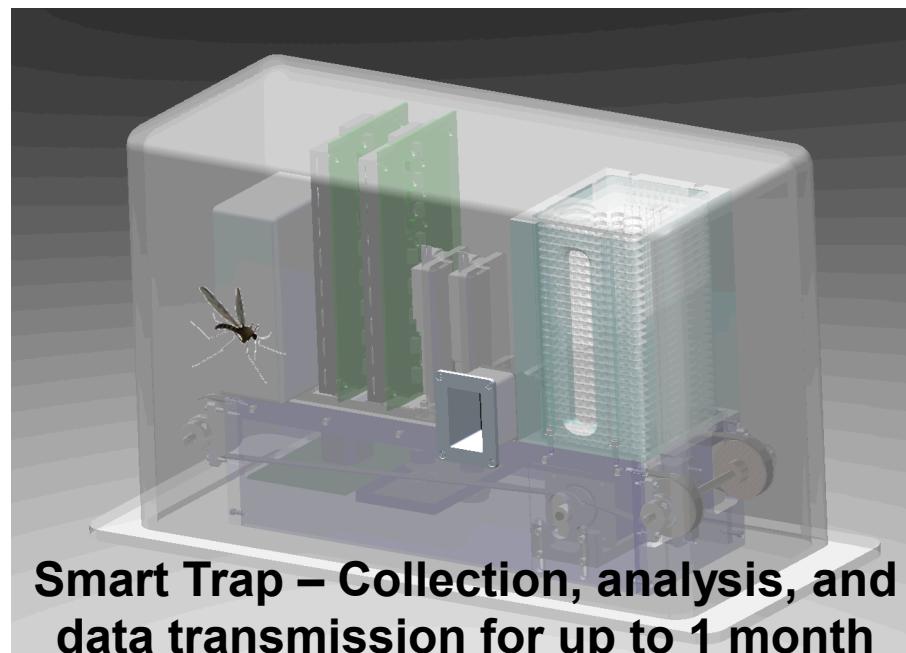
## Adulticide spraying





# Sugar-based surveillance

- Besides blood, mosquitoes feed on sugar for energy.
- Infected mosquitoes deposit virus while sugar-feeding.
- We propose to exploit sugar-feeding in an *autonomous field-based detector*: the **Smart Trap**
- Needs to do a complex job, but be cheap enough to deploy in a network for spatial surveillance

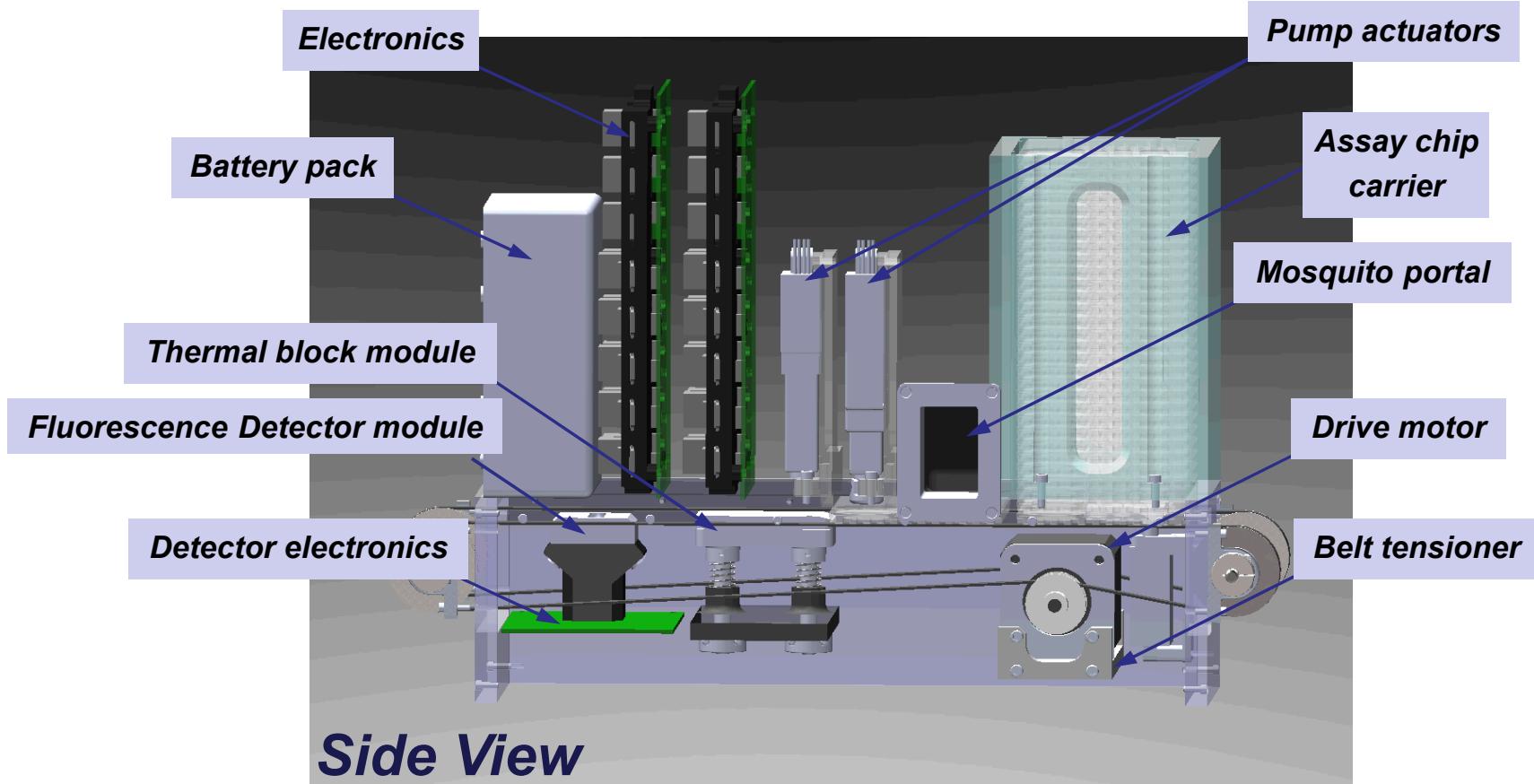


Sugar baits used for field collection  
followed by laboratory analysis

Smart Trap – Collection, analysis, and  
data transmission for up to 1 month



# Smart Trap Concept

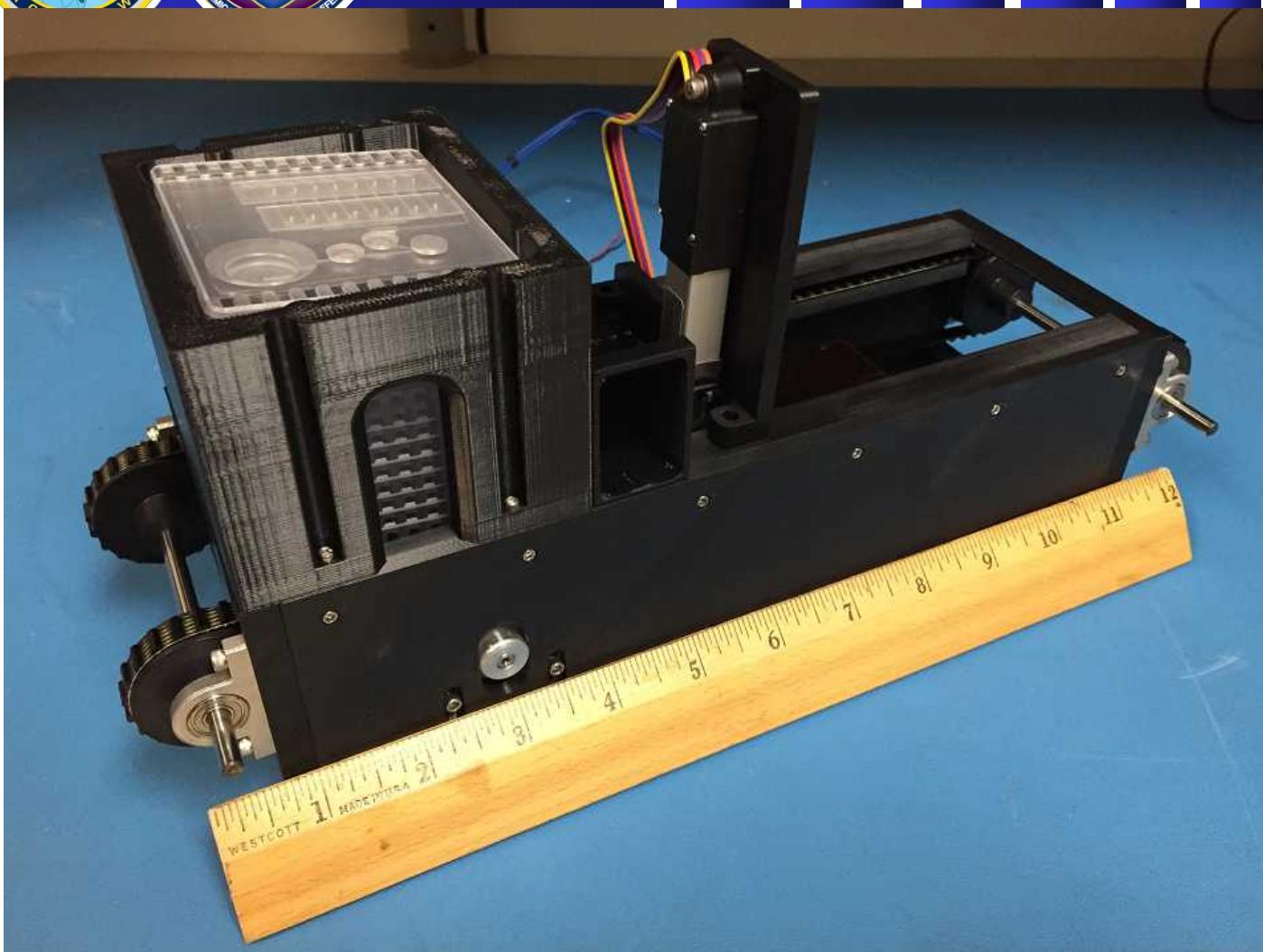


*System cover and side covers removed for illustration purposes*

UNCLASSIFIED



# Smart Trap Prototype



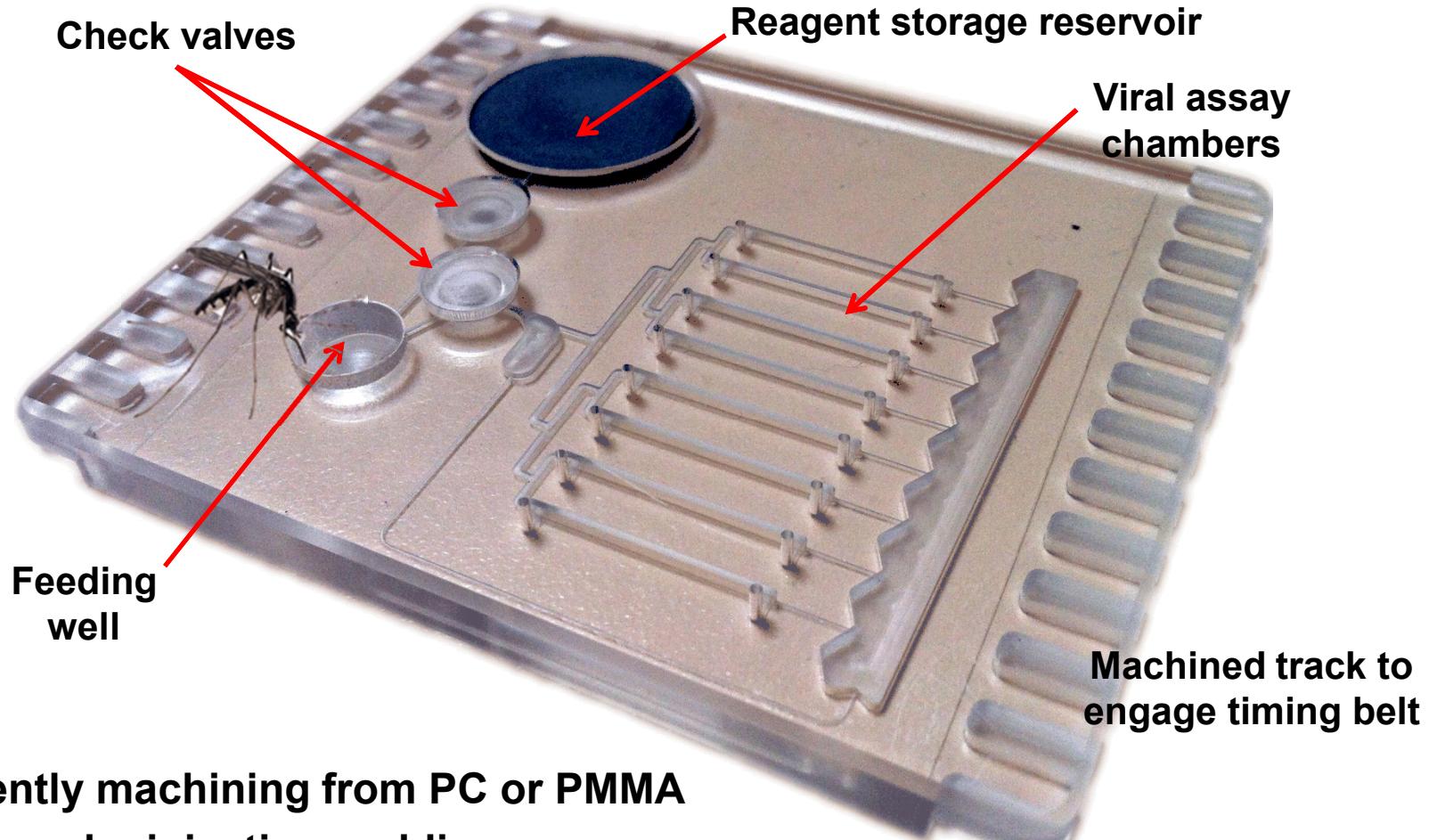
Approximate size : 14 x 6 x 8 inches

UNCLASSIFIED

R.F.Renzi 02/2015



# Daily assay cartridge



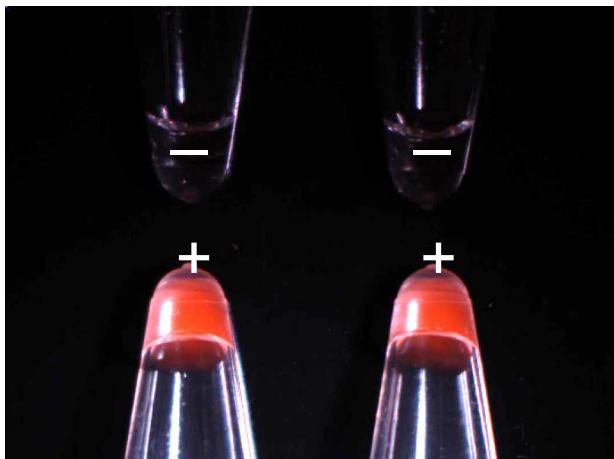
- Currently machining from PC or PMMA
- Scale-up by injection molding



# Viral assays by RT-LAMP

- Isothermal amplification of viral RNA (63 °C)
- Requires minimal sample prep / RNA extraction not necessary
- Detects <0.1 PFU virus in ~30 minutes
- Created novel primer sets for WEEV and SLEV
- Novel Sandia-developed detection chemistry gives bright and distinctive fluorescence endpoint signal

## WNV RT-LAMP endpoint

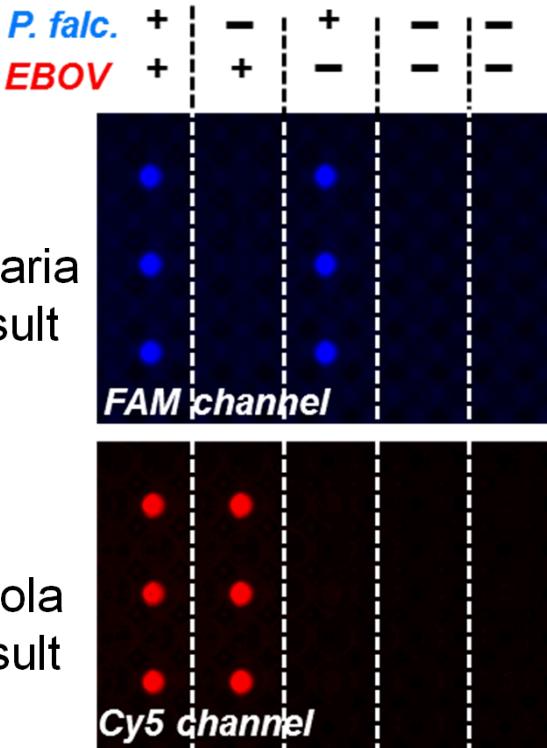


Implemented in Smart trap with a simple LED and photodiode detector  
Also works with an LED flashlight and a color camera (e.g. smart phone) with a red plastic film as an emission filter.



# Potential for clinical diagnostic

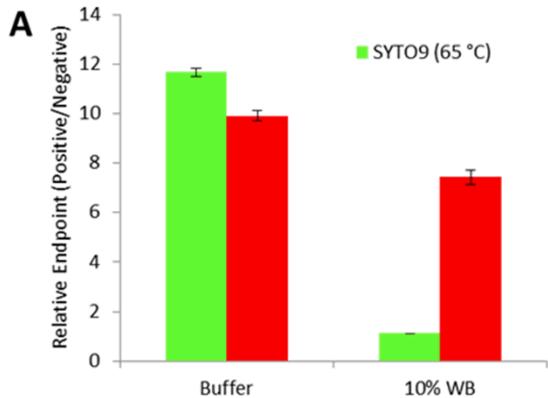
## Single-tube multiplexing Ebola and *Plasmodium*



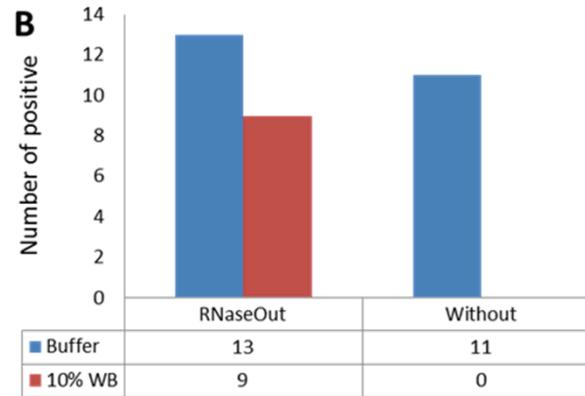
Adaptable to bright visual readout

Newly designed RT-LAMP primers for Ebola GP gene target both historic (1976) and recent (2014) isolates; detect 200 copies in about 20 minutes

RT-LAMP with new detection technique enables detection of Ebola RNA directly in whole blood (10% of total reaction volume, no RNA extraction)



1000 copies EBOV RNA  
(100% detection rate)

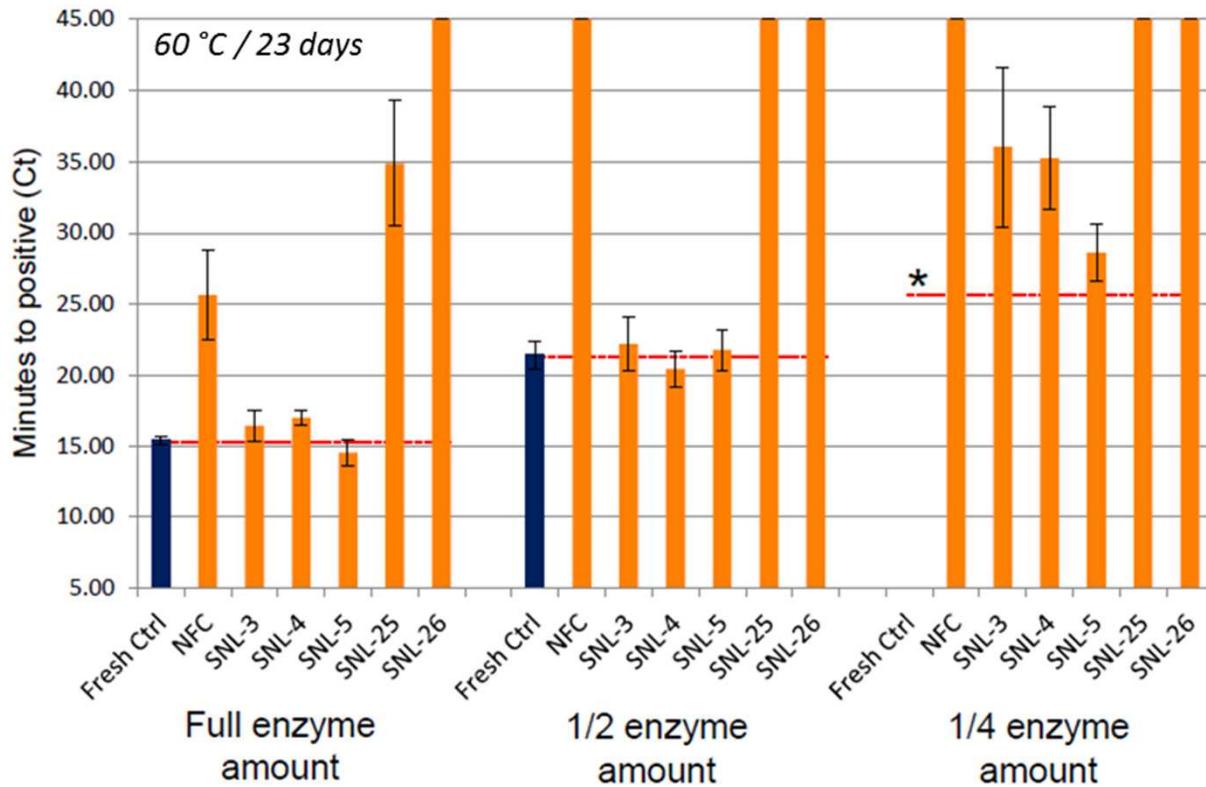


200 copies EBOV RNA ×  
20 replicates per condition  
~50% detection rate



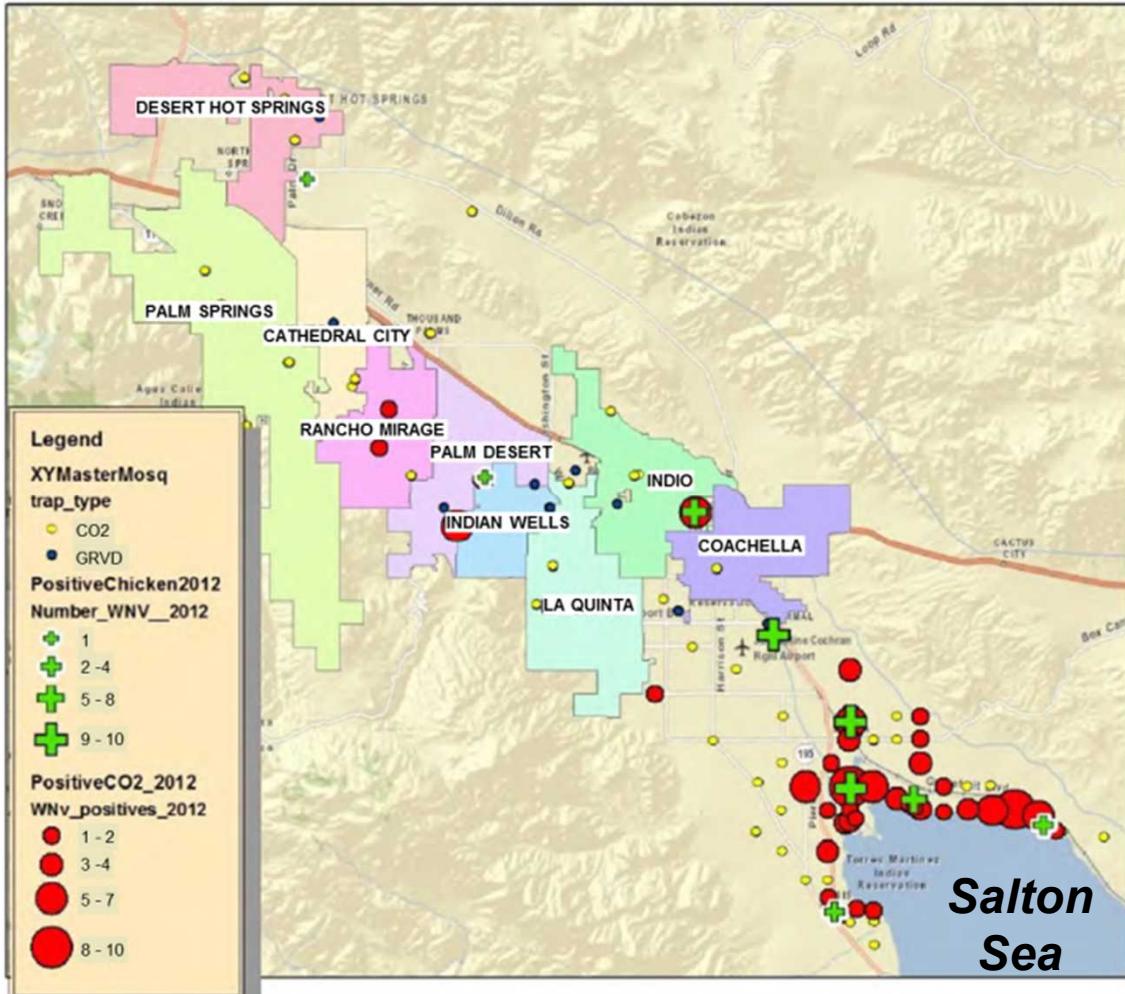
# Field stabilized reagents

- Partnered with Biomatrica to formulate RT-LAMP assay reagents for long-term stability in dry form in assay chip; rehydrate with “bait fluid” to run assay.
- Minimal loss in activity after >3 weeks at 60 C, even with reduced enzyme





# Field test planned 2016

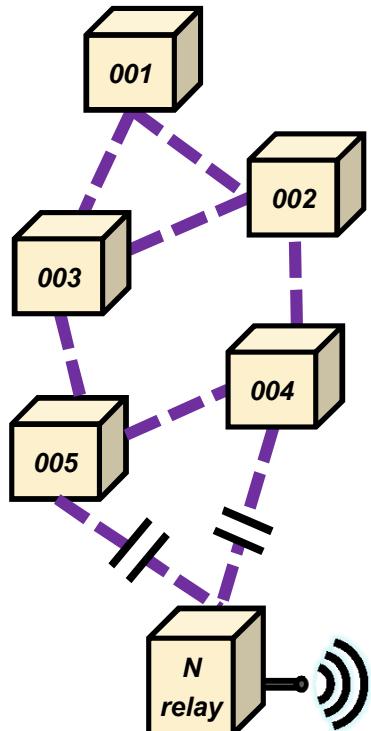


- Will deploy Smart Trap network near Salton Sea
- Perform field test concurrently with conventional vector surveillance for WNV (traps & sentinel chickens)



# Trap wireless networking

## Private wireless trap network

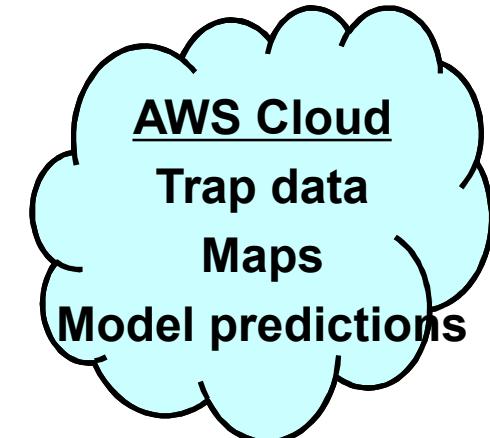


## SMS-coded “result” messages

001A%k...  
002z4m...  
003B\*r...  
004ybA...  
005xcD...  
⋮  
NNNj#z...



## Translate results using BSVE PON diagnostic XML schema



**BSVE**



# AWS screenshot

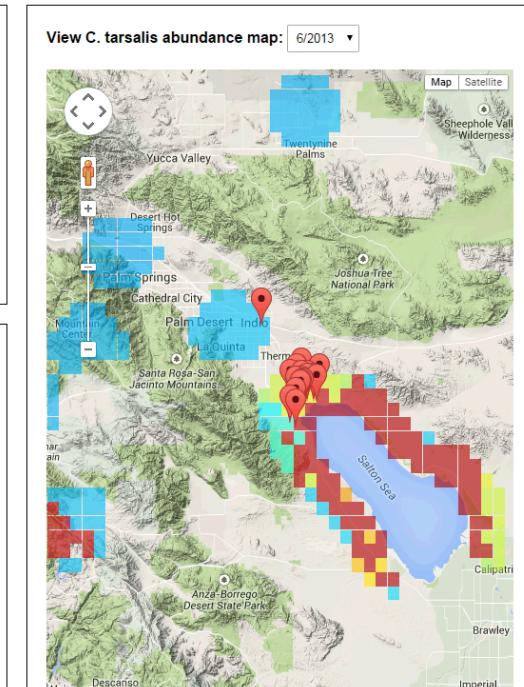
- Trap data stored using PON diagnostic XML schema
- Trap data viewable as time series, or search by location & date
- Behind the scenes: CA vector control data, environmental data, trap data, and statistical model (R script) produce vector abundance map and infection rate map
- Model is anchored at trap locations; interpolates through space



Trap Data

Search:

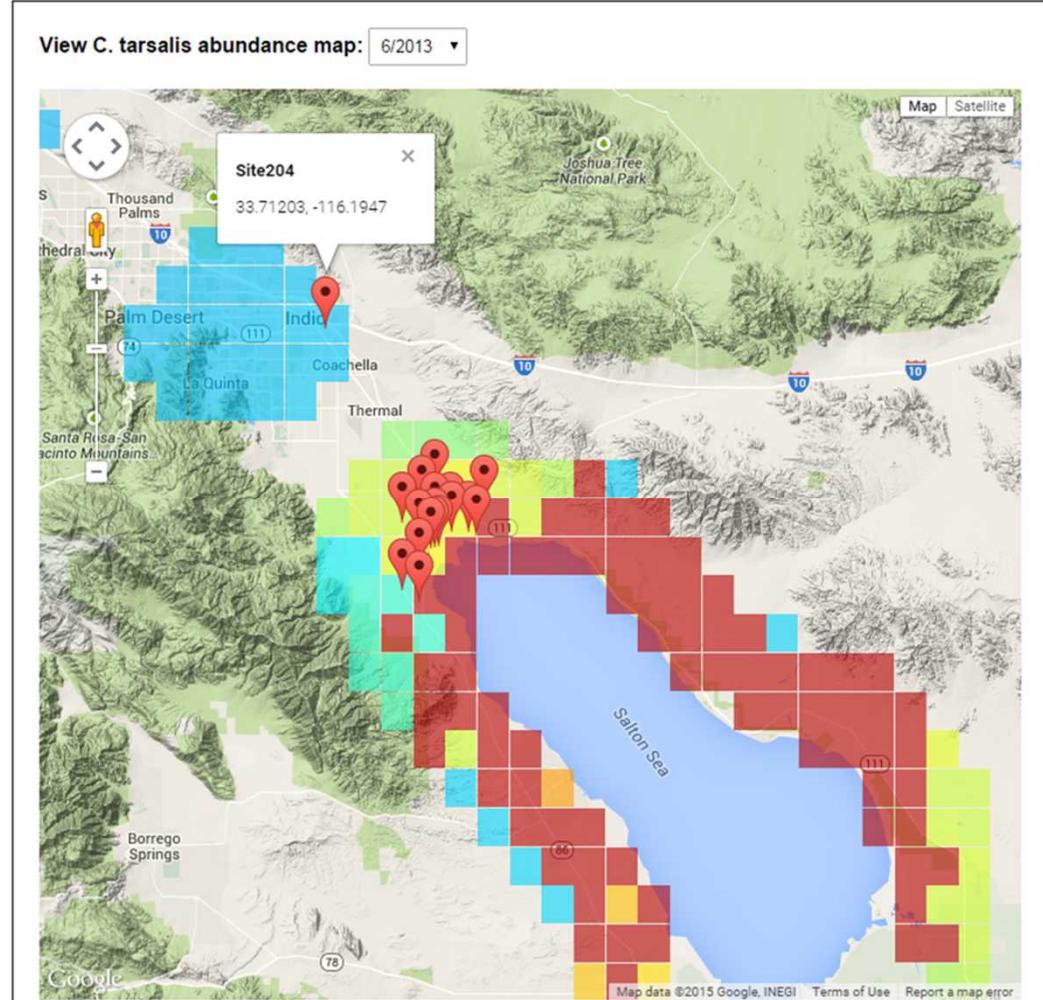
Time	Site ID	West Nile
1/2013	Site121	Negative
1/2013	Site13	Negative
1/2013	Site17	Negative
1/2013	Site204	Negative
1/2013	Site30	Negative
1/2013	Site33	Negative
1/2013	Site34	Negative
1/2013	Site35	Negative





# AWS site visualizations

- Pushpins at trap locations
- Model predictions overlayed on map
- Map results selectable by date





# Future prospects

- Just beginning year 2 of 3-year Smart Trap effort
- Maturing hardware & field testing components
- How to attract the most mosquitoes?
- Correlating sugar-feeding to conventional trapping & surveillance
- Adapting to other targets
  - Other viruses (Dengue, chikungunya, JEV, etc)
  - Plasmodium parasites
  - Other arthropod vectors: flies, ticks, etc?
- RT-LAMP / LAMP assay useful for simple yet sensitive PON test
  - human/veterinary use
  - virus, bacteria, parasite, drug resistance/virulence genes
  - Compatible with blood, saliva, urine, stool, etc.
  - Stabilization without cold chain