

# Visualizing Early Immune Response to Bacterial Infection: Reorganization at the Nanoscale

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# Outline

**Motivation:** Innate Immunity and Toll-Like Receptor (TLR) signaling

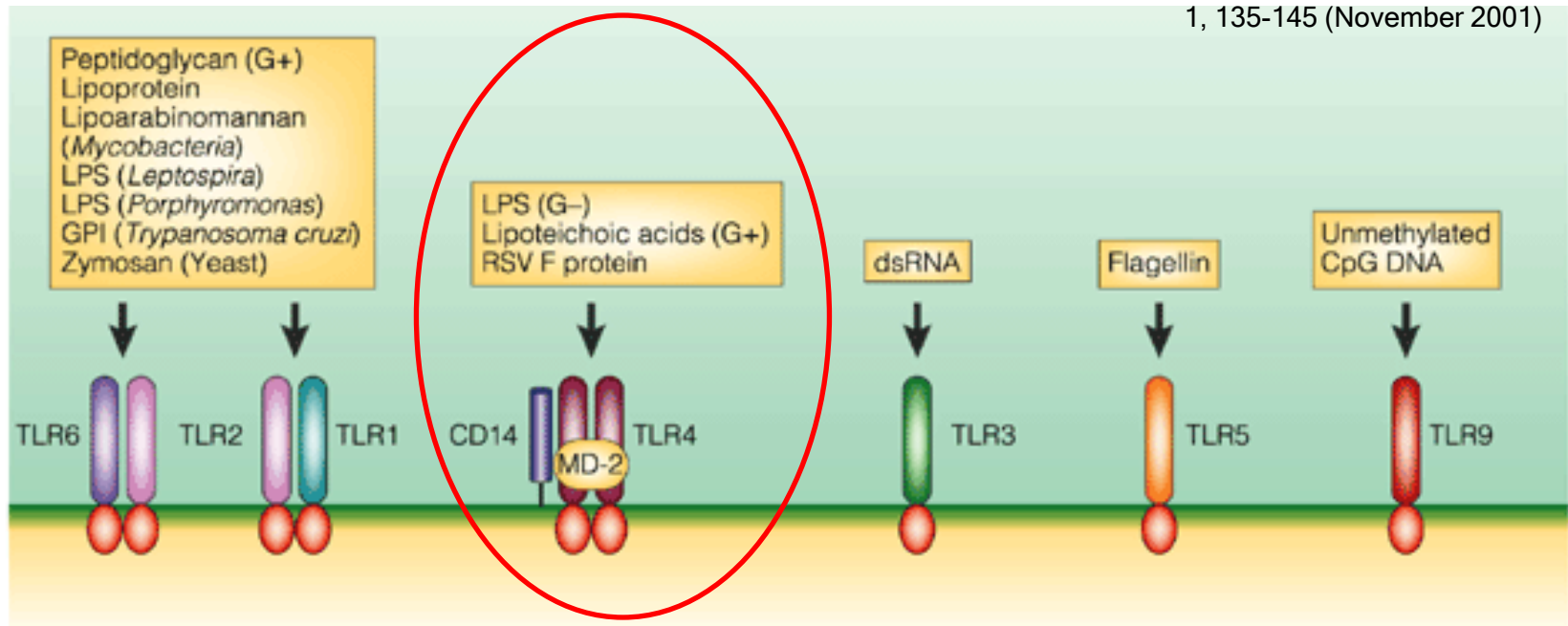
**Methods:** Stochastic Optical Reconstruction Microscopy (STORM)

**Results:** Detection of receptor clustering and co-localization of receptors with antigen

**Conclusions**

# TLRs: Important in Pathogenesis, Biodefense

Nature Reviews | Immunology  
1, 135-145 (November 2001)

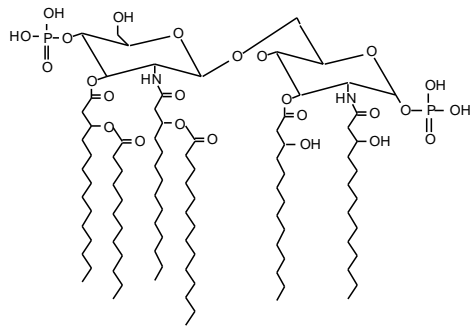


- Important element in mammalian innate immunity
- TLR4 recognizes LPS, starts signaling cascade
- LPS aided by accessory proteins
- Different chemotypes of LPS generate distinct immune responses

# Chemotypes of LPS Exhibit Differential Immune Response

## *Escherichia coli* (control)

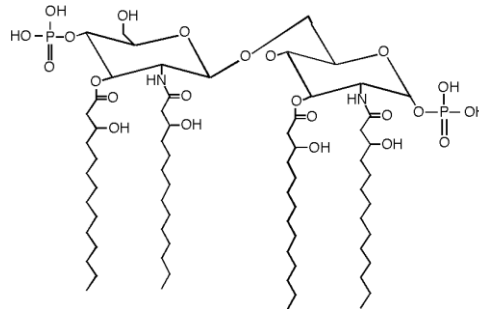
Smooth  
O-polysaccharide



Bind Surface  
+  
↑Stimulatory

## *Yersinia pestis* (37°)

Rough  
O-polysaccharide



Bind Surface  
+  
↓Stimulatory

Differential immune response observed is not fully understood.

- LPS from *E. coli* binds & produces an immune response
- LPS from *Y. pestis* (plague @ 37 °) binds, but does not

Are there clues in the nano-scale arrangement of the early immune response at the membrane interface?

# Receptor Clustering can be Necessary Component of Immune Response

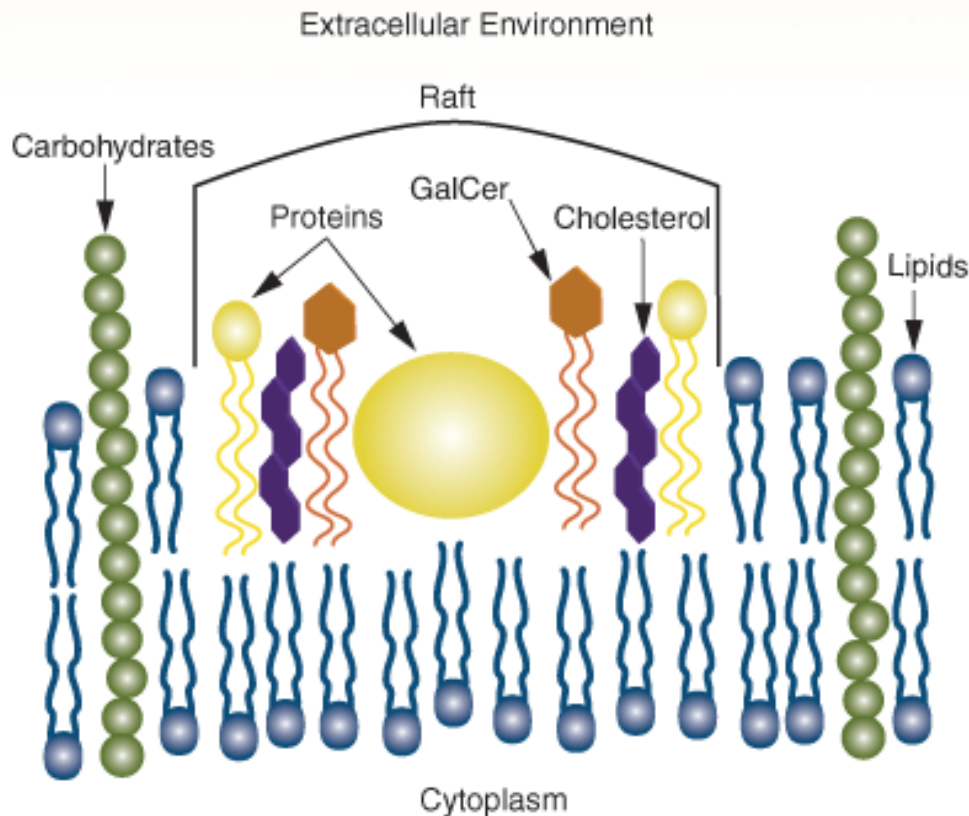


Image Courtesy of Tim Ratto, Lawrence Livermore National Labs

- Domains act as assembly areas
- Aggregation of receptors often follows activation/ligand binding
- Bulk assays have suggested that TLR4 molecules aggregate in lipids rafts within the cell membrane after LPS binding\*
- Visualization at the single cell level has been limited by optical diffraction

\*Triantafilou, et. al, *Biochem. J.* 381(Pt 2): 527-536



# Hypothesis

TLR4 distribution in the membrane changes upon ligand binding. The spatial distribution

*1) Depends on the chemical properties of the LPS,  
and*

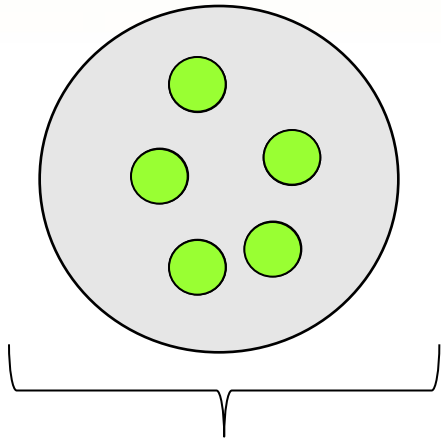
*2) Affects downstream signaling events and  
ultimately cellular response*

Optical super-resolution gives us a way to differentiate TLR4 clustering at a much finer scale than conventional imaging.



# STORM = Subdiffraction Spatial Resolution

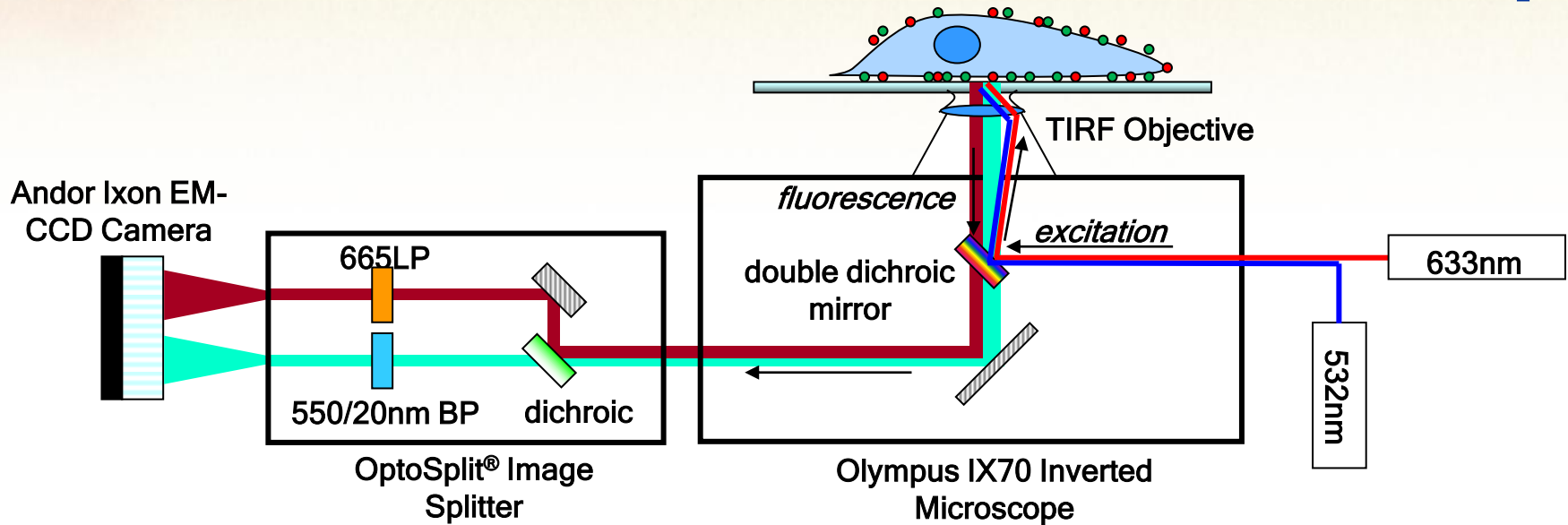
## Stochastic Optical Reconstruction Microscopy



**diffraction-limited spot  
size**

- Assuming  $<1$  fluorophor per diffraction-limited area, it's position can be determined with nanometer precision.
- The Abbe resolution limit can effectively be broken if the fluorophors in a sample can be imaged *independently* from each other.
- In STORM, this means incorporating stochastic “photoswitching”
  - Only a small subset of fluorophors is visible at any given time.
- Photoswitching for organic dyes can occur in buffer containing small thiol (i.e. BME) and oxygen scavenging system. (dSTORM)

# Multicolor STORM Setup



## Unique capabilities:

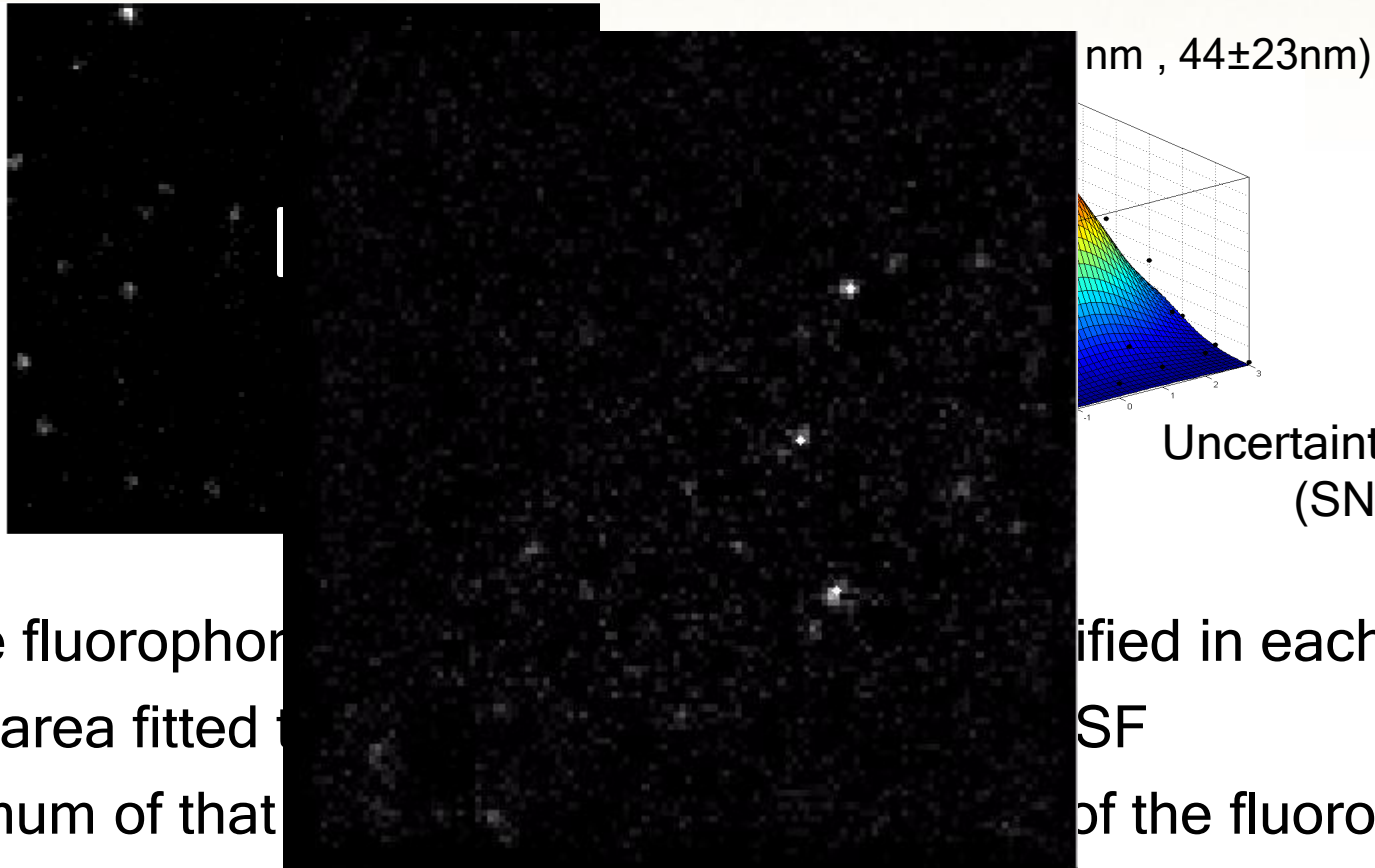
- Four excitation  $\lambda$ 's (405, 488, 532, 633nm), variable angle
- Simultaneous dual-color emission
- Capable of >50fps over 30 $\mu$ m x 30 $\mu$ m FOV

## Advantageous in:

- Receptor reorganization
- Nanoparticle-membrane interactions, uptake
  - Engineered NPs
  - Natural NPs - Viral trafficking

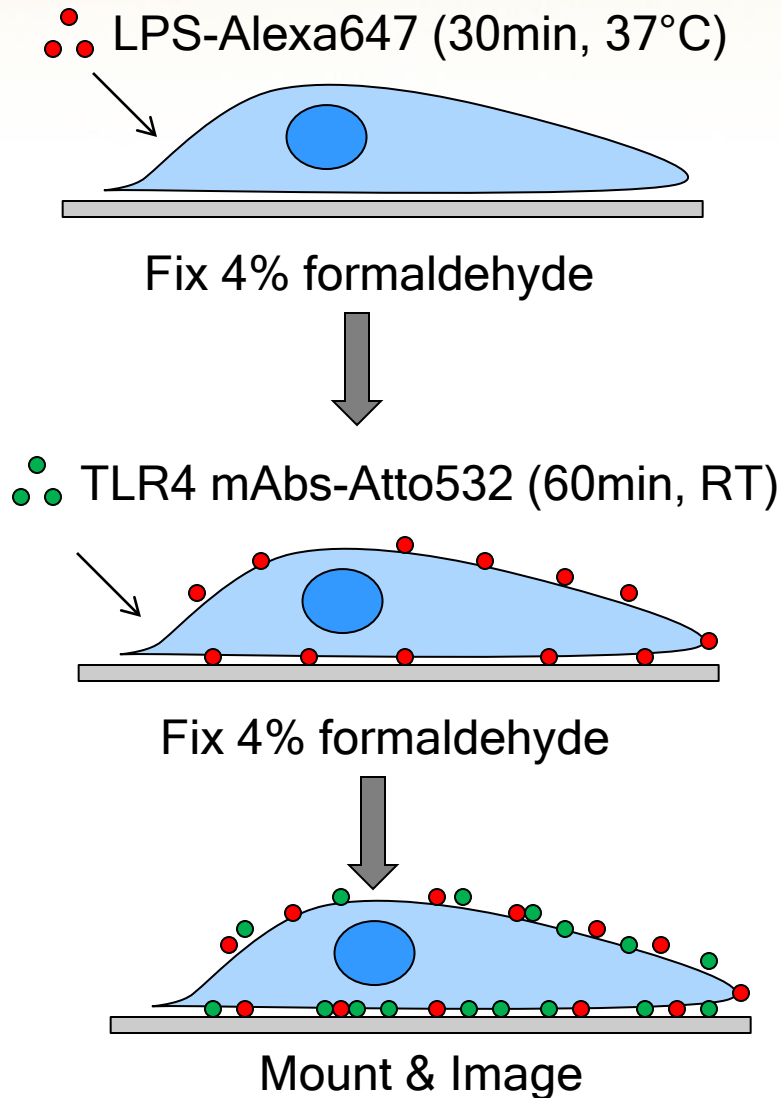


# Fluorophor Localization



- Single fluorophore identified in each frame
- Local area fitted to a 2D Gaussian
- Maximum of that fit is the location of the fluorophore
- Typically, location fit uncertainty 40-60nm
- Process repeated over 1k-10k frames to build STORM image

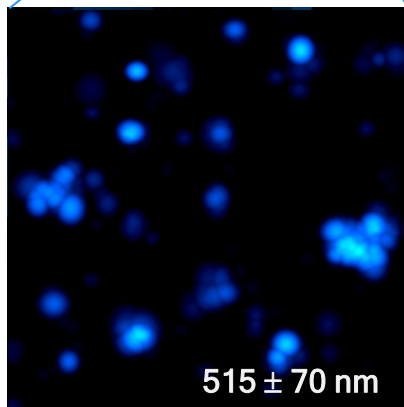
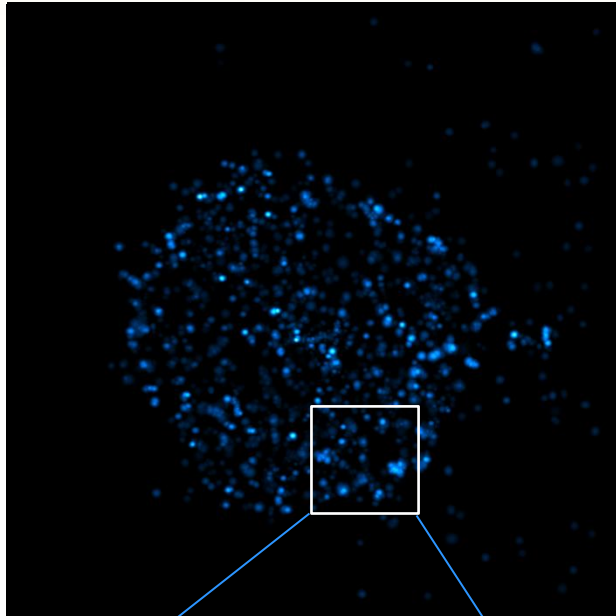
# Experimental Design



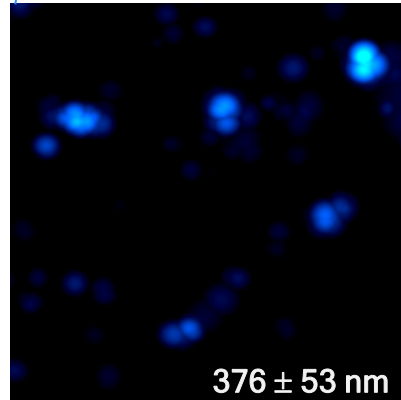
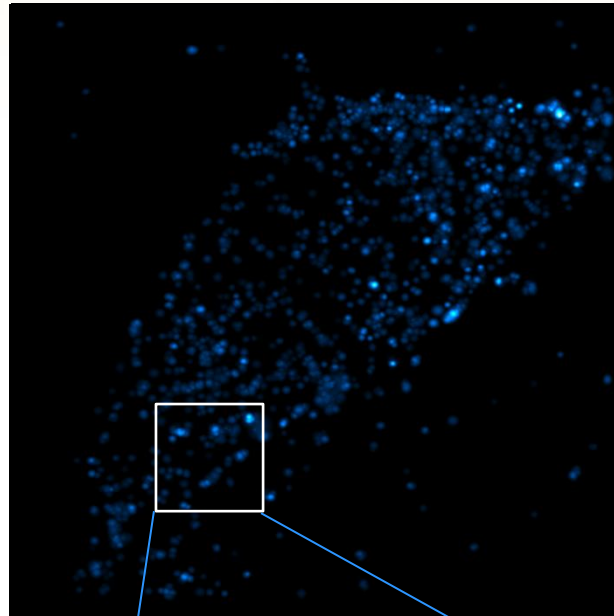
- Mouse macrophage cells (P388D1) incubated with 100nM *E. coli* or *Y. pestis*-derived LPS for 30 min at 37° C and formaldehyde fixed.
- LPS are labeled with Alexa Fluor 647-hydrazide via linkage with core-polysaccharide
- TLR4 receptors visualized via 1<sup>0</sup> antibodies labeled with Atto532
- Cells imaged in O<sub>2</sub>-scavenging buffer containing β-mercaptothiol

# TLR4 Clustering is Specific to LPS

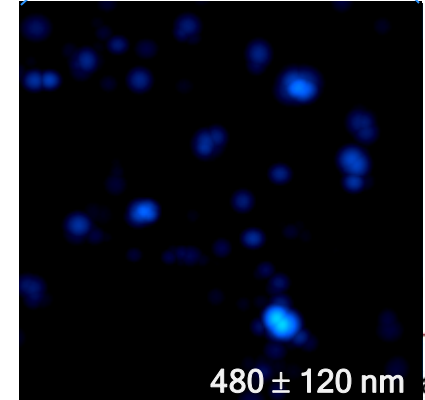
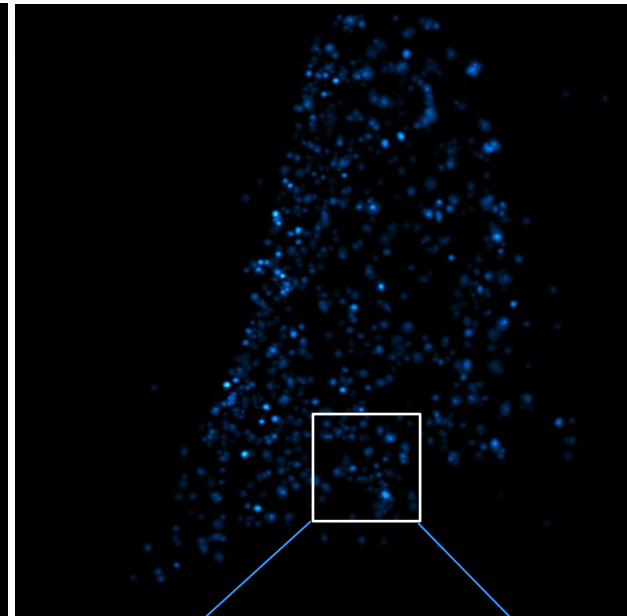
*E. coli* LPS



Flagellin



*Y. pestis* LPS



STORM images are 10 diffraction limited (~400 nm) (40-50 nm)

# Ripley's K-function Analysis

- K-function is a normalized measure of point clustering
- Complete spatial randomness (CSR)
- Transform to H-function, deviation from CSR at each test radius
- Peaks (or inflection points) in  $H(r)$  indicate characteristic cluster sizes

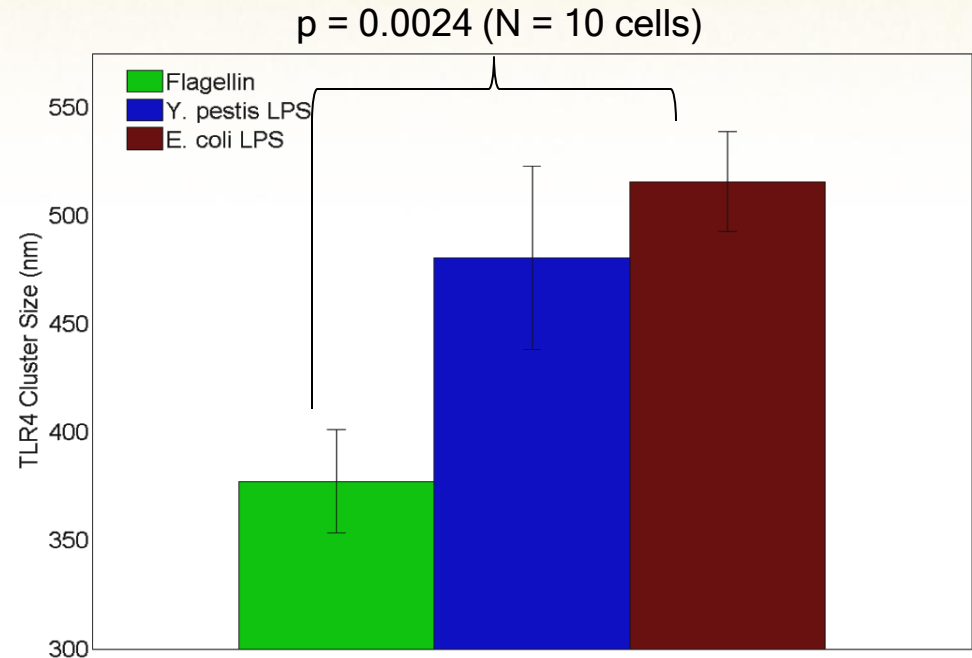
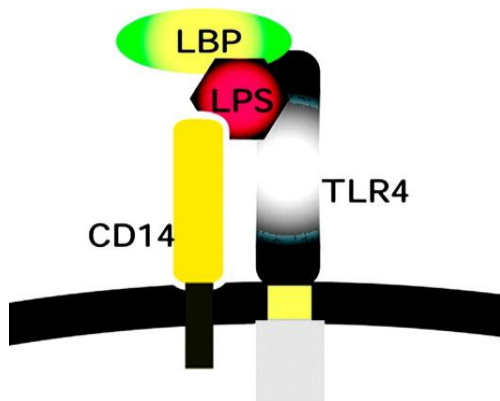
$$K(r) = \frac{A}{N^2} \sum_{j \neq i}^N \sum_{i=1}^N \frac{I(d_{ij} < r)}{w_{ij}}$$

$$K(r) = \pi r^2$$

$$H(r) = \sqrt{\frac{K(r)}{\pi}} - r$$

# Differential TLR4 Clustering is Significant

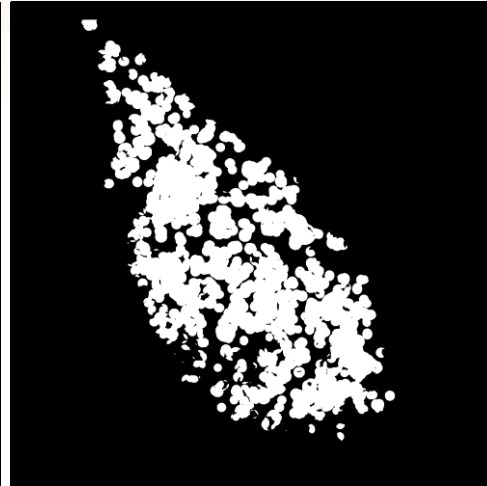
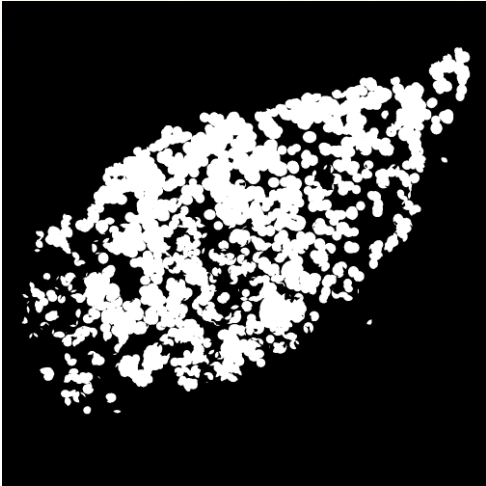
- Ripley's K-analysis indicates that *E. coli* LPS induces significant clustering over negative control (flagellin)
- Suggests that *pestis* induces less clustering, but not significant



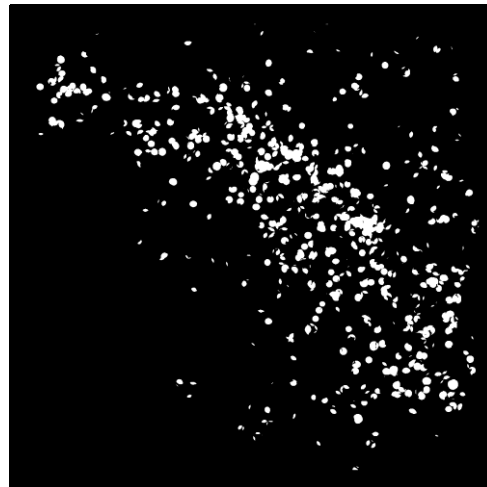
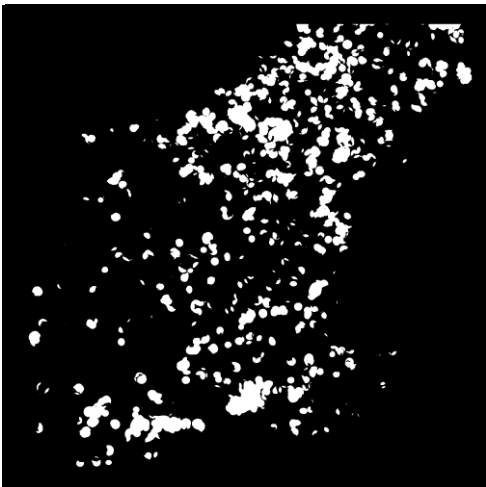
## TLR4-LPS Complex?

# Colocalization of TLR4 & LPS

*E. coli* LPS



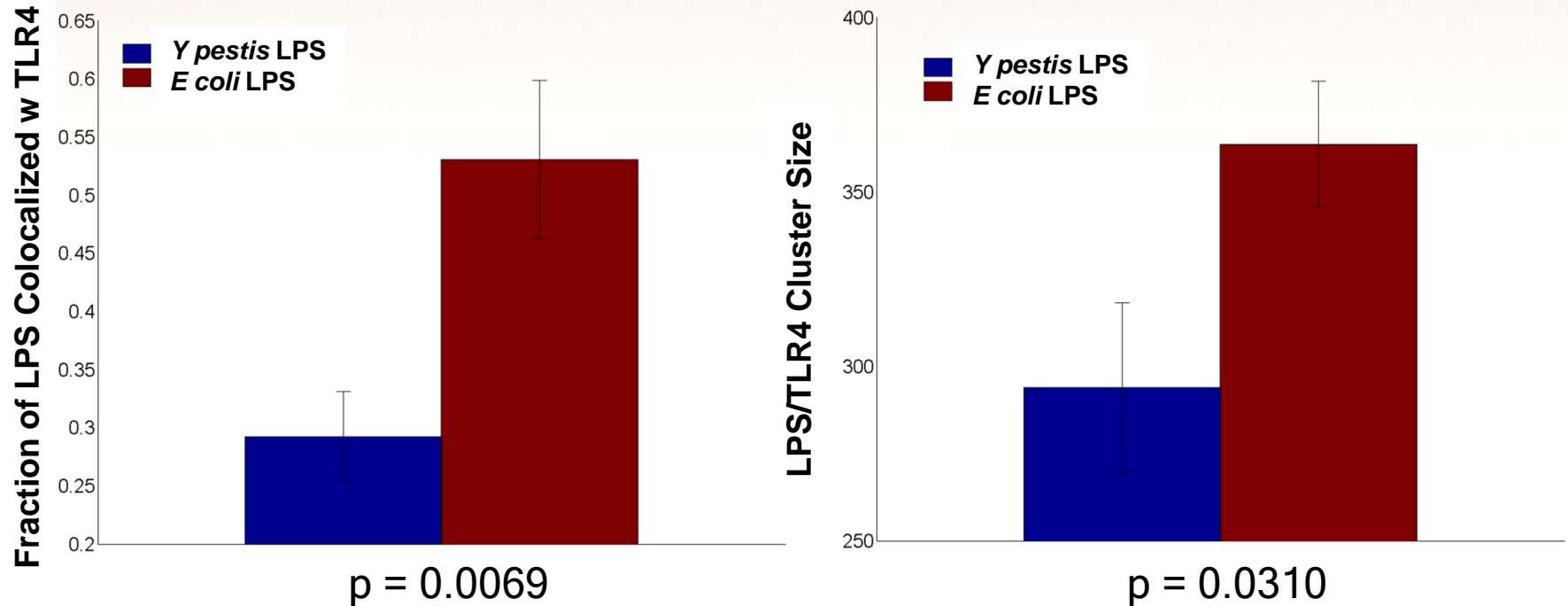
*Y. Pestis* LPS



- Dual-color STORM imaging
  - TLR4 - Atto532
  - LPS - AlexaFluor647
- Image registration via multi-dye PS beads (average error  $\sim 50\text{nm}$ )
- Perform cluster analysis on co-localized points
  - Custom implementation of Ripley's K-Function



# *Y. pestis* LPS is less Efficient at Recruiting TLR4 into Clustered Domains



- Significantly less co-localization of *Y. pestis* LPS with TLR4 compared to *E. coli* LPS
- Significantly smaller *Y. pestis* LPS-TLR4 clusters than *E. coli* LPS-TLR4 clusters





# Conclusions

- Visualization of TLR4 and TLR4-LPS distributions in individual, intact macrophage cells
  - *E. coli* LPS produces a significant increase in TLR4 cluster size within 30 minutes, as compared to a non-specific ligand and non-stimulatory control
  - *Y. pestis* LPS exhibits less co-localization with TLR4 and is less able to recruit TLR4 into clusters as compared to *E. coli* LPS → correlated with down-stream signaling response
  - Role of co-factors ?
- Super-Resolution imaging allows for measuring subtle changes that aren't apparent in conventional microscopy
  - Spatial-temporal behavior of multiple components
  - Broad impact in cell signaling research

# Acknowledgements

- Dr. Roberto Rebeil (NBACC) for isolation of *Y. pestis* LPS

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