

Enhanced Stabilities and Activities of OPH

Part I: Immobilizations → promote stability and activity

Benefits of:

- extra active-site metal
- crowding reagents
- covalent crosslinking
- substrate

Part II: Cell-free production of rational OPH mutants with higher thermal stabilities and activities

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Background: Overcoming enzyme fragility by proper immobilizations

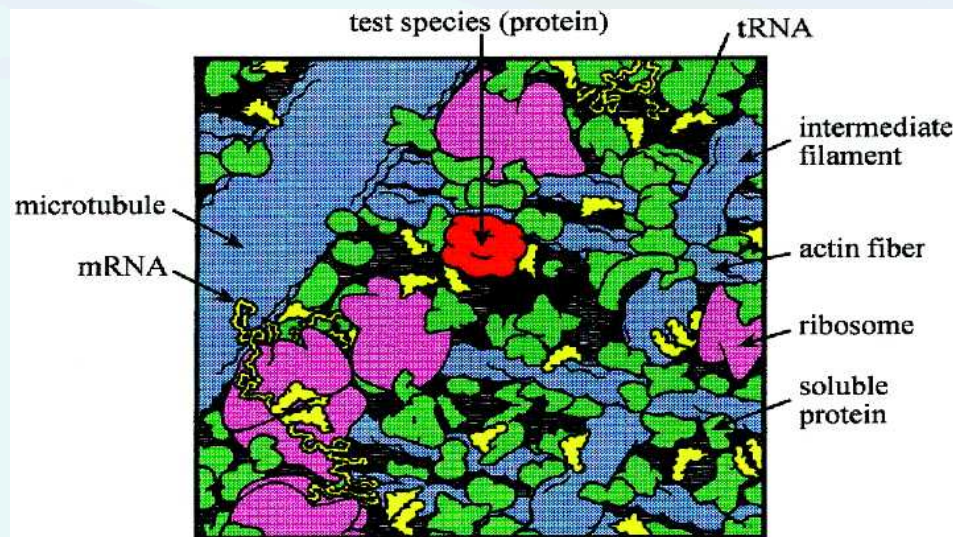
Historical approaches:

- Low levels of immobilized enzyme, and/or
- Immobilization procedure destroys enzymatic activity

Inspiration for this work:

- Cellular architecture
- Molecular crowding
- Rational design of materials
- Rational design of enzymes
- → exceed biological capabilities

Cells are crowded with biomolecules (~400 mg/ml) and structures.

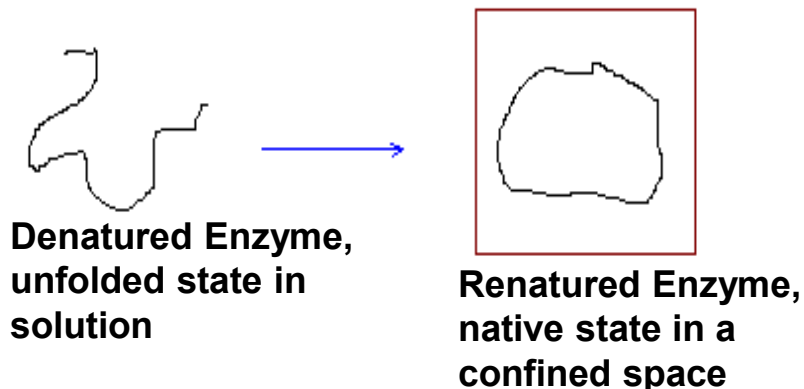


J. Biol. Chem. 2001, 276: 10577

Maintaining and Promoting Enzyme Activity

Confinement can eliminate some expanded configurations of the unfolded chain, shifting equilibrium from unfolded (inactive) to native (active).

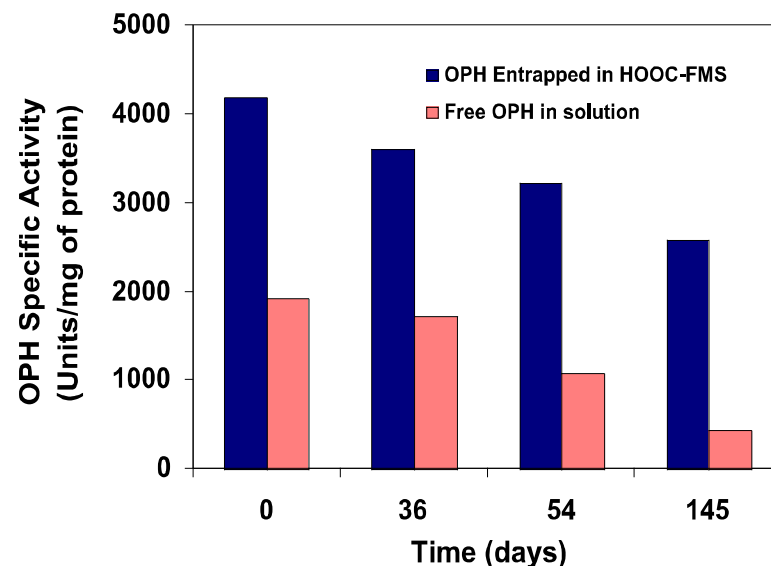
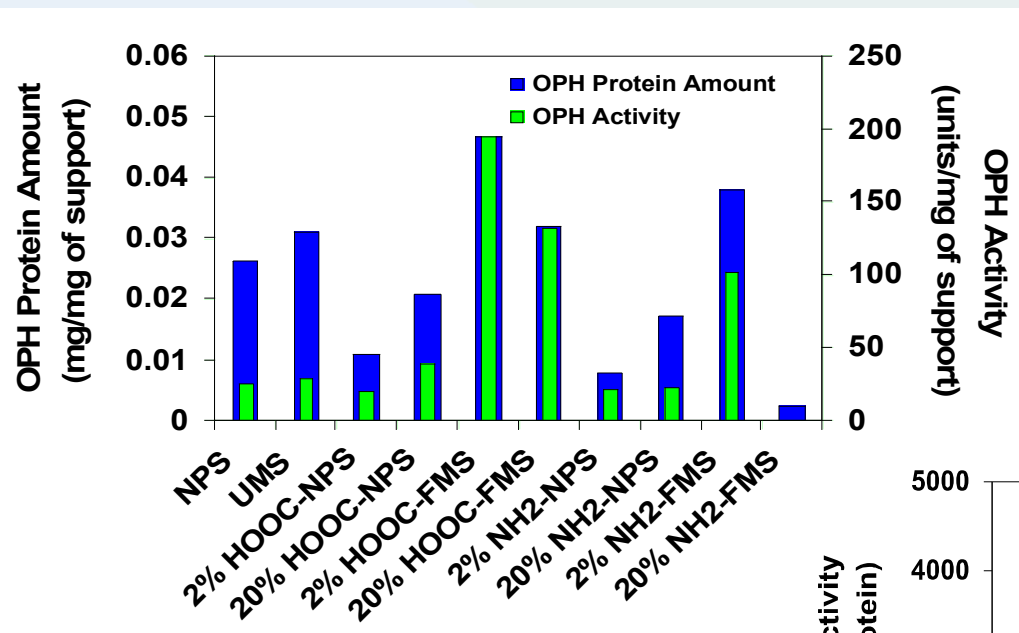
**Cells, molecular crowding,
enhancing specific activity**
Biochemistry 2001, 40: 11289



Samples of OPH solution s	Specific activity of OPH solution (units/mg of protein in solution)	Specific activity of OPH after entrapping in FMS (units/mg of protein in FMS)
Sample 1	1629.70	4286. 29
Sample 2	1327.15	2954. 15
Sample 3	2018.03	3375. 89
Sample 4	1494. 77	3789. 28
Sample 5	782. 04	2233. 08
Sample 6	1863. 56	3357. 55

Lei, C. et al. *Nanotech* 2006 17: 5531 and highlighted
perspectives by Dunker & Fernandez *Trends Biotech* 25: 189

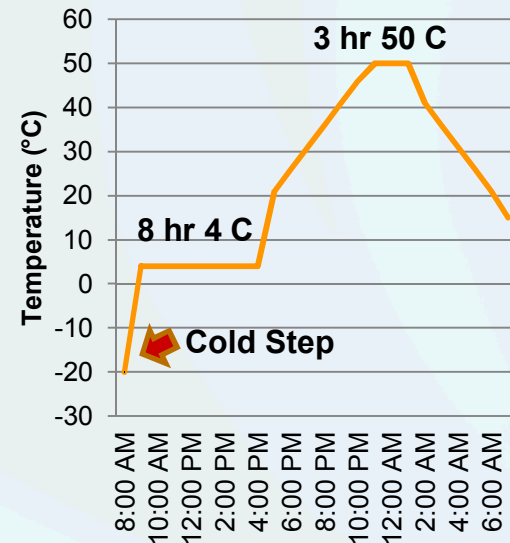
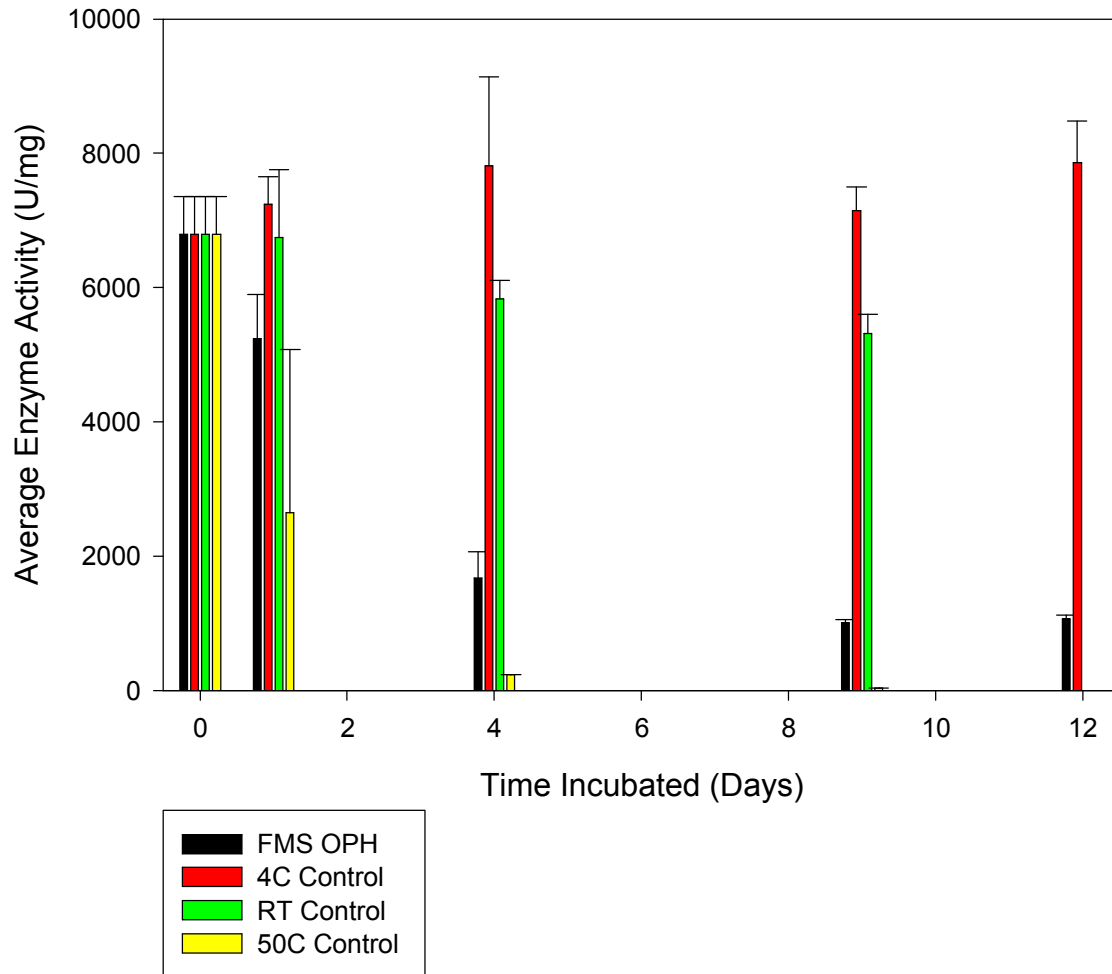
Comparison of different porous silica support for OPH immobilization



Lei, C., Shin, Y., Liu, J.,
Ackerman, E. J. 2002. *J. Am. Chem. Soc.* 124 (38): 11242-11243. Highlighted in *Chem. & Eng. News* [80: 35 (2002)] and *Science* [300: 277 & 290-293 (2003)]

Enhanced Specific Activity & Stability of Immobilized OPH

Diurnal Cycling of FMS OPH with -20C Step



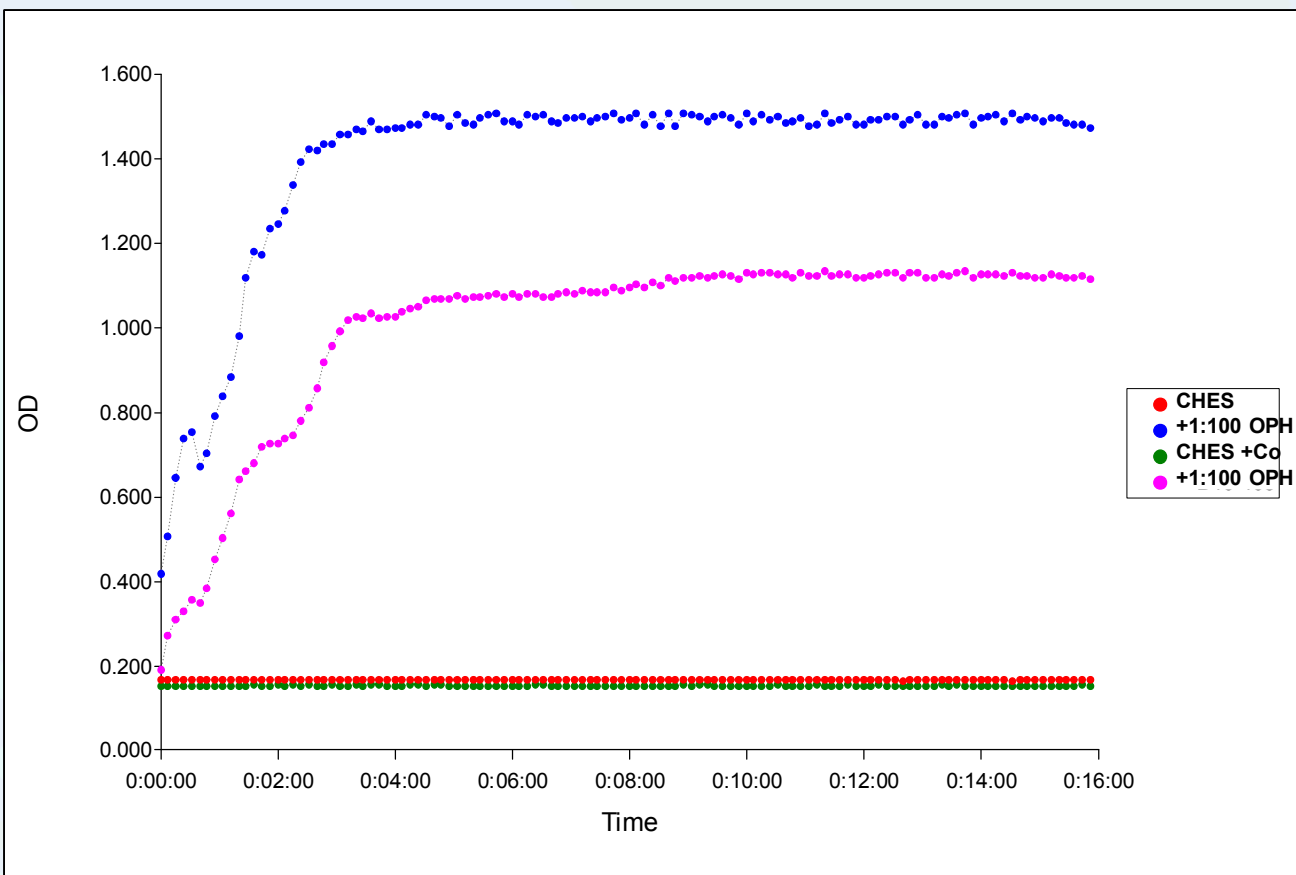
Timeline Diurnal Cycle

Temperature (C)	Hours at Temperature
-20	1
4	8
21	1
26	1
31	1
36	1
41	1
46	1
50	3
41	1
36	1
31	1
26	1
21	1
15	1

Lyophilized OPH stable, but when OPH in solution: FMS OPH > OPH, and

Constant temps > Diurnal cycle. Same with other stressors (pH, peroxides). Detergent inactivates.

OPH activity using plate reader \pm 100 μ M Co acetate



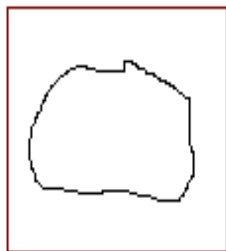
BioTek Eon Plate Reader
Shaking
Temp up to 60 C
~2 min for 96 samples

**Plate Reader faster, better signal-to-noise than stirred spectrophotometer cuvette.
 Extra active-site metal increased activity. (Same for Zn-acetate)**

Will it improve stability in FMS by keeping active site in correct conformation?

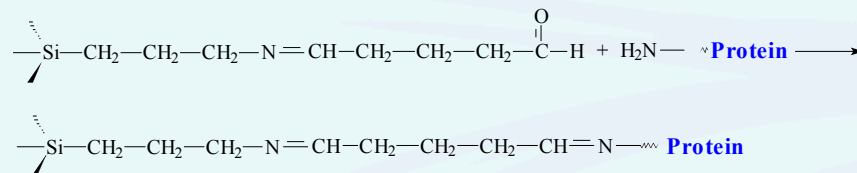
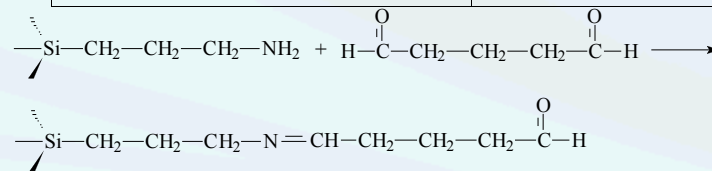
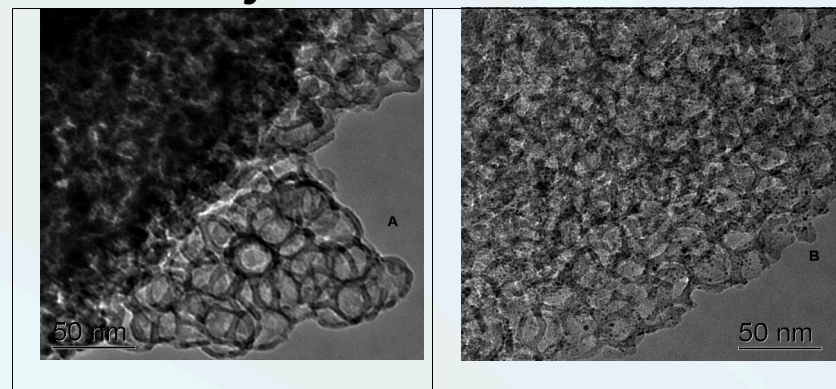
Crosslinking OPH to FMS

Purpose: Test hypothesis that further immobilization by crosslinking OPH to FMS increases specific activity.



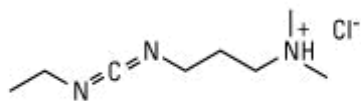
Denatured Enzyme,
unfolded state in
solution

Renatured Enzyme,
native state in a
confined space



Mesoporous silica can be functionalized with amino, carboxyl, or sulfhydryl groups to provide a favorable environment for proteins.

COOH-NH2 Crosslink via EDC

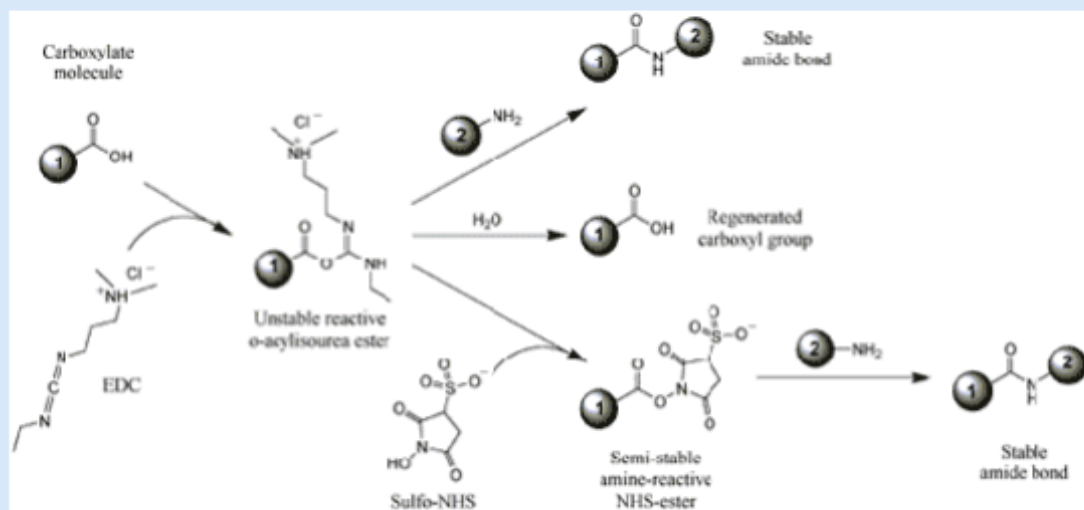


EDC

1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide • HCl

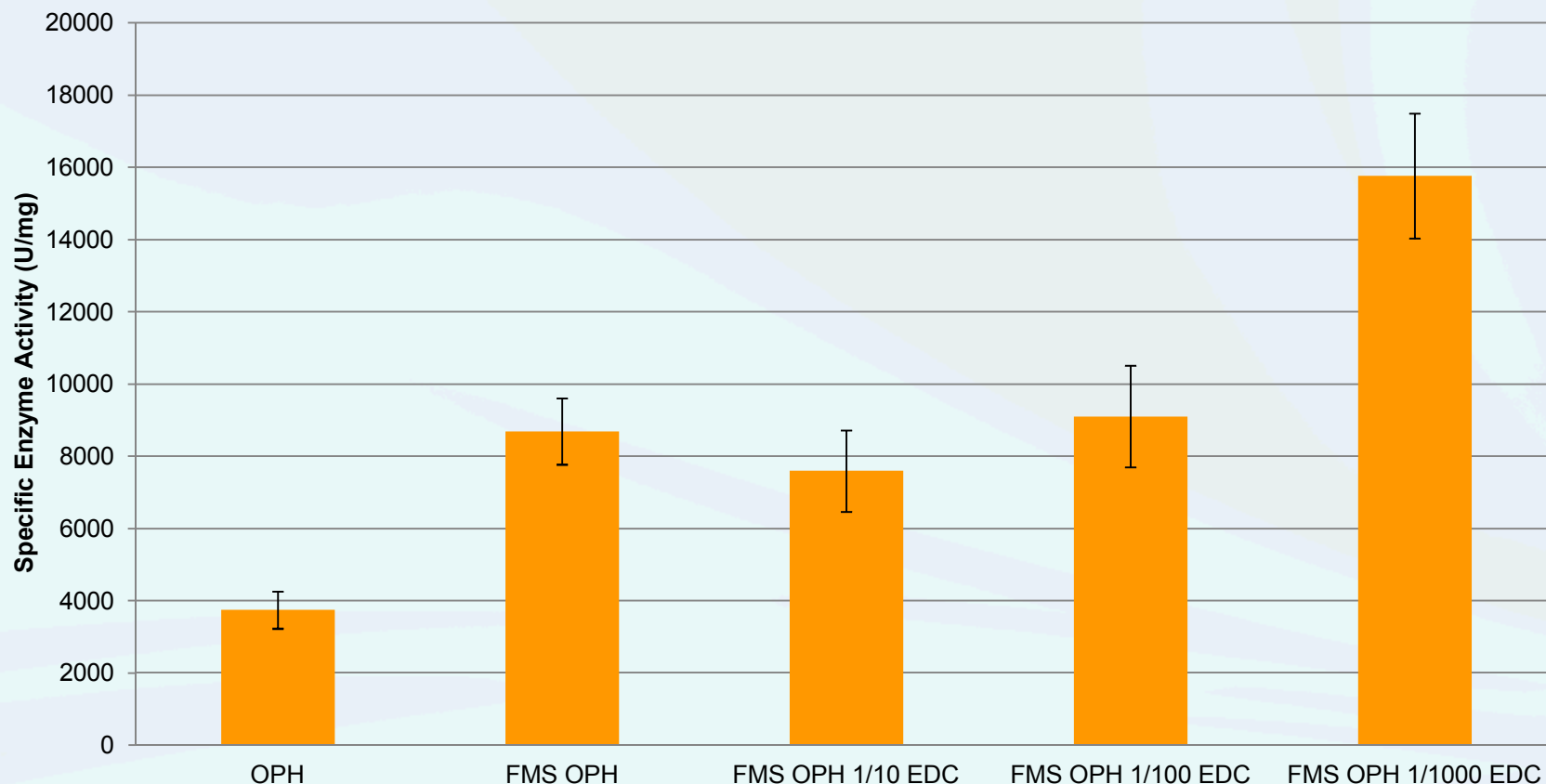
MW 191.70

Spacer Arm 0.0 Å



- Zero-length crosslinking agent.
- Couples carboxyl groups to primary amines.
- EDC that does not encounter an amine rehydrolizes to a carboxyl group.
- EDC can convert carboxyl groups to amine-reactive Sulfo-NHS esters in the presence of Sulfo-NHS (N-hydroxysulfosuccinimide)
- Sulfo-NHS improves coupling efficiency.

Effect of Crosslinking on OPH Activity

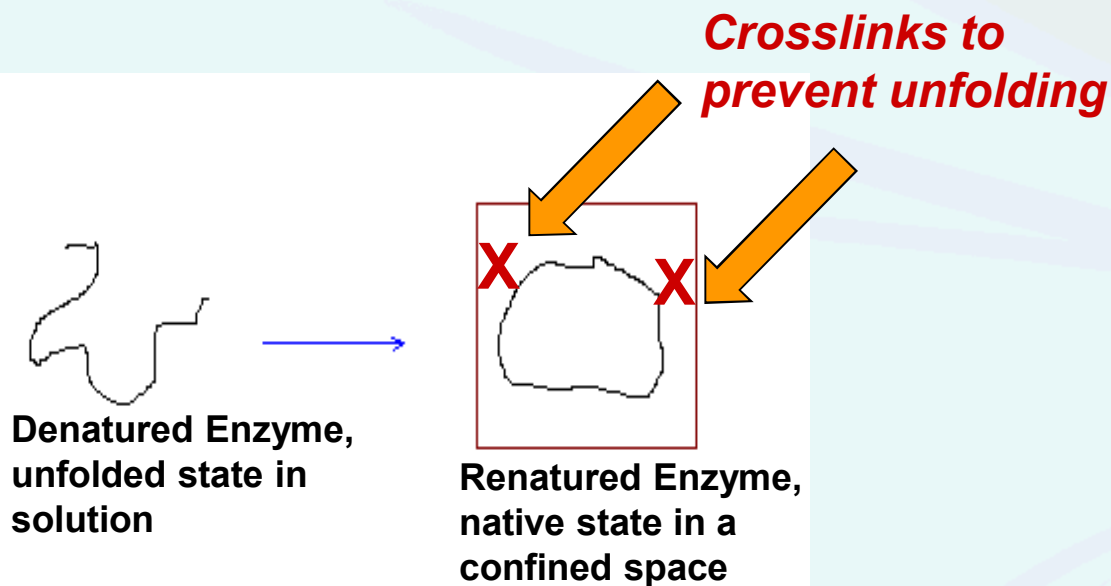


Crosslinking of OPH to FMS can improve specific activity.
Dependence of crosslinker concentration to enhance specific activity.

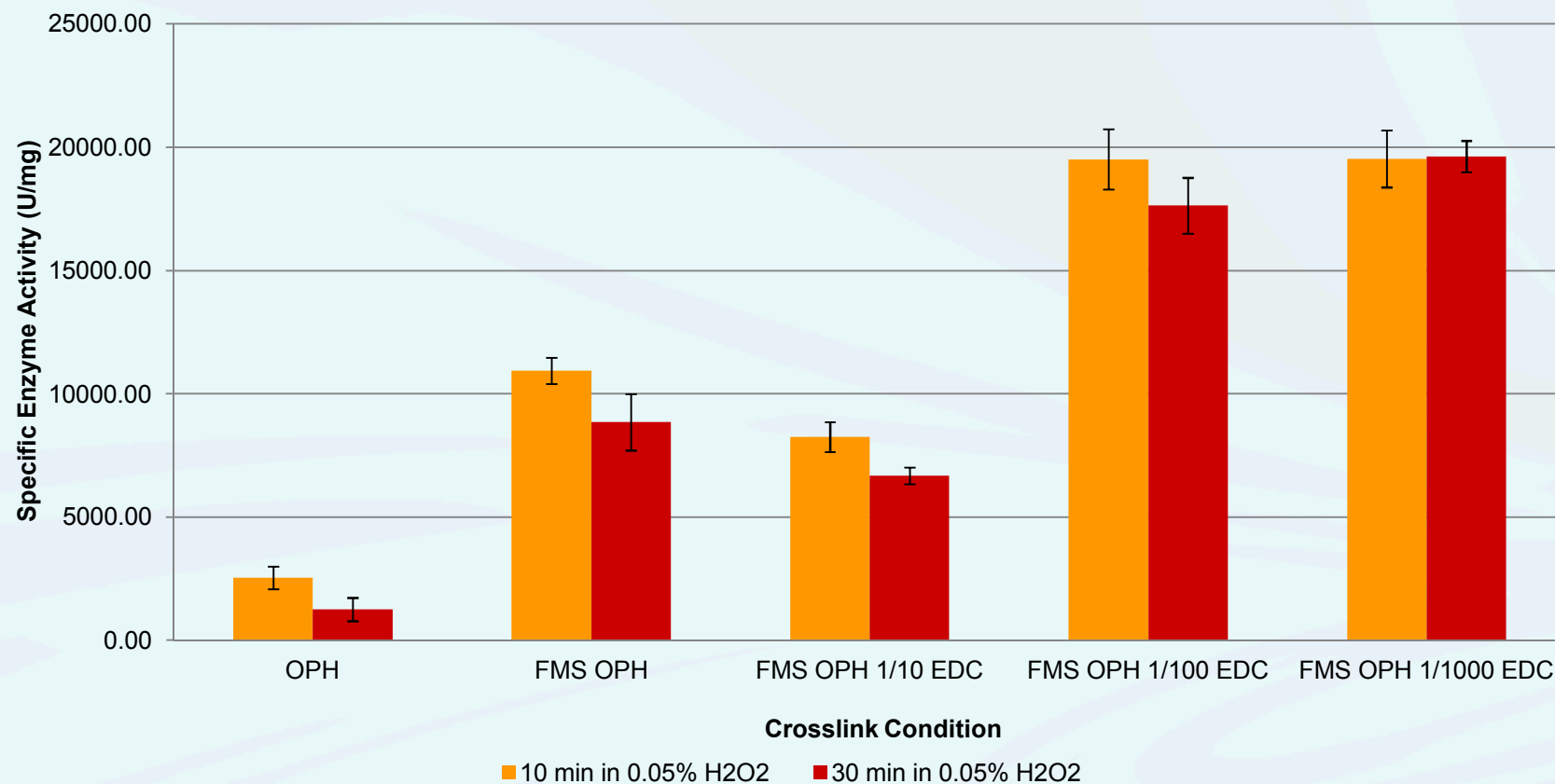
(Not shown: SH-SH maleimide bifunctional crosslinking reagent inhibited activity using FMS with SH groups.)

Tested Stability of Crosslinked OPH in FMS

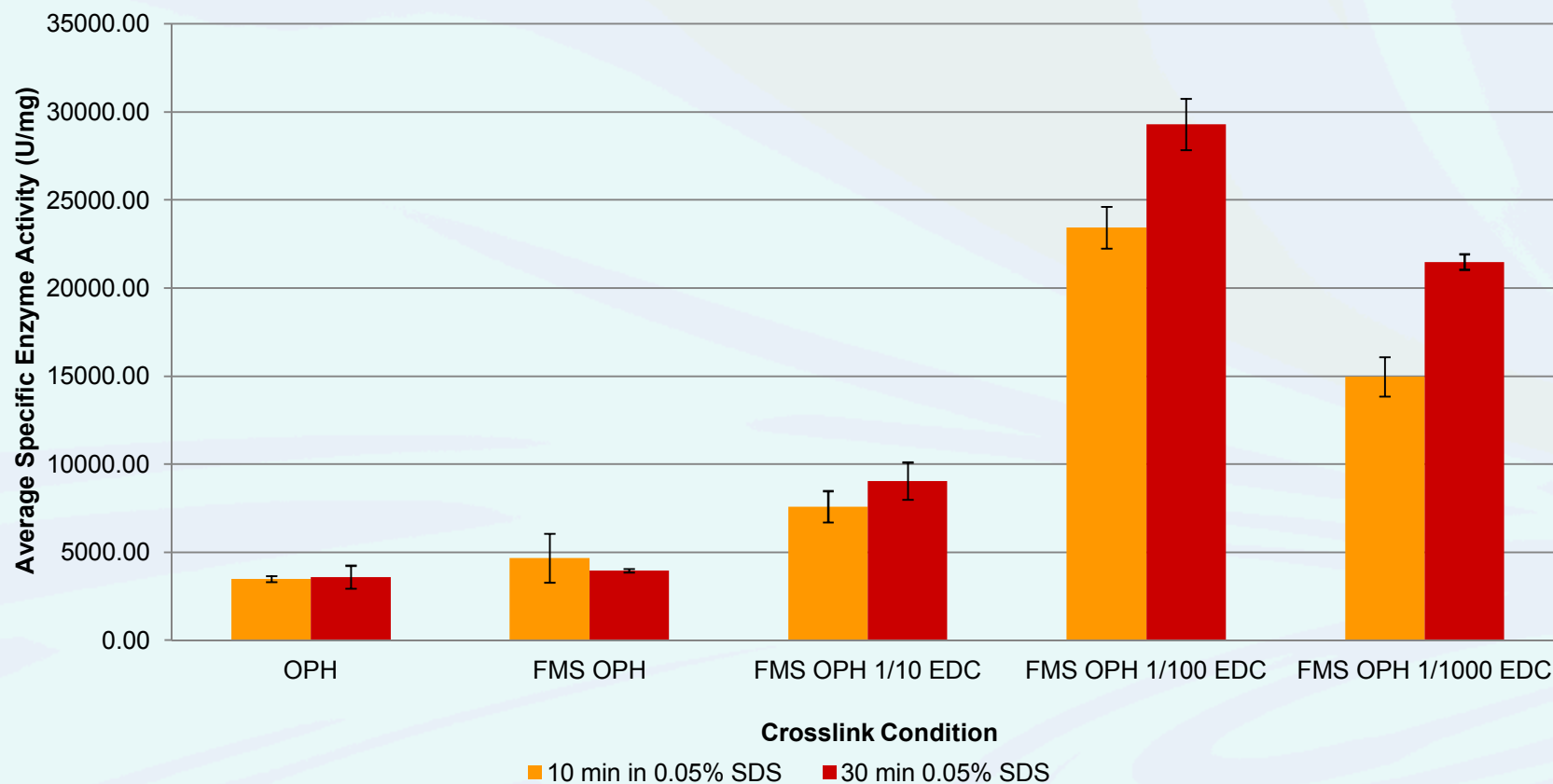
Purpose: Does crosslinking OPH to FMS improve overall enzyme stability?



Stability of Crosslinked OPH in 0.05% H₂O₂

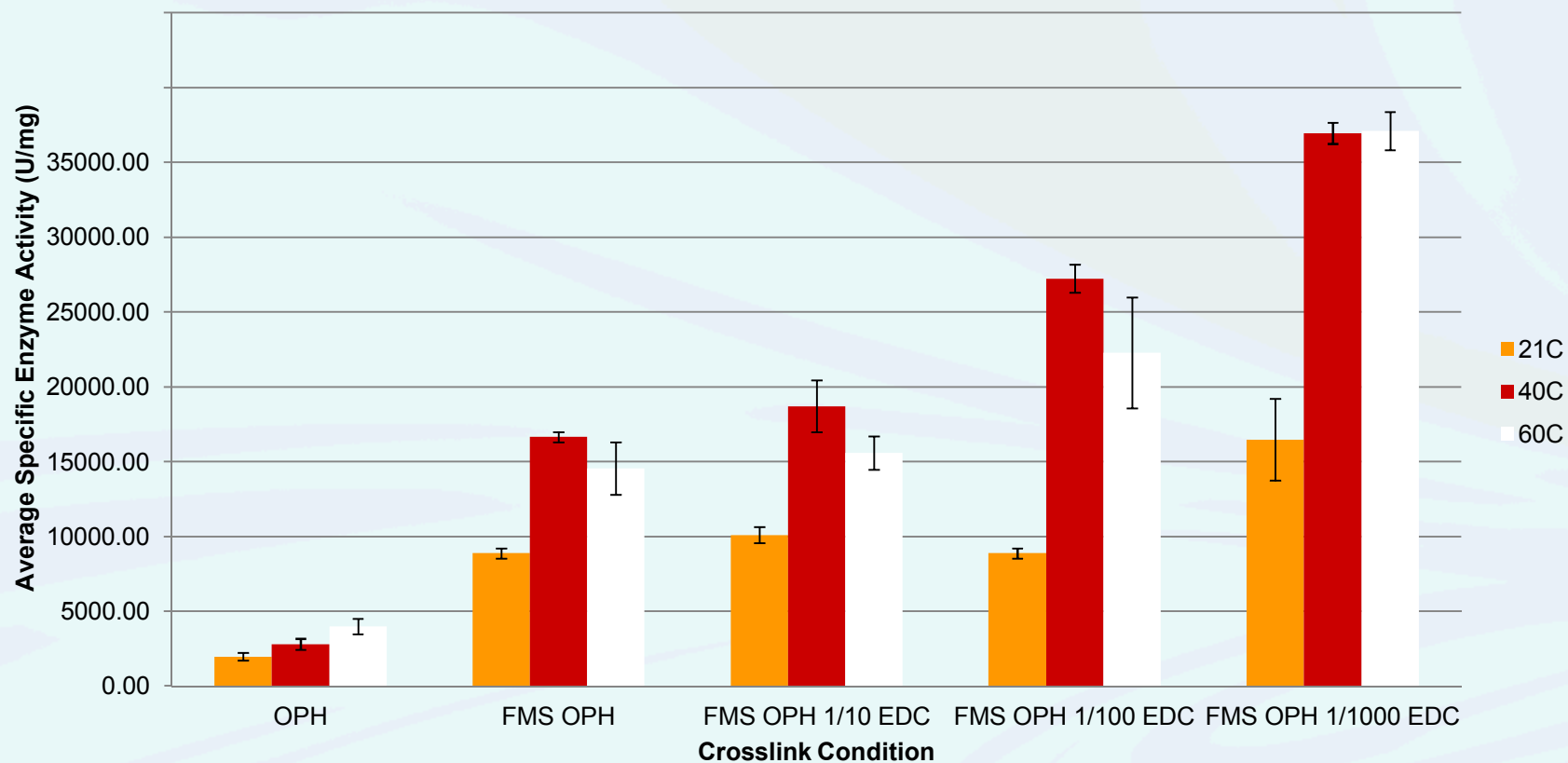


Stability of Crosslinked FMS OPH in 0.05% SDS



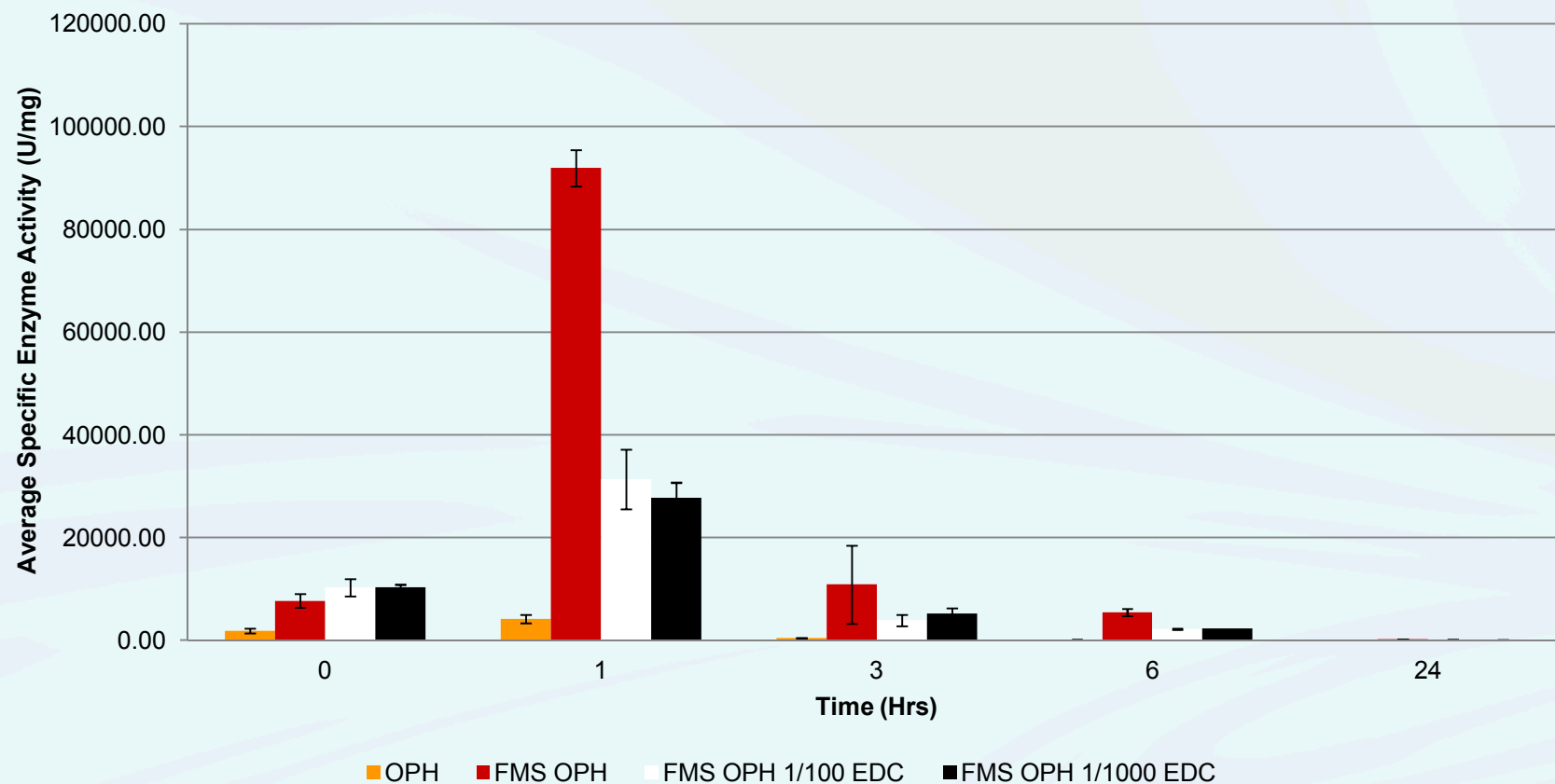
Results

Stability of Crosslinked OPH at High Temperatures



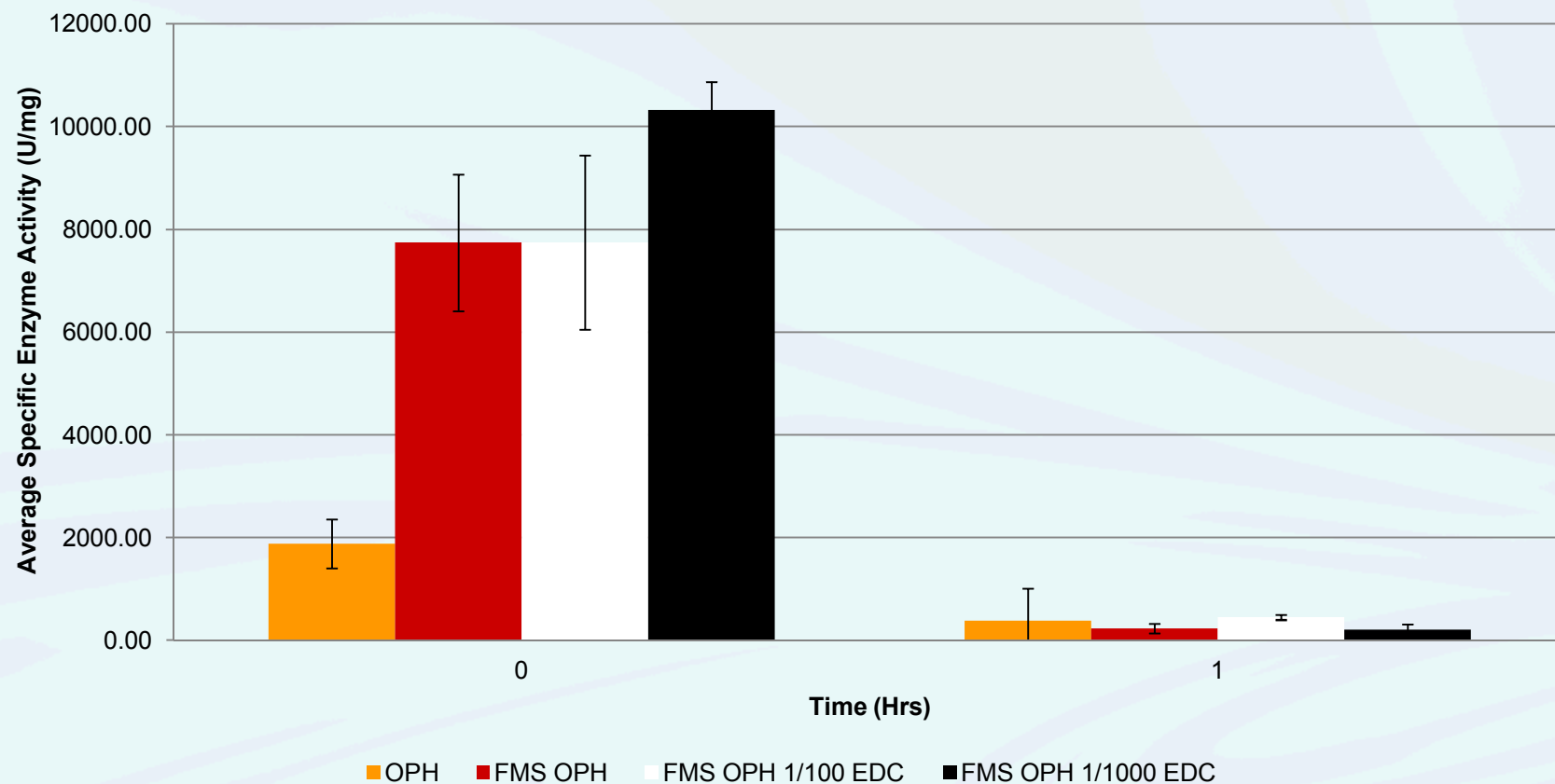
Results

Stability of OPH at 60C

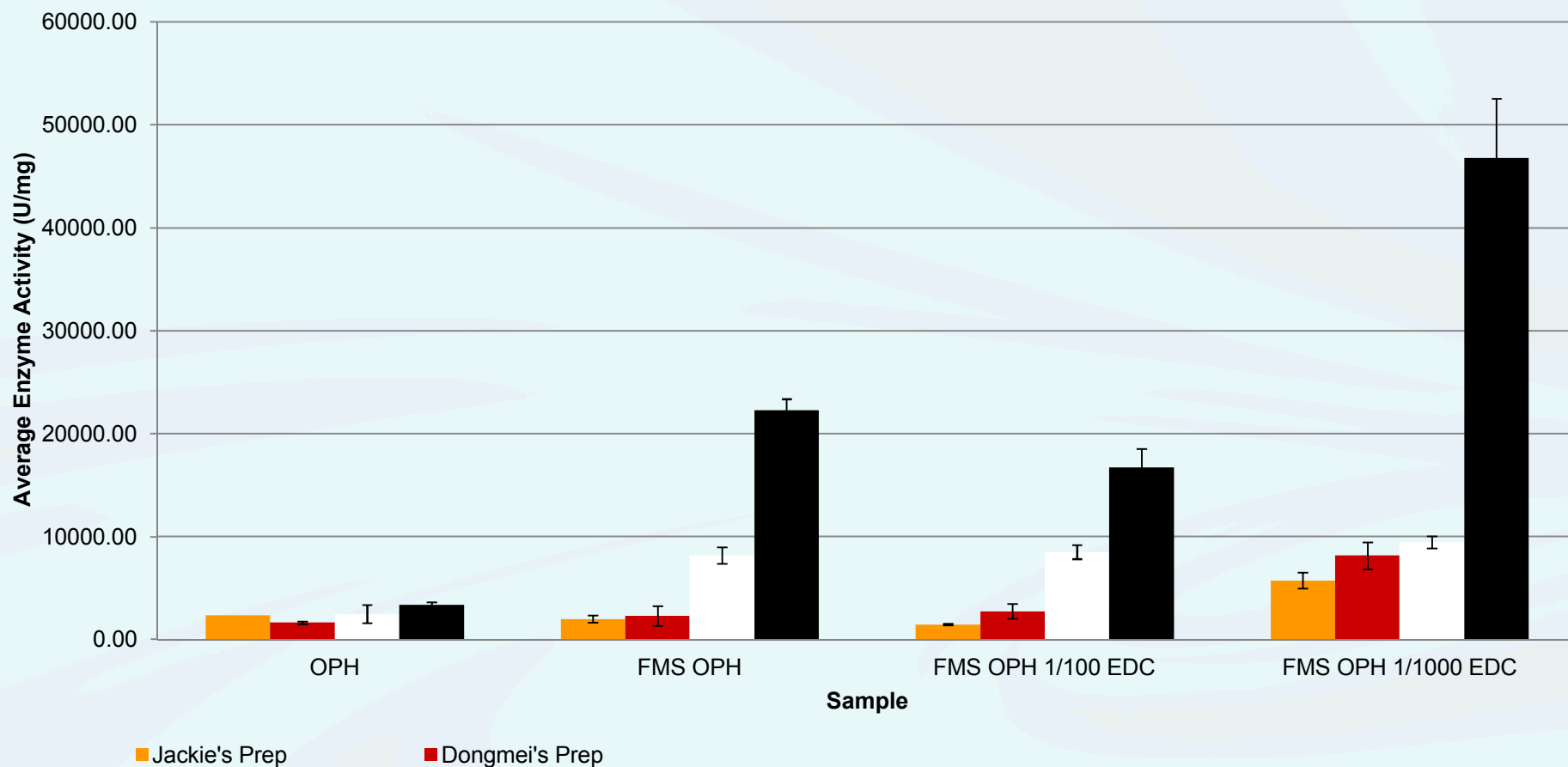


Results

Stability of OPH at 70C



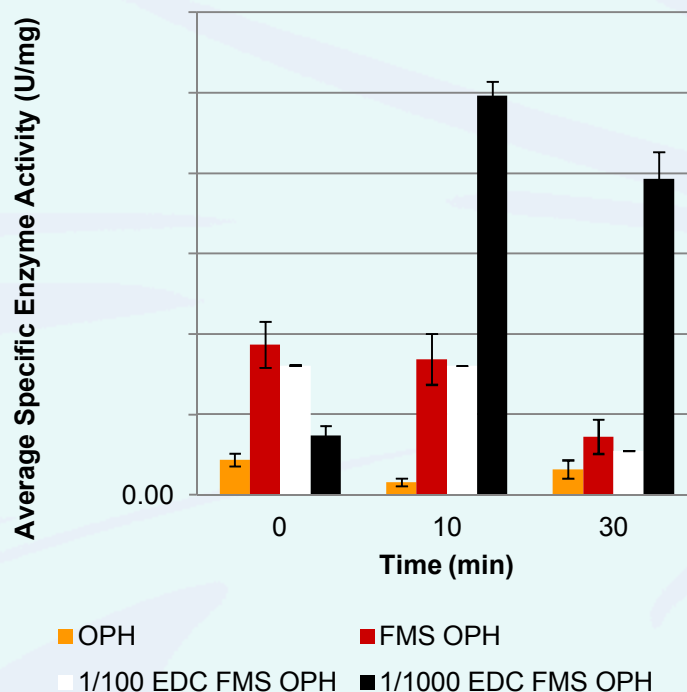
Reproducibility of COOH-NH2 Prep



Conclusions

- Crosslinker significantly improves OPH stability in SuperSoap, SDS and H₂O₂.
- Crosslinker enhances OPH stability over short time at high temperature.
- Same basic results with Doug Cerasoli's OPH sample.

Stability in 1/15 SuperSoap (manufacturer's rec)



Will Crowding Reagents Improve Stability?

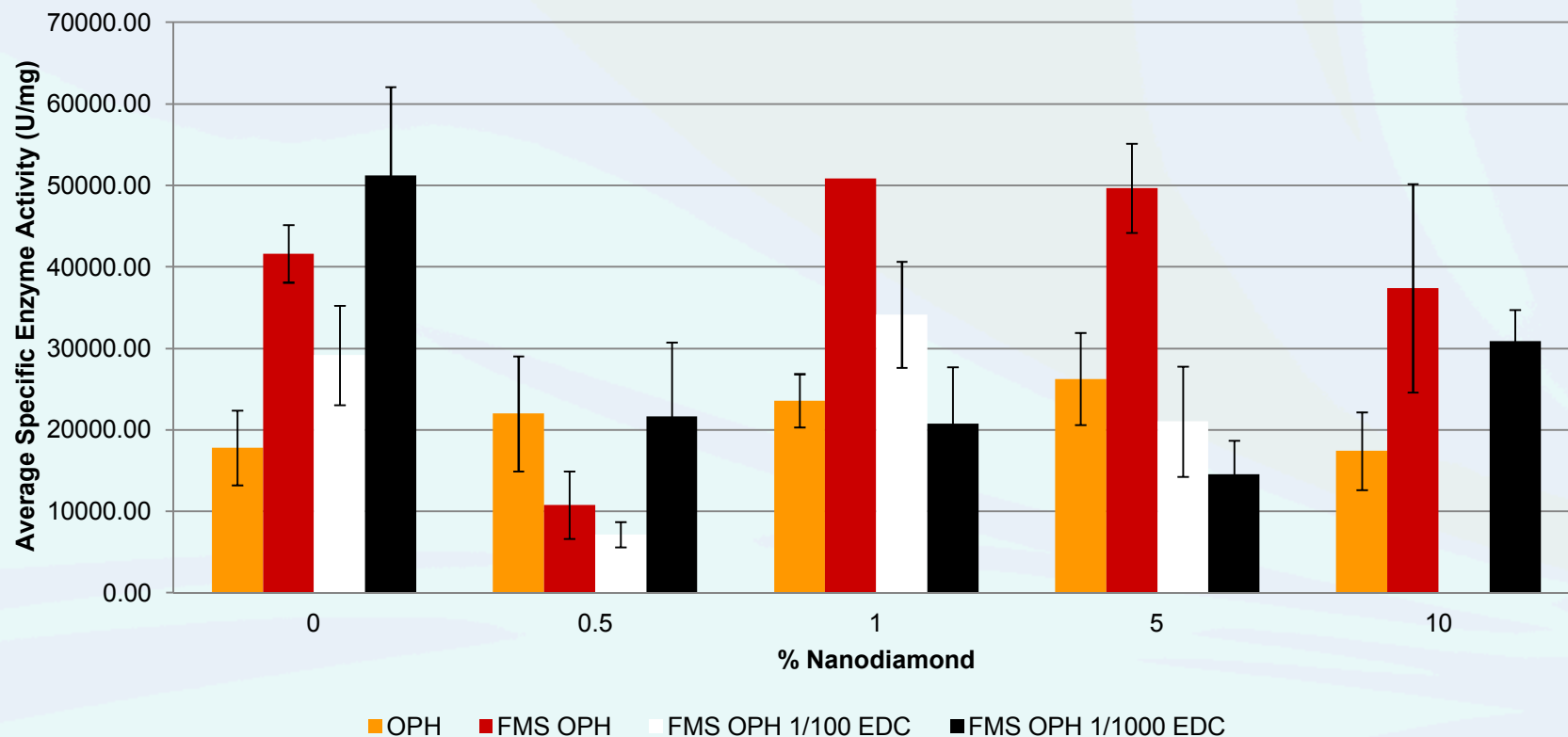
Nanodiamonds or Carbon Nanospheres

■ 4-6 nm Carbodeon Superhard Nanomaterials.

■ Nanodiamond Specifications:

- Content in solid phase ~ 97 wt. %
- Oxidizable carbon content in solid phase $\leq 2.5\%$
- Metallic incombustible impurity content in solid phase ≤ 1.2 wt. %
- Bulk Density ~ 0.5 gm/cm^3
- Pycnometric density ~ $3.1\text{-}3.2 \text{ gm/cm}^3$
- Specific surface area 330 gm/cm^3
- Constant of crystal lattice $0.3573 \pm 0.0005 \text{ nm}$
- Graphitization in vacuum, starting at ~ 1100°C
- Oxidation in air, starting at ~ 450°C

Stability of USAMRICD OPH in Suspension with Nanodiamonds

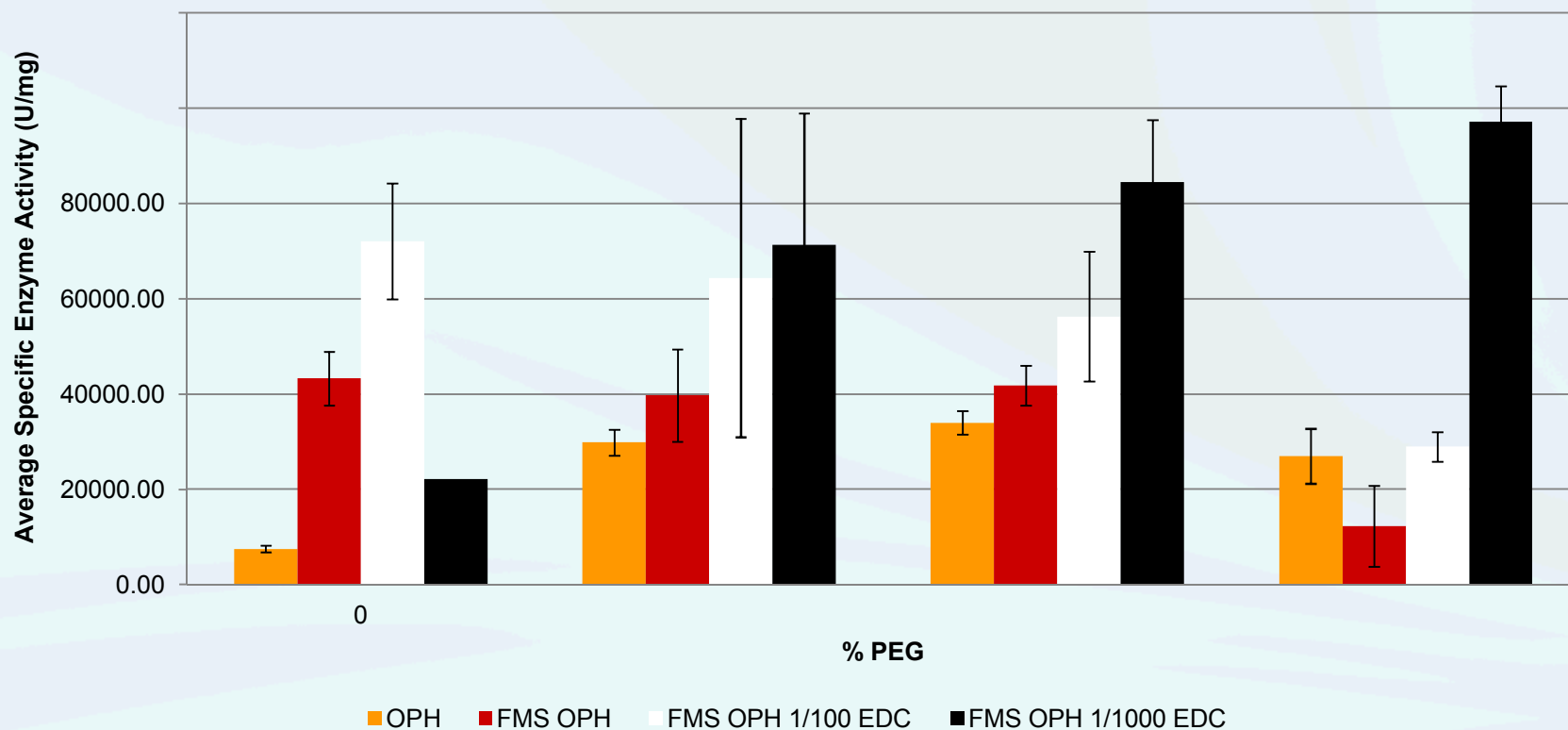


- Nanodiamonds have somewhat inhibitory effect on specific activity.
- There was a slight improvement with FMS OPH.

Polyethylene Glycol (PEG) Protocol

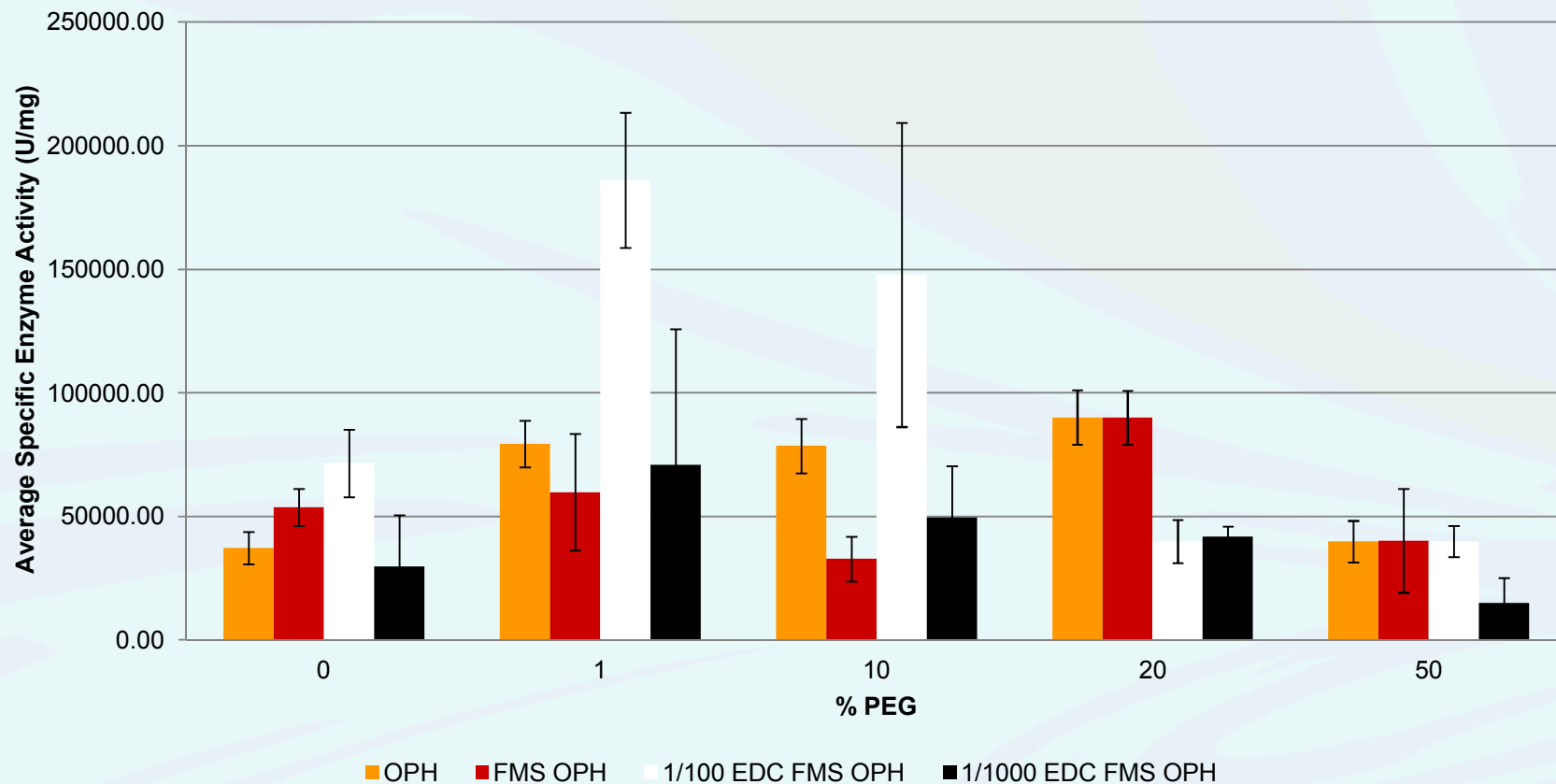
- Prepared different concentrations of PEG (0, 1, 10, 20%) and different MW PEGs (15-20,000, 400, 4,000, & 10,000) in 100 mM CHES (pH 9) with 50 uM zinc acetate.
- Made 1/500 dilutions of OPH, FMS OPH, FMS OPH 1/100 EDC, and 1/1000 FMS OPH in each of the PEG solutions.
- Vortexed briefly and measured enzyme specific activity

Stability of USAMRICD OPH in Suspension with MW 10,000 PEG



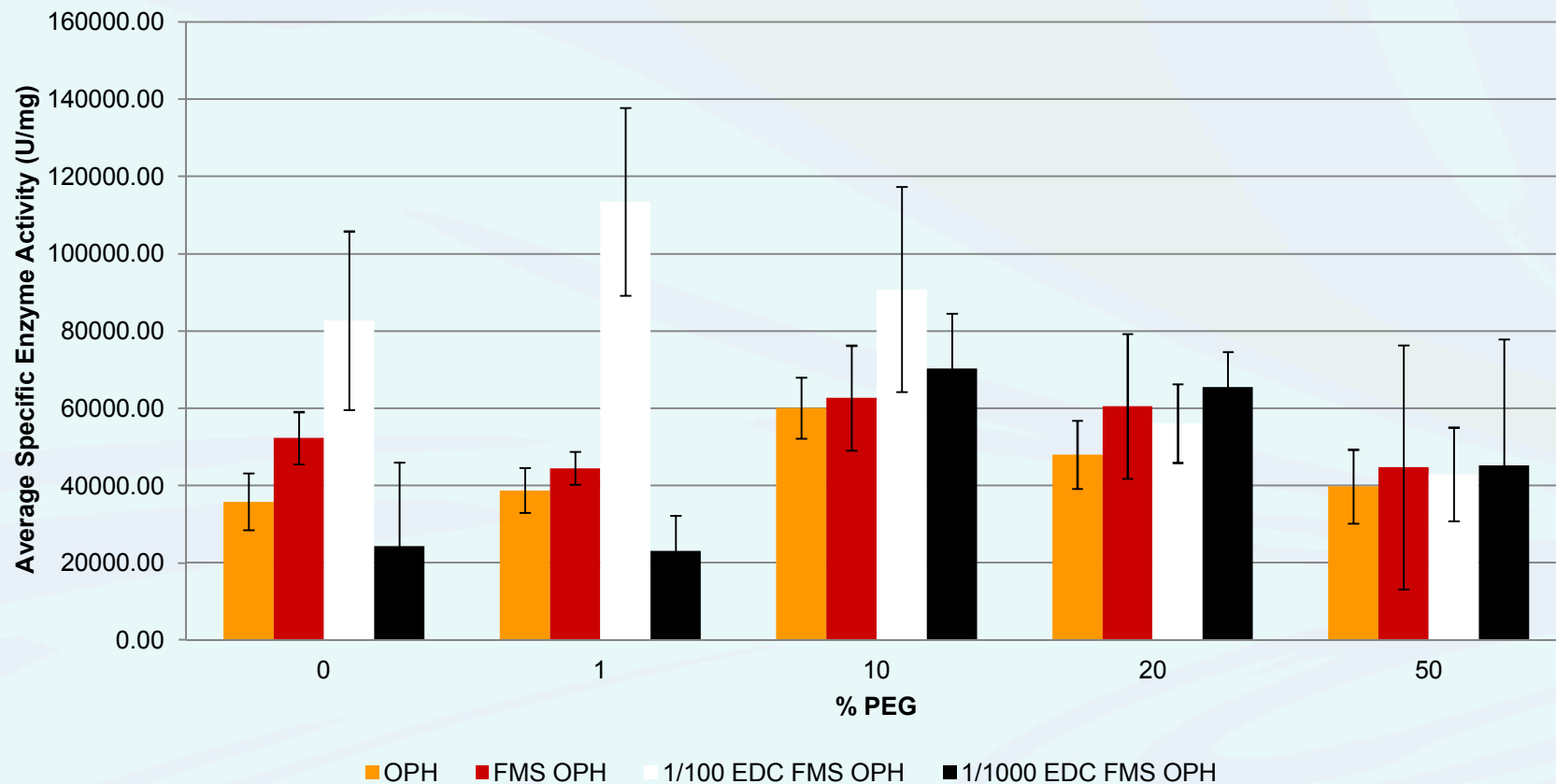
Results

Stability of USAMRICD OPH in 15-20,000 MW PEG



Results

Stability of USAMRICD OPH in 400 MW PEG



Conclusions

- Glycerol does not have much of an effect (not shown).
- Nanodiamonds or carbon nanospheres inhibitory for crosslinked protein but ~stimulatory for uncrosslinked.
- 15-20,000 MW PEG shows some promise in further improving immobilized enzyme activity.

Controlling OPH Conformation During Immobilization Crosslinking

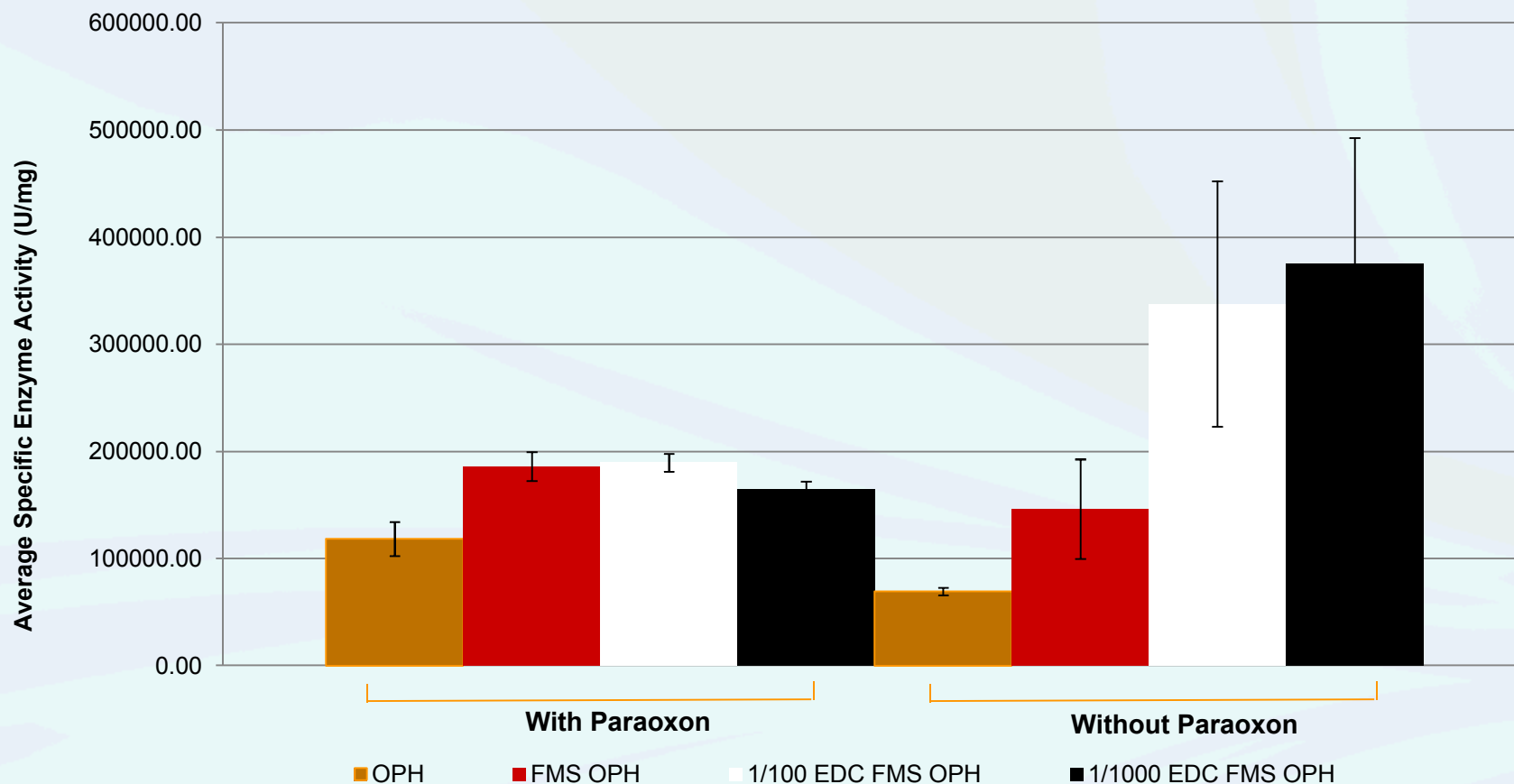
Purpose: Assess whether immobilization in the presence of Paraoxon enhances specific activity.

Hypothesis arose because presence of excess paraoxon during temperature stability measurements led to greater thermal stability.

Procedure

- Prepared COOH-NH₂ crosslinking reaction as described previously using the USAMRICD OPH.
- Prepared a second batch following protocol except added paraoxon to OPH before crosslinking; *i.e.*, add OPH + paraoxon to activated FMS.

Efficiency of OPH Specific Activity when Immobilized with FMS in Paraoxon



Conclusions

- Paraoxon added to OPH before crosslinking reaction to FMS can be inhibitory under initial conditions.
- Is there is an optimum concentration that will not be inhibited by Paraoxon?

and demonstrate benefits of maintaining active-site conformation during crosslinking?

- Overall, OPH alone:

< OPH FMS

< “ “ + Zn (or Co)

< “ “ “ + crosslinking

< “ “ “ “ + excess substrate

Part II: Cell-free translation of rational OPH mutants with higher thermal stabilities and activities

Advantages of Cell-free Approach:

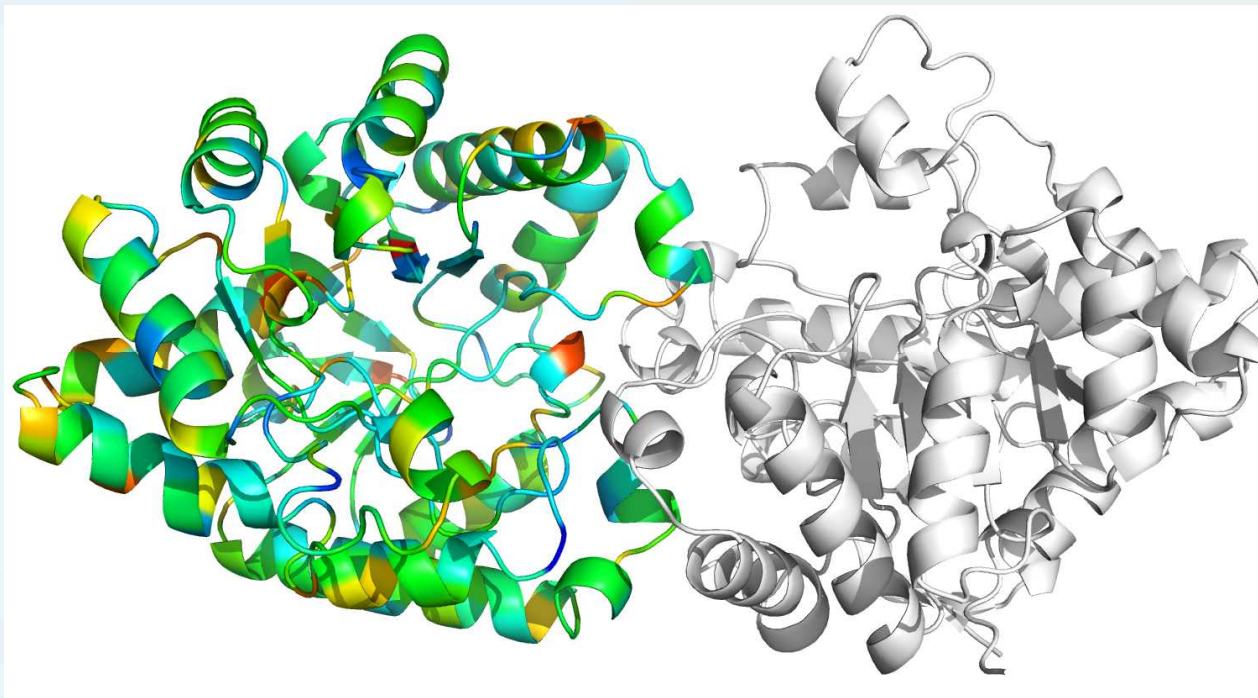
- Faster, less expensive than traditional cellular-based methods
- Proteins can be made directly from PCR templates, eliminating costly, time-consuming steps for plasmid cloning
- Multiple conditions can be screened simultaneously
- Multiple proteins can be expressed simultaneously in one reaction to facilitate production of active protein complexes
- Proteins can be characterized directly from the translation reactions.



**CFS GenDecoder robot
capable of producing
384 proteins/day.**



Putative hotspots for stabilizing mutations

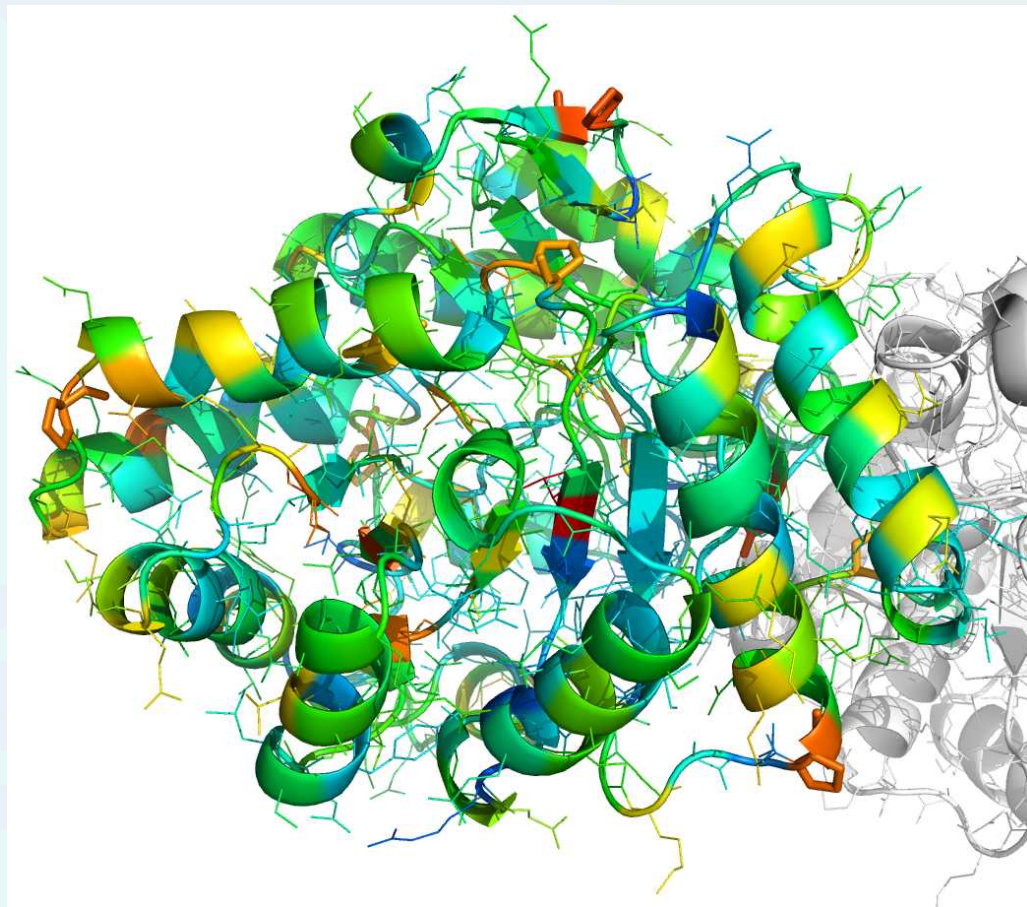


OPH dimer (PDB: 1DPM)

1. Residues are color-coded (from blue to red) according to the $\Delta\Delta G$ upon mutations.
2. Red corresponds to high stabilization effect.
3. The metal-coordinating and ligand-binding residues are not selected for mutation.

- ✓ The majority of the positions does not have a strong stabilizing mutations.
- ✓ There are a few hotspots that can be mutated to increase the stability.

Hotspots: Proline?



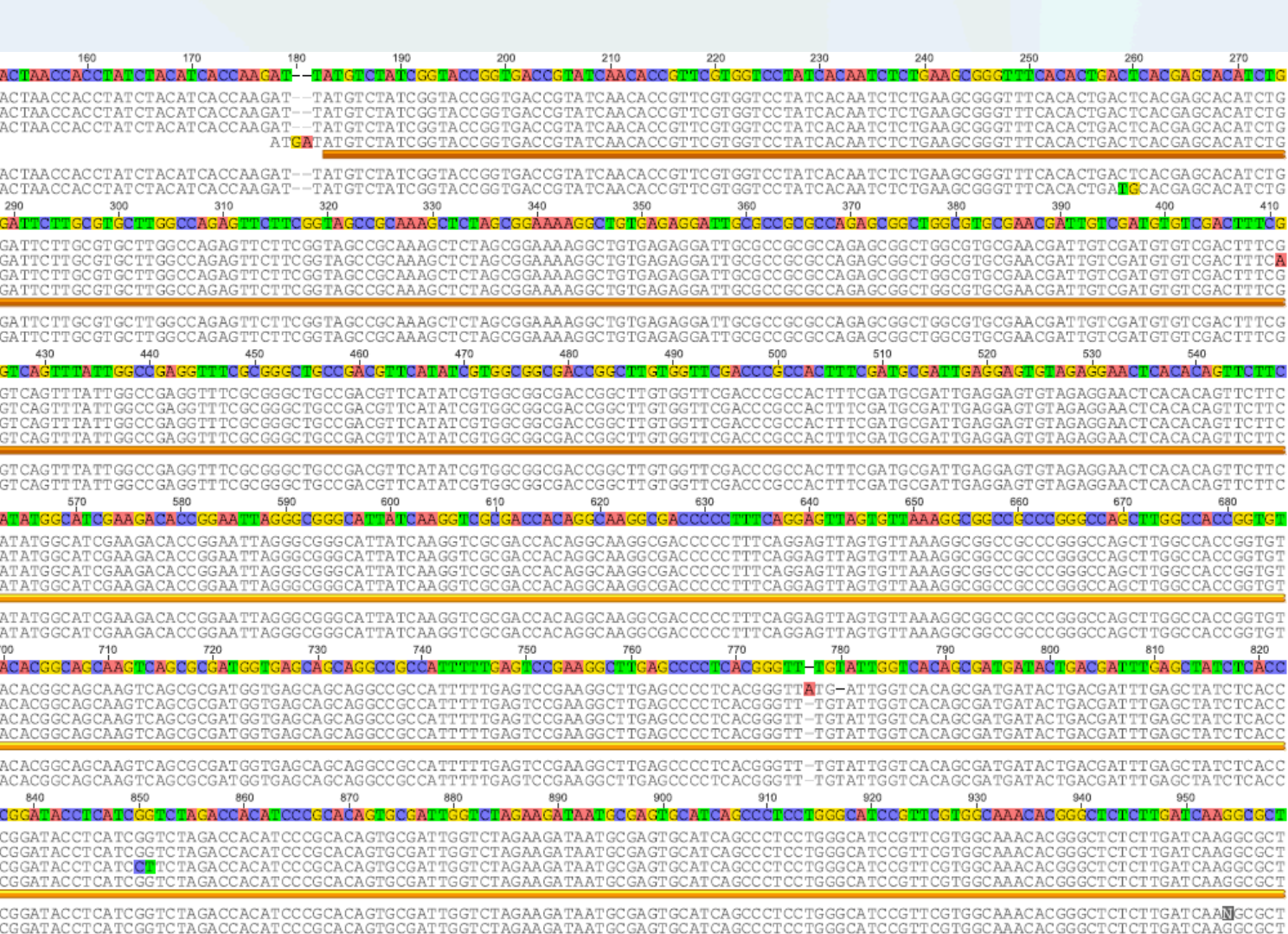
Many conserved prolines, which are picked up by Medusa/Eris as possible stabilizing hotspots.

These proline residues function as the capping of secondary structures, which might not be amenable for mutations without affecting the structure and folding dynamics.

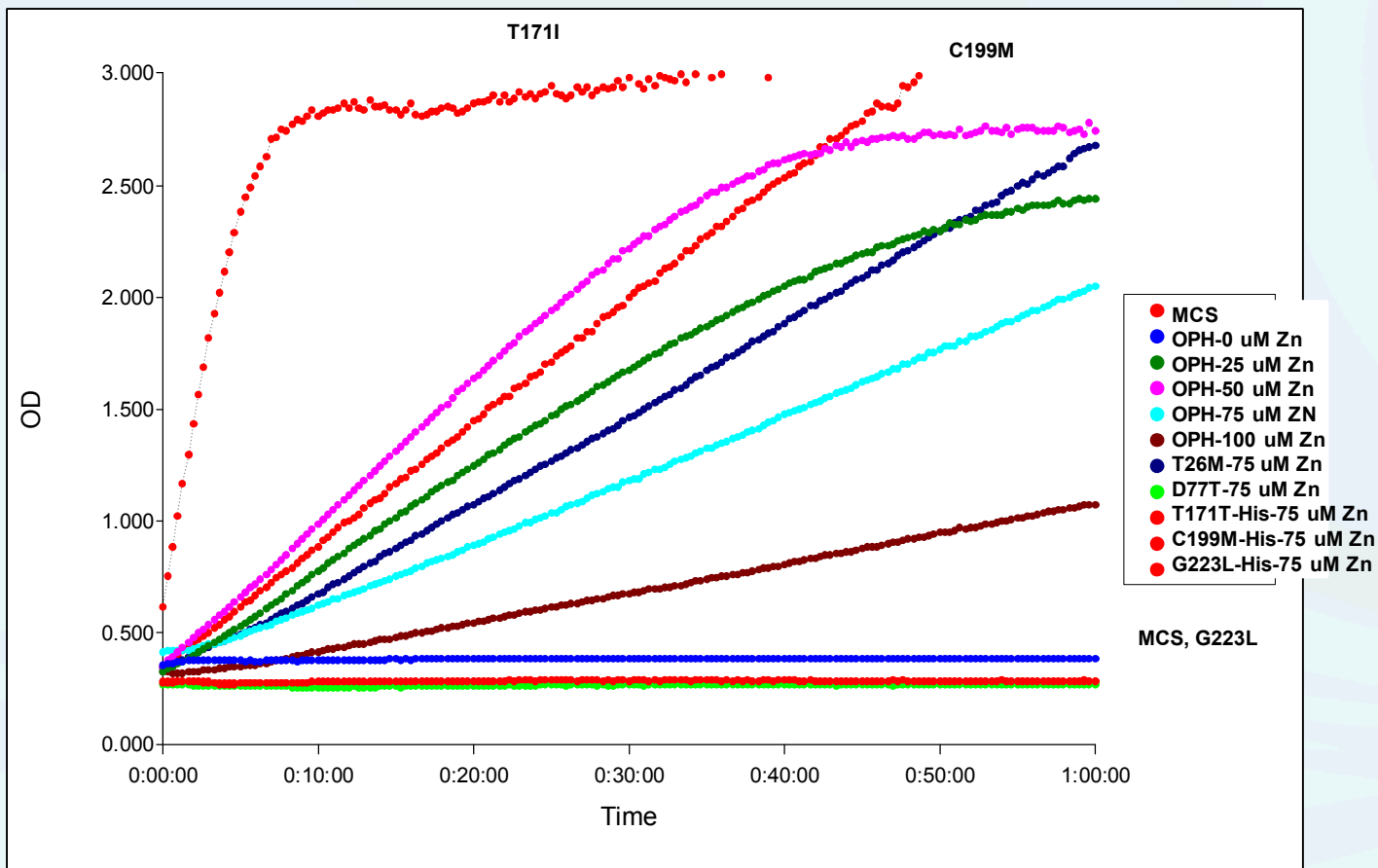
Predicted candidates

Residue	Mutation	Predicted ddG (kcal/mol)
T54	T53Met	-5.0 (Z = -2.0)*
D105	D105Thr	-6.7 (Z = -2.7)
T199	T199Ile	-8.1 (Z = -3.3)
C227	C227Met	-3.8 (Z = -1.5)
G251	G251Leu	-6.7 (Z = -2.7)

***Z-score is estimated by 100 rounds simulation for both wild type, and mutant**

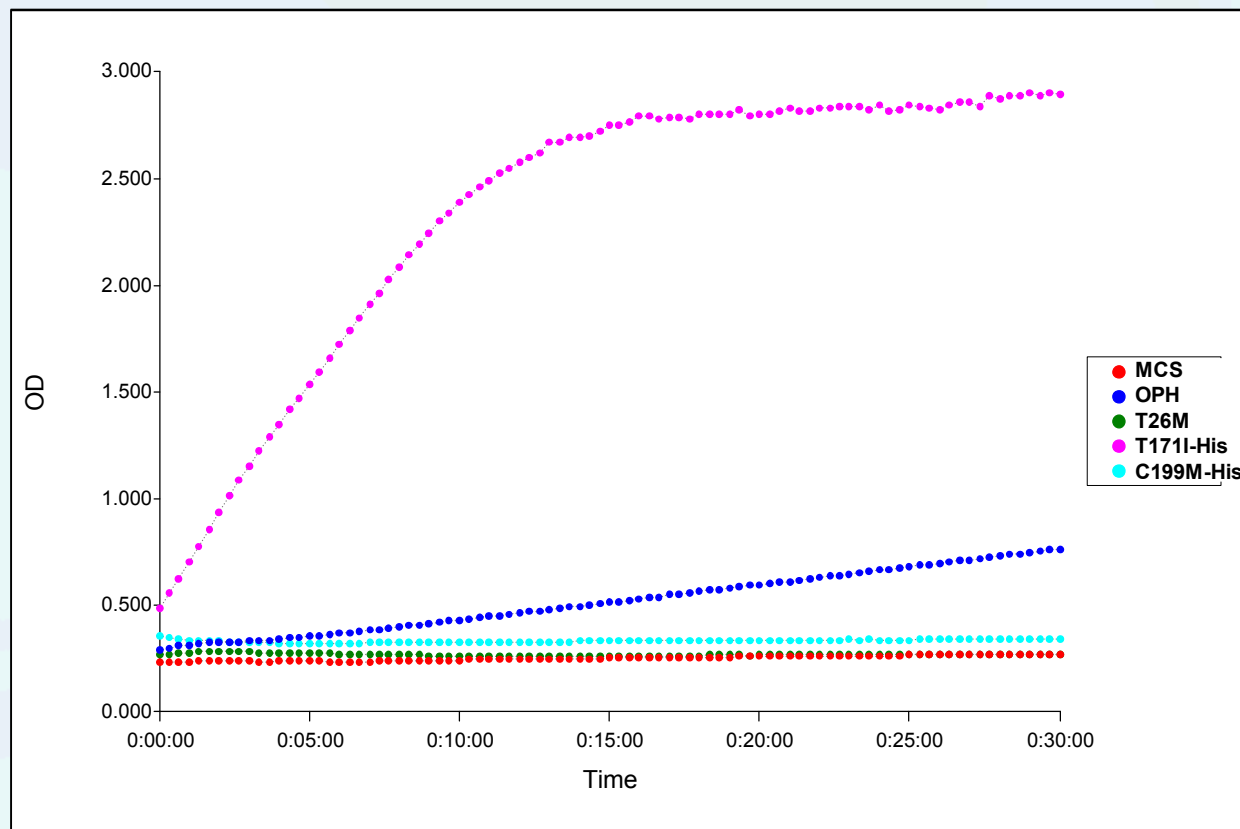


Different $[\text{Zn}(\text{OAc})_2]$ used in CFS translation of WT OPH; 75 μM used for OPH mutant.
2 ul of CFS reaction was used for one activity assay.



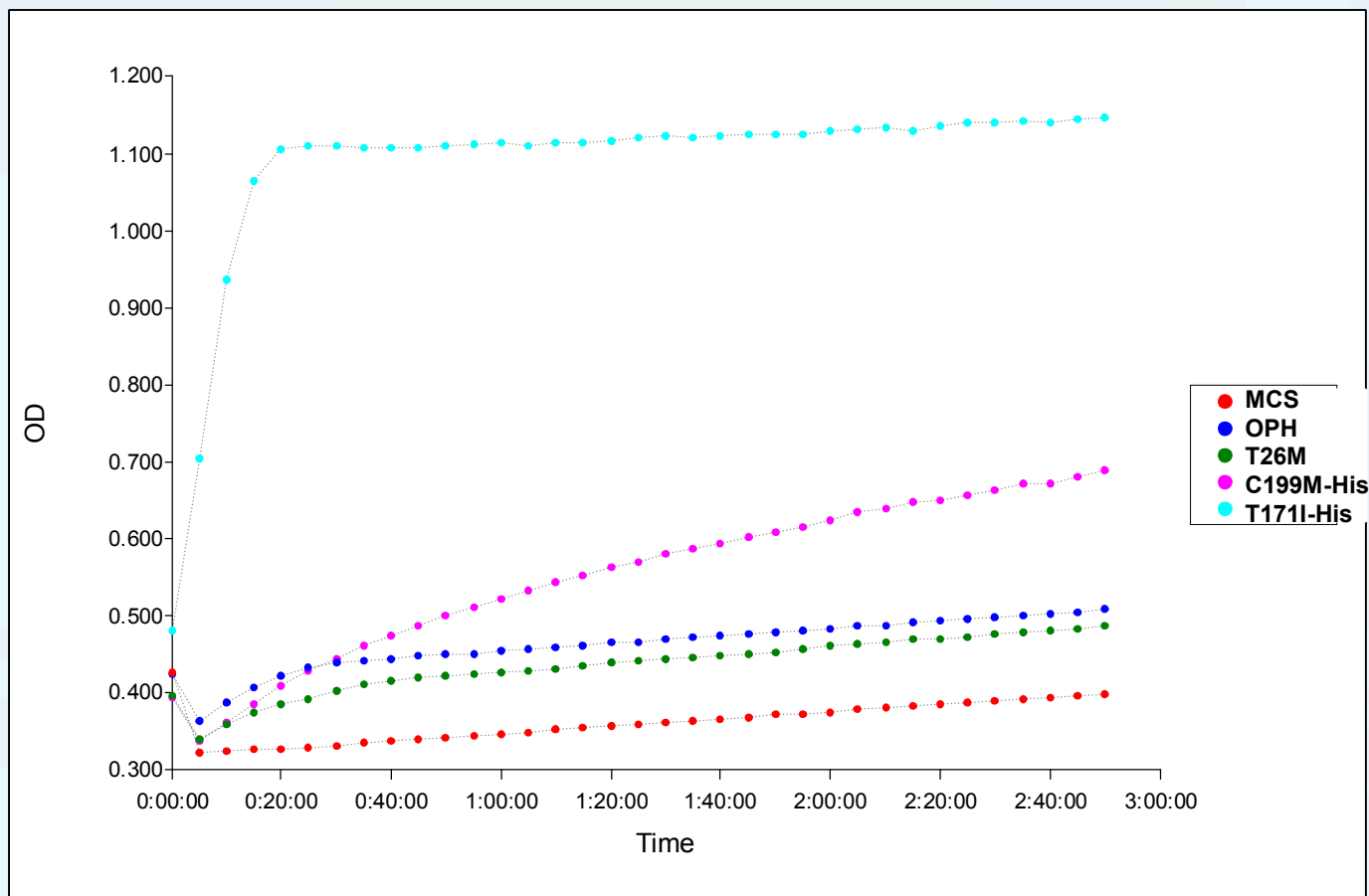
Assay condition: 2 ul of CFS OPH or mutant was added to 200 ul of CHES buffer containing same conc. of Zn as in it's CFS translation, activity was determined: 5 s mix, O.D. 405nm was read every 20 s for 1h.
T171I has much higher activity than WT OPH, T26M and C199M has similar activity as WT OPH. D77T and G223L are not active.

Large scale CFS reaction, 50 μM $\text{Zn}(\text{OAc})_2$ during translation.

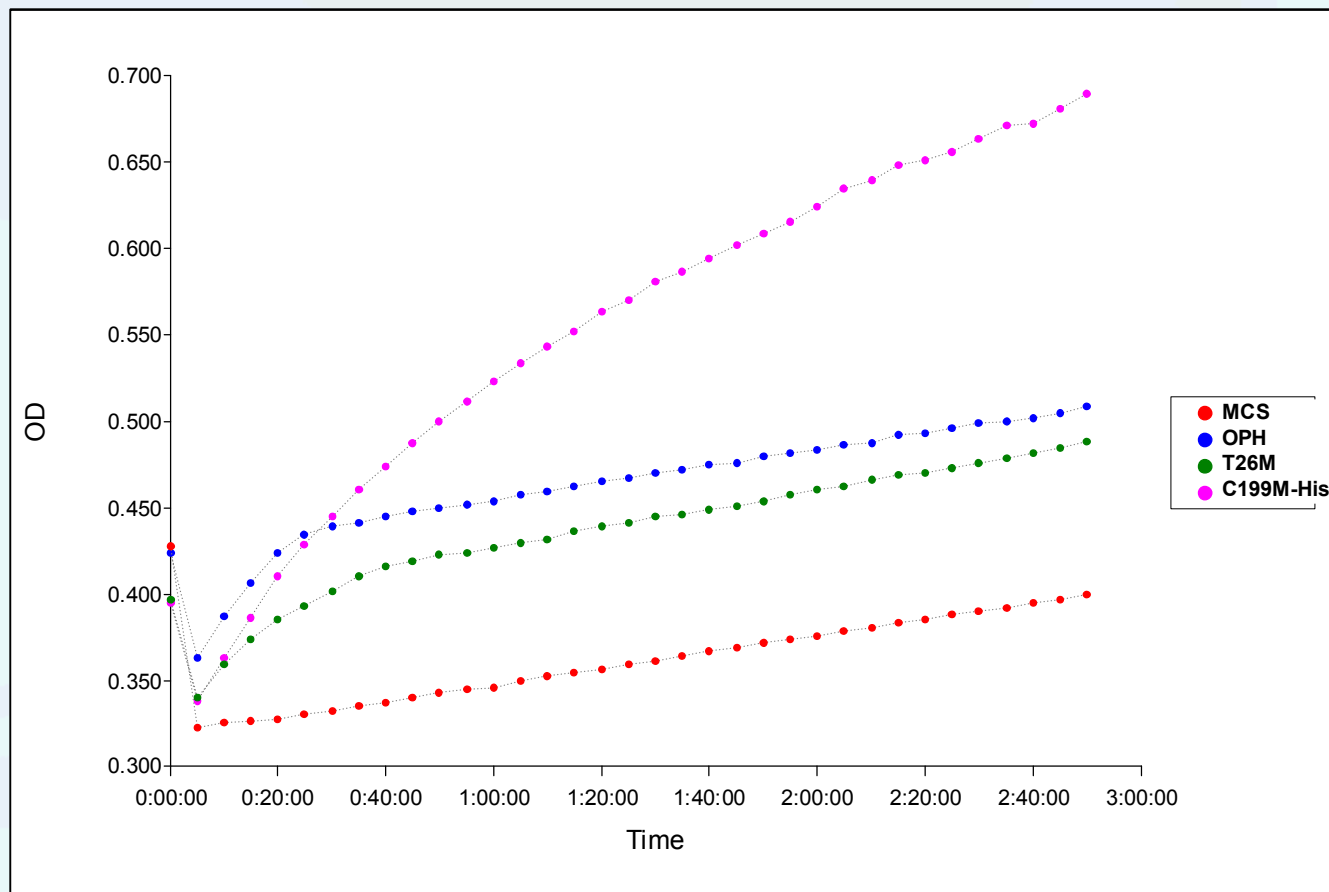


Assay condition: 2 μl of CFS OPH was added to 200 μl of CHES buffer containing 50 μM of $\text{Zn}(\text{OAc})_2$; activity: 5 s mix, O.D. 405 nm was read every 20 s for 30 min.

Use optically-clear adhesive film for long temp expts. to prevent condensation volume losses and damage to plate reader.



Assay condition: CFS OPH and mutants were diluted in CHES buffer at 1:20 ratio, then add 2ul into 200 ul pre-heated CHES buffer containing Zn. Eon was pre-heated to 60C as well. Paraoxon was added to a final conc of 1mM. Activity was determined: at 60C, 5 s mix, O.D. 405nm was read every 5 min for 3 h.

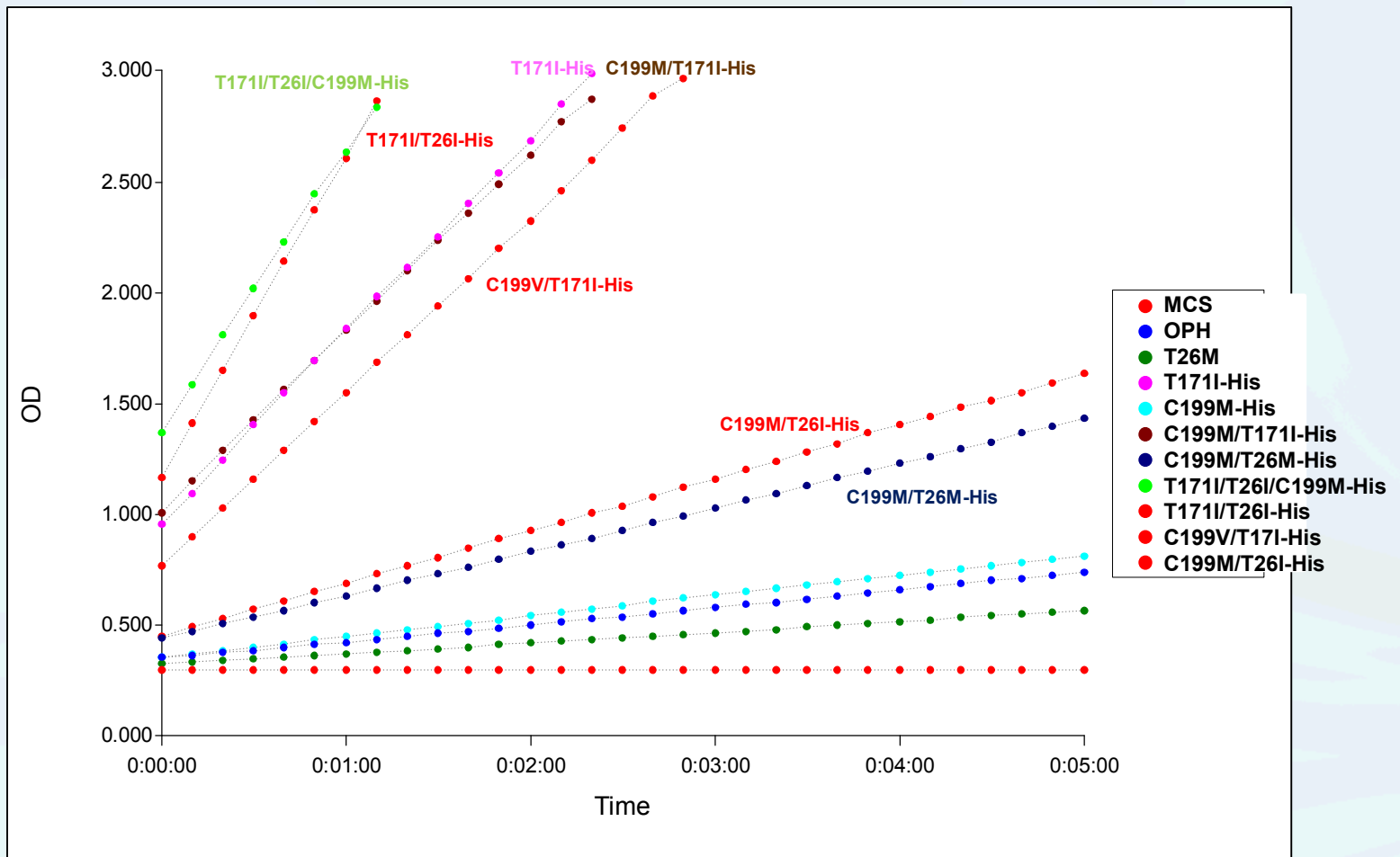


Same figure except T171I-His is not included.

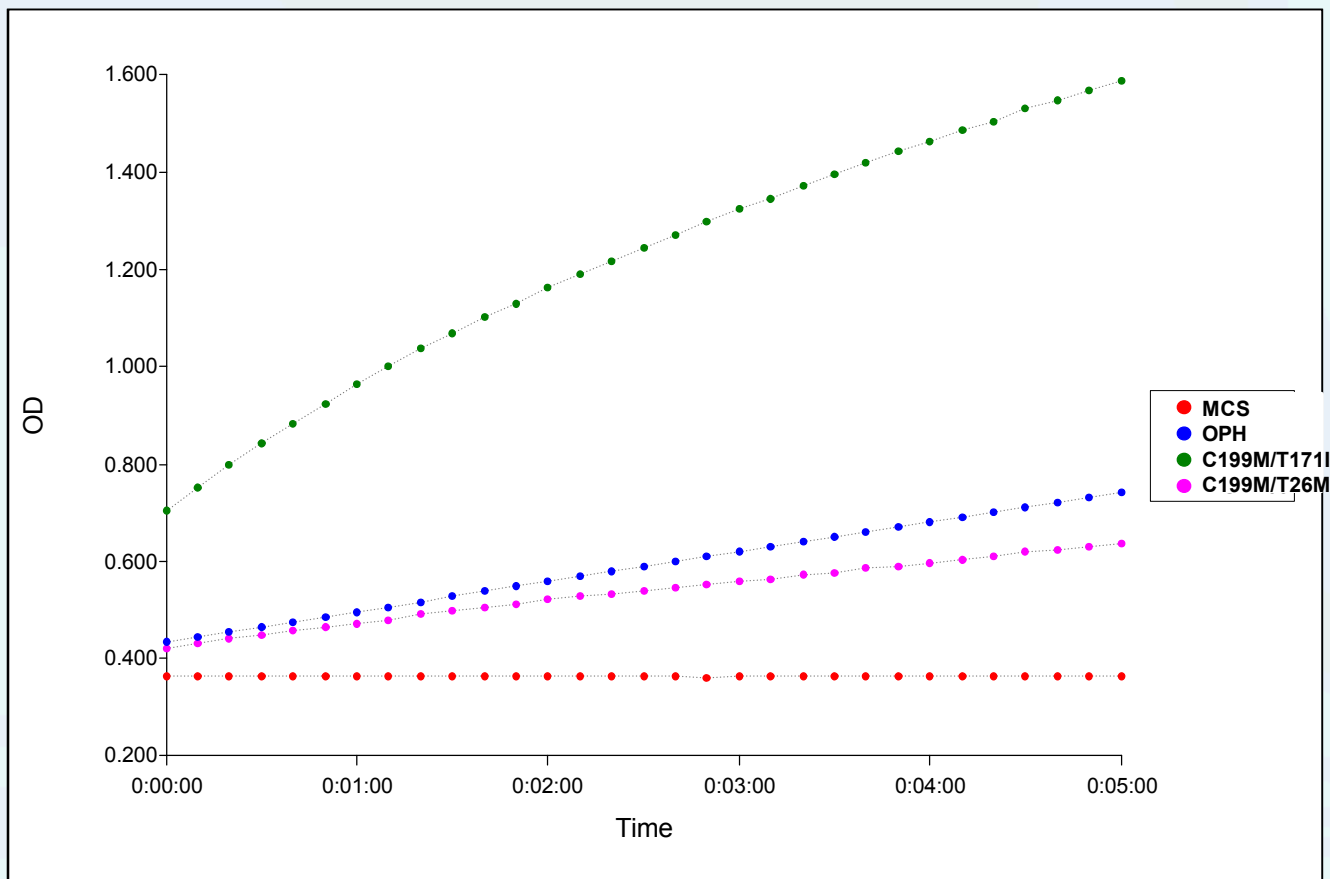


Are beneficial effects of mutations additive?

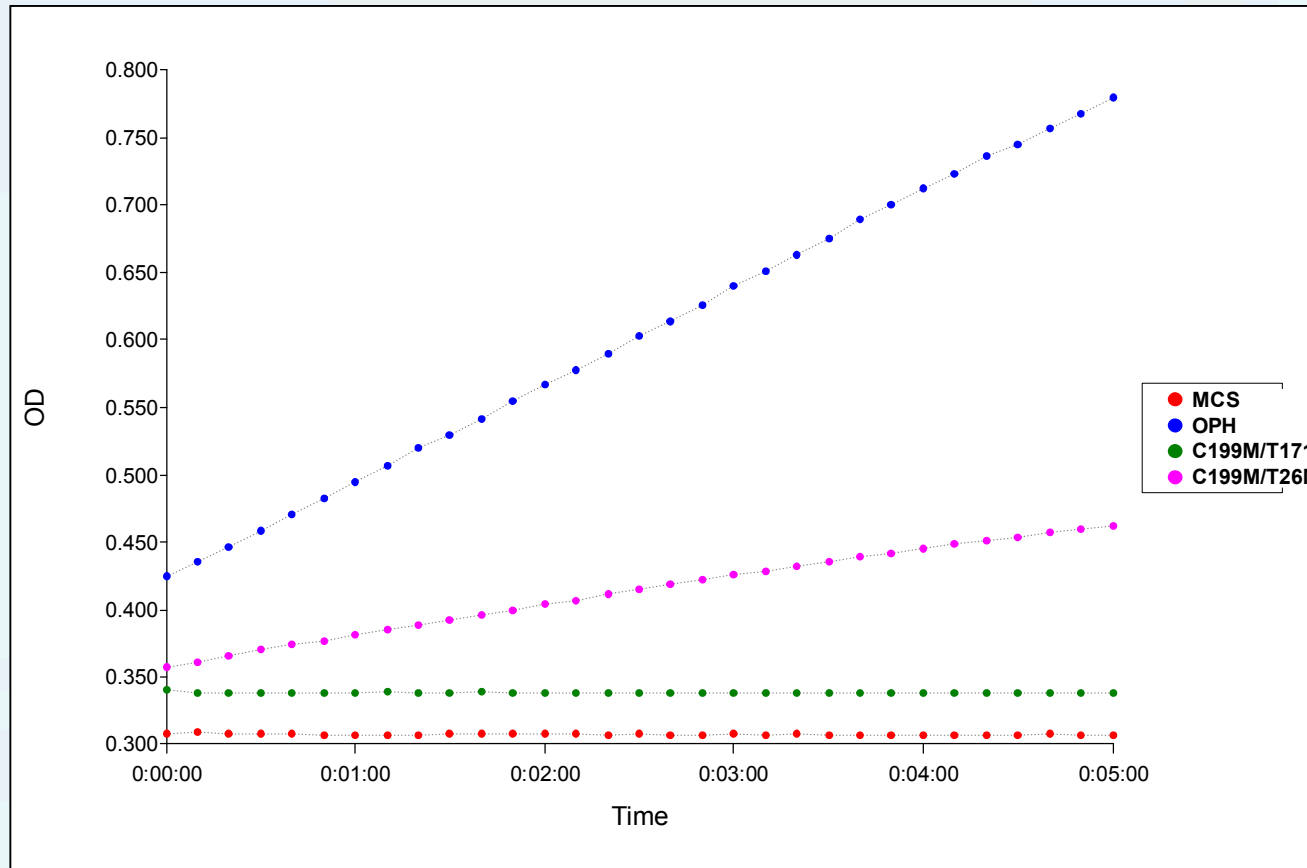
	Expressed	No Expression
1. Single mutants:	T26M D77T T171I-His C199M-His G223L-His	T26M-His D77T-His T171I C199M G223L
2. Double mutants:	C199M/T171I-His C199M/T26M-His	T171I/T26M-His
3. Triple mutant:		T171I/T26M/C199M-His



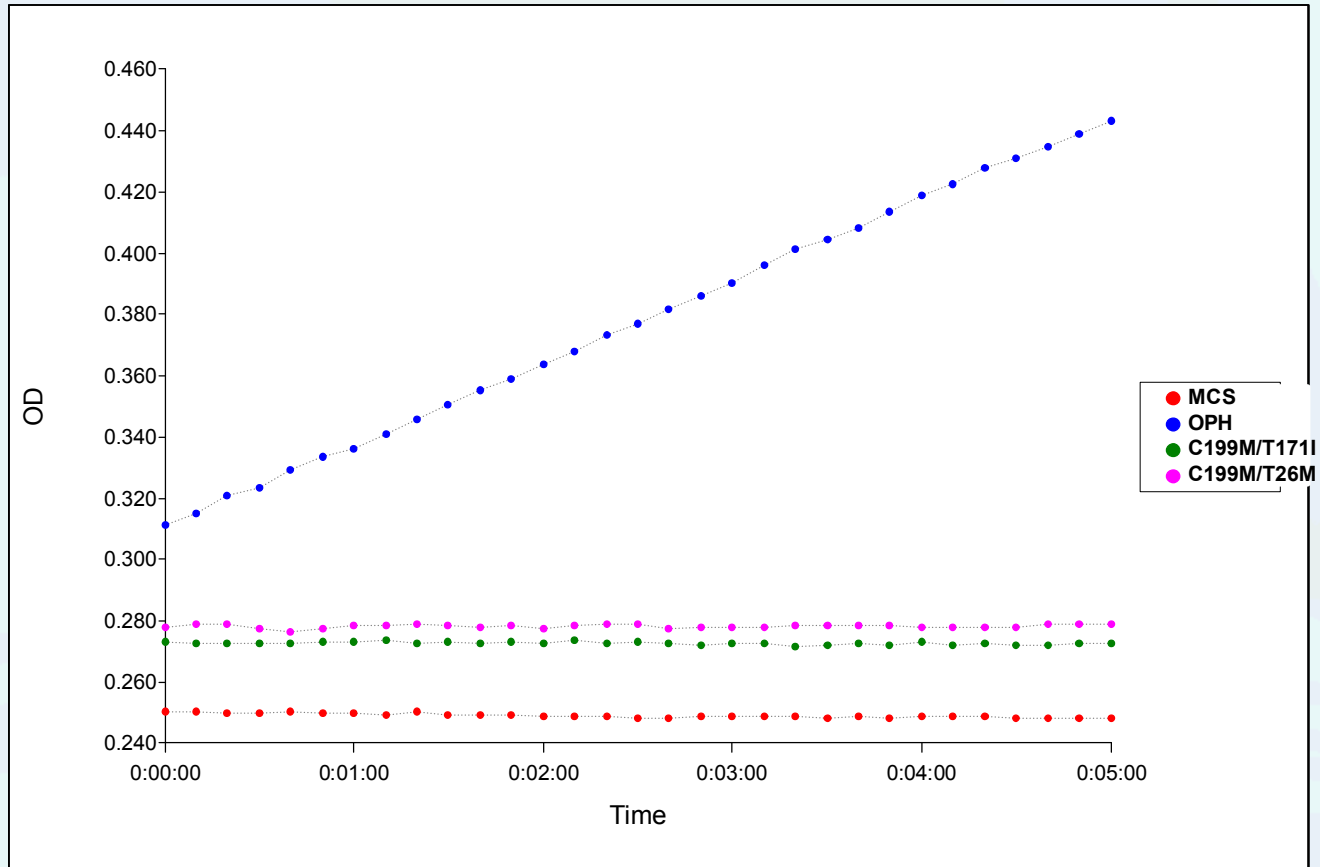
Time course of cell growth on C199M/T171I and C199M/T26M activities



At time 0 min, C199M/T171I is more active than WT OPH, but the activity is not as high as T171I single mutation. C199M/T26M is slightly less active than WT OPH by a tiny bit.



At time 10 min, C199M/T171I lost activity. C199M/T26M is still active but with a much lower activity than WT OPH.



**At time 30 min, C199M/T171I and C199M/T26M both lost activity.
WT OPH remains active.**

New mutants: T171I/T26I/C199M-His

T171I/T26I-His

C199V/T171I-His

C199M/T26I-His

T26I works much better than T26M. Usage of T26I instead of T26M in double-mutation combination increased overall activity.

Double mutants (C199M/T171I-His and C199M/T26M-His) don't decrease activity vs. their single mutants. All samples made using same master mix and activities were measured simultaneously.

Confirmed sequences of Saumil's mutants listed below. They are all correct.

**R13E, R13E-His,
R13E/R63E, R13E/R63E-His
R13E/R63E/D93K, R13E/R63E/D93K-His
R13E/R63E/D93K/K266E, R13E/R63E/D93K/K266E-His**

**H95K, H95K-His
H95K/R136E, H95K/R136E-His
H95K/R136E/K311E, H95K/R136E/K311E-His
H95K/R136E/K311E/R328E, H95K/R136E/K311E/R328E-His**

Amino acid sequence alignment of OPH5.1 vs. wt OPH. First five amino acids from N-term are replaced by his-tag in OPH 5.1.

OPH5.1	1	M	GHHHHHHHHH	10	
WT OPH	1	M	SIGT	5	
OPH5.1	11	GDRINTVRGP	IISKAGFTLT	THEHICGSSAGFLRAWPEFFGSRKALAEKAVRGLRE	70
WT OPH	6	GDRINTVRGP	ITISEAGFTLT	THEHICGSSAGFLRAWPEFFGSRKALAEKAVRGLR	65
OPH5.1	71	GV	ETIVDVSTFDIGRDV	SLLAEVSRAADVHIVAATGLWFDPPLSMRLRSVEELTQFFLRE	130
WT OPH	66	GV	RTIVDVSTFDIGRDV	SLLAEVSRAADVHIVAATGLWFDPPLSMRLRSVEELTQFFLRE	125
OPH5.1	131	IQYGI	EDTGIEAGIIKVATTGKATPFQELVLKAAARASLATGVPVTTHTAASQRDGEQQA	190	
WT OPH	126	IQYGI	EDTGIRAGIIKVATTGKATPFQELVLKAAARASLATGVPVTTHTAASQRDGEQQA	185	
OPH5.1	191	AIFE	EEGLSPSRVCIGHSDDTDDLSYLTALAARGYLIGLDHIPHSAIGLEDNASASALLG	250	
WT OPH	186	AIFE	SEGLSPSRVCIGHSDDTDDLSYLTALAARGYLIGLDHIPHSAIGLEDNASASALLG	245	
OPH5.1	251	IRSWQTRA	KLIKALIDQGYKKQILVSNDWLF	GFSSYVTNIMDVMDRVNPDGMAFIPK	310
WT OPH	246	IRSWQTRAL	LLIKALIDQGYMKQILVSNDWLF	GFSSYVTNIMDVMDRVNPDGMAFIP	305
OPH5.1	311	PFLRE	EGVPQETLAGITVTNP	AEF	LSPTLRAS 342
WT OPH	306	PFLRE	KGVPQQTLAGITVTNP	AR	FLSPTLRAS 337

Initial efforts to express Saumil's mutants failed due to presence of extra N-term amino acids and/or His-Tag. Recloned all mutants into our OPH gene and removed His-Tag. Stay tuned . . .

Conclusions and Future

Immobilizations → promote stability and activity

- Improvements: extra active-site metal & presence of substrate, crowding reagents, covalent
- Design materials for enzyme and enzyme for materials
- Design crosslinking sites into enzyme for max substrate delivery and product removal

Cell-free translation + modeling → promote stability and activity

- Improvements: thermal stability and activity
- Some mutations additive
- Cycles of correlating expression data with model to improve model and OPH.

Relevance to Nerve Agents?

Sent to Steve Harvey:

MCS (i.e., cell-free extract control), OPH, T26M, T171I-His, C199M-His, C199M-T171I-His and C199M-T26M-His.

**Synergy of coarse-grained modeling + Quantum Mechanical
Modeling of Active Site?**