

Nutrient Recycling

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Sandia National Laboratories

Motivation – scale of energy consumption exceeds nutrient production

- To meet 10% of liquid fuel needs (roughly 30 BGY):
 - Algae biomass: 200 - 500 M mt/yr.
 - Phosphorous: 2.4 - 6 M mt/yr
 - Compare 4.1 M mt in 2006: **61 – 146% of recent consumption.**
 - Nitrogen (nitrate, ammonium, etc.) 18 - 45 M mt/yr
 - Compare 14 M mt in 2006: **130 – 320% of recent consumption.**
- Food-vs-fuel concerns for nutrients.
- Nutrients (fertilizer)
 - Needed for biological productivity, not for fuel.
 - Phosphorous: mined resource, essentially nonrenewable.
 - Nitrogen: Haber-Bosch process has own energy requirements.

“The Achilles’
Heel of Algae
Biofuels: Peak
Phosphate”

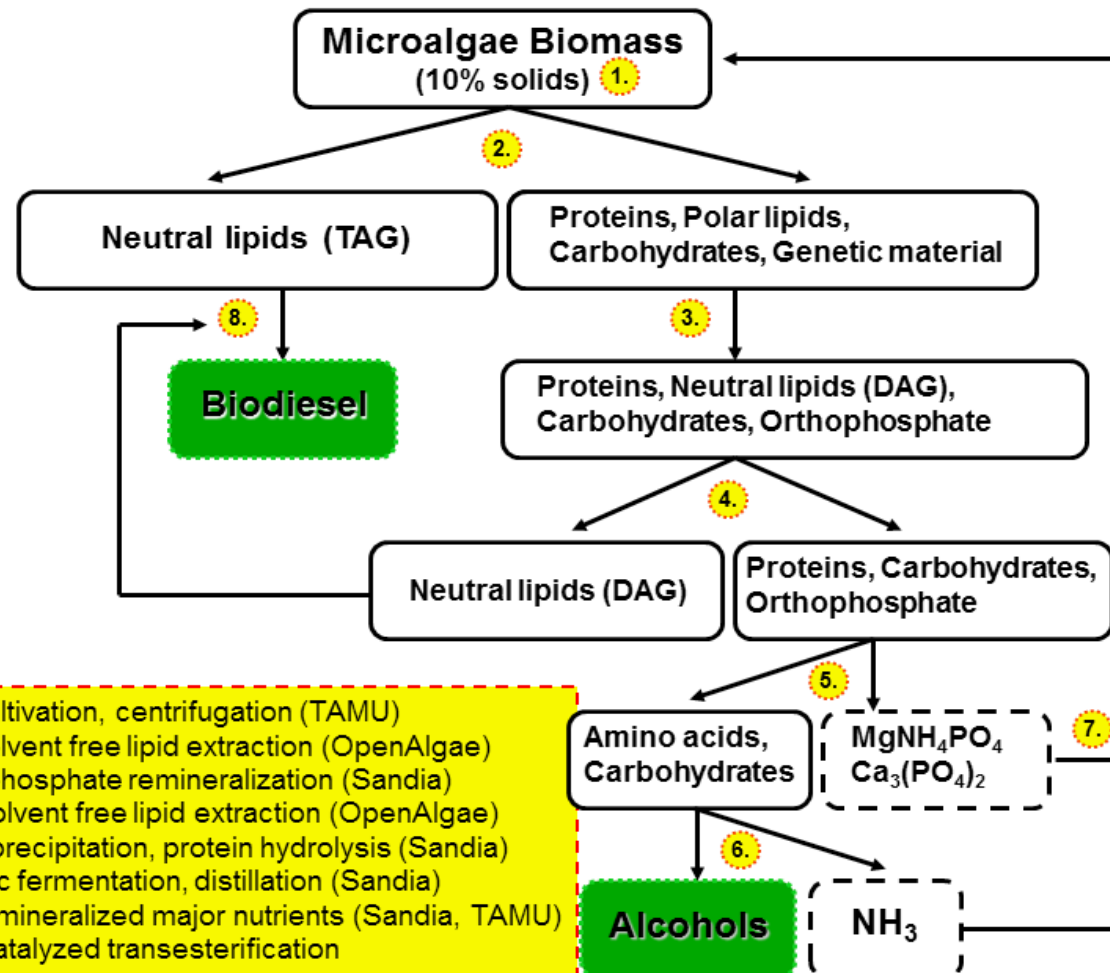
Forbes, Feb 2012

Need to recycle nutrients.

Cannot afford to pass through one time only.

Pate, Klise, Wu, “Resource demand implications for US algae biofuels production Scale-up,” Applied Energy, 88:3377-3388 (2011).

DOE Nutrient Recycling Project



- 1.) Raceway cultivation, centrifugation (TAMU)
- 2.) 1st stage, solvent free lipid extraction (OpenAlgae)
- 3.) Enzymatic phosphate remineralization (Sandia)
- 4.) 2nd stage, solvent free lipid extraction (OpenAlgae)
- 5.) Phosphate precipitation, protein hydrolysis (Sandia)
- 6.) Microaerobic fermentation, distillation (Sandia)
- 7.) Return of remineralized major nutrients (Sandia, TAMU)
- 8.) Acid/base catalyzed transesterification

DOE funded project is a partnership between national lab, university and industry

- Sandia National Labs
 - Project Lead
 - Biochemistry
 - Precipitation Science
- Texas Agrilife:
 - biomass production
 - pilot scale field trials
 - *Nannochloropsis salina*
 - *Phaeodactylum tricornutum*
 - (NAABB strains)
- OpenAlgae
 - TAG extraction
 - DAG extraction
 - Converted phospholipids



Staged approach to development of a one pot reaction

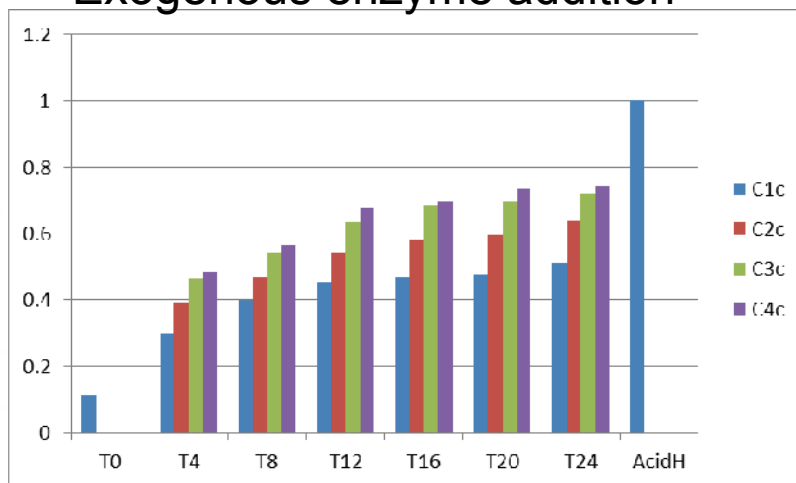
- Test different cocktails for remineralization of phosphate and phospholipid conversion.
 - Determine rate of remineralization
 - Optimize reaction conditions (extraction of nutrients from solid phase)
 - Identify recalcitrant pools
 - Minimize reaction time
- Develop microbial consortia with appropriate enzymatic activities: test culture supernatants.
 - Identify candidate genes, clone, overexpress
 - Test for protein and activity level
- Grow microbial consortia on residual algal biomass—expressing enzymes *in situ* and converting amino acids to ammonium.
 - Optimize growth conditions (limit conversion to microbial biomass)
 - Optimize enzyme production on residual biomass
 - Limit uptake of inorganic phosphate by microbial consortium

Majority of cellular phosphate rapidly remineralized in non-denatured algal biomass

| Biochemical fraction | % cell mass of fraction | gm P per gm DW of fraction |
|----------------------|-------------------------|----------------------------|
| RNA | 3-15 | 0.091 |
| DNA | 0.5-3 | 0.095 |
| Phosphoglycerides | 5-15 | 0.043 |
| ATP | <0.1 | 0.18 |

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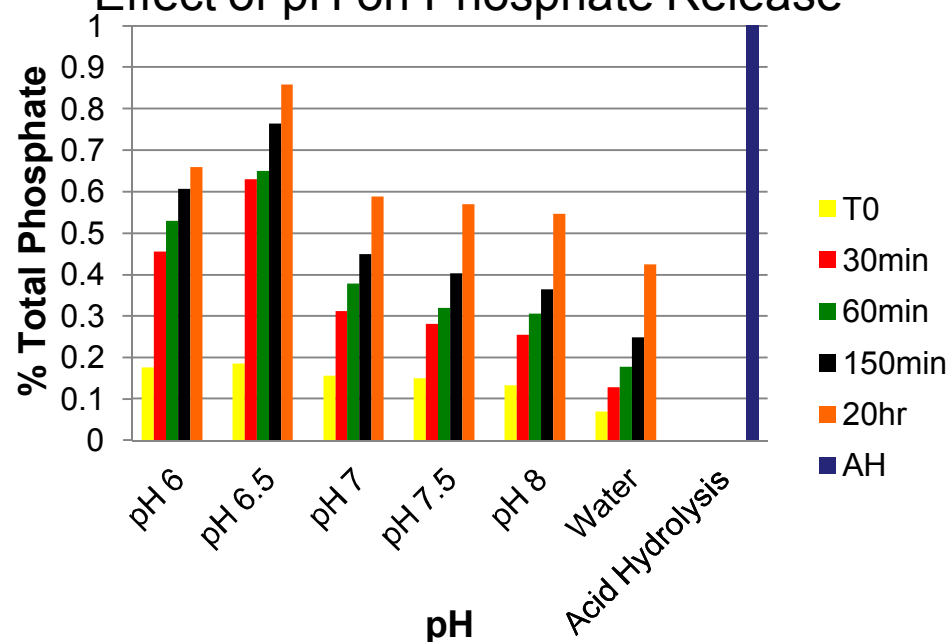
Exogenous enzyme addition



1. No enzyme
2. AP
3. AP + B
4. AP + B + PL-D

AP = alkaline phosphatase
B = benzonase
PL-D = phospholipase D

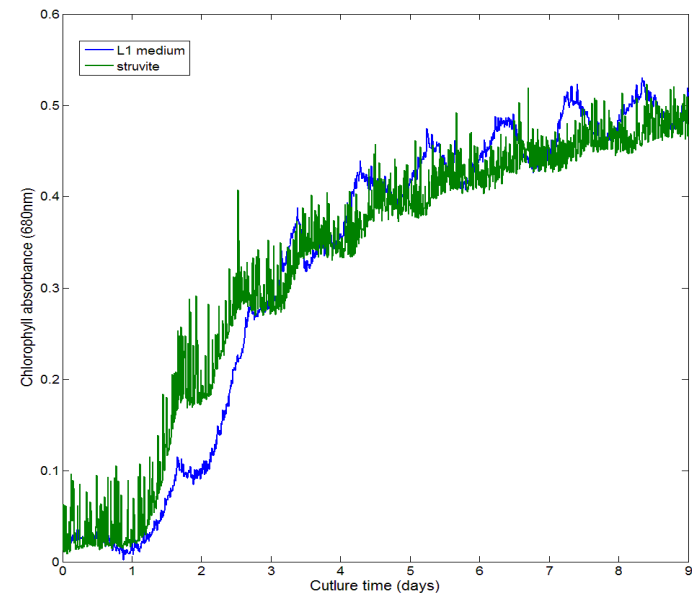
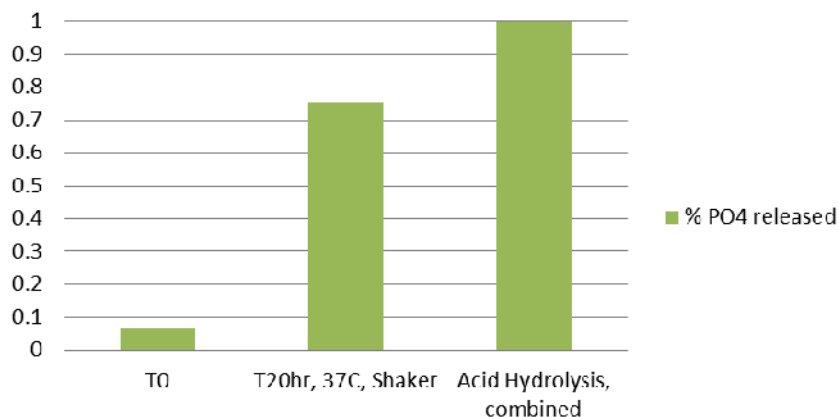
Effect of pH on Phosphate Release



Growth on recycled phosphate

- ~50 gm of 20% solids. *N. salina*
- Diluted to 2% solids pH 6.5, 37°, 20hrs
- Liberated phosphate used to replace total phosphate in lab cultures
- Growth of *P. tricornutum* and *N. salina*

% PO₄ released

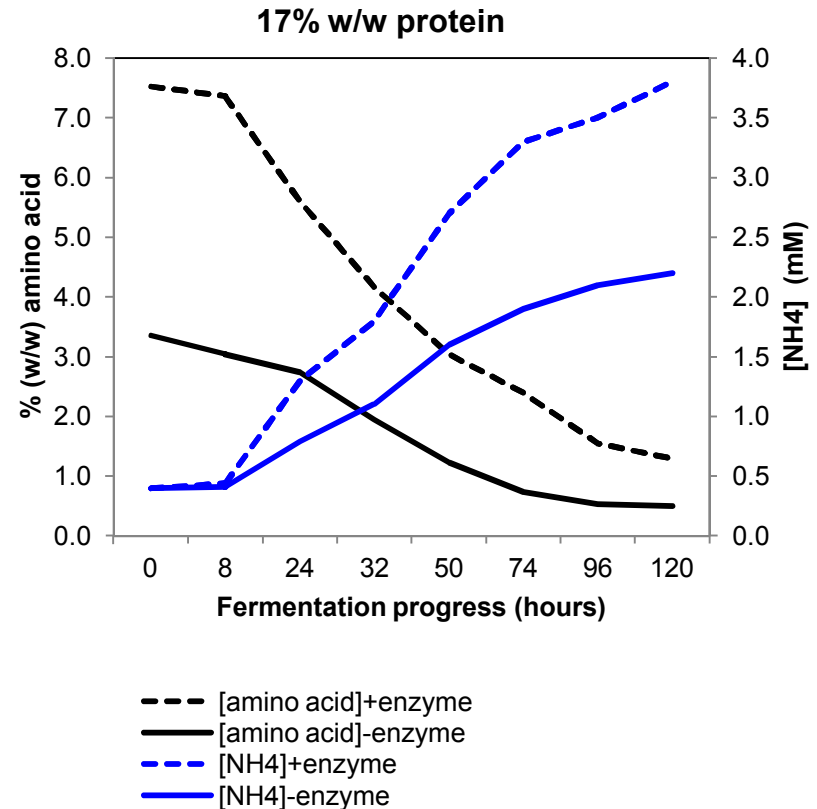


We will take advantage of NH_4 production by Previously developed fermentation process

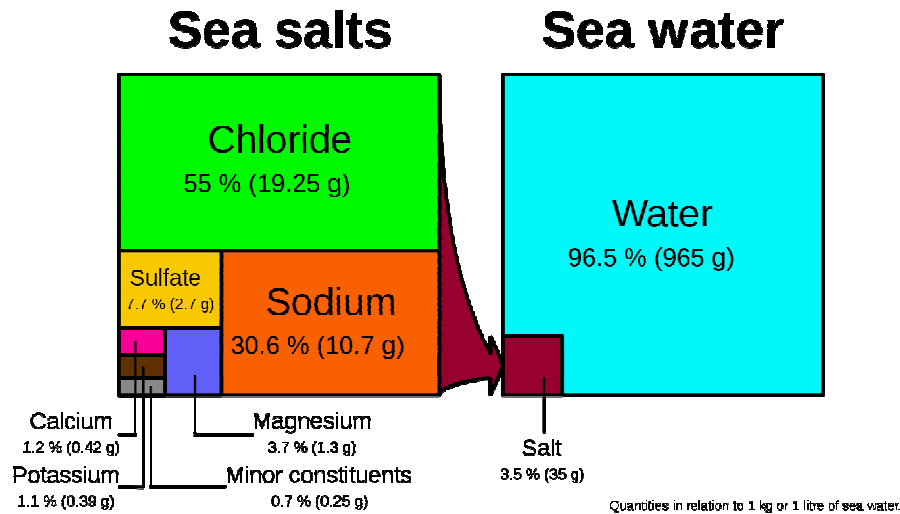
Subject lipid extracted biomass to microaerobic fermentation using an alcohol tolerant metabolically engineered *E. coli* strain, (Xin Huo, et al *Nature Biotech*, 2011)

- 1) Mixed alcohols = 50% protein yield using 5-step process:
- 2) Alcohol components do not significantly vary with biomass type
- 3) Accumulation of alcohols proceeds in distinct temporal phases: isobutanol \rightarrow n-methyl-butanol \rightarrow phenylethanol and n-butanol
- 4) Ammonium is accumulated in fermentation

liquor as amino acid breakdown product



In current system, Mg carried over with biomass may result in struvite fomation



Depending on chemical makeup of growth medium significant extracellular Mg may be carried over with biomass

Internal Mg stores may also be significant: Macronutrient

The resulting P/Mg ratio may promote the formation of struvite: MgNH_4PO_4

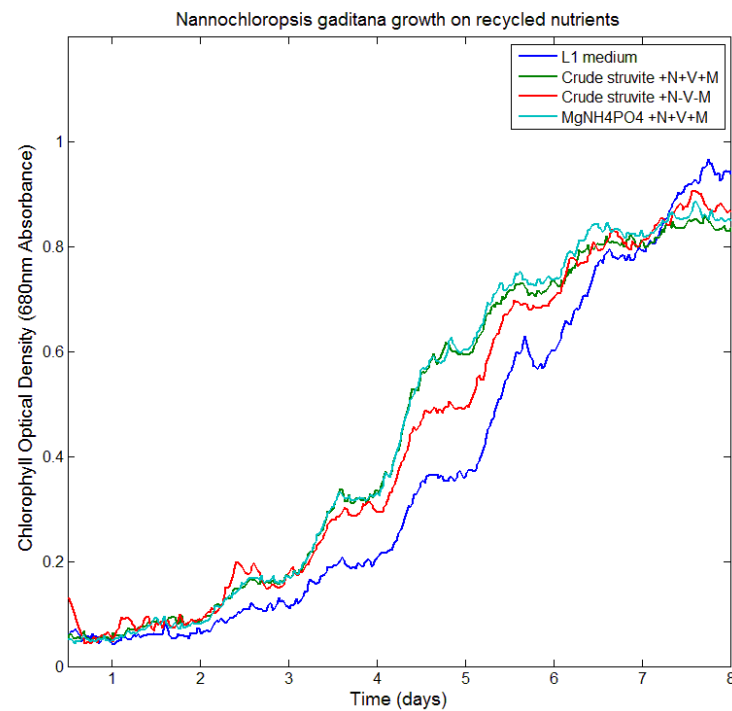
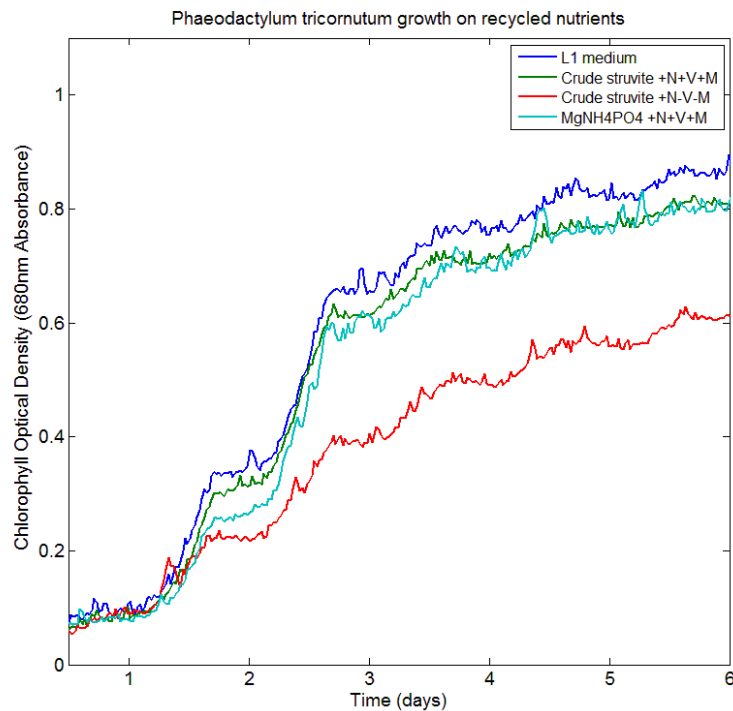
We will not alter the chemistry of the pond or biomass to form or use struvite

No requirement for new Mg

| NaNO_3 (M) | KH_2PO_4 (M) | Fe (mg/g) | Mg (mg/g) |
|------------------------|---------------------------------|--------------|--------------|
| 0.006 | 0.0003 | 3.37 | 77.3 |
| 0.006 | 0.0003 | 4.75 | 67.1 |
| 0.006 | 0.001 | 3.86 | 78.1 |
| 0.006 | 0.001 | 3.81 | 98.3 |
| 0.003 | 0.0003 | 2.83 | 82.0 |
| 0.003 | 0.0003 | 2.91 | 93.5 |
| 0.003 | 0.001 | 3.46 | 74.8 |
| 0.003 | 0.001 | 2.41 | 74.4 |

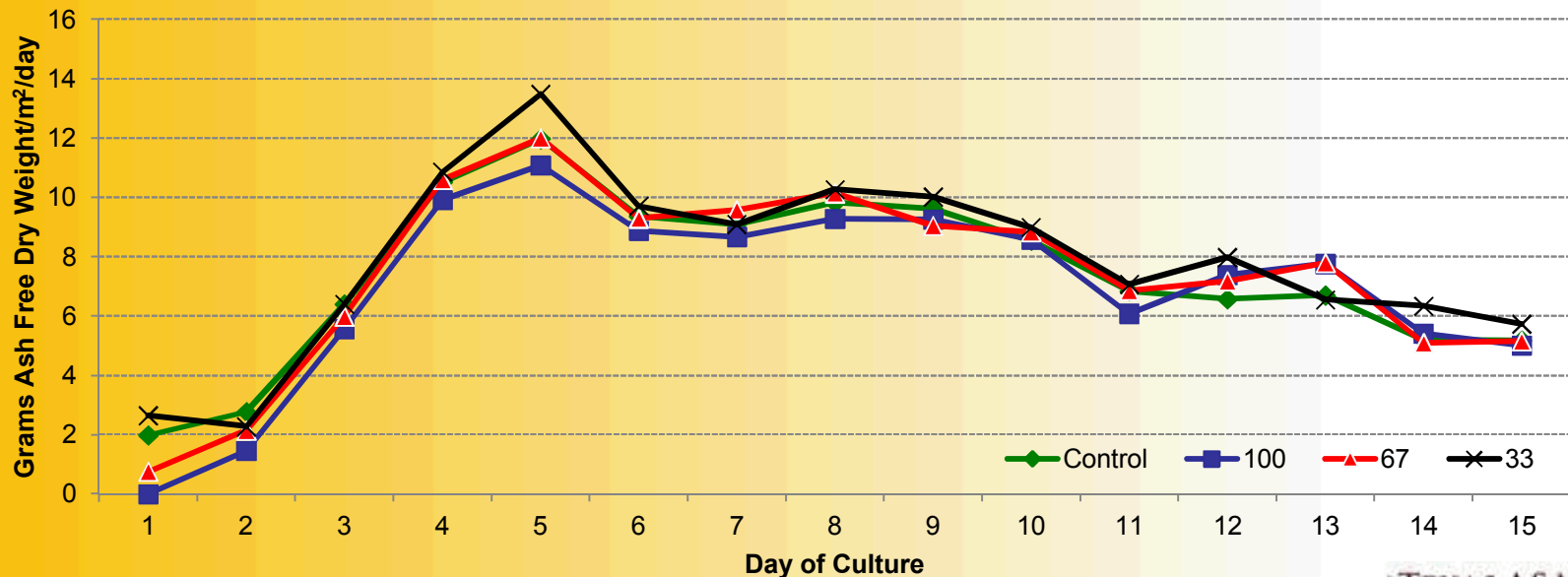
3-4 mMol Mg/gm AFDW

Struvite can replace “new” nutrients in microalgal culture



Struvite Supplementation in raceway cultures: (*Nannochloropsis salina*)

Daily biomass productivity (g AFDW/m²/day) of *Nannochloropsis salina* (CCMP 1776) cultivated with phosphorus replacement (% of control) using commercial struvite



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Summary

- Phosphate can be remineralized from non denatured *N. salina* biomass by enzymatic digest or mild pH treatment
- Remineralized phosphate can provide 100% of phosphate required for growth of *N. salina* or *P. tricornutum*
- Crude struvite can provide 100% of phosphate and large fraction of nitrogen for the growth for the growth of *N. gaditana*, *N. salina* and *P. tricornutum* at laboratory scale
- Crude struvite can provide 100% of phosphate and large fraction of nitrogen for the growth for the growth of *N. salina* in pilot scale outdoor raceways.

Acknowledgments

DOE EERE BioEnergy Technology Office

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Texas Agrilife

- Anthony Siccardi

Open Algae

- Peter Kipp
- Hoyt Thomas



Spare slides

Experimental Design (Mono-Culture)

- Stocking: Cultures of *Nannochloropsis salina* (CCMP 1776) were stocked into 12 outdoor 3 m² fiberglass raceways to achieve an initial stocking density of ~0.15 g/L afdw at 5 cm depth
- Nutrient Mix: “ODI” mix composed of ammonium sulfate, phosphoric acid and ferrous sulfate
- Experimental Design:
 - Control: supplemented with ODI nutrients at a 16:1 N:P ratio
 - Struvite: supplemented with commercial struvite to replace 33, 67 and 100% of the phosphorus in the control treatment
 - Water depth in each raceway was gradually increased to a final depth of 20 cm providing a total working volume of 550 L
- Parameter Monitoring: Raceways were monitored daily for solar radiation, rainfall, wind-speed, pH, temperature, salinity, afdw, ammonia, nitrite, nitrate, and phosphate

Day 5 Biomass Productivity (g AFDW/m²/day)

Day 5 biomass productivity (g AFDW/m²/day) of *Nannochloropsis salina* (CCMP 1776) cultures with phosphorus replacement using commercial struvite^{1,2,3}

| Phosphorus Replacement Level (% Control) | Day 5 biomass productivity (g AFDW/m ² /day) |
|--|---|
| Control | 11.98±2.22 ^a |
| 100 | 11.08±0.50 ^a |
| 67 | 12.00±1.25 ^a |
| 33 | 13.48±0.70 ^a |

^aMeans with similar superscript in the same column are not statistically different ($p > 0.05$)

²N = 3 raceways

³Standard deviation

Final Biomass Productivity (g AFDW/m²/day)

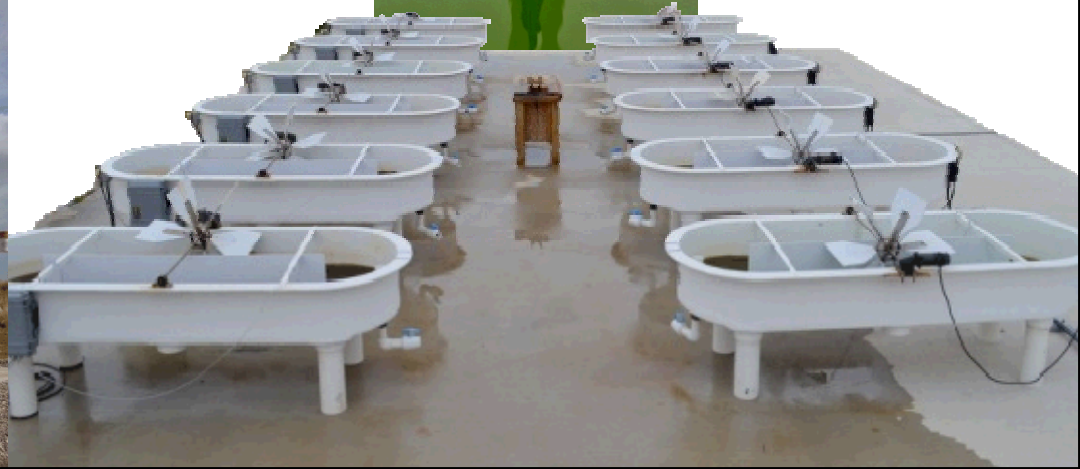
Final biomass productivity (g AFDW/m²/day) of *Nannochloropsis salina* (CCMP 1776) cultures with phosphorus replacement using commercial struvite^{1,2,3}

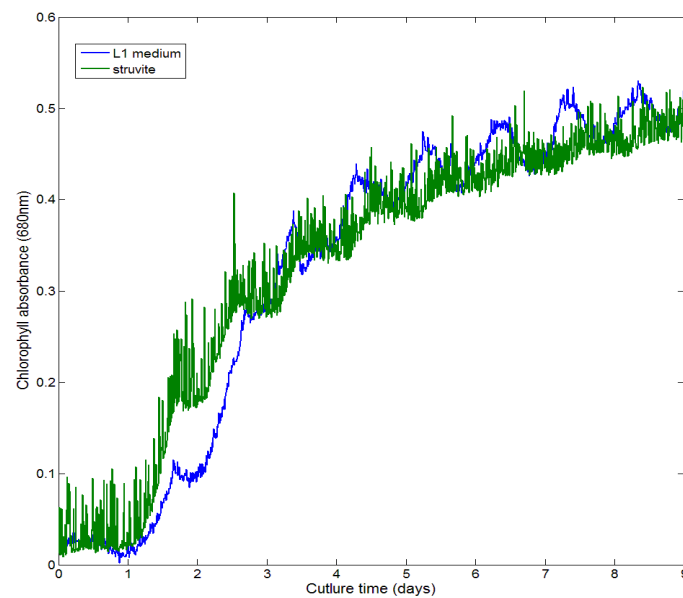
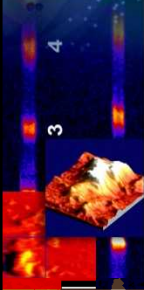
| Phosphorus Replacement Level (% Control) | Final biomass productivity (g AFDW/m ² /day) |
|--|---|
| Control | 5.18±0.32 ^a |
| 100 | 5.01±0.38 ^a |
| 67 | 5.16±0.61 ^a |
| 33 | 5.73±0.53 ^a |

¹ Means with similar superscript in the same column are not statistically different ($p>0.05$)

²N = 3 raceways

³Standard deviation

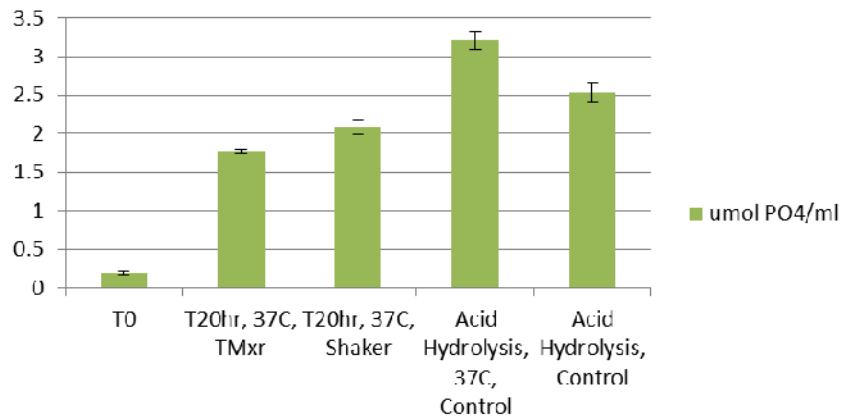




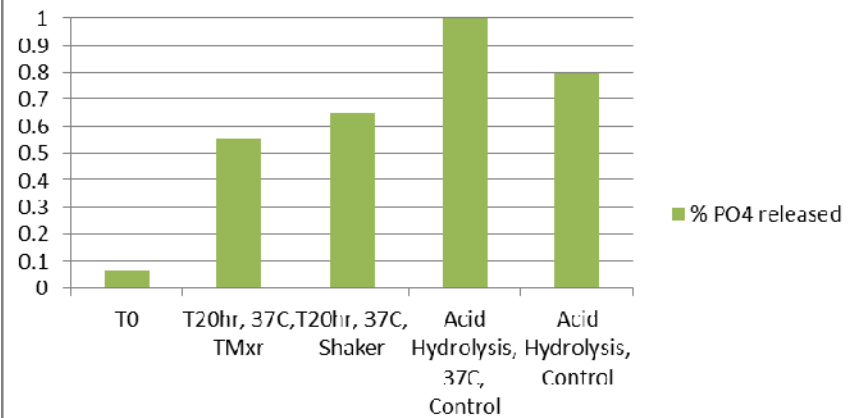
umol PO₄ released from 1ml of TX
biomass (2% solids); umol/ml = mM

% PO₄ release TX biomass (2% solids)

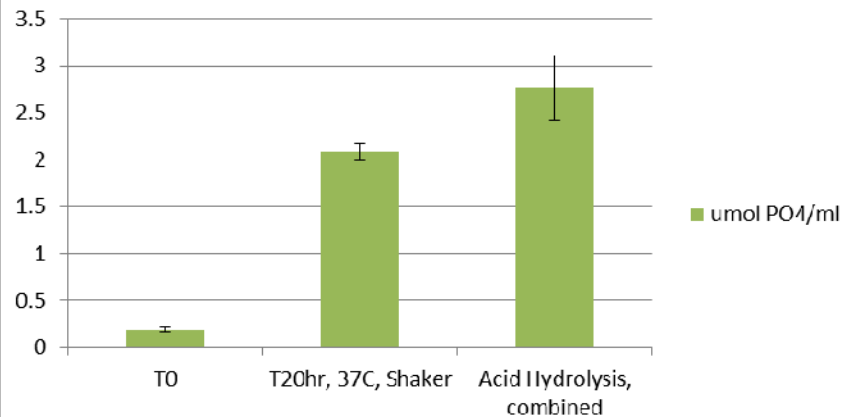
umol PO₄/ml



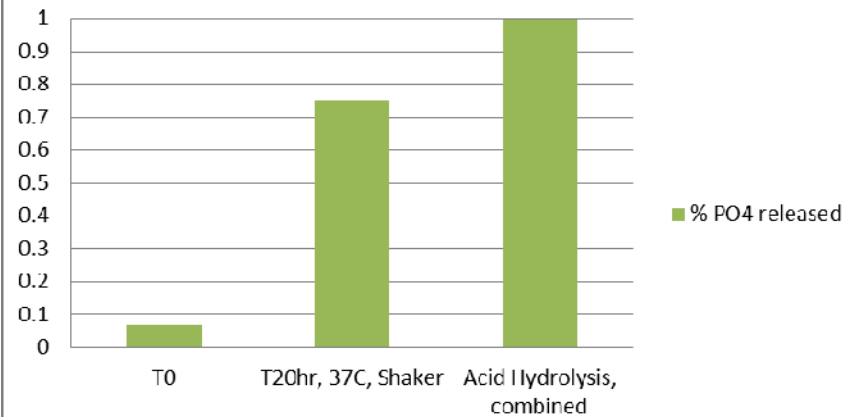
% PO₄ released



umol PO₄/ml

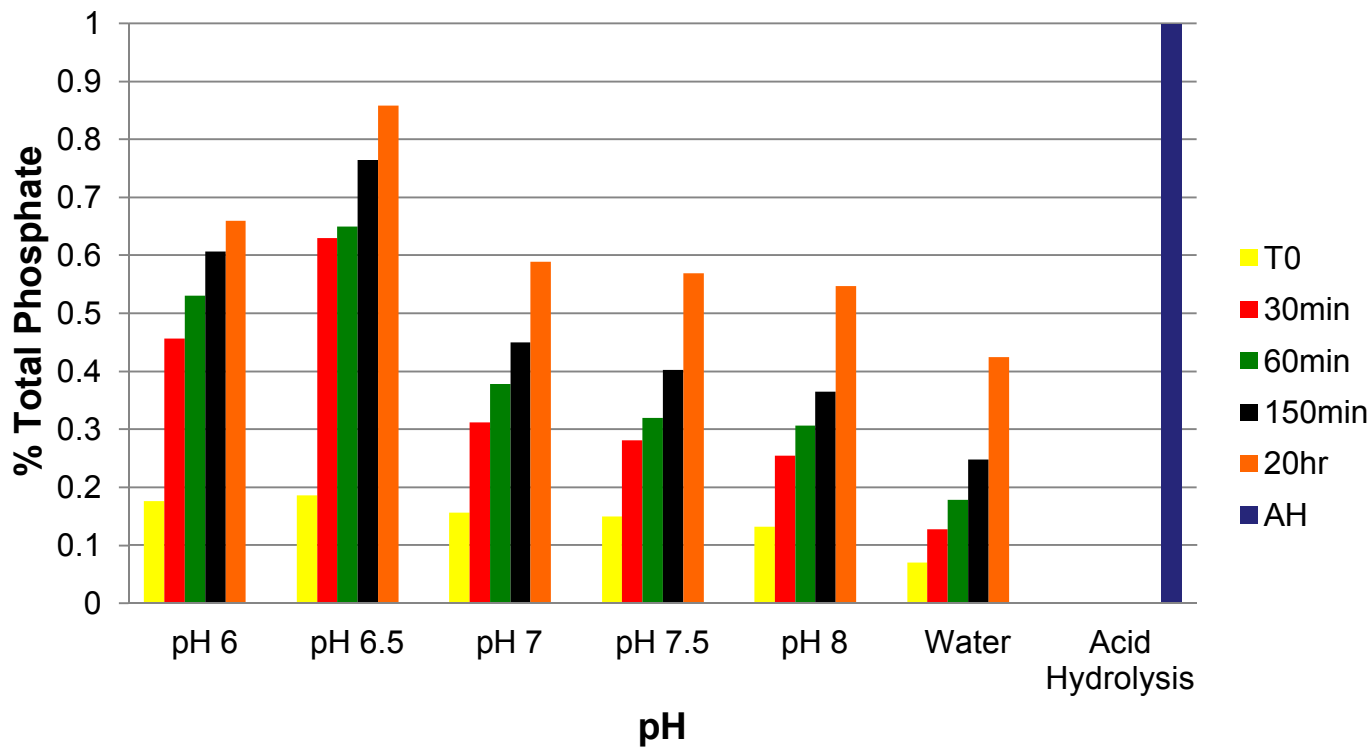


% PO₄ released





Effect of pH on Phosphate Release



pH/EDTA effect on phosphate released

