

# Applications of Super Resolution Microscopy: Imaging Nanoscale Particles and Phenomena

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*Presented at:  
SNL/UNM Symposium on Nanoparticle Human Interactions, June 2-3, 2011*

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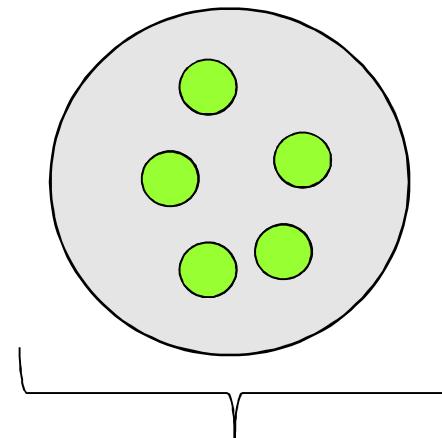


# Outline

- Introduction to Super-resolution Microscopy
  - Methods Overview
  - Advantages & Challenges
- Applications:
  - Receptor reorganization during early immune response
  - Cellulase enzyme dynamics
- On the horizon

# Background

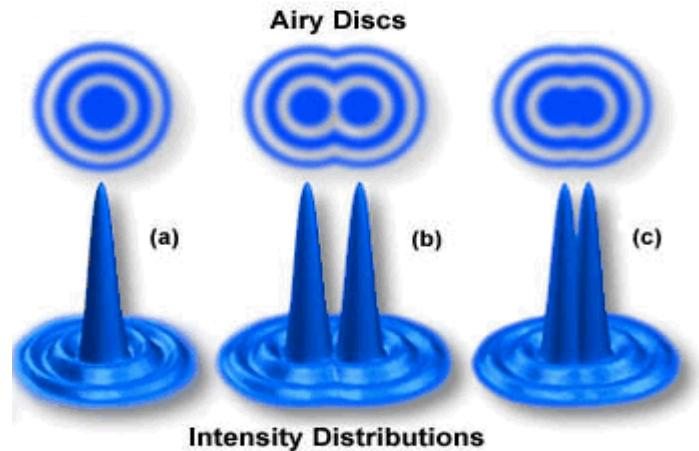
- Optical microscopy is powerful tool for live cell interrogation
  - Interactions
  - Dynamics
  - Kinetics
- Fundamental biological processes occur on a spatial scale below that of optical microscopy



~300nm diffraction-limited spot size

# Breaking the Diffraction Barrier

1873: *Abbe's discovery* Objects in a microscope, closer than  $\sim 1/3$  wavelength of light can't be distinguished



Light doesn't focus to a point, but instead forms an Airy disk and this limits resolution

$$D = 1.22 \lambda N$$

1994: *Enter Stefan Hell* with landmark paper, pioneered a “practical” method to reduce the effective point-spread-function of the microscope to smaller than the dimensions set by diffraction

*Current:* Probe and illumination-based methods for imaging at resolutions below the diffraction of light *in the far field*

=> Super-resolution (SR) microscopy



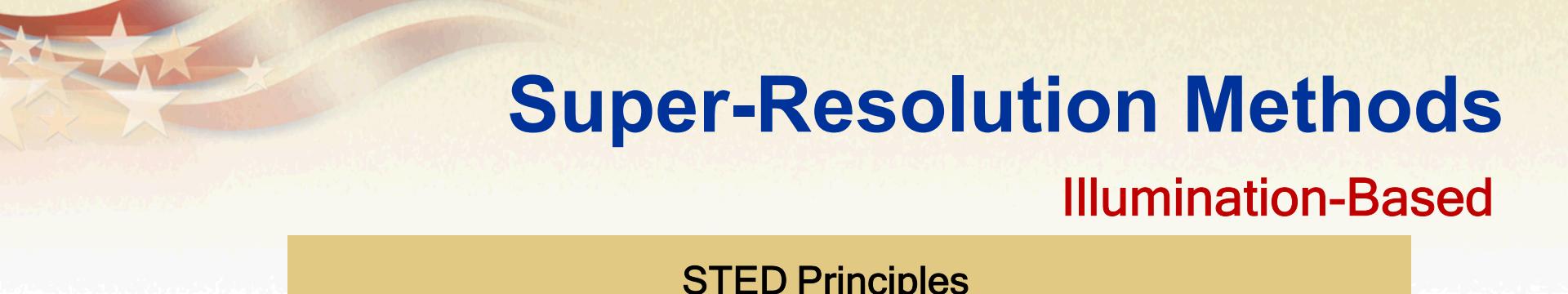
# Super-Resolution Methods

## Probe-Based

- Like pointillism
  - Stochastically activate & deactivate single molecules
  - Fit each measurement to a Gaussian (high precision)
  - Reconstruct image molecule-by-molecule
- Many different (yet highly similar) subtypes (STORM, PALM, PALMIRA, fPALM, dSTORM)
- Basic premise for all of these methods: photoswitching or photoactivating a probe molecule

## Illumination-Based

- Alter the properties of the illumination light to either
  - Permit excitation of signal from sub-diffraction area
  - Permit reconstruct with sub-diffraction resolution
- Stimulation-Emission-Depletion (STED), Ground State Depletion (GSD) and Structured Illumination microscopy (SIM)



# Super-Resolution Methods

## Illumination-Based

### STED Principles

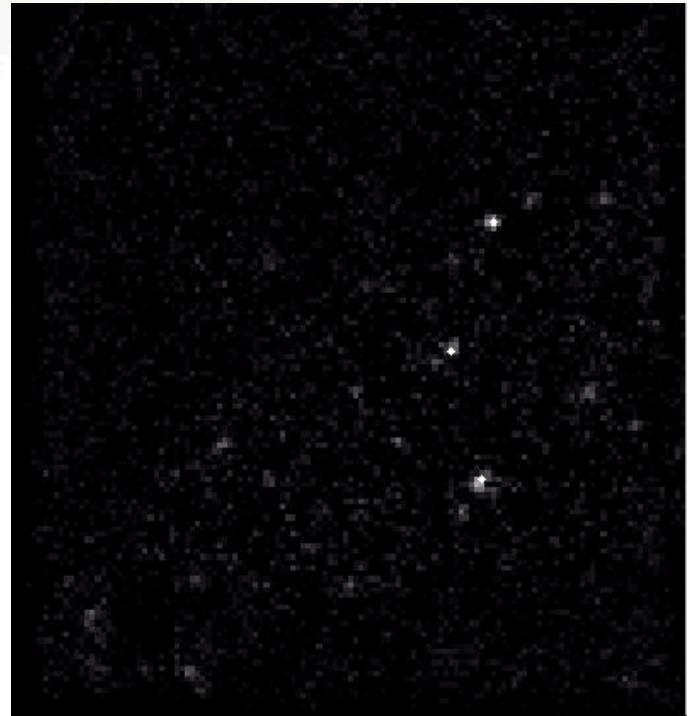
- Live cell compatible, speeds on the order of confocal
- High excitation powers
- Difficult alignment, can be expensive



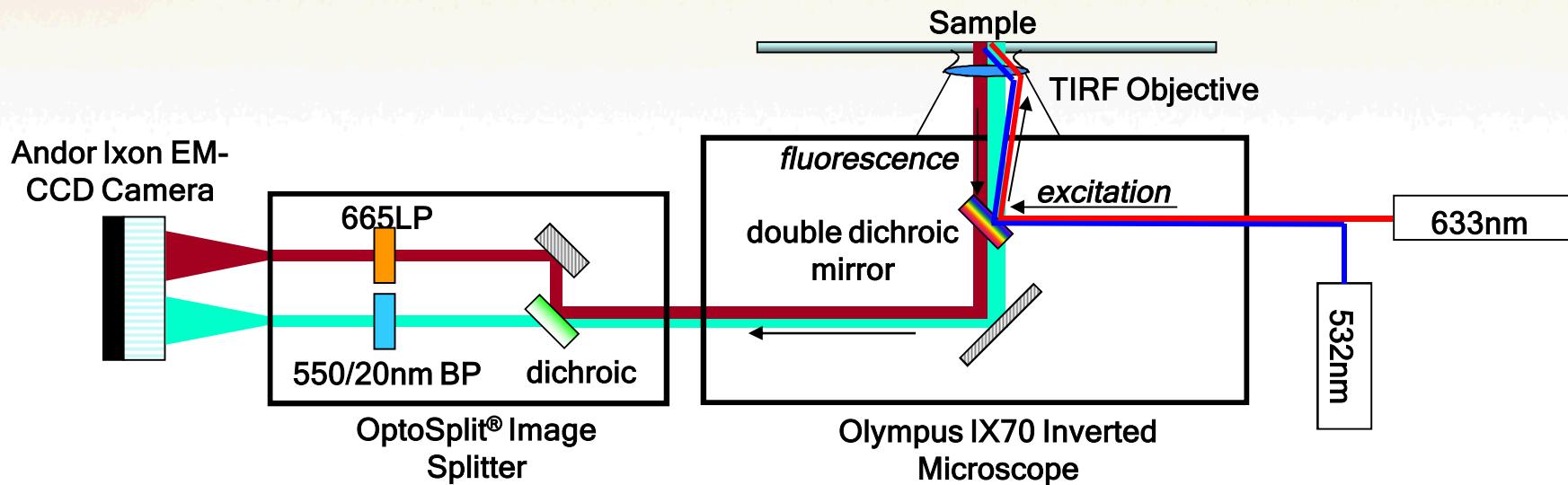
# Super-Resolution Methods

## Probe-Based

- Very good for interactions occurring at interfaces
- Compatible with single molecule applications
- Straightforward, relatively inexpensive implementation
- Repetitive imaging / reconstruction can be slow
- Limited compatibility with live cells, cytoplasmic processes
- Success is often dependant on the dye



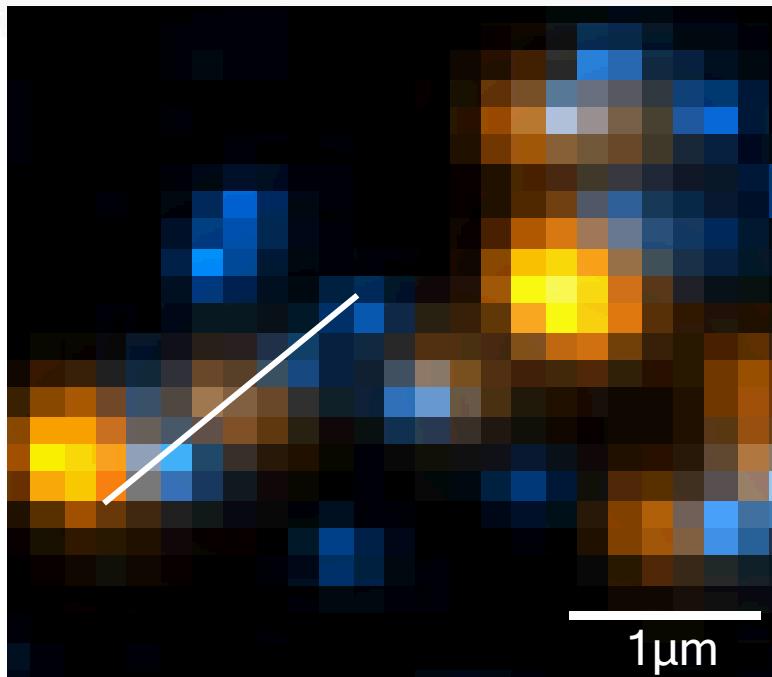
# Imaging Setup



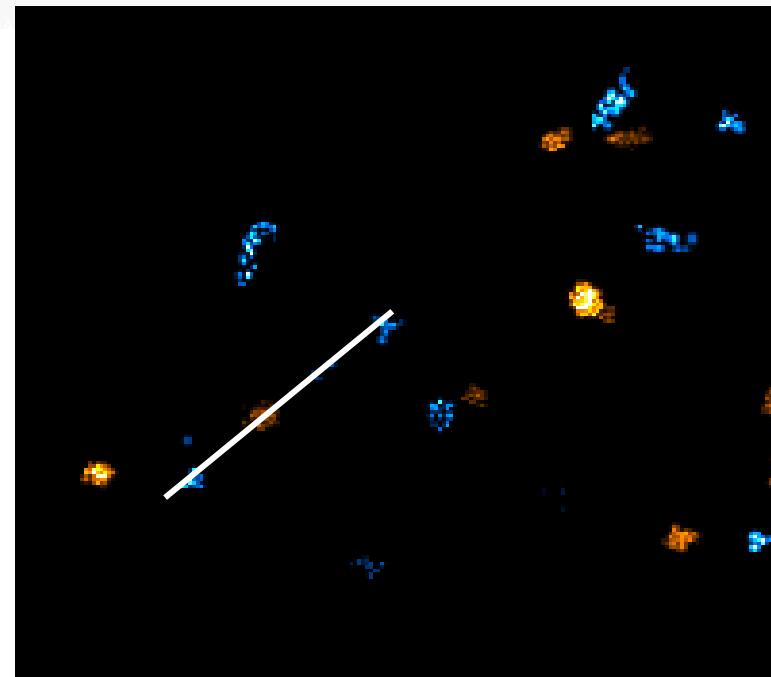
- Olympus IX-71, 60x, NA 1.45 TIRF objective
- Capable of up to four excitation wavelengths (choose among 405, 488, 532, and 633nm), variable angle
- Optosplit® image splitter projects multiple emission wavelengths simultaneously onto EMCCD (Andor iXon)
- Capable of >50fps over 30 $\mu$ m x 30 $\mu$ m FOV

# Multi-Color Validation with Quantum Dots

TIRF

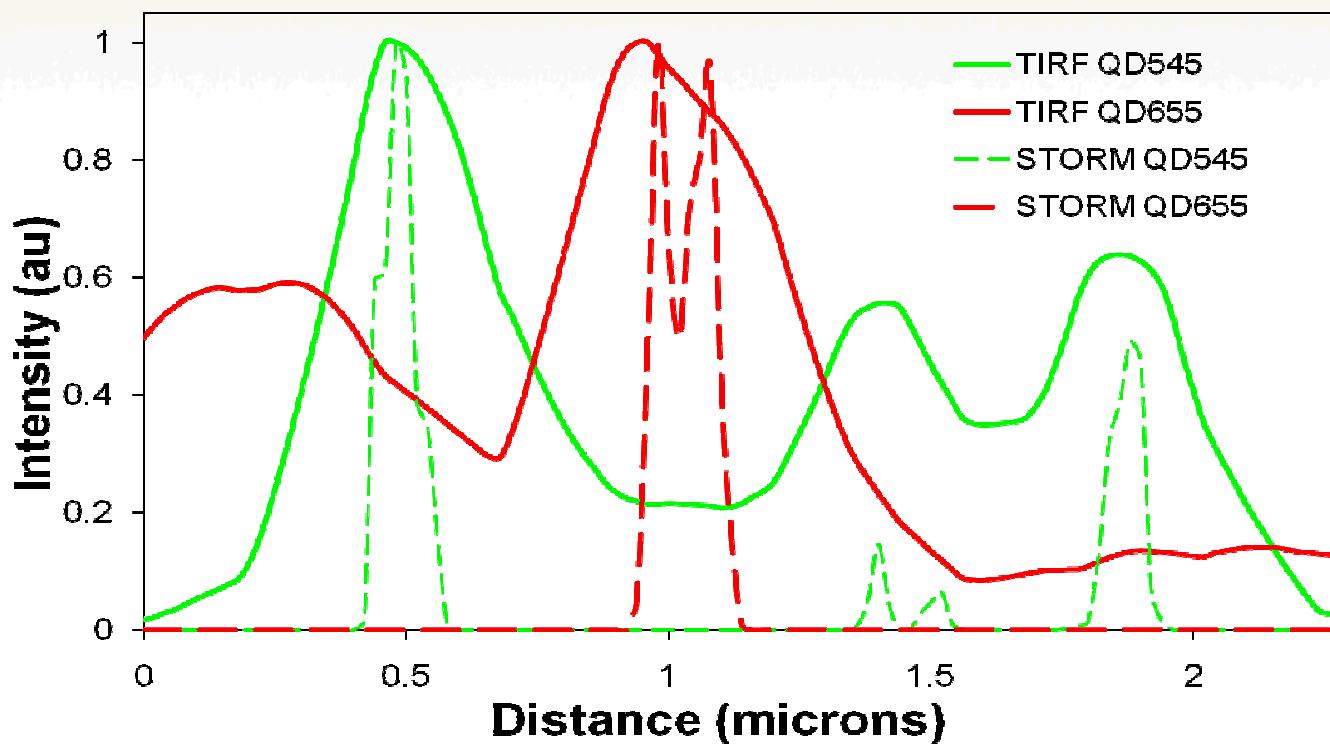


STORM



- QD “blinking” acts as efficient photoswitch
- 2 QD types: 585nm (blue) & 655nm (green) emitting

# Multi-Color Validation with Quantum Dots



- FWHM <40nm (95% confidence) in STORM vs. 400-500nm for conventional imaging



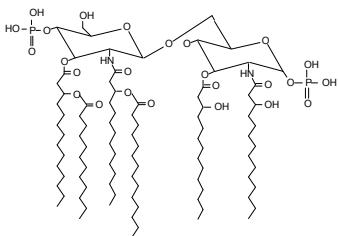
# Imaging Receptor Reorganization During Early Immune Response **APPLICATIONS**

*Jesse Aaron, Bryan Carson, Jerilyn Timlin  
SNL NM: Bioenergy & Defense Technologies*

# TLR4 Receptor Clustering

## *Escherichia coli* (control)

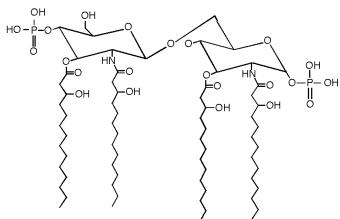
Smooth  
O-polysaccharide



Bind Surface  
+  
↑Stimulatory

## *Yersinia pestis* (37°)

Rough  
O-polysaccharide



Bind Surface  
+  
↓Stimulatory

- Bulk assays have suggested that TLR4 molecules aggregate in lipids rafts within the cell membrane after LPS binding

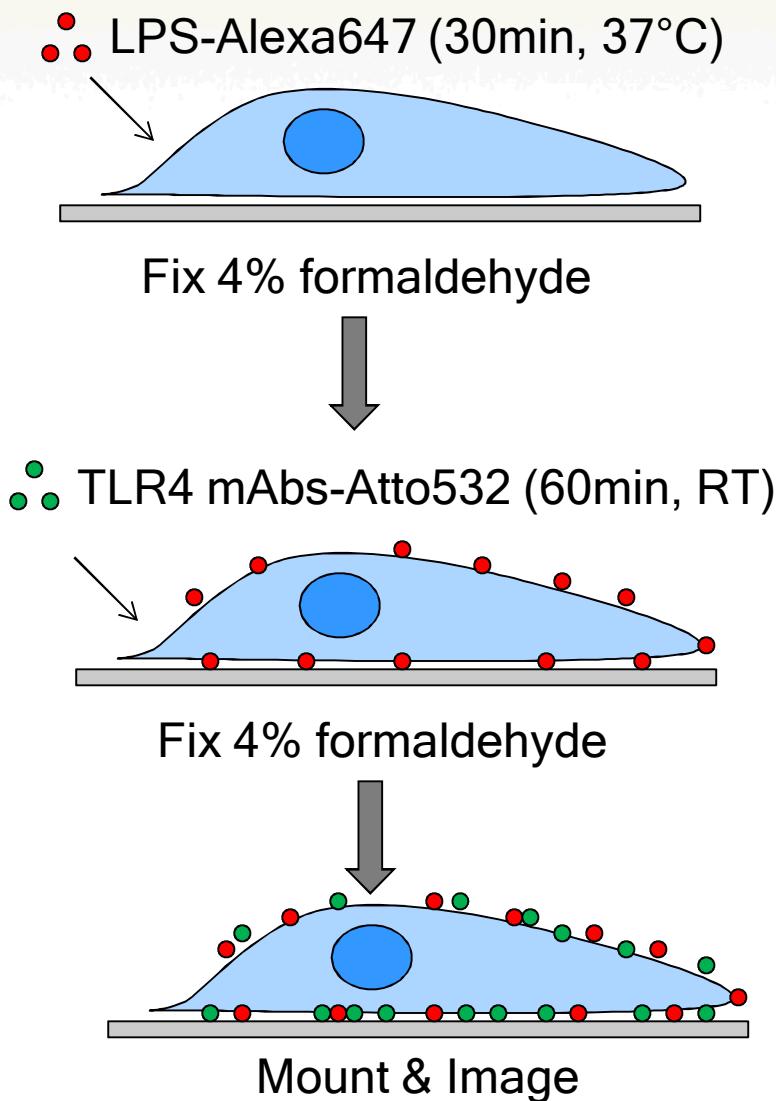
(Triantafilou, et. al, *Biochem. J.* 381(Pt 2): 527-536)

- Differential immune response observed with chemotypes of LPS is not fully understood.

- LPS from *E. coli* binds & produces an immune response
- LPS from *Y. pestis* (plague @ 37 °) binds, but does not

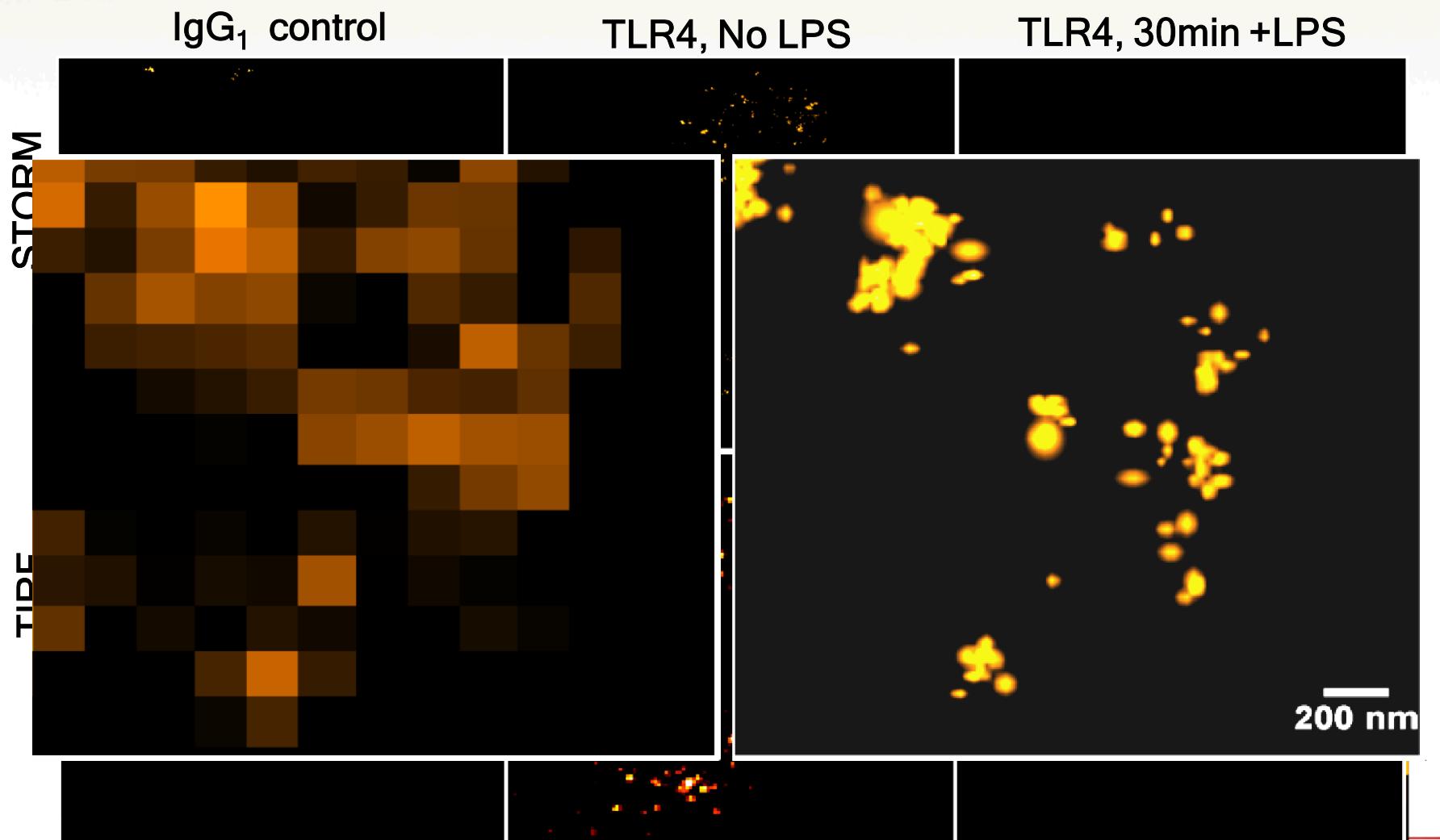
Are there clues in the nano-scale arrangement of the early immune response at the membrane interface?

# Experimental Details

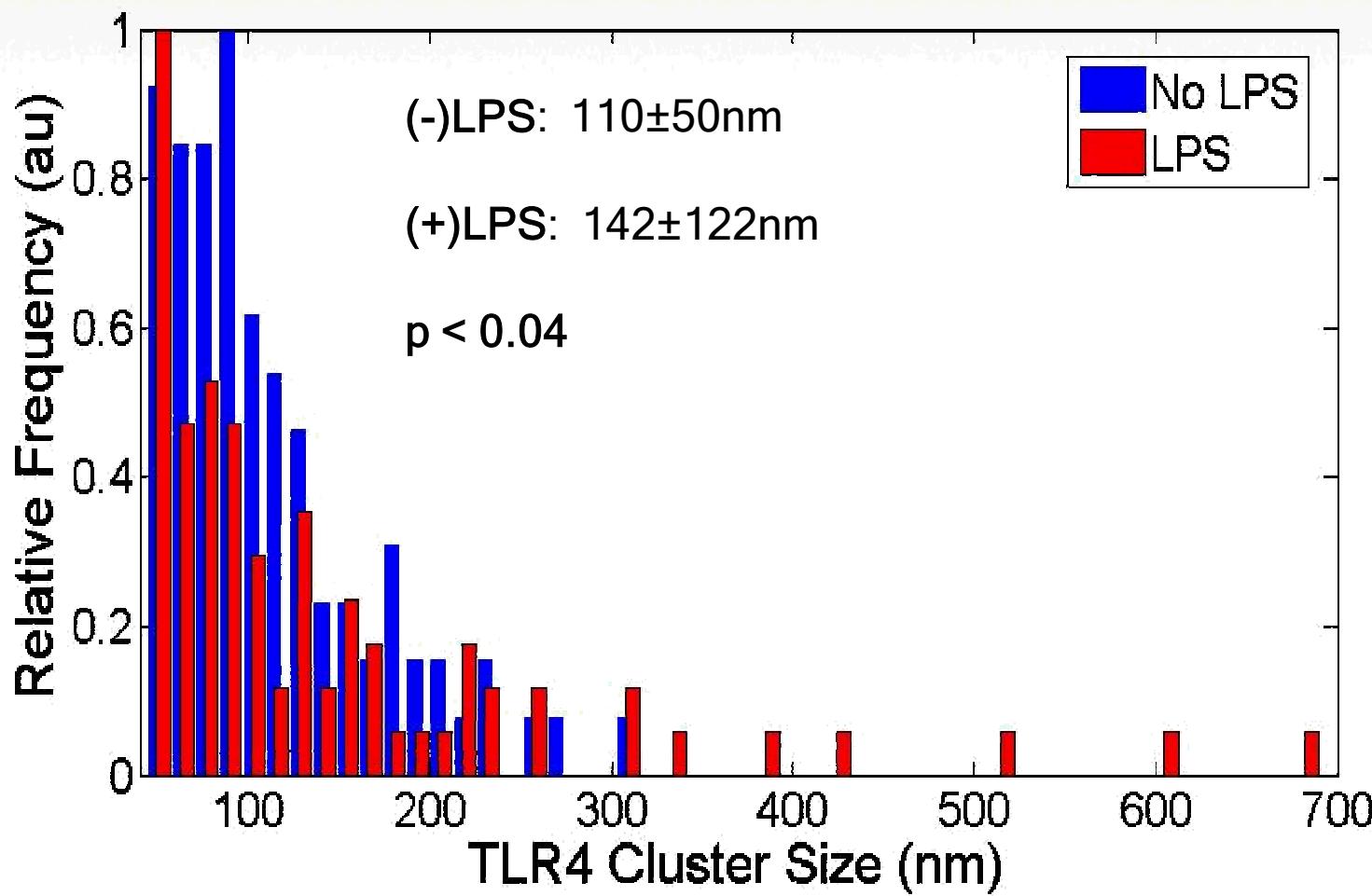


- Mouse macrophage cells (P388D1) incubated with 100nM *E. coli* or *Y. pestis*-derived LPS for 30 min at 37°C and formaldehyde fixed. LPS are labeled with Alexa Fluor 647-hydrazide via linkage with core-polysaccharide
- TLR4 receptors visualized via  $1^{\circ}$  antibodies labeled with Atto532
- Cells imaged in  $O_2$ -scavenging buffer containing  $\beta$ -mercaptothiol using dSTORM

# TLR4 Receptor Localization

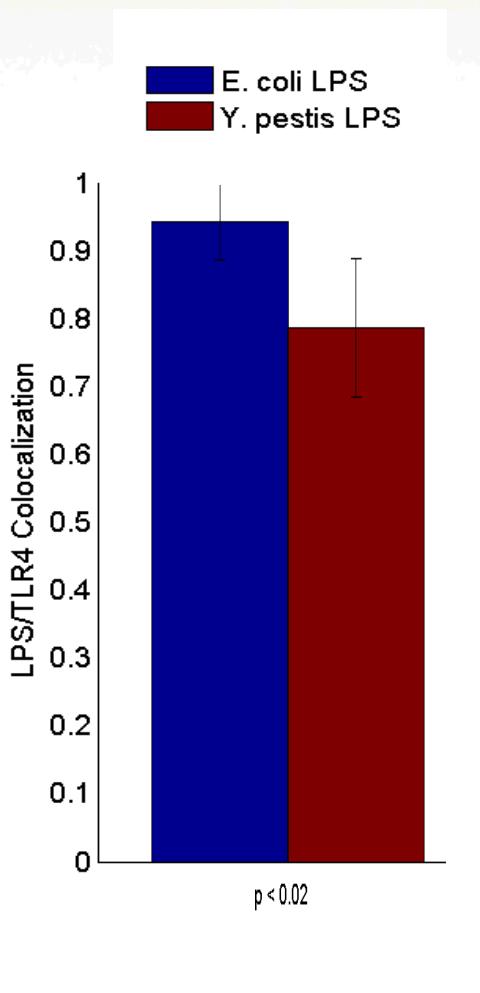
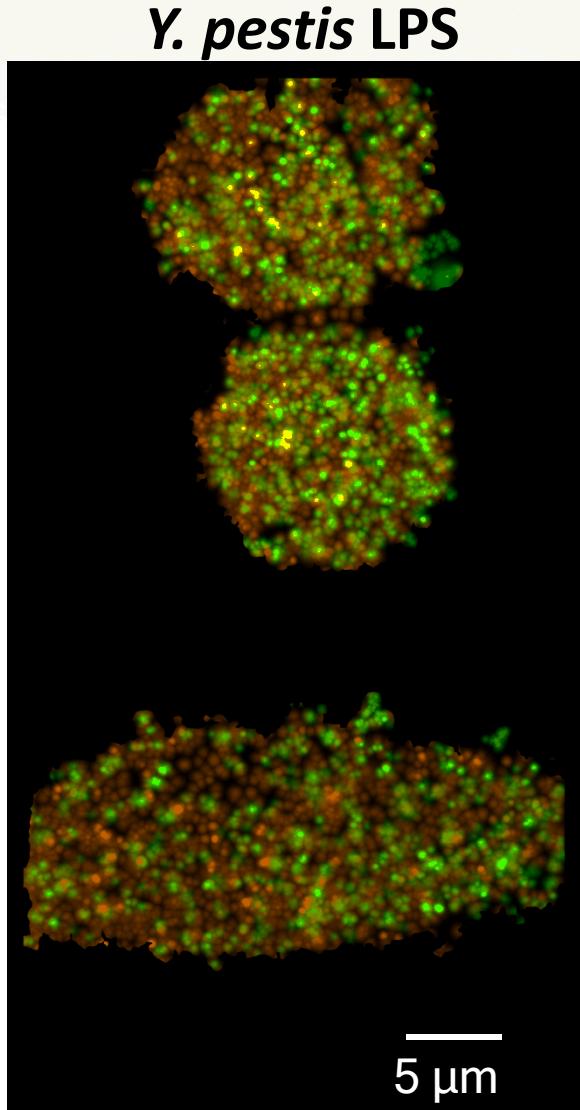
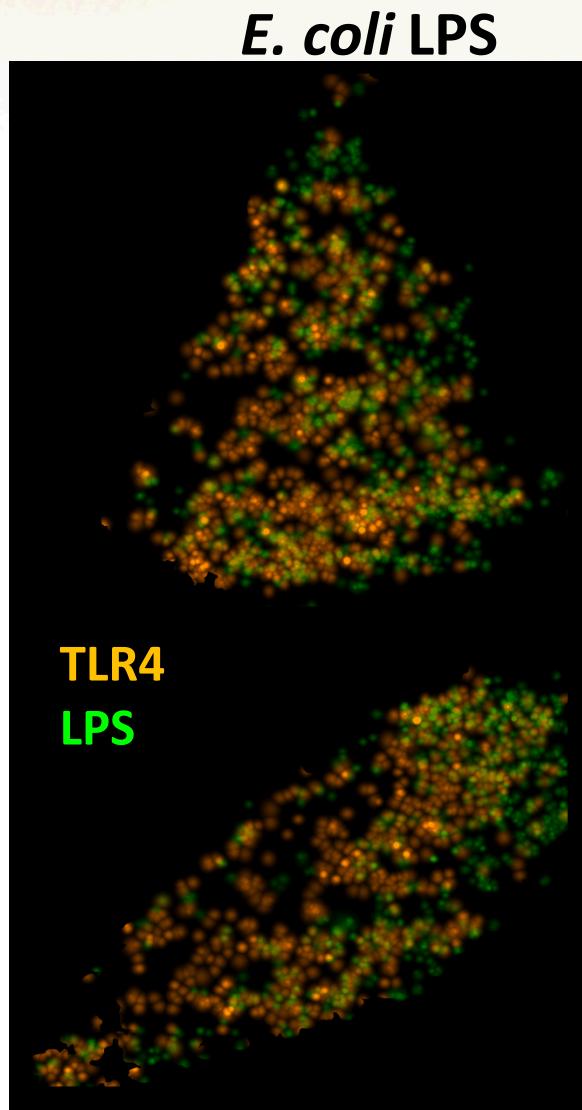


# TLR4 Cluster Size



\*\* Differences not apparent in TIRF ( $p = 0.7$ )

# Simultaneous Visualization of TLR and LPS

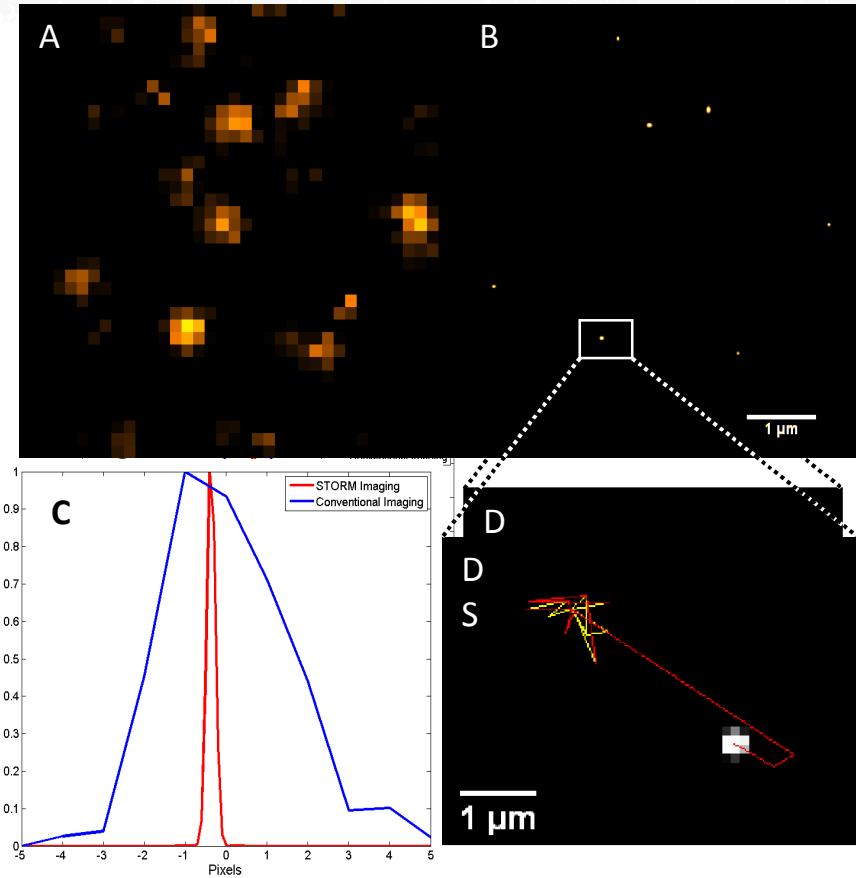




# Cellulase Enzyme Kinetics APPLICATIONS

*Jesse Aaron, Michael Kent, Jerilyn Timlin  
SNL NM: Bioenergy & Defense Technologies  
JBEI: Deconstruction Team*

# Real-time protein displacements on the nanometer scale



- Single cellulase enzymes on amorphous cellulose substrate
- Yellow and red trajectories correspond to movements over 100 ms and 200 ms, respectively
- Broad implications in many fields



# Challenges of the Future

- Probe-based methods:
  - Temporal resolution, imaging speed
  - Live cell compatibility
  - Algorithms for image reconstruction
- Illumination-based methods:
  - Cheaper, more flexible, easier to use
- Extension to multicolor
- Probes, probes, and more probes!
  - Brighter
  - More photostable
  - Easier to get in
  - Larger variety

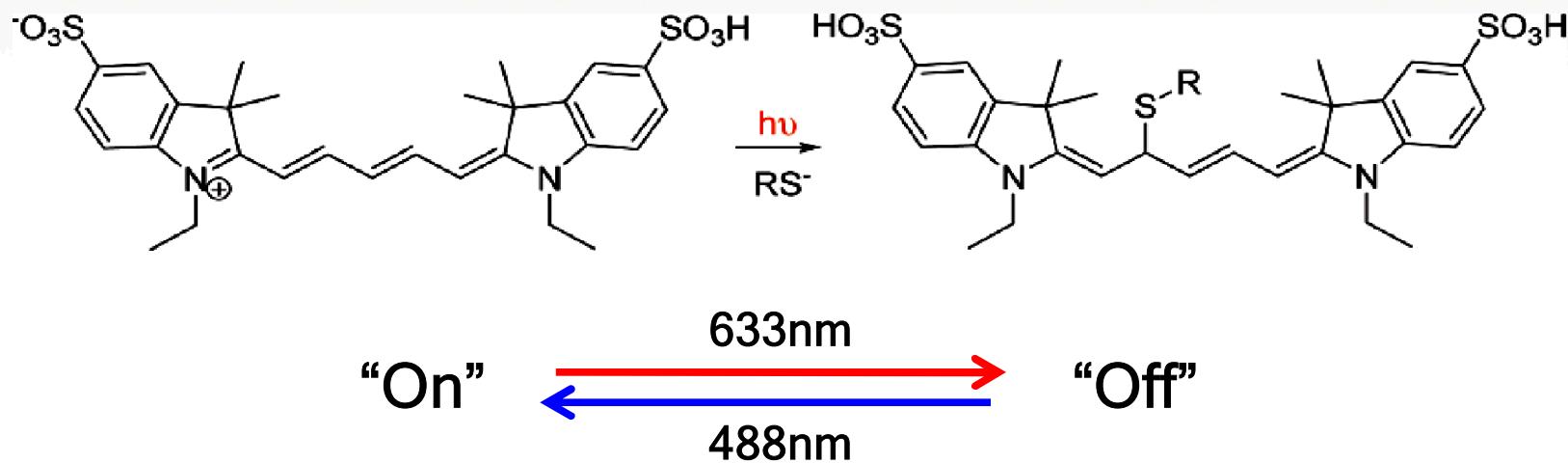
# Acknowledgements

This work was supported in part by:

- Laboratory Directed Research and Development program at Sandia National Laboratories.
- The National Institutes of Health NIH Director's New Innovator Award Program, 1-DP2-OD006673-01



# “Direct” Photoswitching (d-STORM)



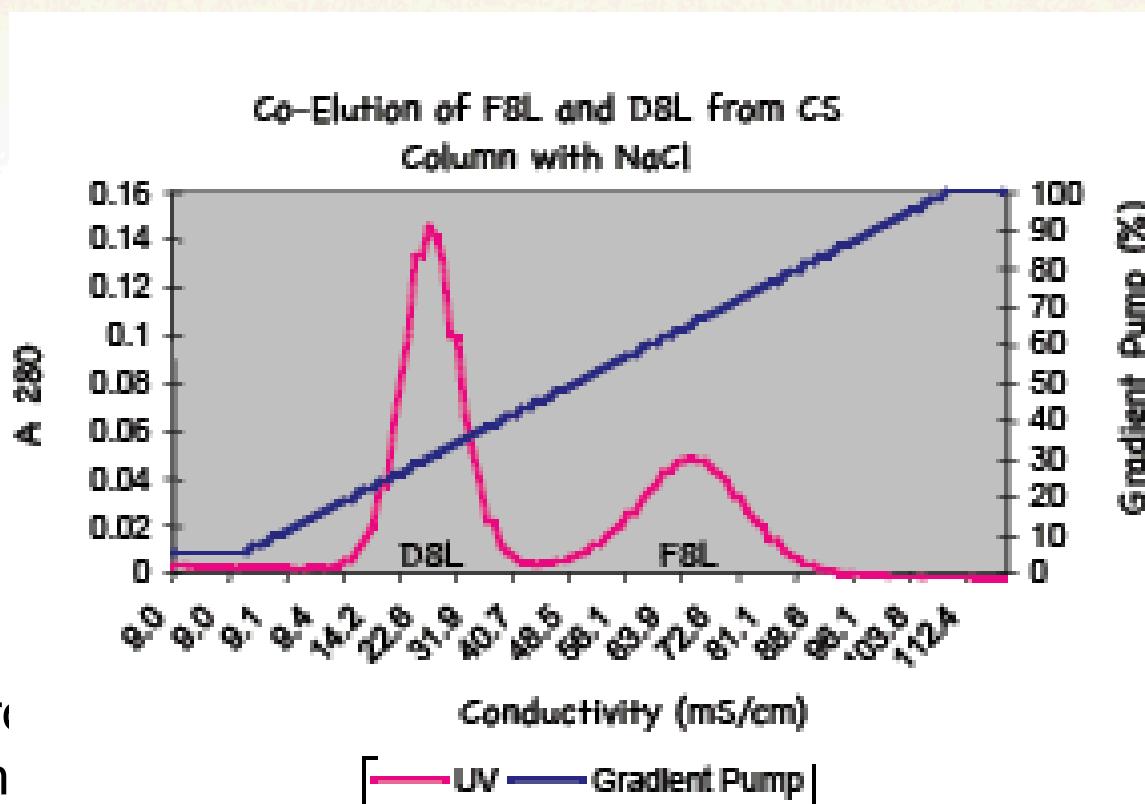
- Cyanine-based dyes have been shown to switch between “on” and “off” states in the presence of small thiol-containing molecules and an oxygen scavenging system
- Process is photon-dependent, and the rate can be adjusted via relative intensities of a probe (633nm) and activation (UV-488nm) beam



# Characterization of Orthopoxvirus Protein Affinity to Chondroitin Sulfate APPLICATIONS

*Jesse Aaron, Jerilyn Timlin, Masood Hadi  
SNL NM: Bioenergy & Defense Technologies  
SNL CA: Biomass Science & Conversion*

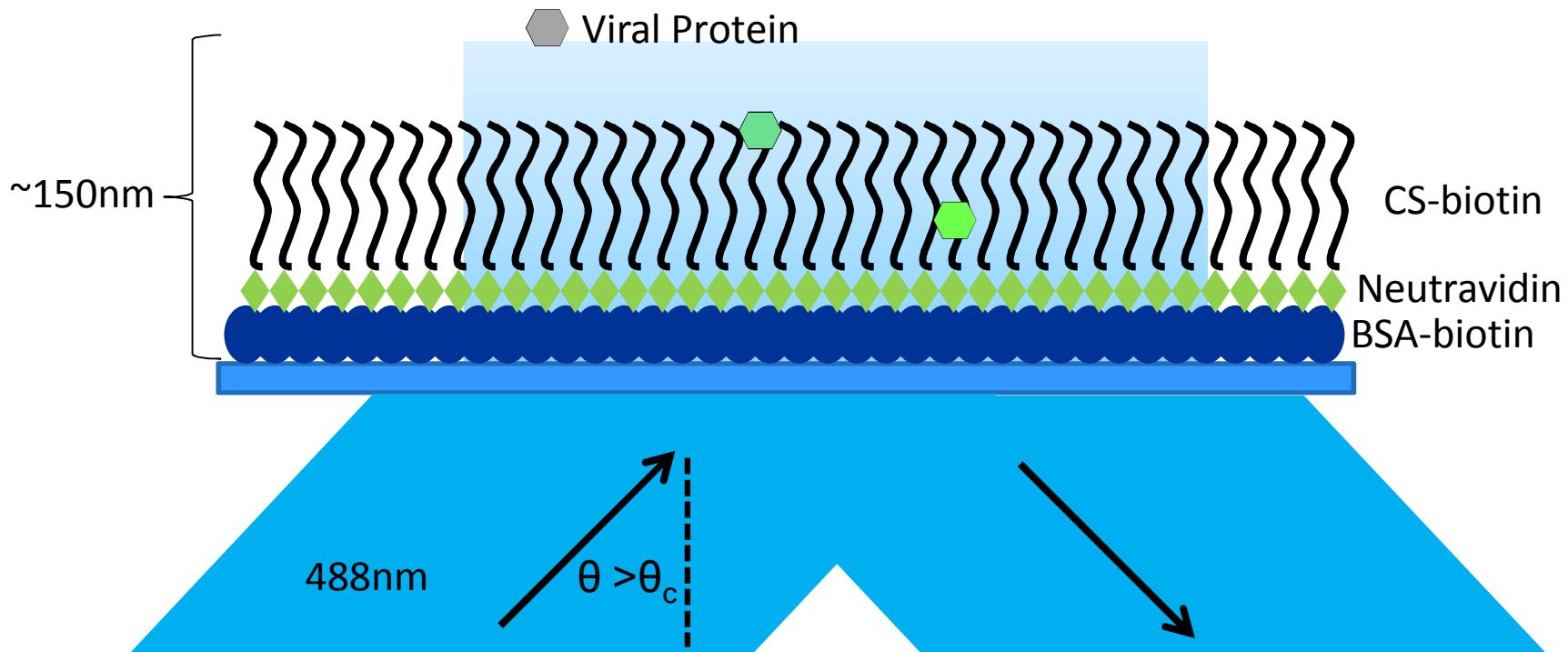
# F8L-Like Proteins



- F8L-Like proteins common among
- Very close to D8L protein
- D8L shown to bind chondroitin sulfate on host cell surface
- D8L(-) mutants <10% infectivity of wild-type
- F8L shows higher affinity to CS than D8L

# TIRF imaging of viral protein binding to CS

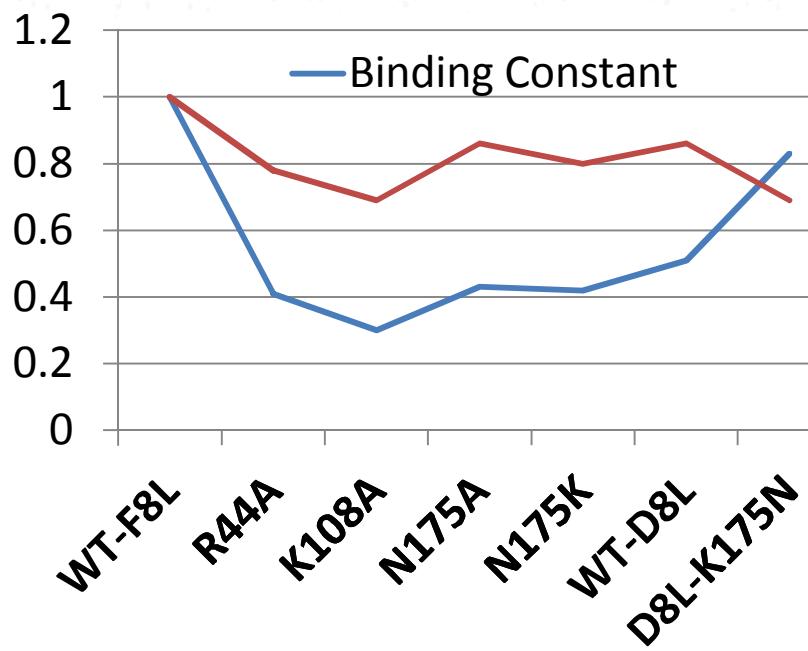
- Affinity column, SPR measurements give bulk/average measurements
- Total internal reflectance fluorescence microscopy =*single molecule* behavior on functionalized surface



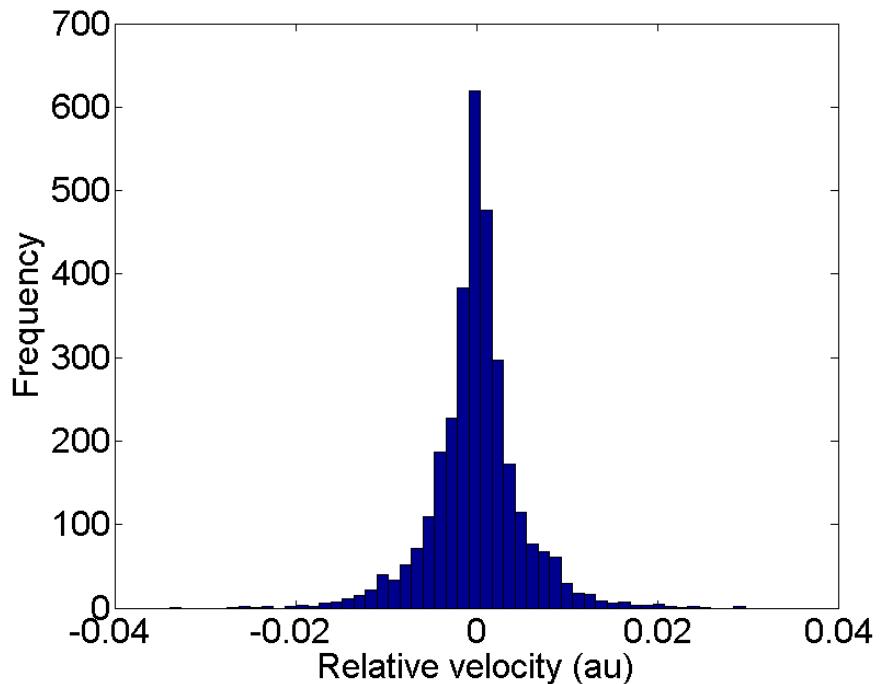
- Evanescent field decays exponentially, i.e.  $I \sim \exp(-z/d)$ , Log(Signal) proportional to axial (z) position
- Proteins not bound to CS are not visible, proteins appear brighter nearest glass substrate

# Results

## Comparison of Binding Kinetics



## Viral protein movement along CS molecules



Consistent with SPR/bulk measurements

Movement does not appear to be directed/unidirectional, Ave velocities centered around zero