

Meeting User Needs: Developing Field-Deployable Biodetection Systems Using a Micro-Separations Approach

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Developing fieldable instruments has challenges in addition to meeting laboratory requirements

Laboratory requirements:

- High sensitivity
- High specificity
- User-friendly or automated
- Robust



Field-portable requirements:

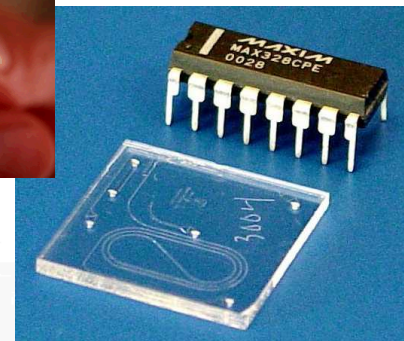
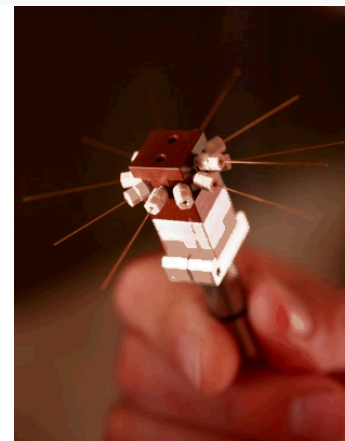
- High sensitivity
- High specificity
- User-friendly or automated
- Robust
- Portable
- Low-power
- Stable reagents
- Low cost



Simply shrinking lab-based equipment for field use is not always feasible, and may lead to a loss in performance and high costs

Sandia “Born Small” approach: Portable systems using miniaturized assays & components

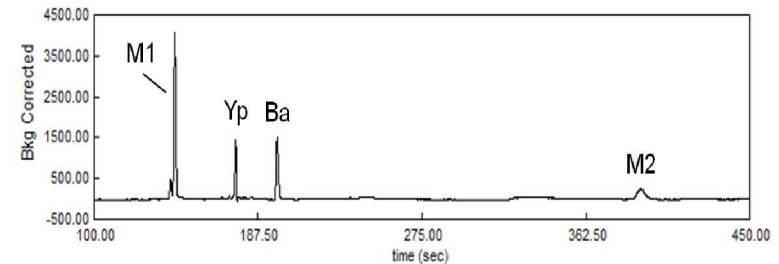
- Sandia began developing systems for microfluidics-based analysis in the late 1990s
 - At that time, the “lab-on-a-chip” concept was just emerging
 - Componentry needed to interface to microfluidic chips and create portable systems **did not exist**
- Out of this effort grew a capability for developing field-portable systems for biodetection and medical diagnostics
 - Programmatic focus arose from our national security mission
- Major program sponsors: Internal Sandia LDRD (start), DOE, DHS, DoD, NIH



Our approach exploits microseparations for detecting biological threat agents

To meet field-portable requirements:

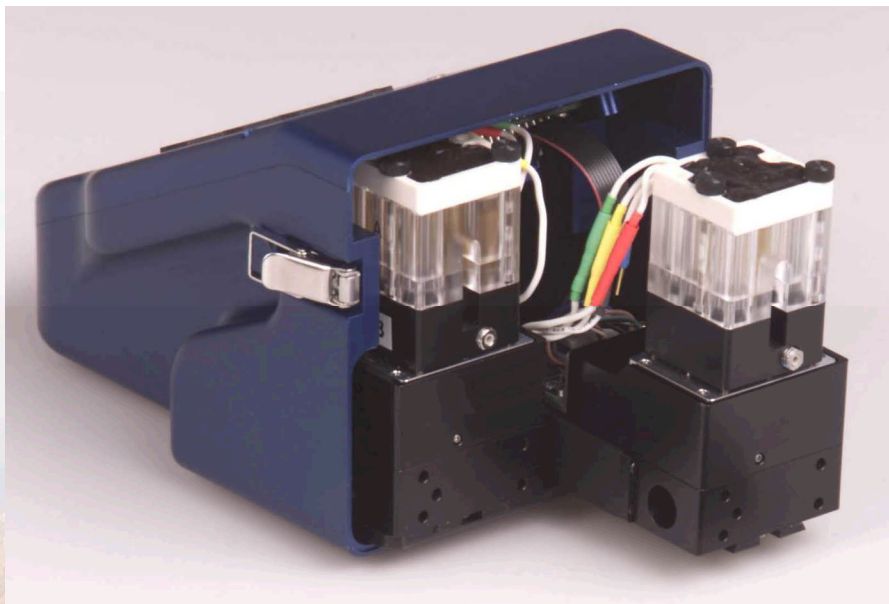
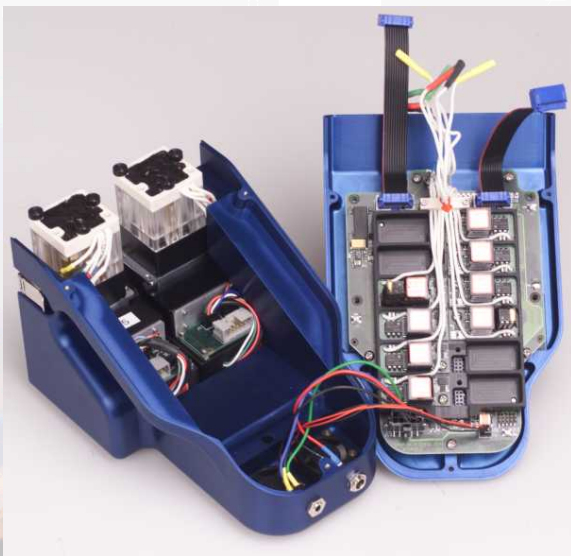
- For high sensitivity, utilize LIF and preconcentration techniques
- For high specificity, utilize orthogonal analyses and develop microseparations-based nucleic acid and antibody assays
- Develop automated sample preparation appropriate for a wide variety of threats
- Incorporate engineering controls to provide reliable system performance
- Utilize components designed to be low power
- Utilize lower cost components (lasers, plastic chips, reagents)
- Buffers and dyes are stable and cheap; very small volumes needed



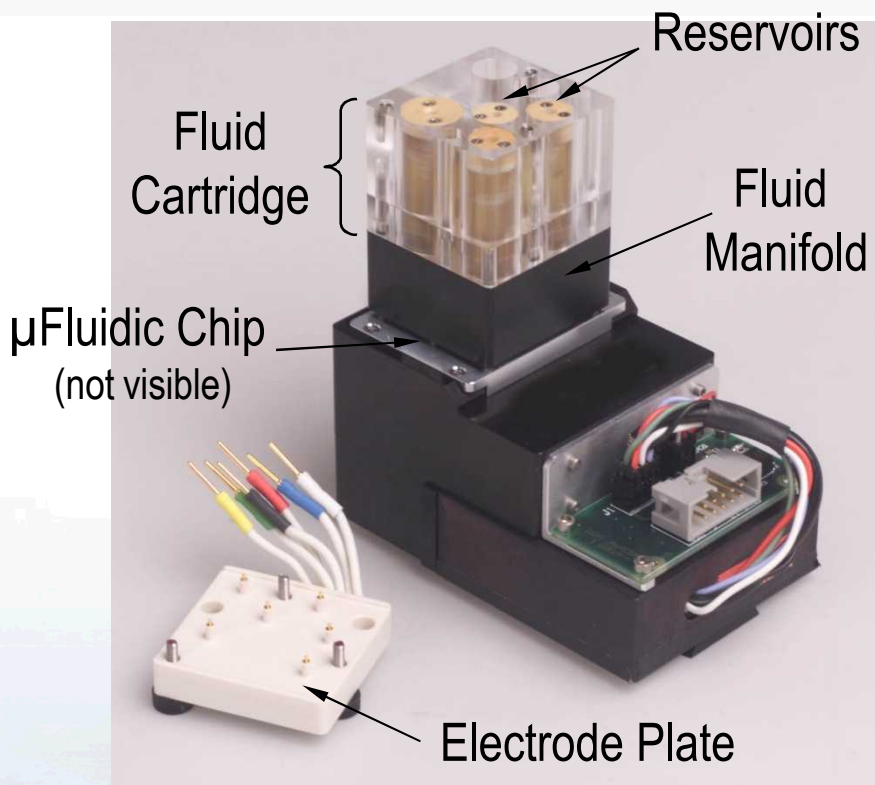
Integrated Biodetection Platforms – μ ChemLab™



- Handheld instrument for first responders
- Modular packaging
- Two analysis modules
- Integrated control and data analysis



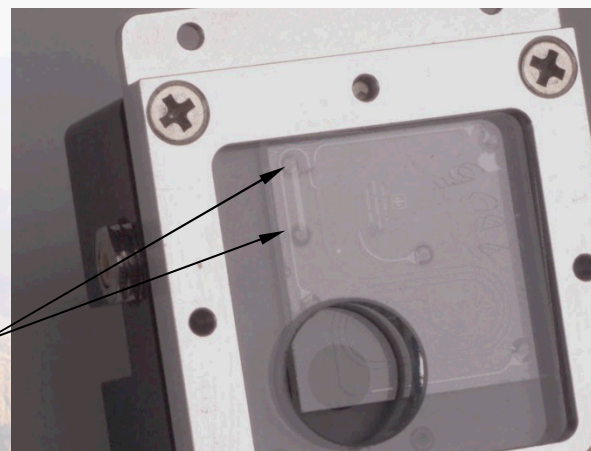
Polymeric fluid manifold provides reproducible leak-tight interface to the microfluidic chip



Separation Module

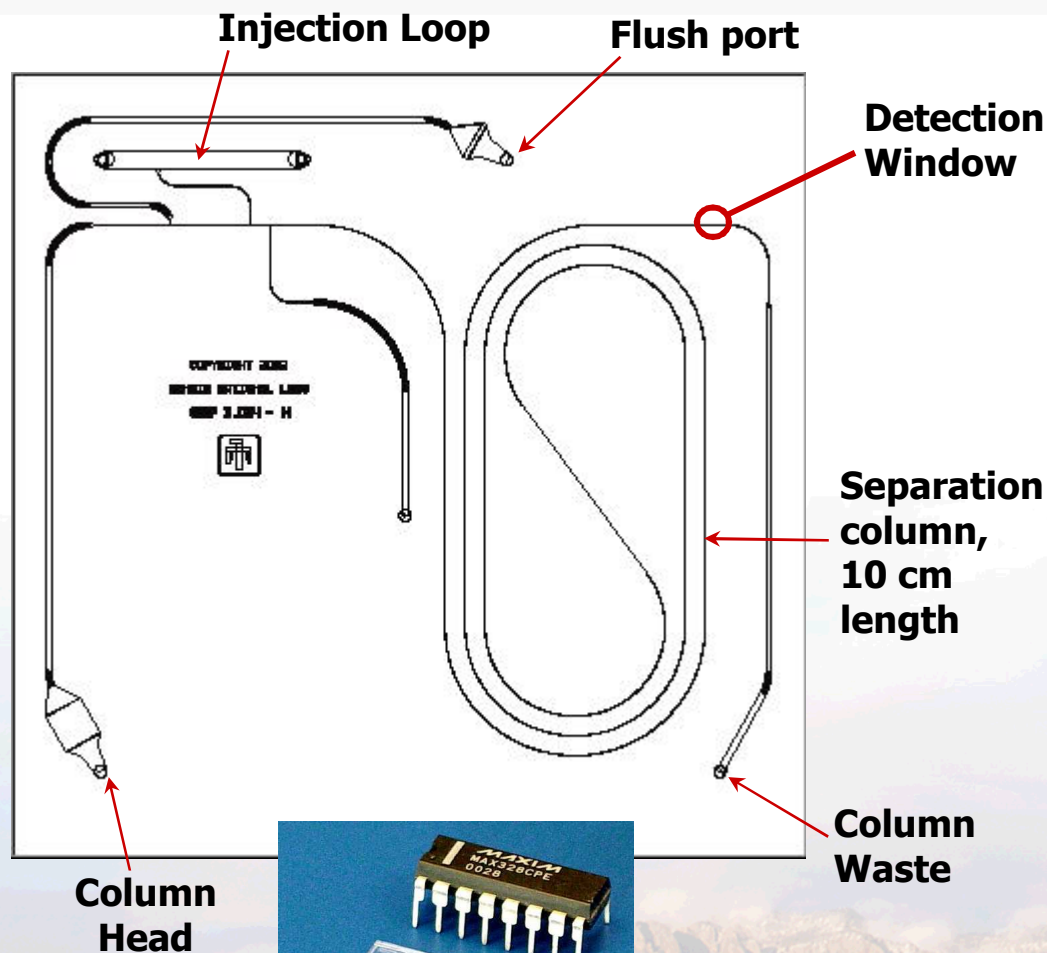
- O-ring face seal enables simple chip installation
- Reservoirs housed in a fluid cartridge hold running solutions
- Capillaries in fluid manifold provide fluid connection between reservoir and chip
- Electrode plate connects high voltage source to solutions

O-ring Face Seals

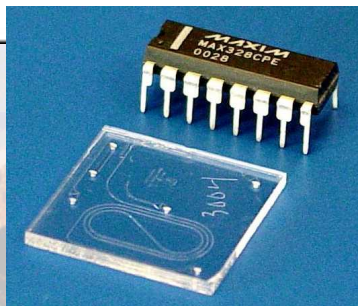
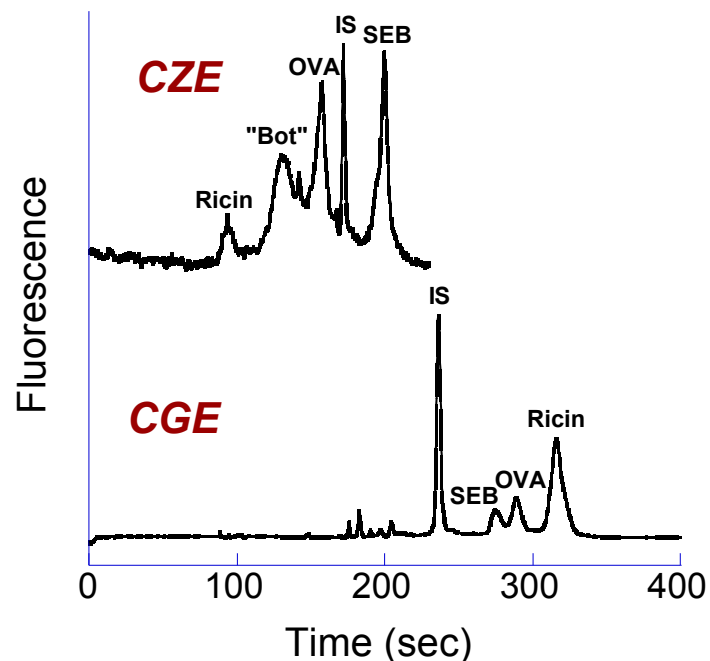


Renzi *et al*, *Anal. Chem.* **2005**, 77, 435-441

Dual CE/LIF platforms enable rapid parallel chip-based analyses at nano-molar sensitivities

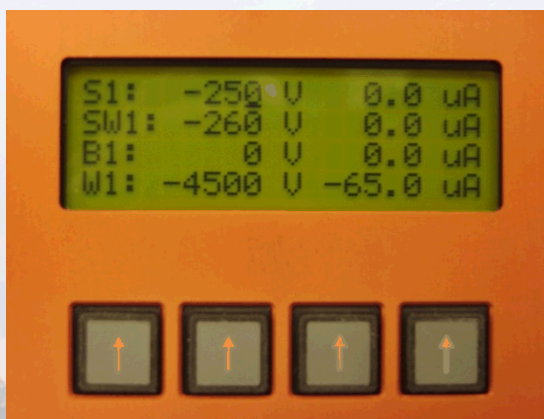
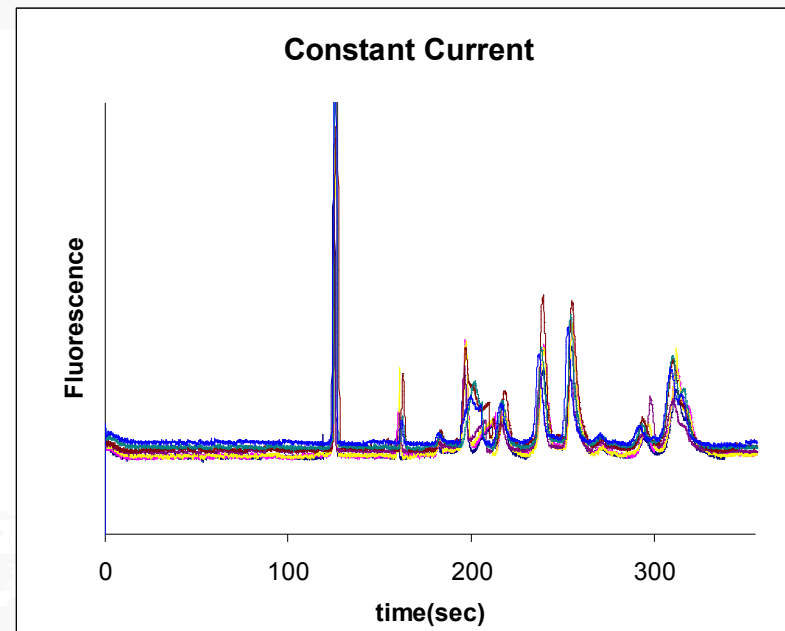
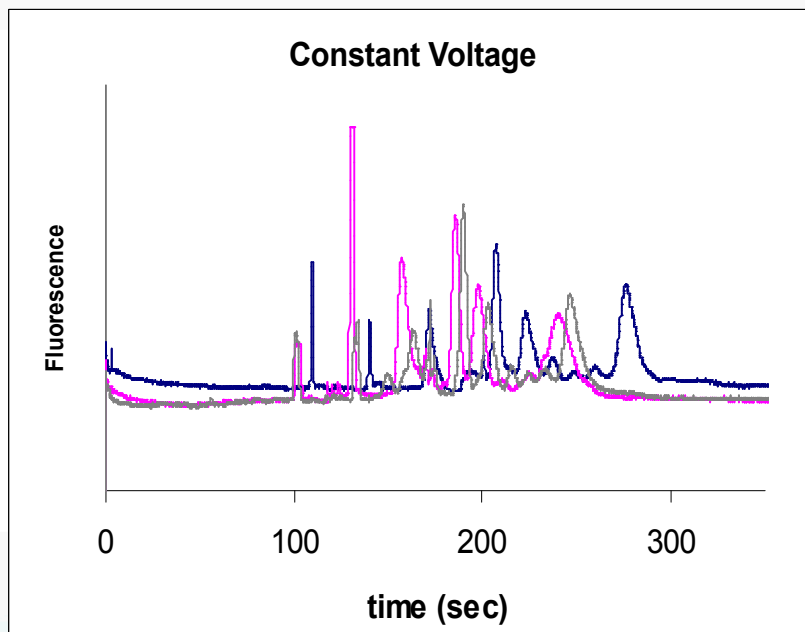


Biotoxin detection uses two separation methods for improved detection reliability



Fruetel et al, *Electrophoresis* **2005**, 26, 1144–1154
Renzi et al, *Anal. Chem.* **2005**, 77, 435–441

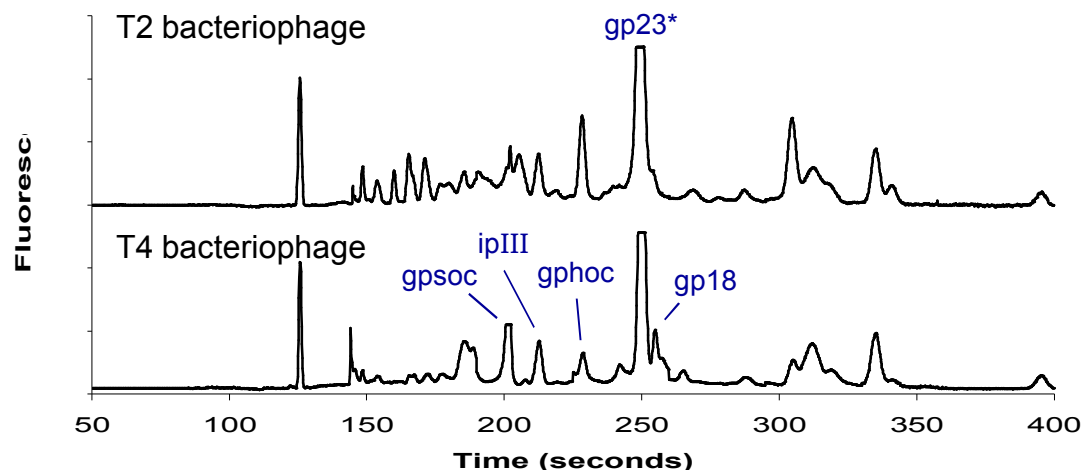
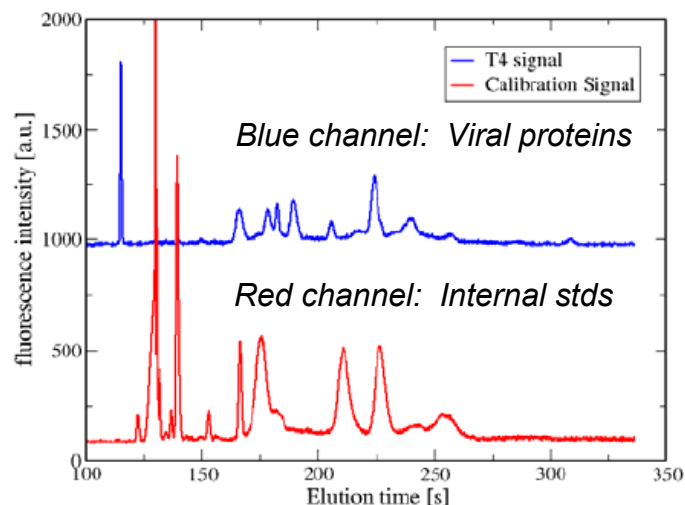
Current monitoring and control improves run-to-run reproducibility and provides system health



run #	cck	lact	CA	OVA	BSA	IgG
1	162.2	199.5	216.7	236	251.7	308.5
2	161.7	199.5	216	236.5	252.6	308.5
3	162.3	201.5	216.6	238.1	254	309.5
4	162.5	196.5	218	238.8	254.5	310
5	160.5	196	216	237.5	253.7	309.5
6	161.2	196.5	217.5	238.8	254.7	311
7	160.5	196.5	217.8	239.2	255	311
8	160	196.6	217.8	239.3	255	312
9	159.5	196	217	239	254.5	311.5
Average	161.1556	197.45	217.04	238.07	254.07	310.1667
S.D.	1.086406	1.967655	0.727553	1.160508	1.119573	1.274755
RSD	0.674	0.997	0.335	0.487	0.441	0.411

Identification of viruses using microfluidic protein profiling and Bayesian classification

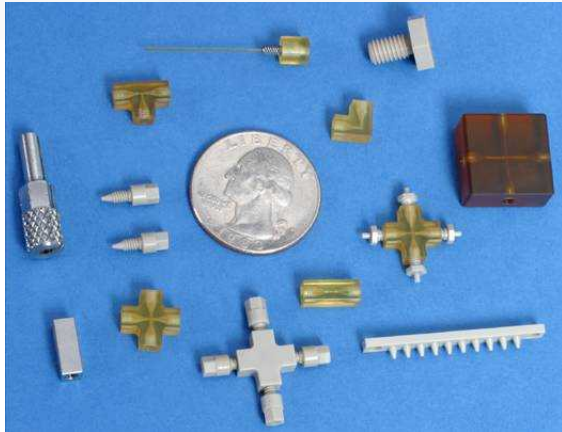
Two-color fluorescence detection enables molecular weight assignments through use of internal standards



Performance of the Bayesian classifier on non-training samples in terms of the number of runs where the Bayes factor >5 (considered decisive evidence of the agent being present)

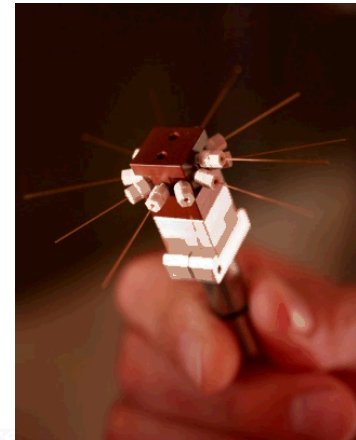
virus	total number of runs	no. of runs used as training data	classification results: no. of runs, not used for training	
			Bayes factor > 5	Bayes factor < 0 ("other")
T2	11	6	4	1
T4	18	9	9	0
vaccinia	29	12	16	1
MS2	41	17	24	0
EBV	13	8	4	1
RSV	14	5	9	0
total	126	57	66	3

Enabling Technologies - Microfluidics

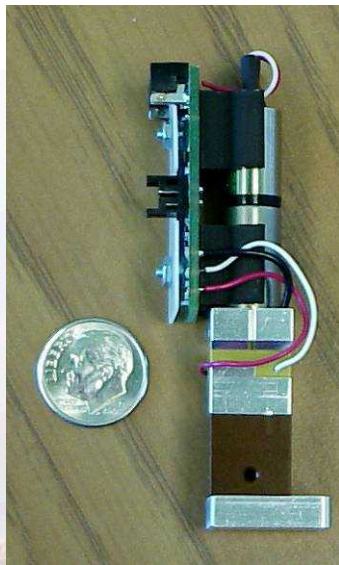


CapTite™ Fluidic Fittings

- One piece design
- Reusable
- Finger or tool tightened
- Up to 10 KSI

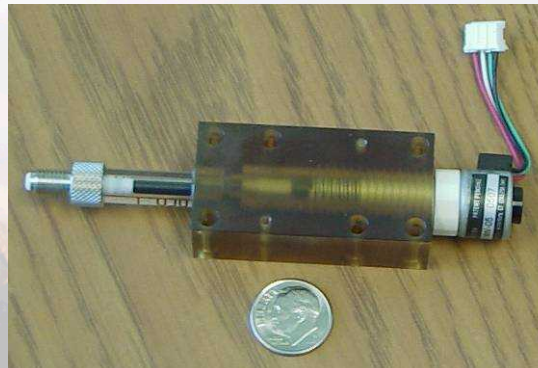


10-port valve



Miniature valves

- 10 KSI capability
- Electrically actuated
- Low swept volumes (~ 10 nL)



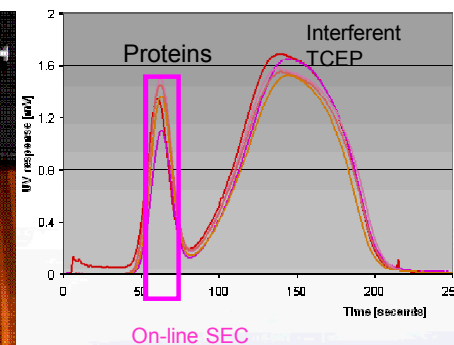
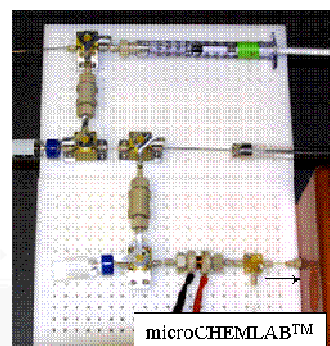
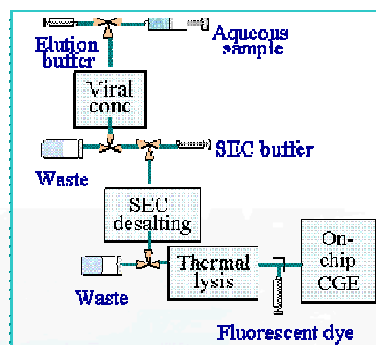
Syringe pumps

- Linear stepper motor
- Controlled fluid metering
- Bidirectional

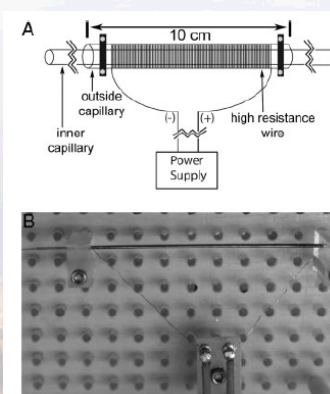
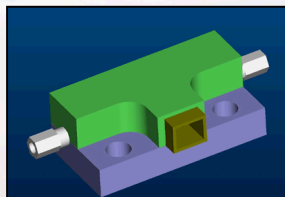
Enabling Technology: Modular microscale sample processing for ease of use and robustness

- **Need:** Bridge the gap between mL-sized samples and nL-sized chip-based analysis in a format that enables portability and unattended operation
- **Developed cartridges** for processing μL sample volumes in seconds to minutes
 - Concentration
 - Fractionation
 - Filtering
 - Sieving
 - Desalting
 - Contaminant removal
 - Digestion
 - Buffer exchange
 - Cell lysis
- **Has been applied to proteins and intact organisms**

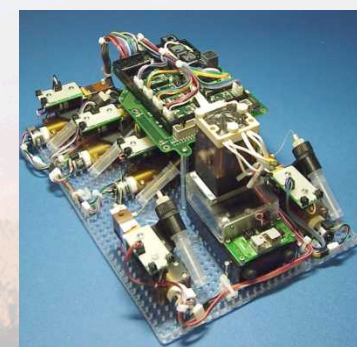
Packed-bed cartridges



Thermo/chemical lysis



Integrated systems



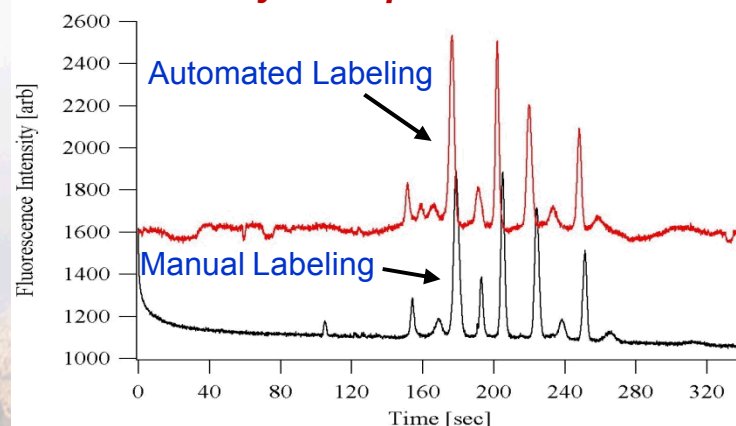
Chirica *et al*, *Anal. Chem.* **2006**, 78, 5362-5368
Hukari *et al*, *Electrophoresis* **2010**, 31, 2804-2812

Integrated Biodetection Platforms – Unattended Water Sensor



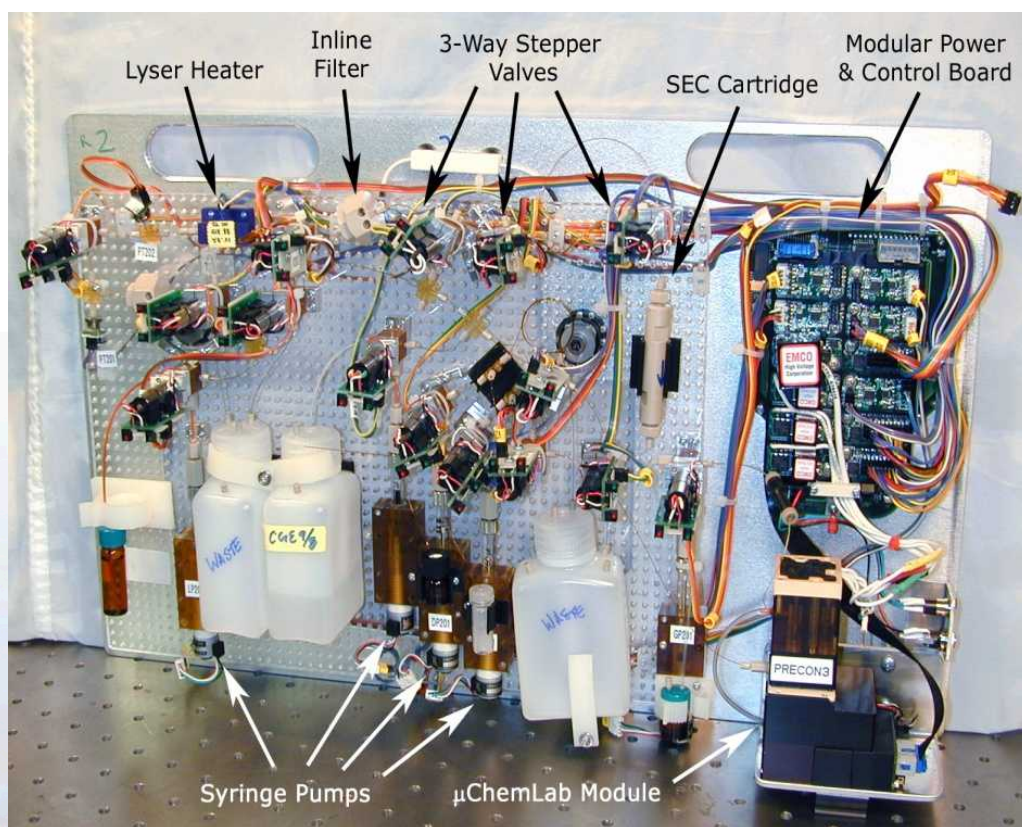
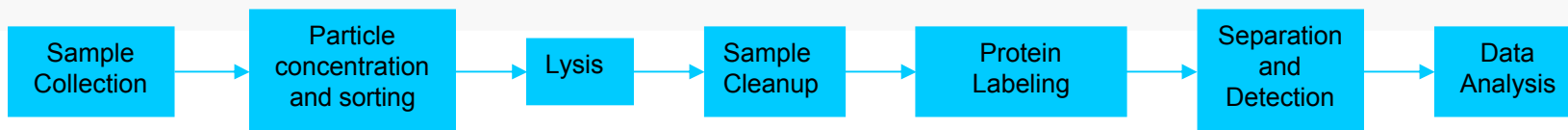
- **Continuous water monitoring for utilities**
 - 30 day unattended operation
 - Analysis every 30 minutes
 - Detects biotoxins
- **Funded through CRADA with Tenix and CH2MHill**
- **Automated: sampling of water main, fluorescent labeling of proteins, and data analysis**

CE analysis of protein standards



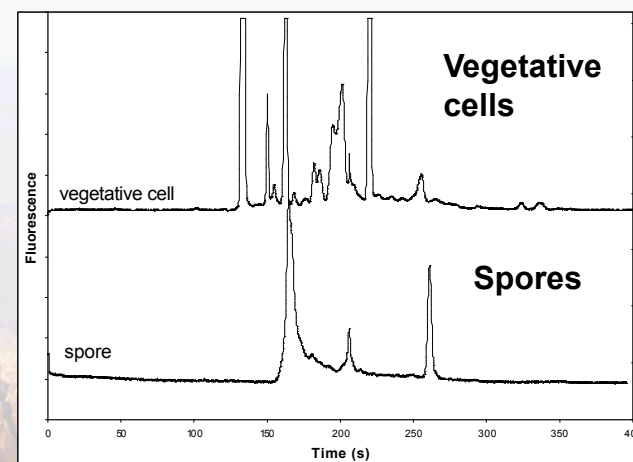
VanderNoot *et al*, *Electrophoresis* **2010**, 31, 2632–2640

Integrated Biodetection Platforms – Automated Microfluidic Protein Profiling System for bacteria



Stachowiak *et al*, *Anal. Chem.* **2007**, 79, 5763-5770

- **Aerosol point detection**
- **Prototype built and tested**
 - Detects toxins, viruses, spores, and vegetative cells
 - 8 hour autonomous operation
 - Rapid response (minutes)
 - Minimal reagents



Integrated Biodetection Platforms – BioBriefcase

■ **Funded by DOE and DHS**

- Developed in collaboration with Lawrence Livermore National Laboratories

■ **Key system requirements:**

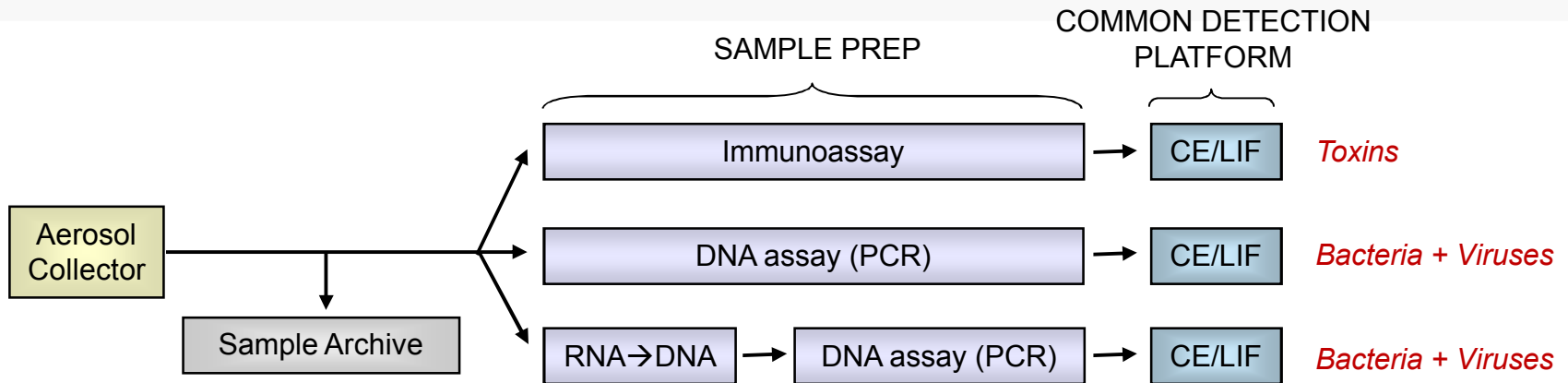
- Continuous, fully autonomous operation in full range of outdoor environments
- Broad agent coverage, >20 agents
- High sensitivity
- Very low false alarm rate

■ **Implemented fluorescent reporter-based assays for PCR and immunoassay detection of bacteria, viruses and toxins**

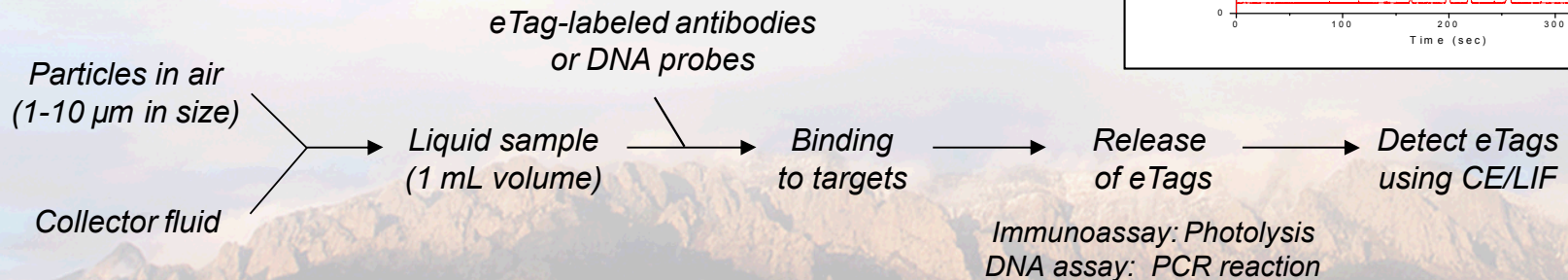
- CE/LIF of reporters provides high multiplexing capability



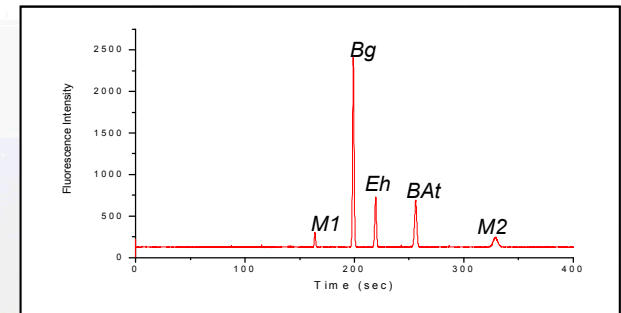
BioBriefcase employs three assay trains to detect the full biothreat spectrum



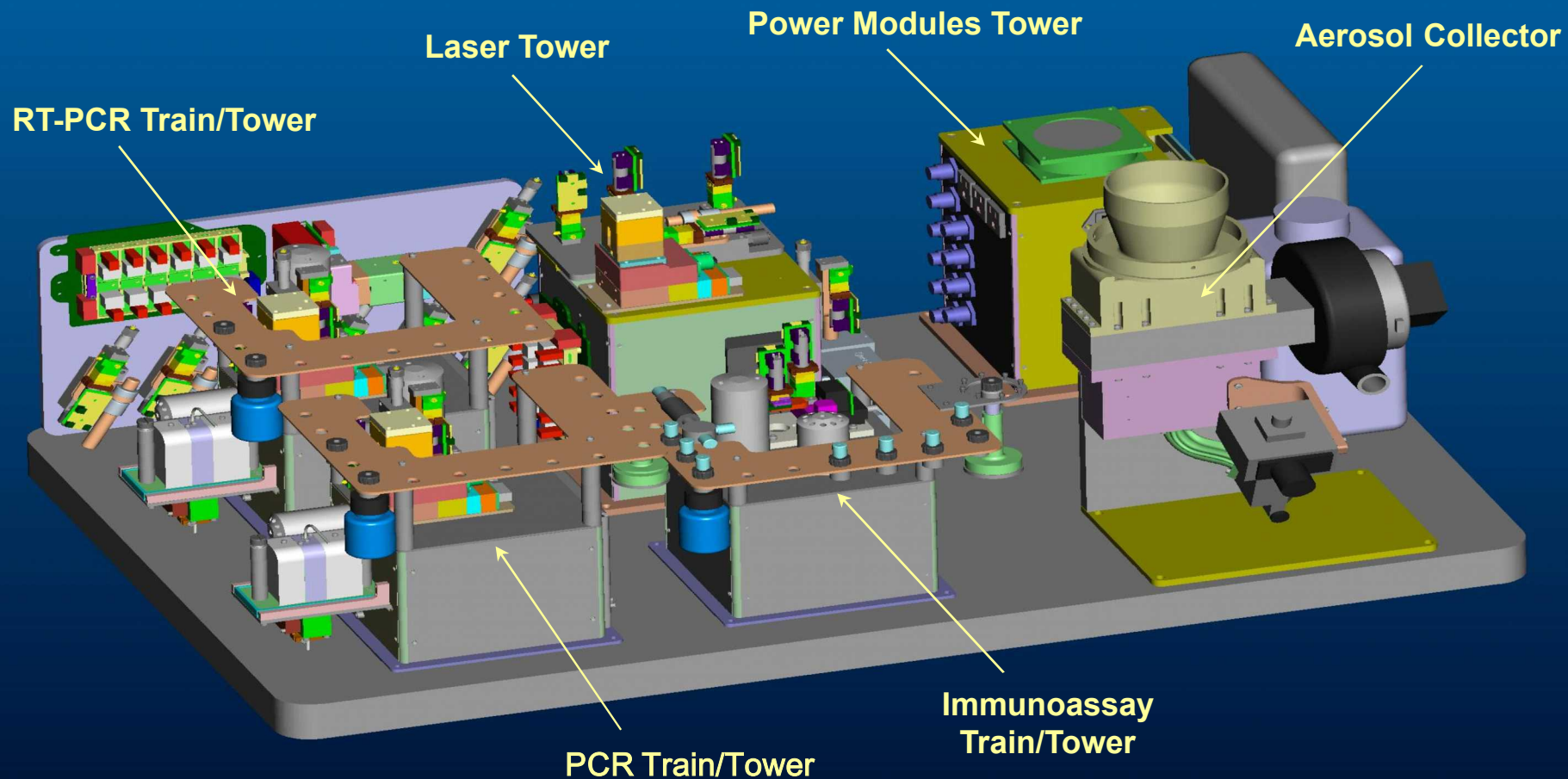
Assays use “eTags” (small fluorescent reporter molecules) to indicate when a pathogen is detected:



CE/LIF analysis of eTags

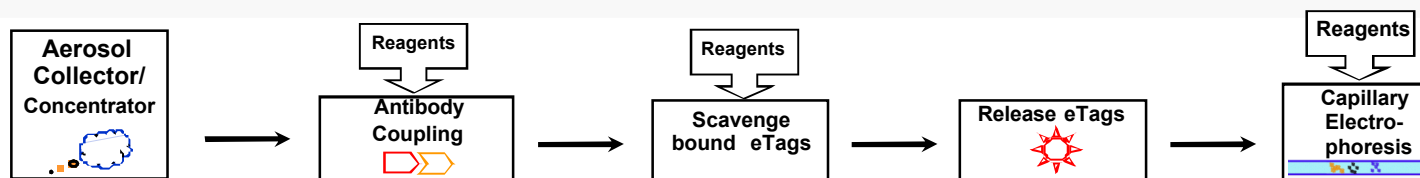


BioBriefcase prototype contains 3 analysis trains, aerosol collector, laser & power distribution towers



Approximate size is 24 x 36 x 14 inches

Integrated hardware can perform automated bead-based immunoassay

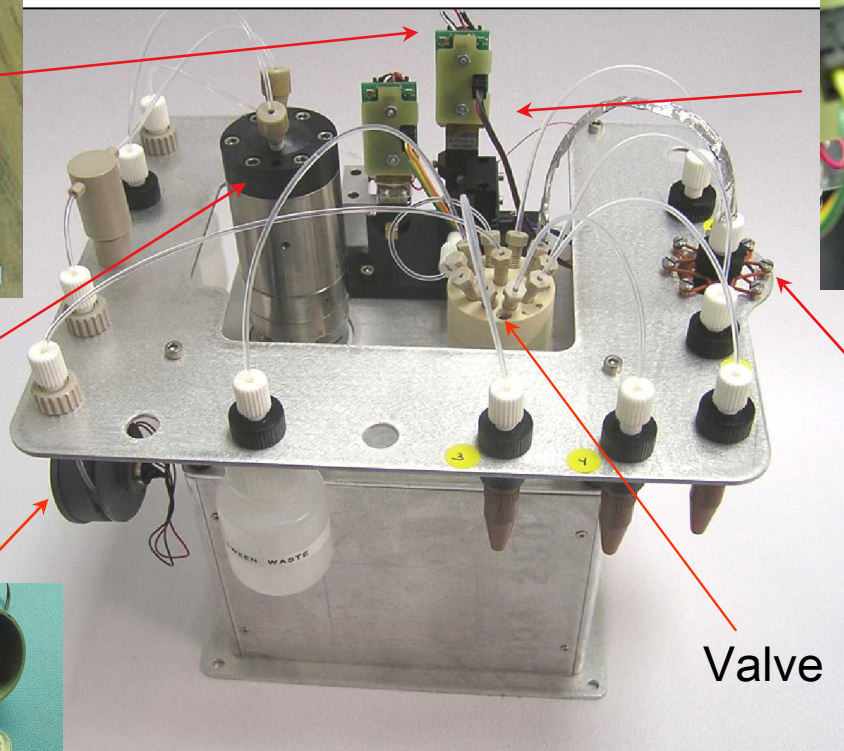
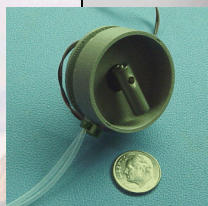


3 way
valve
< 50 nL
dead
volume

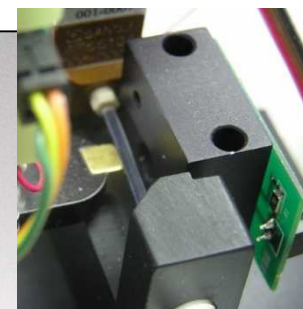


Pump

Tube
bead
mixer



Valve

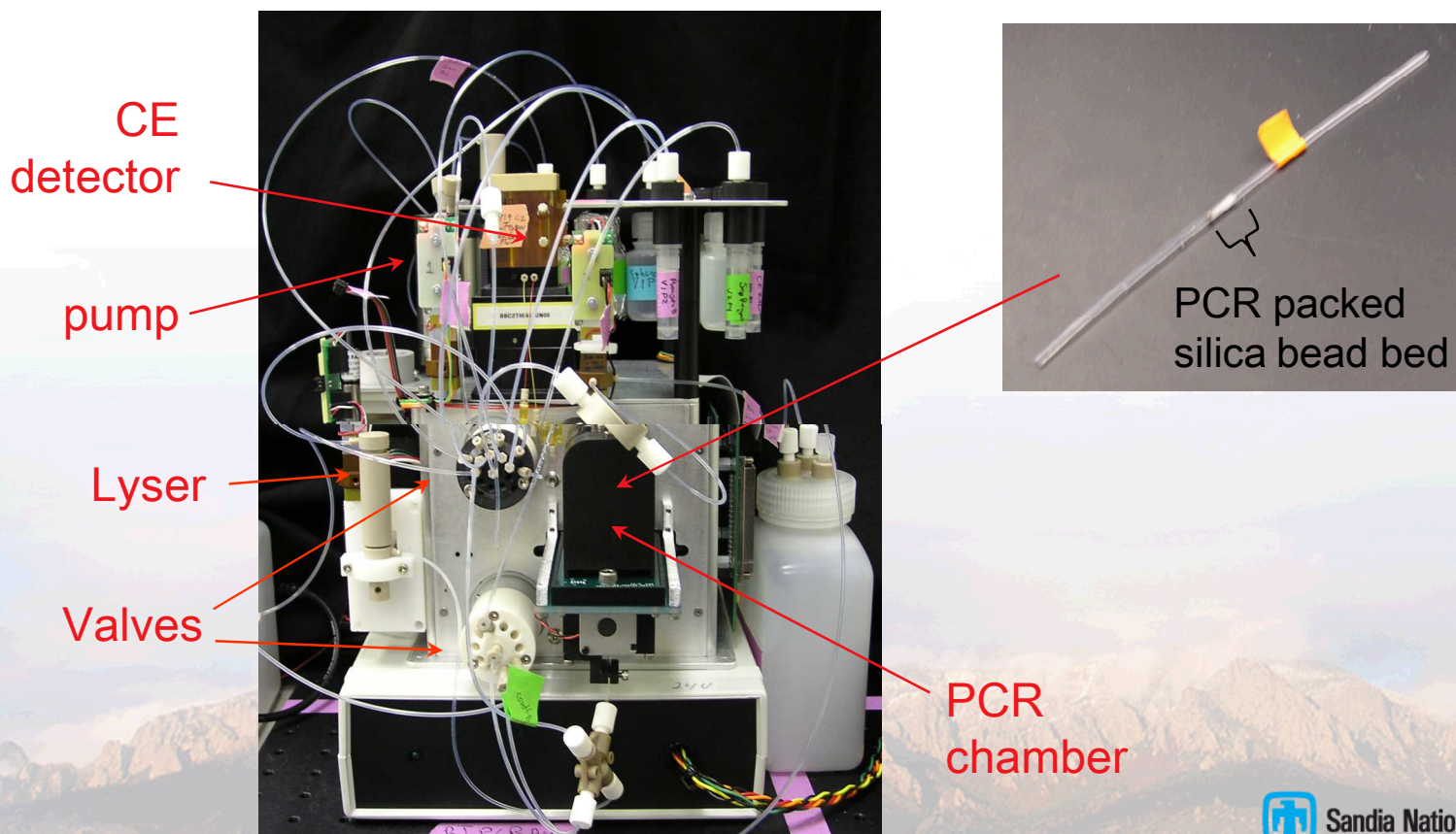
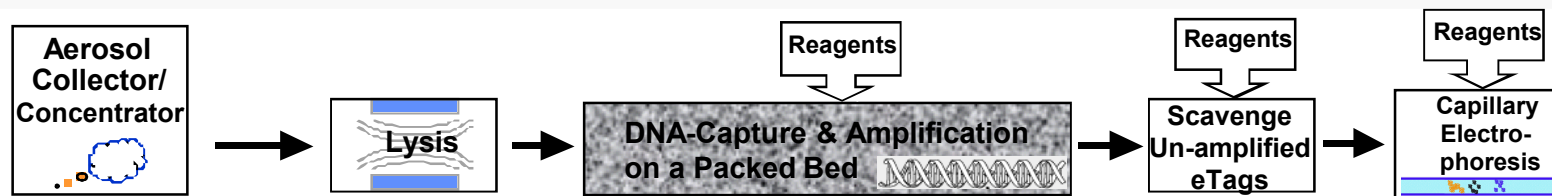


Magnetic
bead trap
and eTag
release



Bead
reagent
mixer

Integrated hardware can perform automated PCR and RT-PCR analyses



Testing conducted at US Army Edgewood Chemical and Biological Center

■ Laboratory testing (20 blind samples) –

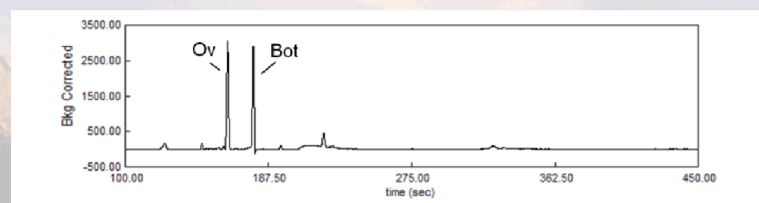
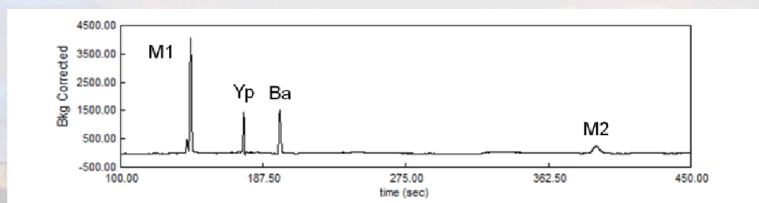
- Liquid sample was drawn automatically into the instrument and analyzed
- Data analysis software generated the detection calls

■ Aerosol chamber testing (18 blind samples) –

- Aerosolized sample was produced directly into the BBC aerosol collector
- Collection continued for 5 additional minutes, then the collection bowl fluid was pumped into the instrument and analyzed (as above)

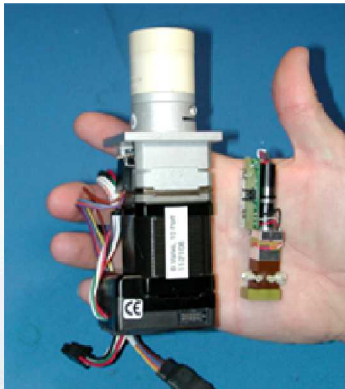
■ Summary of results:

- Correctly identified 87% of samples over the two series of blind challenge tests
 - ♦ Reasons for incorrect calls were immediately identified and correctable
- Although ambient temperature variations shifted CE peak times, we could account this for using internal standards
 - ♦ Independent study of temperature effects indicates a minimum of two internal standards are required for accurate peak calling
 - ♦ Fielded instrument will be environmentally controlled



Conclusions

Next gen fluidic hardware



- Comparison of a commercial valve (left) and Sandia 10-port valve (right)
- The commercial valve is 5 times larger, 20 times heavier, and requires 40 times more power to operate than the Sandia valve

- We have demonstrated capability to successfully build and test portable biodetection instruments using miniaturized assays and componentry
- Our modular design approach enables rapid prototyping of instruments and flexibility to meet new user needs
- By miniaturizing robust analytical methods and instrumentation, system performance is competitive with benchtop systems while meeting field portability requirements
- We are currently extending this technology into medical diagnostics, bioanalysis of precious samples and other areas including biodetection, and welcome opportunities to collaborate