

# Visualizing Early Immune Response: Bacterial Specific Reorganization at the Nanoscale

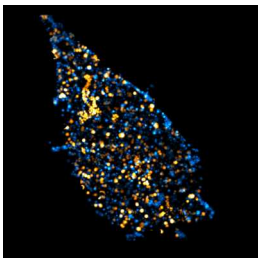
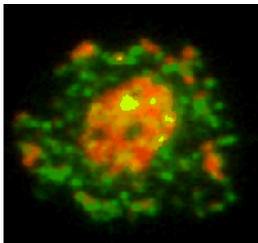
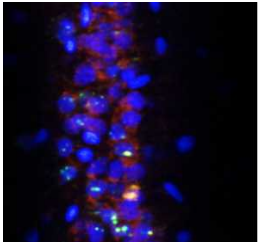
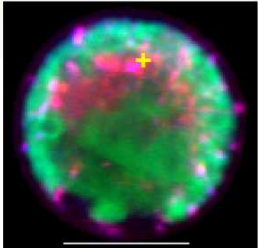
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Albuquerque, New Mexico

*Presented at:  
ASM Biodefense, Washington, DC  
Feb 26-29, 2012*

# Research Focus

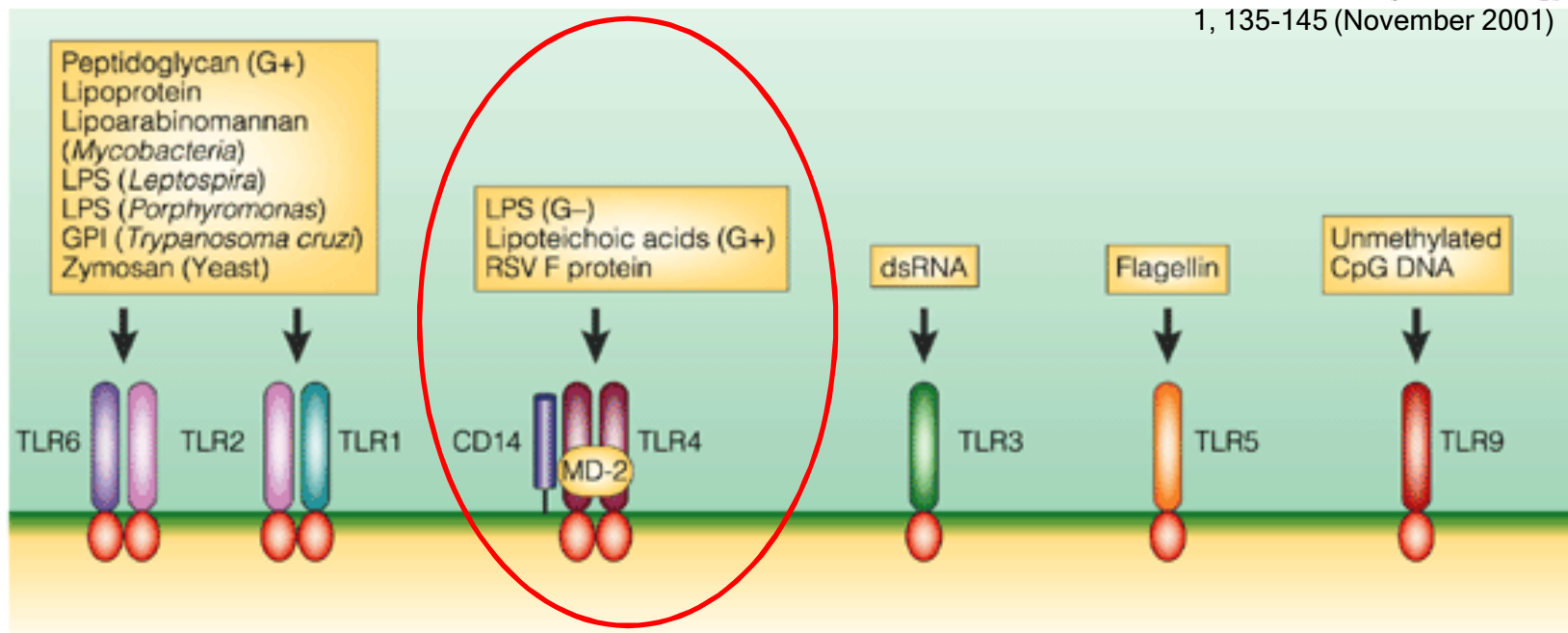
<http://bio.sandia.gov/people/timlin.html>



- Advanced spectroscopy
- Innovative imaging technologies
- Chemometric data analysis tools
  - Multidisciplinary
  - Cell biology, immunology, and microbiology
  - Multicomponent biological systems
  - Biodefense and Bioenergy
- *Multiplexed super resolution microscopy for deciphering complex cellular events*
  - Within the cell: *Work in progress...*
  - At the plasma membrane: *Receptor clustering at the nanoscale*

# TLRs: Important in Pathogenesis, Biodefense

Nature Reviews | Immunology  
1, 135-145 (November 2001)

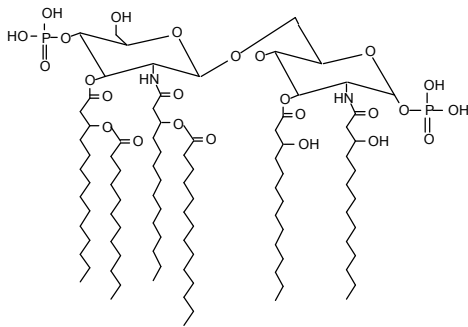


- Important element in mammalian innate immunity
- LPS recognition by TLR4 is aided by accessory proteins
- Close tie between receptor cluster formation, actin cytoskeleton rearrangement, and immune response
- Different chemotypes of LPS generate distinct immune responses

# Chemotypes of LPS Exhibit Differential Immune Response

## *Escherichia coli* (control)

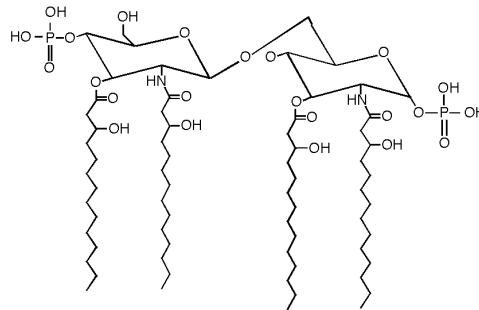
Smooth  
O-polysaccharide



Bind Surface  
+  
↑Stimulatory

## *Yersinia pestis* (37°)

Rough  
O-polysaccharide



Bind Surface  
+  
↓Stimulatory

Differential immune response observed is not fully understood.

- LPS from *E. coli* binds & produces an immune response
- LPS from *Y. pestis* (plague @ 37 °) binds, but does not

Triantafilou, *J Cell Sci* 2002  
Triantafilou, *J Cell Sci* 2004  
Triantafilou, *Biochem J* 2004  
Netea, *Trend Immunol* 2002

# Receptor Clustering Can be Necessary Component of Immune Response

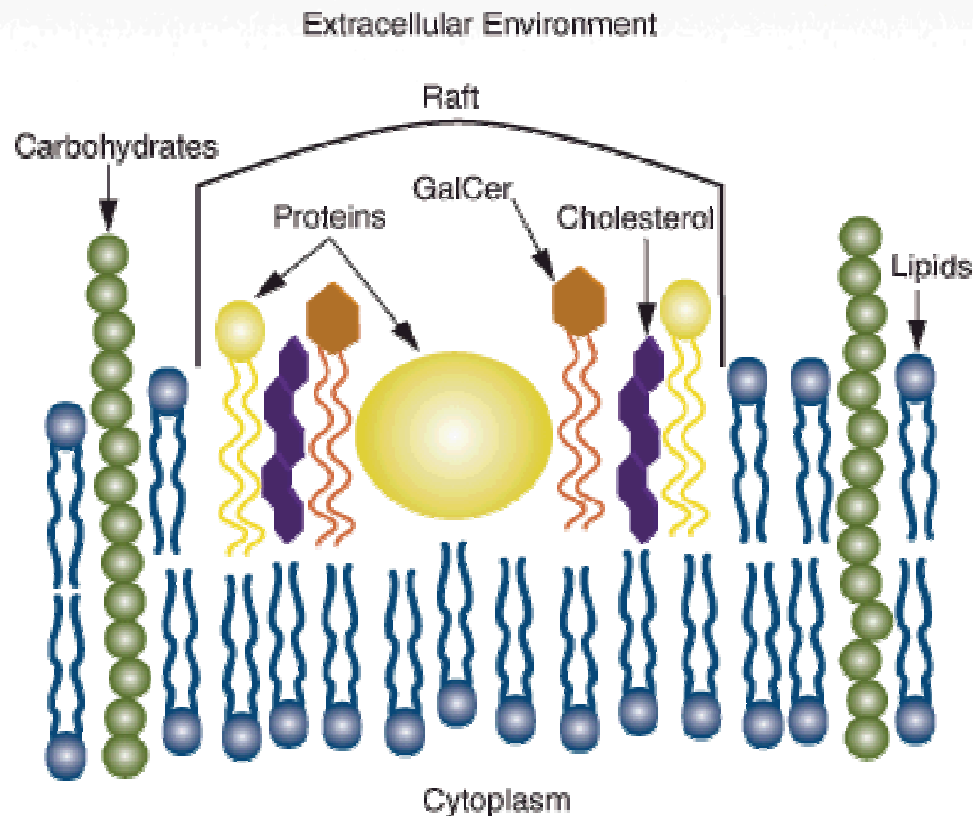


Image Courtesy of Tim Ratto, Lawrence Livermore National Labs

- Domains act as assembly areas
- Aggregation of receptors often follows activation/ligand binding
- Bulk assays have suggested that TLR4 aggregates in lipids rafts within the cell membrane after LPS binding\*
- Link between receptor cluster formation and actin cytoskeleton rearrangement
- Visualization at the single cell level has been limited by optical diffraction

\*Triantafilou, et. al, *Biochem. J.* 381(Pt 2): 527-536



# Hypothesis

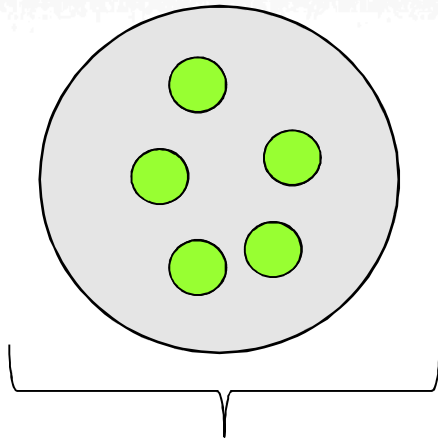
TLR4 receptors cluster at the membrane and the characteristics of this cluster are

- 1) *dependant on the properties of the LPS engaged, and*
- 2) *tied to downstream signaling events*

Optical super-resolution gives us a way to differentiate TLR4 clustering at a much finer scale than conventional imaging.



# Stochastic Optical Reconstruction Microscopy (STORM)



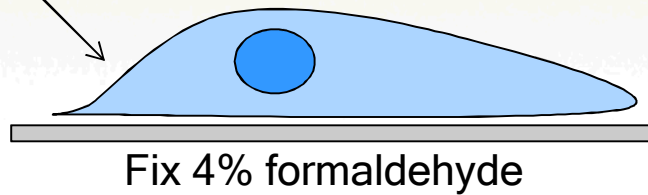
**diffraction-limited spot  
size (~300 nm)**

- The Abbe resolution limit can effectively be broken if the fluorophores in a sample can be imaged *independently* from each other.
- Assuming  $<1$  fluorophor per diffraction-limited area, it's position can be determined with nanometer precision.
- In STORM, this means incorporating stochastic “photoswitching”
- Photoswitching for organic dyes can occur in buffer containing small thiol (i.e. BME) and oxygen scavenging system. (dSTORM)

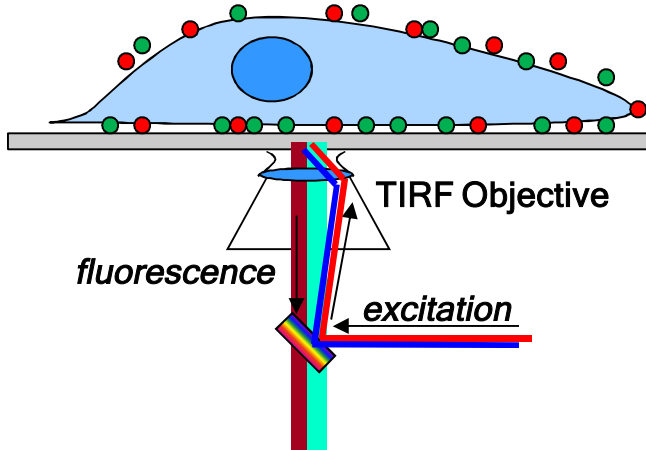
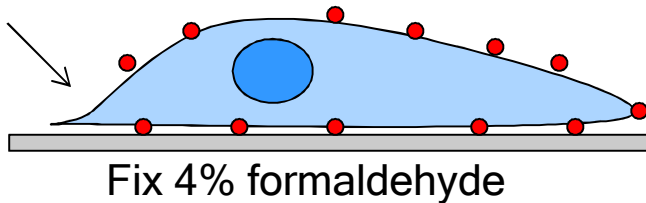
*Rust, et. al, Nat. Meth. 3: 793 - 796 (2006)*

# Experimental Design

• LPS-Alexa647 (30min, 37°C)



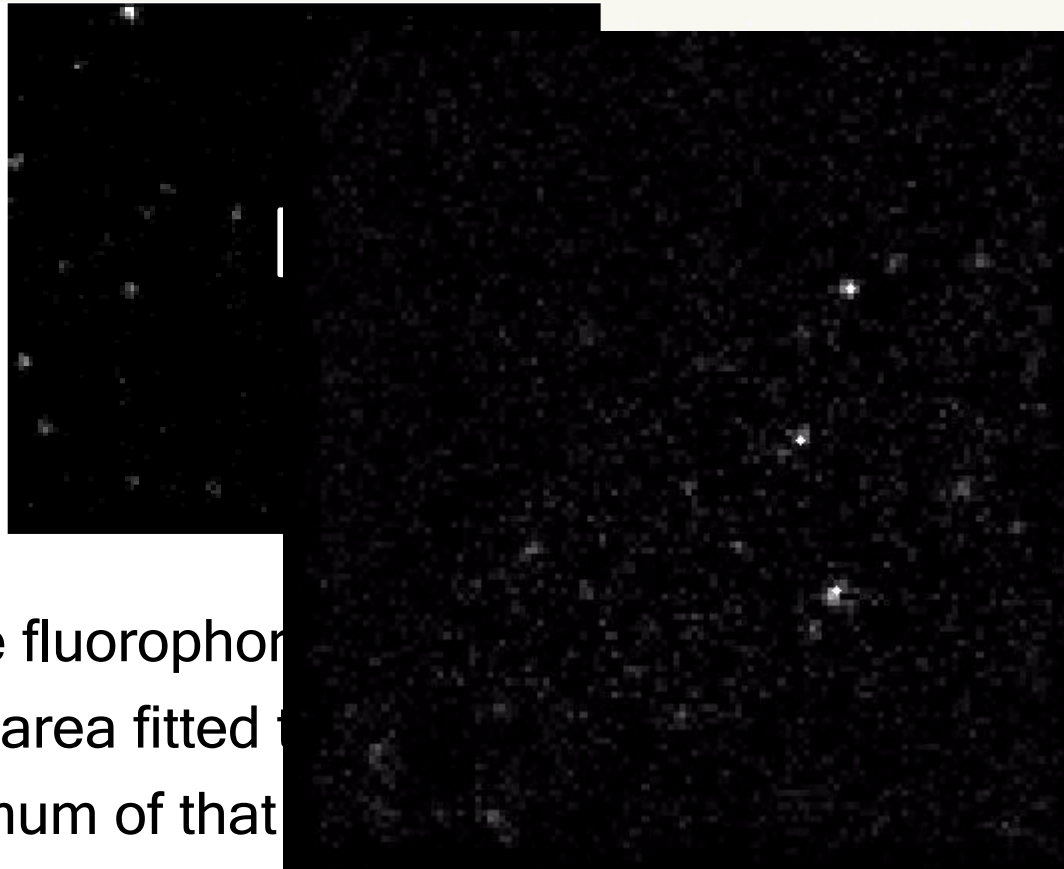
• TLR4 mAbs-Atto532 (60min, RT)



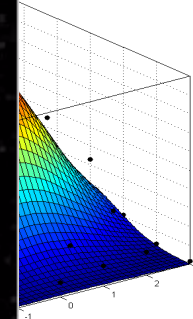
- Mouse macrophage cells (P388D1) incubated with 100nM *E. coli* or *Y. pestis*-derived LPS for 30 min at 37°C and formaldehyde fixed.
- LPS are labeled with Alexa Fluor 647-hydrazide via linkage with core-polysaccharide
- TLR4 receptors visualized via 1<sup>0</sup> antibodies labeled with Atto532
- Mount in O<sub>2</sub>-scavenging buffer containing β-mercaptothiol
- Cells imaged using dual-color TIRF excitation and emission modes



# Fluorophor Localization



nm ,  $44 \pm 23$ nm)

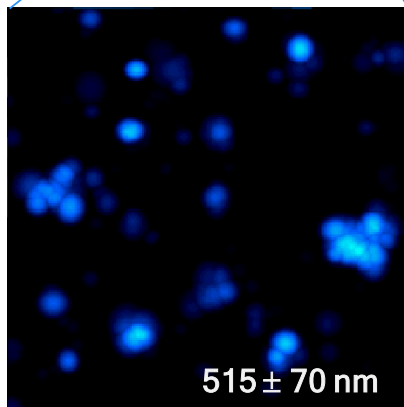
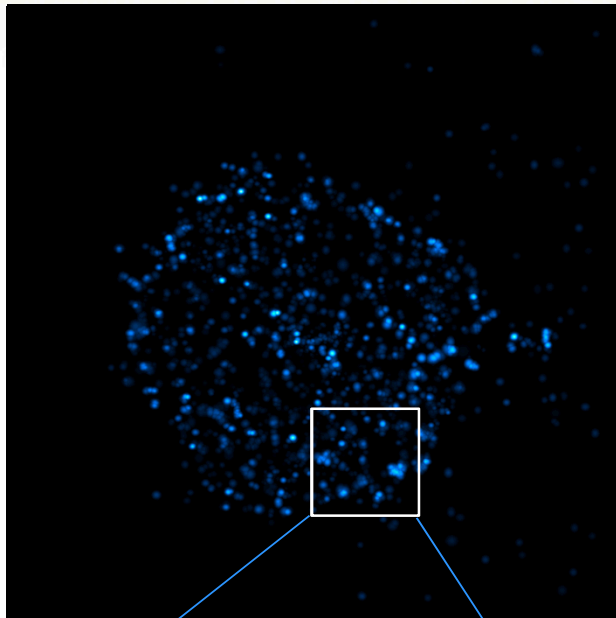


Uncertainty  $\sim 1/\sqrt{N}$   
(SNR)

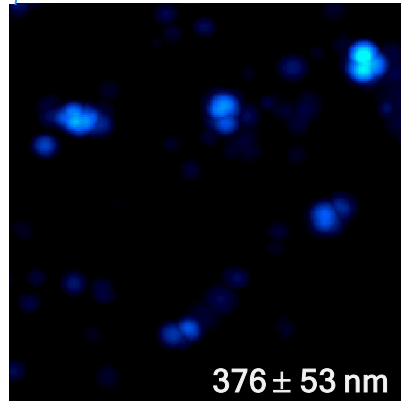
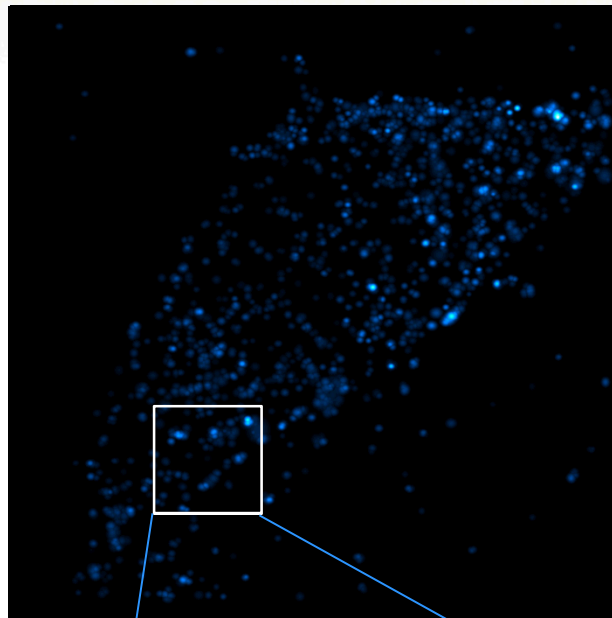
- Single fluorophore identified in each frame
- Local area fitted to PSF
- Maximum of that fit is the location of the fluorophore
- Typically, location fit uncertainty 40-60nm
- Process repeated over 1k-10k frames to build STORM image

# TLR4 Clustering is Specific

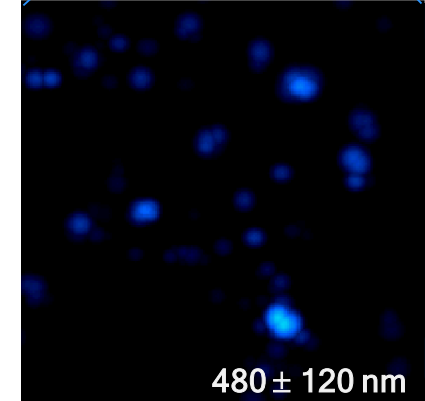
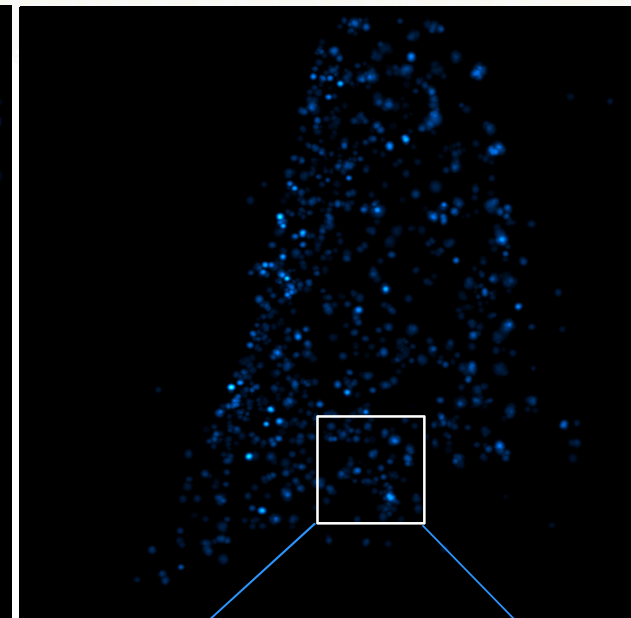
*E. coli* LPS



Flagellin

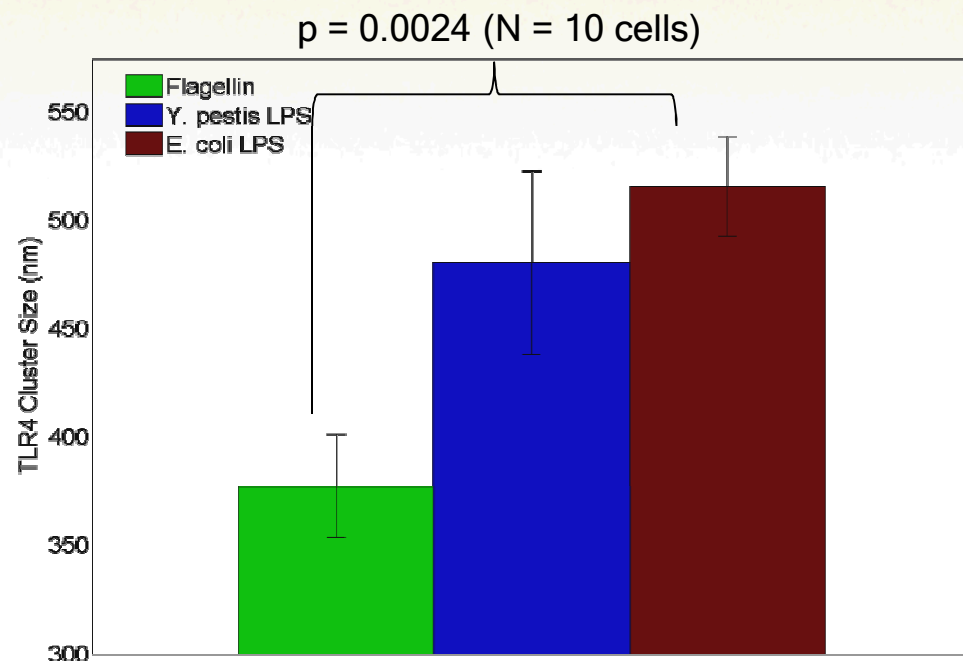
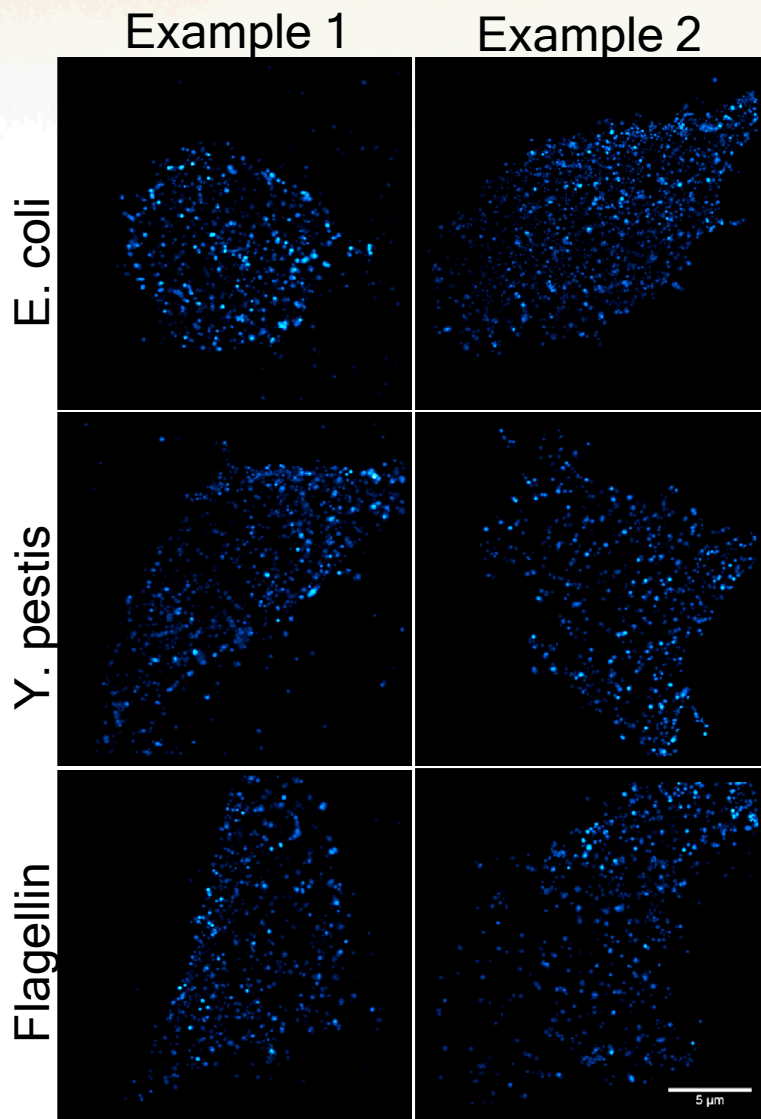


*Y. pestis* LPS



STORM images 8-10 fold increase in resolution

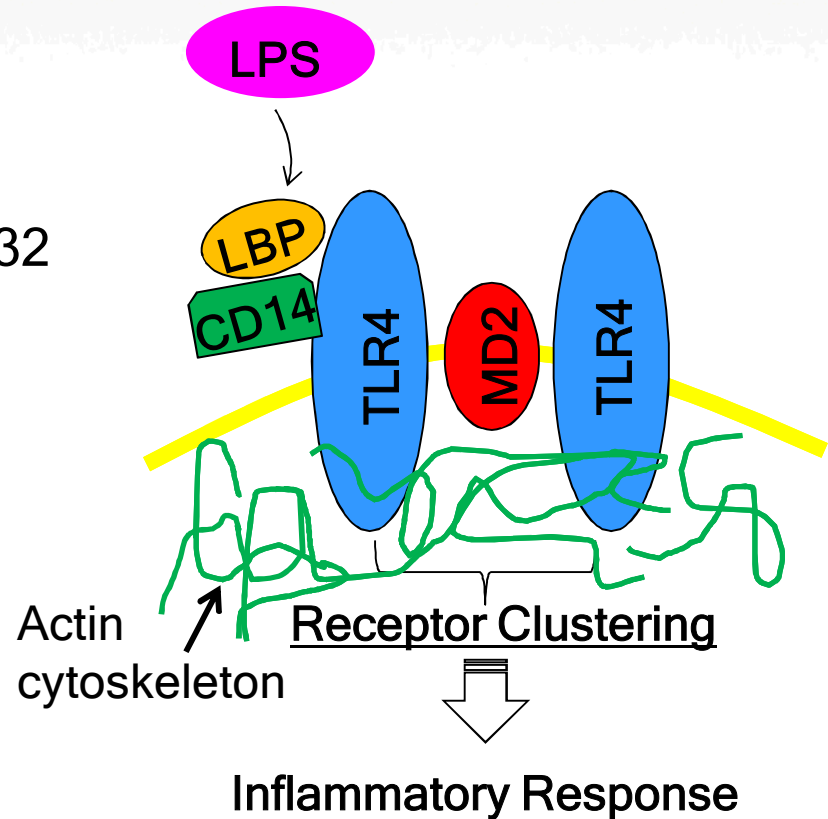
# TLR4 Cluster Analysis



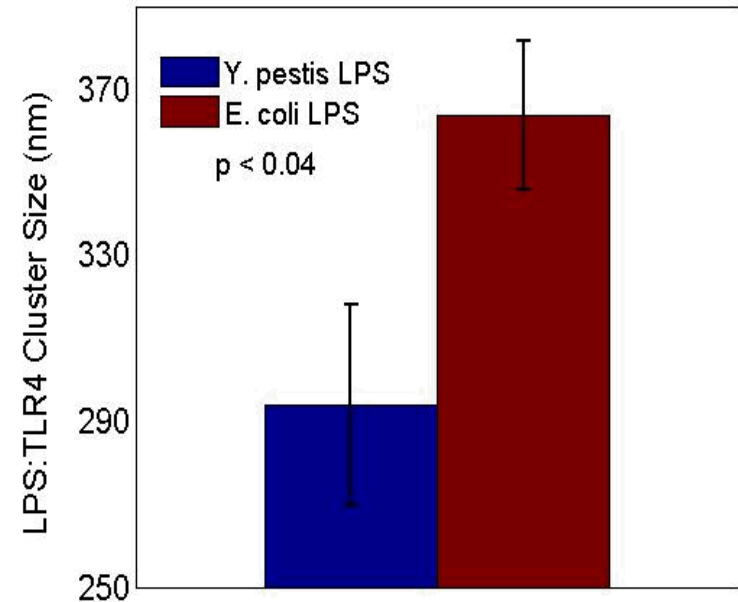
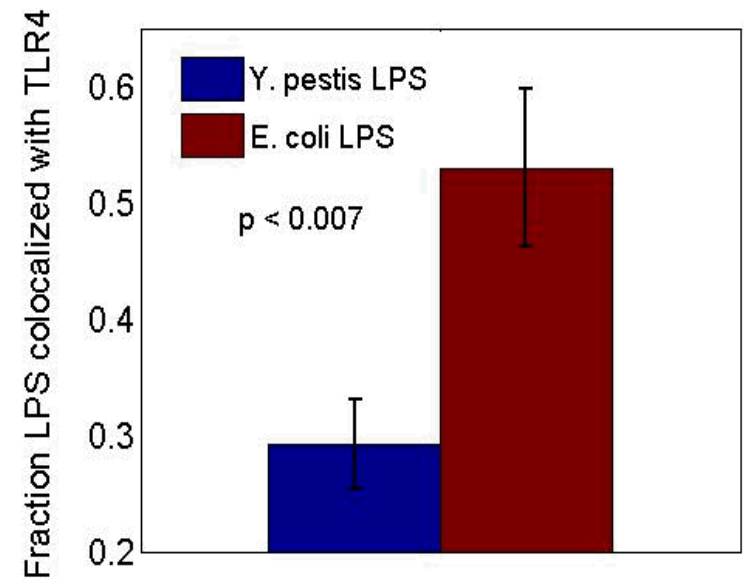
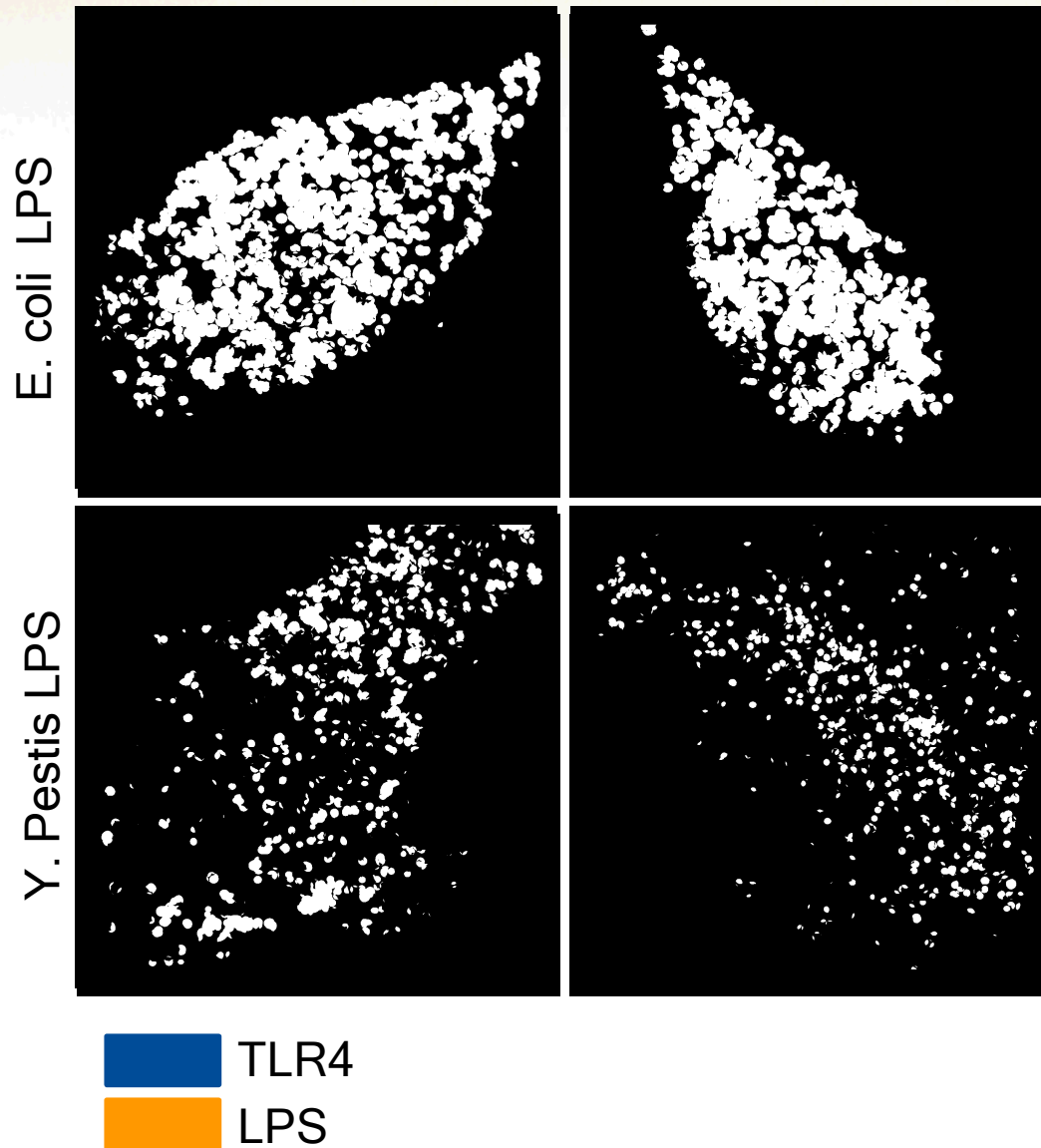
- Ripley's K-analysis indicates that *E. coli* LPS induces significant clustering over negative control (flagellin)
- Suggests that *pestis* induces less clustering, but not significant
- TLR4-LPS complex?

# Imaging the TLR4-LPS Complex

- Dual-color STORM
  - LPS-Alexa647/ TLR4-Atto 532
  - 532 & 638 nm excitation
  - Total irradiance  $\sim 10\text{W}/\text{cm}^2$
  - 0.05-0.1 seconds per frame
- Image registration using multicolor beads
  - Linear transform
  - Errors  $< 50\text{ nm}$



# Colocalization of TLR4 & LPS







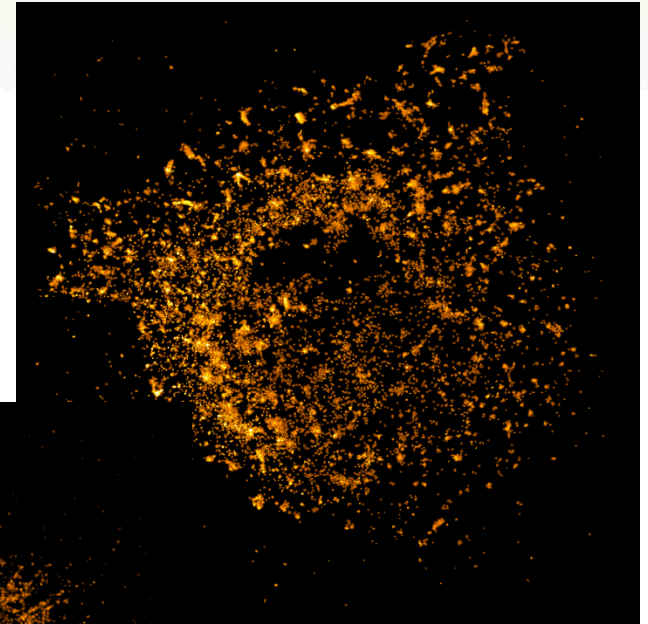
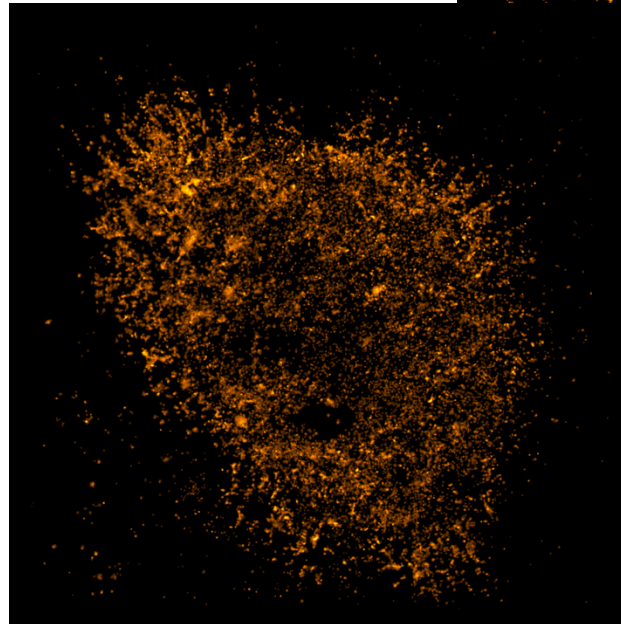
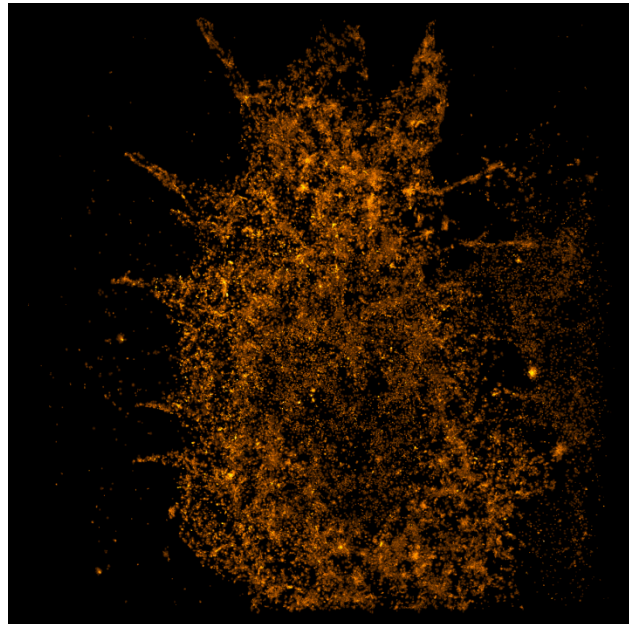
# Conclusions

- Super-Resolution imaging allows for measuring subtle changes that aren't apparent in conventional microscopy
- *E. coli* LPS produces a significant increase in TLR4 cluster size within 30 minutes, as compared to a non-specific ligand (Flagellin) and non-stimulatory control
- *Y. pestis* LPS and *E-coli* LPS exhibit similar degrees of membrane binding
- *Y. pestis* LPS exhibits less co-localization with TLR4 and is less able to recruit TLR4 into clusters as compared to *E. coli* LPS → correlated with down-stream signaling response



# Future Directions

- *Impact of actin cytoskeleton*
- *Fundamental properties of the membrane (e.g. temperature, curvature)*
- *Additional chemotypes of LPS*



# Acknowledgements

## Current Group Members

Dr. Jesse Aaron

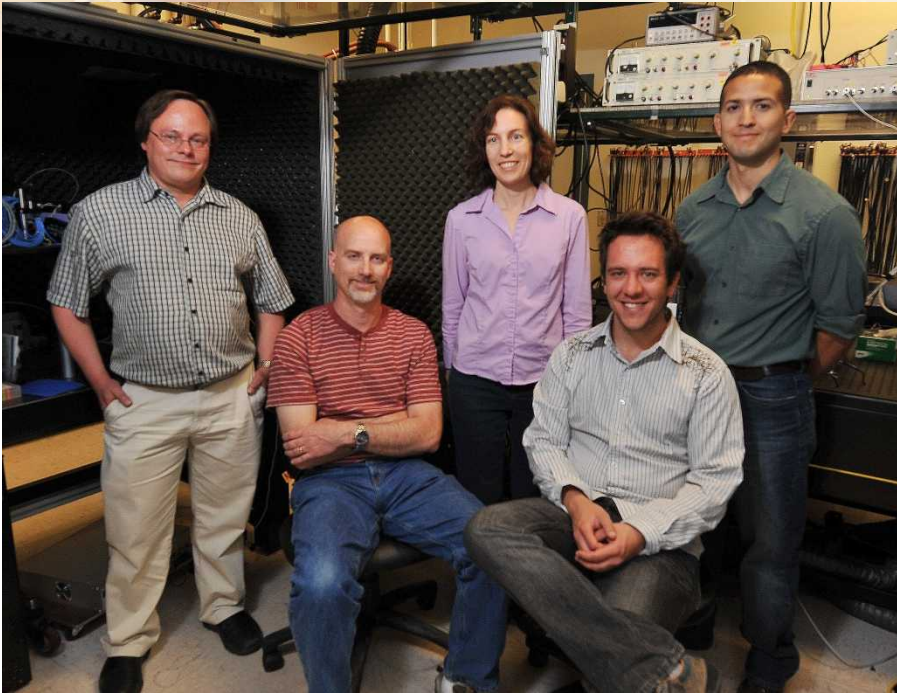
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Amy Powell

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Hanson & Turner Labs, UNM Biology

Blankenship & Pakrasi Labs, WUSTL

Sayre Lab, NM Consortium

Sapphire Energy

Hu Lab, ASU



**STMC**  
SpatioTemporal Modeling Center



**U.S. DEPARTMENT OF  
ENERGY**

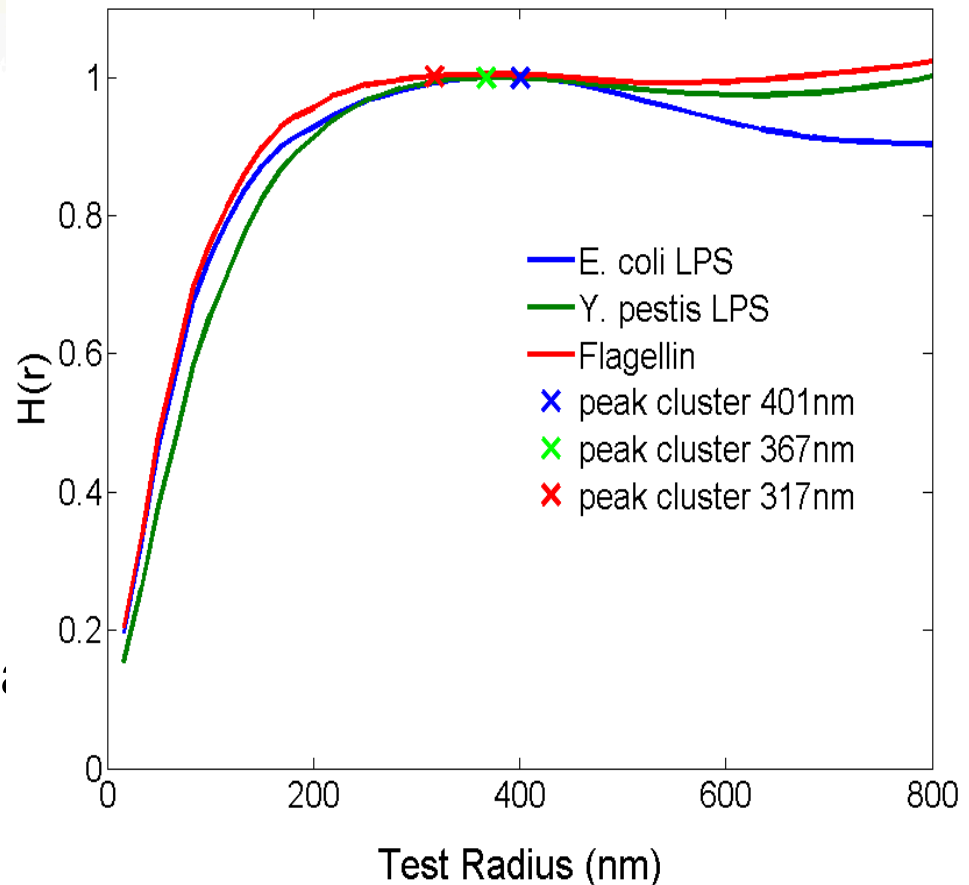
# Extras

# Ripley's K-function Analysis

- K-function is a normalized measure of point clustering

Ripley, B.D. *J. R. Statist. Soc. B*41:368-374 (1979)

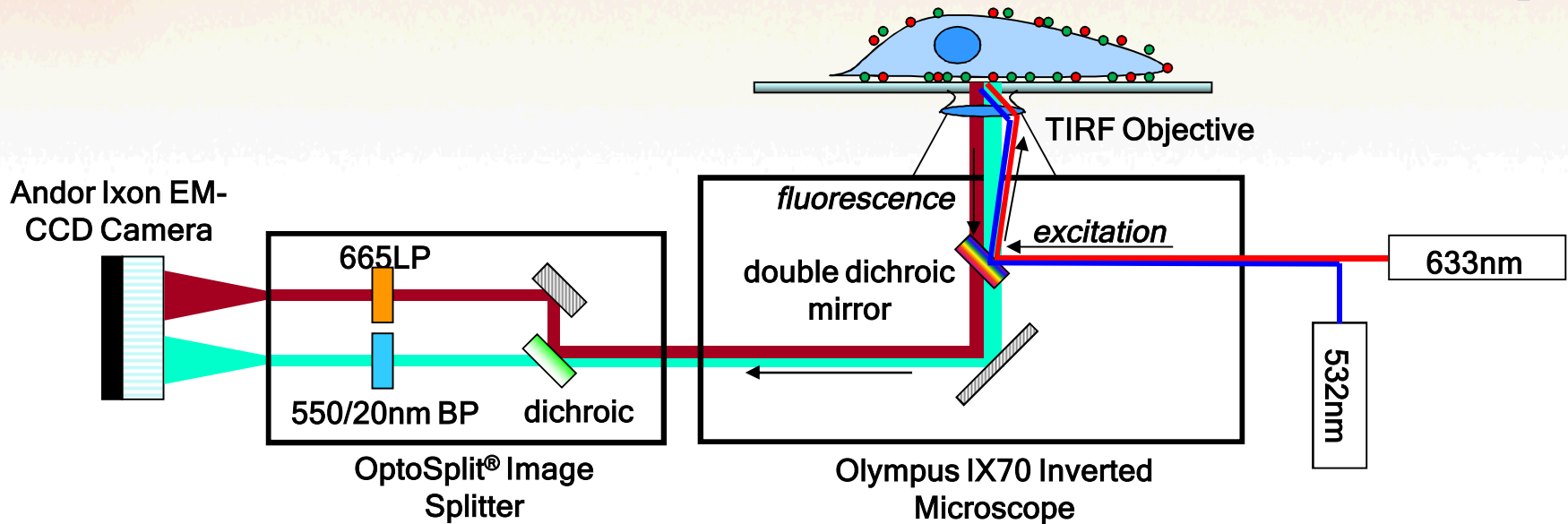
- Complete spatial randomness (CSR)
- Transform to H-function to gauge deviation from CSR at each test radius
- Peaks (or inflection points)  $H(r)$  indicate characteristic cluster sizes



Kriskowski, M.A., et al, *Biophys. J.* 97(4), 1095-1103, (2009)



# Multicolor STORM Setup



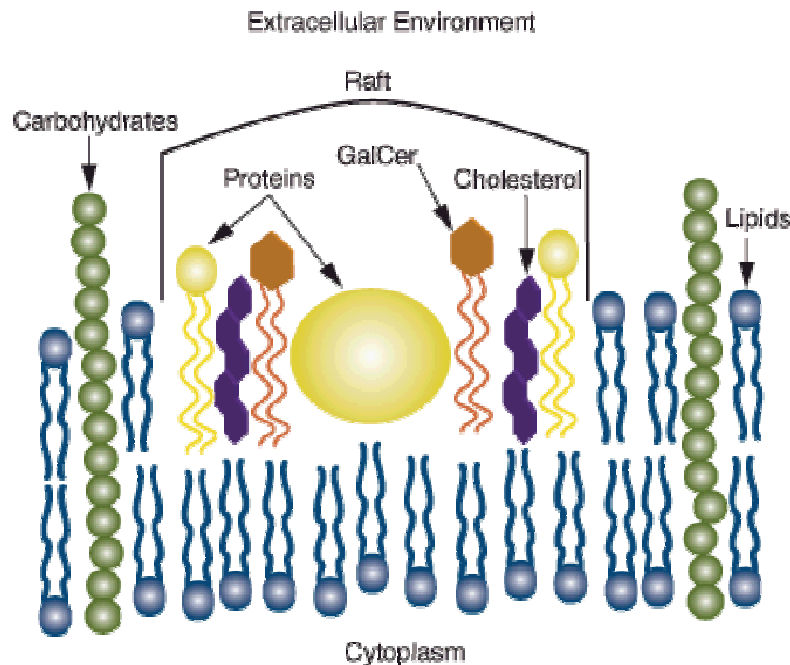
## Unique capabilities:

- Four excitation  $\lambda$ 's (405, 488, 532, 633nm), variable angle
- Simultaneous dual-color emission
- Capable of >50fps over 30 $\mu$ m x 30 $\mu$ m FOV

## Advantageous in:

- Receptor reorganization
- Nanoparticle-membrane interactions, uptake
  - Engineered NPs
  - Natural NPs - Viral trafficking

# Receptor Clustering Can be Necessary in Immune Response



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