

TITLE: Biotic and Abiotic Carbon To Sulfur Bond Cleavage

P.I.: John W. Frost, Associate Professor

DOE/PC/90294--T1

STUDENTS: Kurt Stimpfl and Bernard Heymann, Ph.D. Candidates

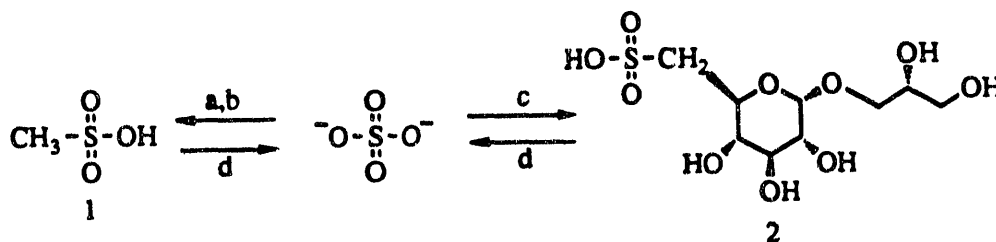
DE92 012640

INSTITUTION: Purdue University, West Lafayette, IN 47907

GRANT NO.: DE-FG22-90PC90294

Cleavage of aliphatic organosulfonate carbon to sulfur (C-S) bonds, a critical link in the global biogeochemical sulfur cycle,^{1,2} has been identified in *Escherichia coli* K-12. Enormous quantities of inorganic sulfate are continuously converted (Scheme I) into methanesulfonic acid 1 and acylated 3-(6-sulfo- α -D-quinovopyranosyl)-L-glycerol 2.^{1,2,3} Biocatalytic desulfurization (Scheme I) of 1 and 2, which share the structural feature of an aliphatic carbon bonded to a sulfonic acid sulfur, completes the cycle. Discovery of this desulfurization in *E. coli* provides an invaluable paradigm for study of a biotic process which, via the biogeochemical sulfur cycle, significantly influences the atmospheric concentration of sulfur-containing molecules.²

SCHEME I



a. biocatalysis (bacteria, plants, algae) c. biocatalysis (plants)

b. photolysis

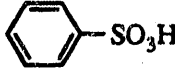
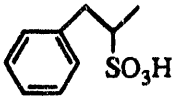
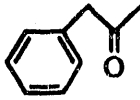









d. biocatalysis (bacteria)

Desulfurizations of aliphatic organosulfonates and aromatic organosulfonates constitute the major classes of biocatalytic organosulfonate C-S bond cleavage processes. Forcing *E. coli* K-12 to use aliphatic organosulfonates (Table I) as a sulfur source revealed capabilities broadly reminiscent of desulfurizations catalyzed by bacteria isolated from soil and harbor water.⁴ Formation of [^{14}C]-carbon dioxide from [^{14}C]-methanesulfonic acid is similar to a recently discovered transformation effected by soil bacteria.¹ However, not all organosulfonate C-S bond cleavage processes are present in *E. coli*. Failure to degrade benzenesulfonate distinguishes *E. coli* from bacteria which are capable of cleaving aromatic organosulfonate C-S bonds.^{5,6}

MASTER

DISTRIBUTION OF THIS DOCUMENT IS UNLIMITED

Table I. Comparison of Biocatalytic Aliphatic Organosulfonate C-S Bond Cleavage Reactions

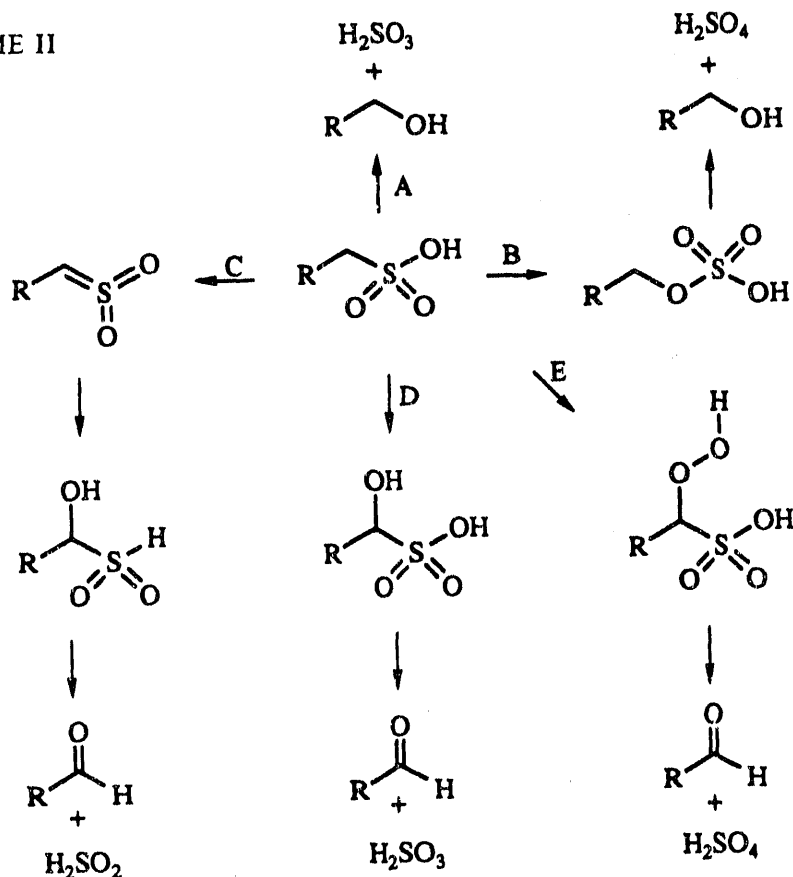
organosulfonate	growth ^a	specific ^b activity 10 ⁻⁴ units/mg	products ^c	
			intact cells	lysate
¹⁴ CH ₃ SO ₃ H	+	0.35	¹⁴ CO ₂	ND ^d
CH ₃ (CH ₂) ₅ SO ₃ H	+	3.3	CH ₃ (CH ₂) ₅ OH	CH ₃ (CH ₂) ₄ CO ₂ H
	-	X	X	X
	-	0.29	X	
 n=1	+	5.4	 n=1	 n=0
 n=2	+	9.0	 n=2	 n=1
 n=3	+	6.4	 n=3	 n=2

^a *E. coli* RB791 (W31110 *lacL819*)⁷ were cultured at 37°C for 14 h in M9 minimal medium⁸ supplemented with thiamine hydrochloride (0.005 g/L) and with organosulfonic acid (0.25 mM) replacing inorganic sulfate. ^b 1 unit = 1 μmol SO₃⁻²/min; Assay solutions (pH 8.8) consisted of protein (10 -14 mg/mL) from desalted cell lysate incubated with trizma·HCl (63 mM), sodium chloride (9 mM), ammonium chloride (19 mM), NADH (10 mM), and organosulfonate (12.5 mM). Aliquots (1 mL) were removed at timed intervals and protein precipitated with 10% (w/v) trichloroacetic acid. Absorbance (412 nm) was measured after incubation of neutralized, deproteinized aliquots with Ellman's reagent. ^c Chloroform extracts of culture supernatants and cell lysates were analyzed by gas chromatography and GC-MS. Structure assignments were based on coinjection with authentic samples and comparison of mass spectra. ^d not determined

Extracellular accumulation of 2-phenylethan-1-ol and trace levels of phenylacetic acid in the supernatant of *E. coli* cultured with 2-phenyl-1-ethanesulfonic acid were characteristic of the carbon fragments formed from the aliphatic organosulfonates (Table I). To ascertain whether the alcohols were the actual carbon fragments *initially* formed by C-S bond cleavage, desulfurization of 2-phenyl-

1,1-dideutero-1-ethanesulfonic acid by intact *E. coli* was examined. Retention of only one deuterium in product 2-phenyl-1-deuteroethan-1-ol implicated aldehyde intermediacy despite the absence of aldehyde products in cell lysate and culture supernatants. This eliminated direct hydrolysis⁹ (Path A; Scheme II) and insertion of oxygen into the C-S bond (Path B; Scheme II) as mechanistic possibilities.

SCHEME II



Although no C-S bond cleavage of organosulfonates was detectable in anaerobic desalted cell lysate, aeration and addition of reduced nicotinamide adenine dinucleotide (NADH) resulted in desulfurization. Formation of phenylacetic acid and phenyl-2-propanone from 2-phenyl-1-ethanesulfonic acid and 1-phenyl-2-propanesulfonic acid were typical (Table I) of the cell-free desulfurizations. The NADH and oxygen requirement for cell-free C-S cleavage activity effectively removed sulfene intermediacy¹⁰ (Path C; Scheme II) from further consideration. Formation of 5-

thio-2-nitrobenzoate when Ellman's reagent was added to deproteinized assay aliquots implicated sulfite formation and was used to determine enzyme specific activities (Table I).¹¹ This eliminates all mechanisms leading to inorganic sulfate¹² (Path E, Scheme II) and leaves α -hydroxylation (Path D, Scheme II) as the only remaining, viable mechanism.

For the present, scrutiny of the products, intermediates, and mechanism of aliphatic organosulfonate desulfurization in *E. coli* has provided key insights into an intriguing yet largely uncharacterized biocatalytic transformation. The next level of appraisal requires purification of the enzyme responsible for aliphatic organosulfonate C-S bond cleavage. Such efforts must contend with the low specific activities in cell lysate and chronic enzyme instability. Fortunately, the relative ease with which soluble enzymes can be overproduced in *E. coli* and observed stabilization of C-S bond cleaving activity with NADH and argon atmosphere¹³ should expedite future manipulations of the enzyme.

Acknowledgment. Research was supported by Department of Energy Contract No. DE-FG22-90PC90294.

REFERENCES

1. Baker, S. C.; Kelly D. P.; Murrell, J. C. *Nature* **1991**, *350*, 627.
2. Kelly, D. P.; Smith, N. A. *Adv. Microb. Ecol.* **1990**, *11*, 345.
3. Harwood, J. L.; Nicholls, R. G. *Biochem. Soc. Trans.* **1979**, *7*, 440
4. (a) Thyse, G. J. E.; Wanders, T. H. *Antonie van Leeuwenhoek* **1972**, *38*, 53. (b) Thyse, G. J. E.; Wanders, T. H. *Antonie van Leeuwenhoek* **197**, *40*, 25. (c) Biedlingmaier, S.; Schmidt, A. *Arch. Microbiol.* **1983**, *136*, 124. (d) Biedlingmaier, S.; Schmidt, A. *Z. Naturforsch.* **1987**, *42c*, 891.
5. For aromatic organosulfonate C-S bond cleavage in cell-free lysate, see: (a) Locher, H. H.; Leisinger, T.; Cook, A. M. *Biochem. J* **1991**, *274*, 833. (b) Thurnheer, T.; Zürrer, D.; Höglinger, O.; Leisinger, T.; Cook, A. M. *Biodegradation* **1990**, *1*, 55. (c) Locher, H. H.; Leisinger, T.; Cook, A. M. *J. Gen. Microbiol.* **1989**, *135*, 1969. (d) Thurnheer, T.; Köhler, T.; Cook, A. M.; Leisinger, T. *J. Gen.*

Microbiol. **1986**, *132*, 1215.

6. For a general review, see: Swisher, R. D. *Surfactant Biodegradation*; Marcel Dekker: New York, 1987.

7. Frost, J. W.; Bender, J. L.; Kadonaga, J. T.; Knowles, J. R. *Biochemistry* **1984**, *23*, 4470.

8. Miller, J. H. *Experiments in Molecular Genetics*; Cold Spring Harbor Laboratory: Cold Spring Harbor, 1972; pp 431.

9. (a) Wagner, F. C.; Reid, E. E. *J. Am. Chem. Soc.* **1931**, *53*, 3407. (b) Brindley, P. B. *J. Chem. Soc. C* **1966**, 163.

10. This process was hypothesized to possibly involve adenylyl transfer to the sulfonic acid catalyzed by ATP sulfurylase,^a subsequent elimination to generate APS along with a sulfene,^{b-d} and final hydrolysis. (a) Leyh, T. S.; Taylor, J. C.; Markham, G. D. *J. Biol. Chem.* **1988**, *263*, 2409. (b) Wallace, T.J. *Quart. Rev.* **1966**, *20*, 67. (c) Opitz, G. *Ang. Chem. Int. Ed.* **1967**, *6*, 107. (d) King, J. F. *Acc. Chem. Res.* **1975**, *8*, 10.

11. Humphrey, R. E.; Ward, M. H.; Hinze, W. *Anal. Chem.* **1970**, *42*, 698.

12. No reduction of inorganic sulfate to sulfite was observed in cell lysates.

13. No appreciable loss of enzyme activity is observed over a 270 h period when cell lysate in the standard assay solution lacking organosulfonate is maintained under argon. Removal of NADH along with exposure to air results in a 90% loss of enzyme activity within 5 h.

DISCLAIMER

This report was prepared as an account of work sponsored by an agency of the United States Government. Neither the United States Government nor any agency thereof, nor any of their employees, makes any warranty, express or implied, or assumes any legal liability or responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product, or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commercial product, process, or service by trade name, trademark, manufacturer, or otherwise does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government or any agency thereof. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States Government or any agency thereof.

END

**DATE
FILMED**

6 / 11 / 92

