

1 ***Suillus*: an emerging model for the study of ectomycorrhizal ecology**  
2 **and evolution**

3  
4 *Lotus Lofgren*<sup>1</sup>, *Nhu H. Nguyen*<sup>2</sup>, *Peter Kennedy*<sup>3,4</sup>, *Eduardo Pérez-Pazos*<sup>4</sup>, *Jessica Fletcher*<sup>5</sup>,  
5 *Hui-Ling Liao*<sup>6,7</sup>, *Haihua Wang*<sup>6,7</sup>, *Kaile Zhang*<sup>6</sup>, *Joske Ruytinx*<sup>8</sup>, *Alexander H. Smith*<sup>5</sup>, *Yi-*  
6 *Hong Ke*<sup>9</sup>, *H. Van T. Cotter*<sup>10</sup>, *Eiona Engwall*<sup>11</sup>, *Khalid M. Hameed*<sup>1</sup>, *Rytas Vilgalys*<sup>1</sup>, *Sara*  
7 *Branco*<sup>5</sup>

8  
9 *1 Department of Biology, Duke University*

10 *2 Department of Tropical Plant and Soil Sciences, University of Hawai'i at Mānoa*

11 *3 Department of Plant and Microbial Biology, University of Minnesota*

12 *4 Department of Ecology, Evolution and Behavior, University of Minnesota*

13 *5 Department of Integrative Biology, University of Colorado Denver*

14 *6 North Florida Research and Education Center, University of Florida*

15 *7 Department of Soil, Water and Ecosystem Sciences, University of Florida*

16 *8 Research Group of Microbiology and Plant Genetics, Department of Bioengineering*

17 *Sciences, Vrije Universiteit Brussel*

18 *9 Department of Ecology and Evolutionary Biology, University of Michigan*

19 *10 University of North Carolina at Chapel Hill Herbarium*

20 *11 Department of Biology, University of North Carolina at Chapel Hill*

21  
22 **ORCID IDs:**

23 *Lotus Lofgren* 0000-0002-0632-102X

24 *Sara Branco* 0000-0002-9890-933X

25 *Nhu H. Nguyen* 0000-0001-8276-7042

26 *Peter Kennedy* 0000-0003-2615-3892

27 *Rytas Vilgalys* 0000-0001-8299-3605

28 *Eduardo Pérez-Pazos* 0000-0002-9662-711X

29 *Joske Ruytinx* 0000-0001-5835-3704

30 *Alexander Smith* 0000-0002-3890-000X

31 *Hui-Ling Liao* 0000-0002-1648-3444

32 *Jessica Fletcher* 0000-0002-8737-1538

33 *Kaile Zhang* 0000-0001-6910-8050

34 *Haihua Wang* 0000-0002-7410-2064

35 *Khalid M. Hameed* 0000-0002-5882-5553

36 *H. Van T. Cotter* 0000-0002-8819-7068

37 *Yi-Hong Ke* 0000-0002-8435-082X

38 *Eiona Engwall* 0009-0002-2092-6495

39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54

55	
56	<b>Keywords:</b>
57	<i>Suillus</i> , ECM, ectomycorrhizal, ecology, evolution, mycology, partner specificity, partner
58	specificity, stress tolerance, model system, genus, symbiosis
59	
60	<b>Contents:</b>
61	Summary
62	<b>I.</b> Introduction
63	<b>II.</b> <i>Suillus</i> as a model genus
64	<b>III.</b> Taxonomy and evolution
65	<b>IV.</b> Ecological interactions
66	<b>V.</b> Environmental (abiotic) adaptation
67	<b>VI.</b> Biological introductions and invasions
68	<b>VII.</b> Conclusions and future directions
69	<b>VIII.</b> Resources and protocols
70	Author contributions
71	Acknowledgements
72	Author's note
73	References
74	
75	
76	
77	<b>Supplemental file S1:</b> Fruit body and spore collection
78	<b>Supplemental file S2:</b> Isolation
79	<b>Supplemental file S3:</b> Culture conditions and storage
80	<b>Supplementary file S4:</b> NaOH extraction
81	<b>Supplementary file S5:</b> CTAB DNA/RNA co-extraction
82	<b>Supplementary file S6:</b> High-molecular weight DNA extraction
83	<b>Supplemental file S7:</b> Host- <i>Suillus</i> bioassays
84	<b>Supplemental Table S1:</b> Host- <i>Suillus</i> associations and genome-sequenced species
85	
86	
87	
88	
89	
90	
91	
92	
93	
94	
95	
96	
97	
98	
99	
100	
101	
102	
103	
104	
105	
106	
107	
108	
109	

110 **Summary**

111

112 Research on mycorrhizal symbiosis has been slowed by a lack of established study systems. To  
113 address this challenge, we have been developing *Suillus*, a widespread ecologically and  
114 economically relevant fungal genus primarily associated with the plant family Pinaceae, into a  
115 model system for studying ectomycorrhizal associations. Over the last decade, we have  
116 compiled extensive genomic resources, culture libraries, a phenotype database, and protocols  
117 for manipulating *Suillus* fungi with and without their tree partners. Our efforts have already  
118 resulted in a large number of publicly available genomes, transcriptomes, and respective  
119 annotations, as well as advances in our understanding of mycorrhizal partner specificity and  
120 host communication, fungal and plant nutrition, environmental adaptation, soil nutrient cycling,  
121 interspecific competition, and biological invasions. Here, we highlight the most significant  
122 recent findings enabled by *Suillus*, present a suite of protocols for working with the genus, and  
123 discuss how *Suillus* is emerging as an important model to elucidate the ecology and evolution  
124 of ectomycorrhizal interactions.

125

126

127 **I. Introduction**

128

129 The ectomycorrhizal (ECM) symbiosis formed between plant roots and soil fungi is characterized  
130 by the obligate exchange of plant derived carbohydrates for fungal scavenged nutrients (Smith &  
131 Read, 2010). These cross-kingdom mutualisms provide both direct and indirect impacts on plant  
132 growth, ranging from increased mineral nutrient access to improved abiotic and biotic stress  
133 tolerance (Smith & Read, 2010; Branco *et al.*, 2022). These benefits have long been recognized as  
134 critical to the health of both natural and managed forest systems, with ECM fungal inoculation  
135 regularly employed by the nursery and forestry industries to enhance and protect seedling growth  
136 (Torres & Honrubia, 1994; Cripps & R., 2011; Mateos *et al.*, 2017). ECM fungi also offer a  
137 multitude of ecosystem services, playing fundamental roles in the functioning and maintenance of  
138 community composition, water relations, and global carbon and nutrient cycling (Smith & Read,  
139 2010; Branco *et al.*, 2022). In recent years, these attributes have attracted increased interest from  
140 diverse fields including ecosystem ecology, evolutionary biology, biogeochemistry, global change  
141 biology, and sustainability studies.

142

143 Despite the recognition of their importance and the growing interest in ECM fungi, there are still  
144 significant gaps in our understanding of how ECM fungi function and interact with their plant  
145 partners, microbial communities, and environments. The cryptic belowground nature of ECM fungi,  
146 coupled to high species diversity, variable tractability under laboratory conditions, lack of  
147 standardized protocols, as well as technical challenges associated with the isolation, sequencing,  
148 and manipulation of host-associated dikaryotic species, have hindered the advancement of the field.  
149 Many questions remain unanswered, including how ECM fungi evolve at different biological scales,  
150 the mechanisms facilitating interactions between ECM fungi and their hosts, and how ECM  
151 communities will be affected by environmental change (Plett *et al.*, 2024; Dauphin & Peter, 2024).

152

153 While recent technological advances, such as the ability to generate and analyze extensive genomic  
154 datasets, promise to improve our ability to tackle some of the aforementioned questions, our  
155 understanding of ECM mutualism remains limited due a scarcity of well-developed model systems.  
156 Establishing ECM models that are diverse, ecologically relevant, and amenable to experimentation  
157 and *in vitro* manipulation is critical to advancing mycorrhizal research. The establishment of models  
158 allows the scientific community to build capacity around common systems, increasing the breadth  
159 and utility of core methodologies, improving reproducibility by using a common set of strains and  
160 procedures, and creating discourse and momentum around novel findings and approaches.

161

162 Model systems are often defined at the species level, with each system offering unique advantages  
163 and limitations. Mycologists have built capacity around the development of systems such as  
164 *Coprinopsis cinerea* for studying fruit body development (Kamada *et al.*, 2010; Plaza *et al.*, 2014),

165 *Schizosaccharomyces pombe* for probing fundamental molecular and cellular biology (Hoffman *et al.*, 2015), *Candida albicans* for investigating human fungal pathogenesis (Kabir *et al.*, 2012),  
166 *Neurospora crassa* for understanding gene silencing and circadian rhythms (Selker *et al.*, 1987;  
167 Roche *et al.*, 2014), and *Saccharomyces cerevisiae* for elucidating biological processes from  
168 neurodegenerative disorders to biological aging (Miller-Fleming *et al.*, 2008; Murakami &  
169 Kaeberlein, 2009; Karathia *et al.*, 2011). The development of these systems has resulted in the  
170 creation of numerous tools, and invaluable insights into eukaryotic biology. The concerted effort  
171 towards developing ECM model systems for fungal ecological and evolutionary research will foster  
172 rapid tool development and the critical mass necessary to cultivate effective collaborations and  
173 propel the field of mycorrhizal research into the future.

174  
175  
176 Several ECM models are in development including *Laccaria bicolor*, *Hebeloma cylindrosporum*,  
177 and *Pisolithus tinctorius*. These species have produced fundamental insights into the ecological,  
178 physiological, and molecular mechanisms of ECM-host interactions (Cairney & Chambers, 1997;  
179 Marmeisse *et al.*, 2004; Martin *et al.*, 2008). To date, these taxa have been primarily used as species-  
180 level models, but are rapidly accumulating genomic resources at the genus level. As of October  
181 2023, MycoCosm web portal (<https://mycocosm.jgi.doe.gov/>) now includes two species of *Laccaria*  
182 (including 15 genomes of *L. bicolor*), two species of *Hebeloma*, and 10 species of *Pisolithus*. Other  
183 ECM fungal genera quickly accumulating sequenced genomes include *Tuber* (10 species) and  
184 *Lactarius* (12 species). The choice of which ECM lineages should be targeted for model  
185 development should take into consideration diversity and tractability, trait variation, reproducibility,  
186 accessibility and availability, ecological relevance, conservation/invasion status, applicability and  
187 extensibility to other systems, as well as community interest and momentum. Fungi in the genus  
188 *Suillus* fulfill many of these considerations, including laboratory and field tractability, high  
189 ecological relevance, species and trait diversity, accessibility and abundance, and an engaged and  
190 rapidly growing research community. To this end, we have been developing *Suillus* into a genus-  
191 level system to characterize the ecology and evolution of ECM fungal symbioses.

192  
193 *Suillus* is a speciose genus with most species displaying strong partner specificity with members of  
194 the plant family Pinaceae (Kretzer *et al.*, 1996; Nguyen *et al.*, 2016b). *Suillus* produces abundant  
195 fruitbodies, and are some of the most frequently encountered ECM mushrooms in northern  
196 temperate and boreal forest systems. The genus includes pioneer species that are instrumental for  
197 forest establishment across both natural and managed systems, where they have facilitated both the  
198 success of forestry plantations, and extensive ecological damage as invasive host-symbiont pairs  
199 encroach into introduced ranges and disrupt local communities (Policelli *et al.*, 2019). In addition,  
200 the genus includes both stress-tolerant and stress-sensitive species, enabling investigation into  
201 environmental adaptation of both ECM fungi and their associated plant partners (Colpaert *et al.*,  
202 2011; Bazzicalupo *et al.*, 2020; Zhang *et al.*, 2021). Finally, relative to other ECM fungi, *Suillus* is  
203 highly tractable to laboratory manipulation, including but not limited to, the ability to be isolated  
204 axenically and grown apart from their hosts with specific nutritional supplementation, long term *in*  
205 *vitro* culture viability, and the ability to colonize host plants via both long-lived reactive spores or  
206 mycelial extension (Nguyen *et al.*, 2012).

207  
208 Here, we present *Suillus* as an emerging model genus for ECM fungal ecology and evolution. Our  
209 goals are 1) to provide a primer on *Suillus* that synthesizes our current understanding of the biology  
210 and eco-evolutionary theory relevant to this unique genus, and 2) to standardize and disseminate the  
211 protocols developed to study *Suillus* both in the field and the laboratory. This primer is not intended  
212 to be systematic literature review, but rather a broad introduction to the genus and its potential to  
213 serve as a model system. Specifically, we introduce the vision and goals of the International *Suillus*  
214 Consortium (a working group of over 20 independent laboratories collectively dedicated to the  
215 development of the *Suillus* system, <http://www2.hawaii.edu/~nn33/suillus>), consolidate the latest  
216 findings in *Suillus* biology, ecology, and evolution, and compile molecular and experimental  
217 protocols. Sections II-VII cover the development of *Suillus* as a model genus, its taxonomy and  
218 evolution, significant ecological interactions, abiotic environmental adaptations, the biology and  
219 implications of introductions and invasions, and a look forward at outstanding questions, current  
220 challenges, and concentrations areas for system development. Section VIII outlines a series of  
221 protocols for working with the genus which we have included as a series of standalone

222 supplementary documents.

223

224

## 225 **II. *Suillus* as a model genus**

226

### 227 **II.i: The power of genus-level model systems**

228 Unlike model systems that are based on individual species, the strength of the *Suillus* system  
229 lies at the genus level. There are multiple benefits to leveraging genus-level systems. First, they  
230 provide the biological and ecological diversity needed to ask questions and test hypotheses  
231 across broad scales, from the individual to the ecosystem level. Second, they offer a  
232 comparative framework that allows researchers to investigate eco-evolutionary questions about  
233 speciation, adaptation, and how closely related taxa respond to environmental perturbation.  
234 Third, they allow the generalization of research findings beyond individual species to identify  
235 commonalities, trends, and general biological principles that apply to close relatives as well as  
236 other organisms occupying the same guild, lifestyle or ecological niche. Finally, there are  
237 experimental and research community advantages, such as the development of shared tools that  
238 can be used by multiple research groups with minimal protocol optimization. Having shared  
239 genomic and computational resources also allows different research teams to tackle diverse  
240 questions and minimize experimental overlap while increasing collaborative opportunities. The  
241 benefits of using model clades (at the genus level or above) for ecological and evolutionary  
242 research are well recognized in other systems, driving the development of model genera such  
243 as the bivalve *Mytilus* for ecotoxicology and climate change research (Leopold *et al.*, 2019;  
244 Ribeiro *et al.*, 2019), *Mimulus* plants for studying adaptation and speciation (Wu *et al.*, 2008;  
245 Twyford *et al.*, 2015) and *Anolis* lizards for studying adaptive radiation and ecomorphology  
246 (Sanger & Kircher, 2017).

247

248

### 249 **II.ii: *Suillus* is an excellent model for ECM ecology and evolution**

250 The genus *Suillus* is diverse (Fig. 1) and displays a range of host associations and a gradient of  
251 partner specificity responses, possesses unique traits with high ecological relevance, and is both  
252 accessible and tractable, making it an ideal model for studying ECM ecology and evolution.  
253 Generally, members of the genus have strong partner specificity with the plants in the family  
254 Pinaceae (Kretzer *et al.*, 1996), making it a favorite target of fungal partner specificity research.  
255 In natural environments *Suillus* typically associates with three of the four subfamilies of the  
256 Pinaceae (Pinoideae, Laricoieae, and Piceoideae), allowing researchers to study host-symbiont  
257 associations across different taxonomic levels, including among host genera, subgenera, and  
258 species. *Suillus* also facilitates the study of host switching (Lofgren *et al.*, 2021; Zhang *et al.*,  
259 2022), both between typical hosts and outside of these host groups, thanks to several species  
260 that defy the canonical partner specificity patterns demonstrated by most species (Lofgren *et al.*,  
261 2018; Pérez-Pazos *et al.*, 2021).

262

263 *Suillus* contains species that demonstrate a variety of unique ecological attributes: they occur  
264 as pioneers (Peay *et al.*, 2012) and as introduced species (Thompson *et al.*, 2022), demonstrate  
265 variable stress tolerance (Branco *et al.*, 2022; Erlandson *et al.*, 2022), and associate with hosts  
266 across different life stages, ranging from seedlings to mature forests (Rineau *et al.*, 2016).  
267 *Suillus* serve as effective symbionts that assist in host seedling establishment and development  
268 (Jenkins *et al.*, 2018). Further, *Suillus* can act as hosts themselves for a diversity of fungi and  
269 bacteria, including related fungi in the genera *Gomphidius* and *Chroogomphus* (Olsson *et al.*,  
270 2000) (Fig. 1 O-P) and bacteria that both associate with living hyphae and mycorrhizas (Izumi  
271 *et al.*, 2006; Timonen & Hurek, 2011), and actively decompose dead mycelia (Maillard *et al.*,  
272 2022, 2023). Together, these traits offer exciting opportunities to help solve a broad range of  
273 ecological and evolutionary questions.

274

275 *Suillus* is also an accessible and tractable genus that is amenable for study by a diverse array of  
276 research specialties. Species in the genus tend to fruit abundantly, are commonly encountered,

277 and are relatively easy to identify in the field. While many ECM taxa are difficult or impossible  
278 to culture independently from their hosts (e.g., *Cortinarius*, *Russula*, *Lactarius*, *Tomentella*)  
279 (Palmer, 1971; Brundrett *et al.*, 1996; Nygren *et al.*, 2007), obtaining *in vitro* axenic cultures of  
280 *Suillus* is a straightforward process across the genus. Cultures of *Suillus* can be grown on solid  
281 media, in liquid culture, or somewhere in between (for example, on glass beads partially  
282 submerged in liquid media) and are amenable to long-term storage. *Suillus* mushrooms produce  
283 ample long-lived spores, and mycorrhization can be carried out rapidly (in as little as one  
284 month) using either spore-based or mycelial-based inoculation, providing flexibility for either  
285 high-throughput colonization or the ability to control the provenance of specific strains. The  
286 ability to generate mycelial biomass quickly in culture allows for manipulative studies (such as  
287 nutrient use and stress tolerance) either alone, with a plant partner, or with third-party  
288 community members such as other fungi or bacteria. *Suillus* species have relatively small  
289 genomes (~50 Mbp) that are comparatively easy to sequence and assemble. Through  
290 collaborations with the Joint Genome Institute, the International *Suillus* Consortium research  
291 teams have sequenced and assembled 46 draft genomes to date, spanning the taxonomic diversity  
292 of the genus. Sequencing efforts have also resulted in over 300 hundred shallow-depth genomes  
293 for particular species of interest such as *S. luteus*, *S. brevipes*, and *S. salmonicolor*  
294 (*cothurnatus*). The depth and breadth of sequencing across the genus provides genomic tools  
295 for the community to ask questions that can span the spectrum of systematics, ecology, and  
296 evolution. These traits have enabled the development of many *Suillus* optimized protocols  
297 (outlined in section VIII) and make the genus ideal for the development of manipulative and -  
298 omics based approaches, such as functional-omics (both *in vitro* and *in situ*), and gene editing  
299 (discussed in section VII).

300

### 301 **II.iii: The SuilluScope database**

302 While genus-level models are powerful systems for addressing questions that span ecological  
303 gradients, linking response variables back to genomic variability remains a grand challenge for  
304 most biological systems. Conducting genotype-to-phenotype analysis would be greatly aided  
305 by coupling genomic resources to the generation of diverse, reproducible, phenotypic data, and  
306 the broad availability of this data to the community. The *Suillus* Genome Strain Culture  
307 Collection constitutes a living library of fungal isolates used for whole-genome sequencing  
308 projects. To help the community take full advantage of both the genomic resources available  
309 for *Suillus*, and the phenotypic and genotypic diversity across the genus, we present a new  
310 interactive *Suillus* database, SuilluScope (v1.0Beta) available at [www.SuilluScope.com](http://www.SuilluScope.com). The  
311 initial release of the database contains phenotype information on growth rate, optimal growth  
312 temperatures, and culture images on multiple media types for isolates in the *Suillus* Genome  
313 Strain Culture Collection. The goal of the database is to act as a centralized and accessible  
314 community repository and analysis platform for phenotype assays conducted on the same set  
315 of genome-sequenced strains. This resource will allow the community to identify optimal  
316 growth conditions for specific strains, and predict and interpret response variables across the  
317 genus for downstream analysis. Subsequent releases will include additional phenotypic  
318 variables, and new genome strains will be added as they become available. The *Suillus* Genome  
319 Strain Culture Collection has been integrated into the USDA ARS NRRL culture collection and  
320 all strains are available to the community without restriction. Strain accession numbers and  
321 detailed isolate information are available as a living document in the SuilluScope database, and  
322 will be updated as new assays are conducted, and new genome strains are integrated into the  
323 collection.

324

325 One example currently offered by the database is the selection of optimal growth conditions for  
326 specific species and strains. Many species of *Suillus* exhibit symptoms of stress on rich media  
327 including slower growth and increased pigment production (Fig. 2 A-P). This is a particularly  
328 important consideration for preparing material for sequencing and molecular work, as  
329 pigmentation is often associated with decreased extraction efficiency. For this reason, rich  
330 media preparations such as Modified Melin-Norkrans (MMN) and Hagem's (see details in  
331 section VIII) are often prepared with a 50% reduction in carbon (glucose, malt extract, or yeast

332 extract depending on the formulation, Supporting Information S3). However, total carbon  
333 content is not the only determinant of pigment production, and individual species (or strains)  
334 often display unique requirements for pigment reduction, such as varying the availability of  
335 organic and inorganic nitrogen, media pH, or temperature. Media type has additional wide-  
336 ranging effects on culture phenotype, including growth rate, topology, and the production of  
337 aerial hypha (Fig. 2 A-H). Similarly, while most species of *Suillus* will sustain growth at  
338 temperatures between 10-30 °C, growth rates vary among *Suillus* species, and optimal growth  
339 temperatures for a given species can be identified using the database. For example, maximum  
340 growth rates peak between 19 and 33 days after media transfer and vary between 29mm<sup>2</sup>/day  
341 (*S. spraguei* EM44) to 137mm<sup>2</sup>/day (*S. quiescens* FC197) when grown at room temperature  
342 (Fig. 3).

343  
344  
345

### 346 **III. Taxonomy and evolution**

347

#### 348 **III.i: Taxonomy and species divergence**

349 In 1821 S.F. Gray first proposed splitting *Suillus* from the genus *Boletus*, which Linnaeus had  
350 originally defined to comprise all pored fungi (Murrill, 1909). The genus *Suillus* was formally  
351 sanctioned by Elias Magnus Fries (1821-[1832]), but was not widely accepted until the 1950s  
352 (Dahlberg & Finlay, 1999). Perhaps due to the distinct morphological characters typically  
353 associated with *Suillus* mushrooms, the genus *Suillus* Gray has undergone surprisingly little  
354 revision over the years outside of new species additions and occasional species splitting, often  
355 associated with the recognition of taxa occurring in distinct geographic regions. To date, there  
356 are 112 species of *Suillus* recognized as valid in the Catalog of Life Database  
357 (<https://www.catalogueoflife.org/>), a number we expect to increase over the coming decade,  
358 particularly as new taxa are characterized from understudied areas such as Asia and the Indian  
359 subcontinent (Nguyen *et al.*, 2016b). The *Suillus* phylogeny is broadly divided into 3-5 major  
360 phylogenetic sections, and grouped into corresponding subgenera, including *Boletinus*,  
361 *Spectabilis*, *Larigini*, *Douglasii*, and *Suillus* (Zhang *et al.*, 2022). However, this intrageneric  
362 classification has not been formally recognized and the exact number of subgenera remains  
363 controversial.

364

365 Current classification schemes place *Suillus* in the order Boletales, in the suborder Suillineae, which  
366 contains the genera *Truncocollumella*, *Rhizopogon*, *Gomphidius*, *Chroogomphus*, and the  
367 monotypic genus *Psiloboletinus* (Besl & Bresinsky, 1997; Binder & Hibbett, 2006; Wu *et al.*,  
368 2020). The Suillineae are colloquially referred to as suilloid fungi, but represent multiple families  
369 including the Suillaceae (*Suillus*, *Truncocolumella*, and *Psiloboletinus*), Rhizopogonaceae  
370 (*Rhizopogon*), and the Gomphidiaceae (*Gomphidius* and *Chroogomphus*). The Suillineae are  
371 thought to be the first clade in the Boletales to have independently evolved the ECM lifestyle (Sato  
372 & Toju, 2019), a trait consistent across all families in the Suillineae, except the Gomphidiaceae  
373 which contains at least some mycotrophic species (Fig. 1 O-P).

374

375 *Suillus* diverged from other suilloid fungi between 40.2 and 71.1 MYA (Lepage *et al.*, 1997;  
376 Zhang *et al.*, 2022) (Fig. 4). Within *Suillus*, the earliest diverging clades are associated with the  
377 hosts *Larix* and *Pseudotsuga*. The most significant diversification in the genus occurred during  
378 the switch onto *Pinus* and it is within this *Pinus*-associated lineage that we find evidence for  
379 the active divergence of species complexes. However, the pattern of host switching within  
380 *Pinus* (especially between the host subgenera *Pinus* and *Strobus*) is convoluted and not yet fully  
381 resolved (Zhang *et al.*, 2022). We find notable cases of ongoing evolution associated with host-  
382 switching across the genus, both in clades that are actively diverging and in those that appear  
383 relatively stable. For example, *S. brunnescens* diverged from within the apparently stable *S.*  
384 *luteus* clade, concurrently switching from subgenus *Pinus* to subgenus *Strobus*.

385

386 The three most notable actively diverging clades in *Suillus* are the /albivelatus, /flavidus and  
387 /placidus species complexes (Nguyen *et al.*, 2016b). Within these clades, similar morphological  
388 forms are often difficult to distinguish among the members within each complex. For example,  
389 the /albivelatus clade contains a group of Western North American species (*S. albivelatus*, *S.*  
390 *pseudobrevipes*, *S. volcanalis*, and *S. watsatchicus*) that are difficult to tell apart  
391 morphologically. Likewise, the members of the large-pored species within the /flavidus clade  
392 (*S. umbonatus*, *S. megaporinus*, *S. flavidus*, and *S. helenae*) are morphologically similar.  
393 Conversely, the /placidus clade contains a group of species (*S. placidus*, *S. subalpinus*, *S.*  
394 *anomalus* and *S. punctatipes*) which, although morphologically distinct, are not (yet) supported  
395 as monophyletic group (Nguyen *et al.*, 2016b). Among these clades are species such as *S.*  
396 *punctatipes*, which has the ability to associate with alternative hosts such as *Picea* and *Abies*  
397 (Pérez-Pazos *et al.*, 2021), although it should be noted that species outside of these clades such  
398 as *S. glandulosus* and *S. subaureus* also have the ability to associate with alternative hosts. The  
399 active evolution within (and outside of) these species complexes provide novel information that  
400 can be leveraged to understand the molecular and evolutionary mechanisms of host association  
401 and host-switching among plant-associated fungi.

### 402 403 **III.ii: Atypical morphological features**

404 Secotioid and gasteroid fruit bodies of *Suillus*-like fungi have occasionally been observed and  
405 were once classified as belonging to the genus *Gastrosuillus*. However, *Gastrosuillus* was  
406 dissolved after molecular evidence showed that these fungi were polyphyletic in *Suillus* (Baura  
407 *et al.*, 1992; Kretzer & Bruns, 1997). Transitions in fruit body form can happen rapidly and are  
408 likely controlled by a small number of genes (Bruns *et al.*, 1989). It is currently unknown  
409 whether the examples of secotioid/gasteroid *Suillus* found to date represent ongoing speciation  
410 or variants of known species. Such variants could be the result of spontaneous genetic  
411 mutations, malformations resulting from environmental stress during development, or  
412 ecological plasticity in fruit body form in response to environmental conditions. These  
413 scenarios are not mutually exclusive, and it may well be the case that while some  
414 secotioid/gasteroid fruit bodies are aberrations (as is likely the case for the secotioid examples  
415 of *S. grevillei* (Kretzer & Bruns, 1997)), others have undergone speciation and are yet to be  
416 typified, or have previously been misclassified. This is likely the case for *Rhopalogaster* cf.  
417 *transversarius*, a morphologically distinct secotioid species currently considered a member of  
418 the *Rhizopogonaceae*, but likely to be reclassified in *Suillus*, after molecular studies have found  
419 it to be nested deeply in the genus (Smith *et al.*, 2018). As the rapid evolution of truffle-like  
420 basidiocarps is thought to be adaptive in low moisture availability (as seen in the sister genus  
421 *Rhizopogon*) (Bruns *et al.*, 1989), secotioid/gasteroid *Suillus* represent an intriguing system for  
422 future work given the ongoing interest into *Suillus* drought responses (Wang *et al.*, 2021;  
423 Erlandson *et al.*, 2022; Castaño *et al.*, 2023).

424  
425 Another interesting morphological feature demonstrated by some species of *Suillus* and  
426 *Rhizopogon* is the formation of tubercles. These structures are characterized by dense clusters  
427 of mycorrhized root tips surrounded in a hyphal rind (Randall & Grand, 1985). The biological  
428 and ecological function of tubercles is poorly understood, but as they bear resemblance to the  
429 bacterial nodules formed by *Rhizobium* and *Bradyrhizobium* on legumes or *Frankia* on *Alnus*,  
430 it has long been thought that they may play a role in nitrogen fixation. Paul *et al.* (2007) used  
431 an acetylene reduction assay to assess potential N-fixation in *S. tomentosus* tubercles and found  
432 evidence of nitrogenase activity up to 25098.8 nmol C<sub>2</sub>H<sub>4</sub> g<sup>-1</sup> per tubercle over a 24 hour  
433 period, suggesting that the ability to form tubercles could play a significant role in N-fixation.  
434 Follow up work from the same group later identified multiple species of diazotrophic bacteria  
435 in association with the interior tissue of *S. tomentosus* tubercles (Paul *et al.*, 2012), as well as  
436 isotopic evidence for N-transfer to the host tree (Chapman & Paul, 2012). Future work is needed  
437 to assess the diversity and frequency of tuberculate forming suilloid fungi, better characterize  
438 the bacterial communities associated with these structures, and quantify the contribution of  
439 tubercle-associated nitrogenase activity to host nitrogen budgets.

440

441

442 **III.iii: *Suillus* vs. *Rhizopogon***

443 The genus *Rhizopogon* is closely related to *Suillus* and shares many of the traits that make  
444 *Suillus* an attractive model for studying ecology and evolution. Like *Suillus*, *Rhizopogon*  
445 species are obligately ECM, easily isolated, and speciose, currently including 225 recognized  
446 species which are divided into approximately five subgenera (Grubisha *et al.*, 2002).  
447 *Rhizopogon* species exhibit high to moderate partner specificity and typically associate with  
448 host trees in the Pinaceae, primarily *Pinus* and *Pseudotsuga* (Molina *et al.*, 1997). Rarely,  
449 primary host associations have also been observed with *Larix* (Miyamoto *et al.*, 2019), and  
450 secondary colonization is occasionally reported on non-Pinaceae hosts such as *Arbutus* and  
451 *Arctostaphylos* (Massicotte *et al.*, 1994; Kennedy *et al.*, 2012). Unlike the complex history of  
452 host-switching and reversion observed in *Suillus*, partner specificity in *Rhizopogon* is primarily  
453 reflected by fungal subgeneric classification (e.g., the subgenus *Villosuli* associates exclusively  
454 with *Pseudotsuga*, whereas the subgenus *Amylopogon* has a more generalist host range  
455 (Grubisha *et al.*, 2002)).

456

457 Unlike *Suillus*, *Rhizopogon* exclusively forms truffle-like hypogeous fruitbodies, an adaptation  
458 which likely evolved from pileate-stipitate (mushroom) ancestors (Bruns *et al.*, 1989; Sánchez-  
459 García *et al.*, 2020). Like most hypogeous fungi, *Rhizopogon* spp. largely depend on animals  
460 excavate their fruit bodies and distribute their spores (Bradshaw *et al.*, 2022). The clear  
461 differentiation in fruit body forms between *Rhizopogon* and *Suillus* has been cited as evidence  
462 of strong selection against intermediate forms, putatively driven by differences in dispersal  
463 mechanisms (Bruns *et al.*, 1989). Like *Suillus*, *Rhizopogon* species form long-lived spore-banks  
464 in the soil (Shemesh *et al.*, 2023) and are often associated with early successional colonization,  
465 including seedlings and invasion fronts (Policelli *et al.*, 2019), but often demonstrate poor  
466 competitive ability outside of these scenarios (Bruns *et al.*, 2002).

467

468 The hypogeous and cryptic nature of *Rhizopogon* fruitbodies makes them more difficult to survey  
469 and sample compared to mushroom-forming fungi like *Suillus*. Similarly, quantifying the  
470 percent of individual root tips colonized by *Rhizopogon* for field or *in-vitro* assays can be  
471 challenging due to their typically coralloid morphology (Kennedy & Peay, 2007). For  
472 fruitbodies, morphological differentiation of *Rhizopogon* is notoriously complicated by a lack  
473 of variable diagnostic characters, which confound field identification, and contribute to the  
474 instability of species concepts developed in the pre-molecular era (Bidartondo & Bruns, 2002;  
475 Bubrisky & Kennedy, 2014; Koukol *et al.*, 2022; Karlsen-Ayala *et al.*, 2022). Although  
476 *Rhizopogon* was long thought to be sister to *Suillus*, multiple molecular studies have identified  
477 fungi in the family Gomphidiaceae as more closely related to *Suillus* (Grubisha *et al.*, 2002;  
478 Wu *et al.*, 2020). However, given that the Gomphidiaceae are putatively nonmycorrhizal, and  
479 the Suillaceae genera *Truncocolumella* and *Psiloboletinus* while ECM, together contain only  
480 five recognized species (Wu *et al.*, 2020), these genera have a distinct, but limited, use as  
481 comparative genus-level systems. In contrast, work in *Rhizopogon* has contributed a rich  
482 independent body of research to our understanding of ECM ecology and evolution and is the  
483 most ecologically relevant genus for comparative studies with *Suillus*. In addition to many  
484 similarities, *Rhizopogon* also demonstrates traits which are unique from *Suillus*, including a  
485 high propensity to be parasitized by mycoheterotrophic plants (Bidartondo & Bruns, 2002; Dowie  
486 *et al.*, 2012; Grubisha *et al.*, 2014), and the potential for not only heat tolerance, but growth  
487 stimulation in response to fire (Bruns *et al.*, 2019). As we continue to build *Suillus* into a model  
488 genus for ecology and evolution, the concurrent development of other groups with contrasting  
489 ecological and life history strategies, including but not limited to other suilloid fungi, will  
490 complement our understanding of the mechanisms and ecological drivers at play in ECM forest  
491 systems.

492

493

494 **III.iv: Population genetics**

495 Because genetic divergence can often be detected before morphological divergence, deep  
496 sampling of individual species can provide a wealth of information into microevolutionary  
497 processes. To this end, member labs of International *Suillus* Consortium have focused efforts  
498 on continental-to-global sampling of *S. luteus* (the type species of *Suillus*), as well as *S.*  
499 *brevipes*, *S. salmonicolor* (*cothurnatus*), *S. pungens*, and *S. quiescens*. The deep sampling of  
500 these species across both native and introduced ranges has facilitated our ability to define  
501 population structure, characterize diversification events, and identify genetic bottlenecks  
502 associated with global introductions, as well as other microevolutionary processes that can  
503 drive divergence (Branco *et al.*, 2017; Pildain *et al.*, 2021; Ke *et al.*, 2023).

504  
505 Population genetics, the study of intraspecific variation and the forces that result in evolutionary  
506 changes in species over time (Wills, 2007), is a critical tool for understanding fungal evolution.  
507 However, the cryptic nature of fungi makes population studies challenging. Specifically,  
508 accurately counting and characterizing fungal individuals, as well as delimiting and describing  
509 fungal populations and their respective allele pools is notably more complicated in fungi than  
510 for plants or animals. *Suillus* is no exception to this generalization, and technical challenges to  
511 studying microevolutionary changes have historically complicated our understanding of how  
512 populations are structured and distributed, the patterns and processes that drive gene flow, and  
513 how environmental change affects genetic diversity.

514  
515 Despite these challenges, *Suillus* fungi have been targeted for population genetic analysis since  
516 the early 1990s, capitalizing on the new availability and affordability of DNA sequencing.  
517 These studies contributed key insights about genet size (Dahlberg & Stenlid, 1994; Bonello *et*  
518 *al.*, 1998; Zhou *et al.*, 1999), population genetic structure (Dahlberg & Stenlid, 1990; Muller *et*  
519 *al.*, 2007; Burchhardt *et al.*, 2011), and gene flow (Zhou *et al.*, 2001; Burchhardt *et al.*, 2011)  
520 but were limited in scope regarding the number of individuals, the number of populations, and  
521 the number of markers used for population differentiation. Recent population-level sequencing  
522 efforts have facilitated major contributions to the understanding of fungal population genetics,  
523 including key insights relevant to both *Suillus* evolution and to ECM fungal evolution in  
524 general. Many of these advances have been facilitated by the ability to sequence whole genomes  
525 and recent efforts to assemble and annotate high quality reference strains. For example, work  
526 on *S. brevipes* showed that the species is highly outcrossing and includes several distinct  
527 populations across North America that often display restricted gene flow at the regional scale  
528 (Branco *et al.*, 2015, 2017). These studies also revealed that climate and soil chemistry play  
529 important roles in *S. brevipes* population structure. In contrast, the European species *S. luteus*  
530 displays an absence of population structure at small spatial scales, including across highly  
531 contrasting environments (Bazzicalupo *et al.*, 2020).

532

533

### 534 **III.v: Mating systems**

535 Whether fungi produce progeny by selfing, inbreeding, or outcrossing impacts the evolutionary  
536 trajectory of species by influencing genetic diversity, deviation from Hardy–Weinberg  
537 equilibrium, and the relative level of linkage disequilibrium (Nieuwenhuis *et al.*, 2013). The  
538 breeding systems of basidiomycetes are categorized based on the number of loci controlling  
539 mating type, where two functional MAT loci are present in tetrapolar breeding systems, one  
540 functional MAT locus is present in bipolar breeding systems, and a lack of any self-  
541 incompatibility loci results in homothallic (self-fertile) breeding systems (Nieuwenhuis *et al.*,  
542 2013). Generally, basidiomycete mating compatibility and mating types are governed by two  
543 self-incompatibility loci, the HD MAT locus and P/R MAT locus (Heitman *et al.*, 2013). Two  
544 or more self-incompatible allele types at each locus dictate the mating types and the mating  
545 compatibility between gametes (Wang & Mitchell-Olds, 2017). Genomic analysis has shown  
546 that in *Suillus*, the P/R MAT locus contains three pheromone receptors and three or more  
547 pheromone precursors (Mujic *et al.*, 2017) and the HD MAT locus contains a pair of  
548 homeodomain proteins flanked by genes encoding mitochondrial intermediate peptidase (MIP),

549 as well as a gene known as the ‘beta-flanking gene’; a syntenic arrangement that appears to be  
550 conserved across most basidiomycetes (Ke *et al.*, 2023).

551

552 In *Suillus*, breeding systems can be studied by both genomic methods and by constructing  
553 crosses. Crossing experiments require monokaryotic strains, most frequently produced via the  
554 germination of basidiospores (discussed in section VIII). Crossing experiments in *Suillus*  
555 indicate that *S. luteus*, *S. variegatus*, *S. granulatus*, and *S. bovinus* all have bipolar, multi-allelic  
556 mating systems (Fries & Neumann, 1990; Fries & Sun, 1992; Fries, 1994). In these species,  
557 incompatibility between individuals is rare, implying that there are a high number of mating  
558 types maintained at the population level. The highest diversity of mating types recorded to date  
559 are in the species *S. luteus* where eight mating types were recovered from four individuals, and  
560 all mating types were found to be unique (Fries & Neumann, 1990). Although these studies  
561 have added significantly to our understanding of ECM fungal genetics, *Suillus* presents several  
562 challenges to constructing and confirming laboratory crosses. For example, clamp connections  
563 are not a consistent trait among dikaryotic strains of *Suillus*, and therefore the presence of  
564 clamps cannot be used as a proxy for monokaryon compatibility as in other groups (Fries &  
565 Neumann, 1990). Further, the secondarily homothallic binucleate basidiospores produced by  
566 some species of *Suillus* complicate both the determination of breeding systems and crossing  
567 experiments (Jacobson & Miller, 1994). Although the incidence of binucleate spore production  
568 by *Suillus* appears to be low (Horton, 2006), the population frequency and fecundity of these  
569 binucleate spores is not yet resolved.

570

571 Insights into the evolution of breeding systems in the genus have also been supported by  
572 genome-based analyses. In agreement with studies employing laboratory crosses, comparative  
573 genomic based analysis of pheromone receptors suggests that *S. brevipes* has a bipolar breeding  
574 system (Mujic *et al.*, 2017). Allelic analysis of *Suillus* genomes suggest that many species have  
575 a long-term multi-allelic state, and a trans-specific polymorphism at the HD MAT locus (Ke *et*  
576 *al.*, 2023). This result is in agreement with the high diversity of mating types previously  
577 identified in crossing experiments, supporting HD MAT as the primary locus contributing to  
578 the bipolar mating system observed in *Suillus*. The closely related species *Rhizopogon*  
579 *rubescens*, also has a bipolar, multi-allelic breeding system (Kawai *et al.*, 2008). This fact, in  
580 combination with the recognition of the wide-spread bipolar, multi-allelic mating systems  
581 observed in multiple species of *Suillus*, suggests that a bipolar multi-allelic breeding system is  
582 likely the ancestral state for this group.

583

584 Population-level research suggests that *Suillus* generally displays high levels of outcrossing.  
585 For example, studies using molecular markers in *S. pungens* (Bonello *et al.*, 1998) and *S.*  
586 *spraguei* (Burchhardt *et al.*, 2011) have highlighted an excesses of heterozygosity at the  
587 population level, suggesting that outcrossing is more predominant than random mating.  
588 Similarly, population genomic analysis of *S. brevipes* has shown a sharp decay in linkage  
589 disequilibrium, which also indicates a high level of outcrossing (Branco *et al.*, 2017). The  
590 mechanisms associated with the extensive outcrossing observed in *Suillus* have not been tested  
591 experimentally but may be the result of self-incompatibility and heterosis. Crossing studies  
592 have found mixed results for the presence of a functional somatic incompatibility system, which  
593 may be present only in certain species such as *S. luteus* (Fries & Neumann, 1990). Identifying  
594 the molecular mechanisms responsible for the extremely high levels of outcrossing observed in  
595 the genus, including characterizing the identity and frequency of self-incompatibility systems,  
596 is a key future direction for understanding *Suillus* genetics.

597

### 598 **III.vi: Genomics**

599 While light-coverage genome sequencing of several key taxa has added important insights  
600 about population-level processes in *Suillus*, light-coverage sequencing is not adequate to  
601 capture nuanced information about gene and genome diversity. To this end, member labs of the  
602 International *Suillus* Consortium have spearheaded large-scale initiatives to sequence and  
603 annotate a phylogenetically and ecologically diverse set of *Suillus* fungi, using deep-coverage,

604 whole genome sequencing of both DNA and RNA, with an emphasis on long-read technologies.  
605 Many of these efforts have been carried out in collaboration with the US Department of  
606 Energy's Joint Genome Institute, resulting in a genome set comprising more species from a  
607 single genus than any other ECM group to date. Currently, this genome set includes 46 *Suillus*  
608 genomes, representing approximately 40 distinct species. Many of these genomes have been  
609 released publicly on the MycoCosm Web Portal at <https://mycocosm.jgi.doe.gov> (Grigoriev *et*  
610 *al.*, 2014), facilitating community access and further work into *Suillus* biology, such as the  
611 ability to compare and contrast *Suillus* with other fungal groups (Lofgren *et al.*, 2019; Miyauchi  
612 *et al.*, 2020; Wu *et al.*, 2022).

613

614 Comparative genomic studies using these genomes have revealed the extent of genetic diversity  
615 in the genus, identifying a dynamic genomic landscape involving many gene families, including  
616 a higher number of rapid gene family expansions and a higher number of rapid gene family  
617 contractions than any other ECM group investigated to date (Lofgren *et al.*, 2021). These  
618 analyses have identified several genomic features that appear to be characteristic of the genus,  
619 including a diversity of gene clusters involved in secondary metabolism, and genes involved in  
620 the processing of reactive oxygen species (Lofgren *et al.*, 2021). The ecological roles that these  
621 genes play in *Suillus* biology is an active area of research. While secondary metabolic clusters  
622 likely function in a variety of biological processes in *Suillus*, the genus-wide expansion of  
623 terpene and non-ribosomal peptide synthetase (NRPS)-like clusters hint at roles in interspecific  
624 interactions such as microbial and host communication and are ideal targets for investigating  
625 the remarkable partner specificity displayed by the genus. Recently, LC/MS based untargeted  
626 metabolomic analysis of three genome-sequenced strains of *Suillus* identified hundreds of  
627 unique secondary metabolites, including 116 putative terpenes (Mudbhari *et al.*, 2023). The  
628 most abundant terpene classes were identified as terpene lactones, sesquiterpenes and di-  
629 terpenes, including several compounds induced exclusively when grown in co-culture with  
630 other species of *Suillus*. These results further suggest that these metabolites play a role in  
631 interactions between *Suillus* and other community members, but further work is needed to  
632 characterize these compounds and their ecological functions. Similarly, the processing of  
633 reactive oxygen species is known to be an important part of host colonization (Liao *et al.*, 2016),  
634 as well as mitigating host-mediated stress responses such as the oxidative bursts characteristic  
635 of plant drought stress (Zou *et al.*, 2021), or the unique ability of *Suillus* to withstand high levels  
636 of heavy metal exposure and to facilitate metal sequestration (Bazzicalupo *et al.*, 2020). Future  
637 work linking the genomic landscape of *Suillus* to the functions displayed by the genus will be  
638 key to developing a deeper mechanistic understanding of *Suillus* biology.

639

640 Efforts to leverage the benefits of both deep-coverage whole genome sequencing and  
641 population-level sampling are underway, with projects designed to characterize genome  
642 variation in target species such as *S. quiescens*, *S. luteus*, *S. brevipes*, *S. salmonicolor*, *S.*  
643 *pungens*, and *S. tomentosus*. We expect these projects to yield novel insights into intraspecific  
644 variation in *Suillus*, including genome structure, noncoding and repeat regions, and improved  
645 frameworks for linking genotypes to phenotypes. For example, comparative genomic analysis  
646 including 12 strains of *S. brevipes* found that rDNA copy number varied from 72 to 156 within  
647 the species, only slightly less variation than was estimated across the genus as a whole (44 to  
648 198 copies) (Lofgren *et al.*, 2019). Leveraging functional-omics technologies on genome-  
649 sequenced strains will produce further insights into natural variation, and the ecology of *Suillus*-  
650 community interactions. Continuing efforts to produce more reference-quality *Suillus* genomes,  
651 the increased sampling breadth of target species for pangenomic analysis, improved pipelines  
652 for *de novo* assembly and annotation of unique strains, and an increase in the use of long read  
653 sequencing, will all propel and direct future insights into ECM fungal genomics.

654

655

656

## 657 **IV. Ecological interactions**

658

659 The unique intrinsic traits discussed above have enabled many studies on ecological  
660 interactions between *Suillus* and their surrounding communities. There has long been a focus  
661 on *Suillus* in the context of ECM partner specificity as well as many investigations of *Suillus*  
662 species as both competitors of other fungi and symbionts mediating plant responses to changing  
663 environmental conditions. Below we outline some of the major findings that have emerged  
664 from these ecologically focused studies.

665

#### 666 **IV.i: Partner specificity**

667 Generally, co-occurring ECM host species exhibit large overlaps in ECM species associations,  
668 suggesting that partner specificity is low for most ECM taxa (Peay *et al.*, 2015; Peay, 2016;  
669 Arraiano-Castilho *et al.*, 2021). While recent work has highlighted the possibility that cryptic  
670 species may inflate estimates of host generalism by artificially combining taxa that are actually  
671 distinct, a phenomena known as cryptic specificity (Sato *et al.*, 2007; Wilson *et al.*, 2017; van  
672 der Linde *et al.*, 2018), how often this occurs in practice is unclear. Importantly, partner  
673 specificity is not a binary classification of specialists versus generalists, but rather a spectrum  
674 of interactions, ranging from fungal associations with a single host species to those spanning  
675 hundreds of host species. While it is likely that some genera such as *Laccaria*, *Thelephora* and  
676 *Russula*, are largely composed host generalists (Roy *et al.*, 2008; Smith *et al.*, 2009; Cho *et al.*,  
677 2021), others such as *Lactarius* and *Strobilomyces* probably exhibit gradients of partner  
678 specificity or host preference (unequal colonization when multiple host taxa are available) that  
679 warrant more nuance than they have been historically afforded (Looney *et al.*, 2018; Tang *et al.*,  
680 2021).

681

682 Early on, it was recognized that a small number of ECM fungal groups exhibited remarkable  
683 and conspicuously high partner specificity with certain host taxa. These specialists include  
684 *Alnicola*, *Alpova* and other basidiomycetes associated with the actinorrhizal host genus *Alnus*  
685 (Tedersoo *et al.*, 2009; Rochet *et al.*, 2011), the genus *Leccinum* with various species of broad-  
686 leaf trees (Den Bakker *et al.*, 2004), and the genera *Suillus* and *Rhizopogon* with the plant family  
687 Pinaceae (Dahlberg & Finlay, 1999). Each of the major host genera (*Pseudotsuga*, *Larix*, and  
688 *Pinus*) are associated with a suite of *Suillus* species that appear to form mycorrhizas almost  
689 exclusively with that host (Table S1, Nguyen *et al.*, 2016b). While partner specificity in ECM  
690 fungi is most well recognized at the level of host genus (Molina *et al.*, 1992), *Suillus* species  
691 tend to be specific to host subgenera, particularly within the genus *Pinus*. The most detailed  
692 account of this level of specificity was presented by Liao *et al.* (Liao *et al.*, 2016), who  
693 combined seedling bioassays with metatranscriptomics to critically assess specific plant-fungus  
694 associations. The authors compared colonization and gene expression patterns across 5 species  
695 of *Suillus* and 10 species of *Pinus* and showed that several *Suillus* species, including *S.*  
696 *americanus*, *S. granulatus*, and *S. spraguei* only formed compatible associations with *Pinus*  
697 species in a single subgenus. This subgenus-scale specificity appeared to be associated with the  
698 expression of similar genes and metabolic pathways previously identified in pathogenic plant-  
699 fungal interactions.

700

701 While close pairings of particular *Suillus* species with specific hosts is the most common  
702 framing of this ECM symbiosis, recent studies indicate that patterns of *Suillus* partner  
703 specificity are strongly mediated by ecological context. Within the plant host family Pinaceae,  
704 multiple tree genera have long been considered non-hosts of *Suillus*, including *Picea* (a genus  
705 in the subfamily Piceoideae nested within a clade of known *Suillus* host genera; (Gernandt *et al.*,  
706 2016) as well as *Abies* and *Tsuga* (in the subfamily Abietoideae, a clade that does not contain  
707 *Suillus* host genera) (Fig. 4). Based on sporadic field reports of *Suillus* mushrooms being  
708 collected in *Picea* or *Abies/Tsuga* forests with no known *Suillus* host genera present (Doudrick  
709 *et al.*, 2011), Pérez-Pazos *et al.* (Pérez-Pazos *et al.*, 2021) combined field root tip sampling and  
710 experimental seedling bioassays to assess the partner specificity of two *Suillus* species; *S.*  
711 *glandulosus* and *S. punctatipes*. The authors found that *Picea*, *Abies*, and *Tsuga* root tips were

712 colonized by *Suillus* species in the field and that both *Suillus* species were capable of colonizing  
713 alternative hosts in the laboratory. Importantly, however, the colonization of alternative hosts  
714 only occurred when a known host (a species of *Pinus* or *Pseudotsuga*) was co-planted in the  
715 same container. This outcome matched prior studies of ECM partner specificity (Massicotte *et*  
716 *al.*, 1994), where it was shown that certain ‘primary’ host species are required to trigger spore  
717 germination, but that additional ‘secondary’ hosts could be colonized once the ECM fungus  
718 was growing as mycelium. Even greater host phylogenetic breadth was shown for *S. subaureus*  
719 by Lofgren *et al.* (2018), who demonstrated through a combination of field sampling and  
720 seedling bioassays that this species was capable of colonizing both *Pinus* and *Quercus* hosts.  
721 This unusual association with *Quercus* (an angiosperm) appeared to be associated with the local  
722 extirpation of *Pinus* hosts during intensive logging decades earlier, resulting in a unique  
723 ecological legacy effect. Differences in host range between primary and secondary colonization  
724 such as those identified for *S. glandulosus*, *S. punctatipes* and *S. subaureus* may seem like  
725 curious exceptions but could have significant consequences for ecosystems. Secondary  
726 colonization is likely responsible for reports of *Suillus* on other non-canonical hosts, including  
727 *Betula* (Nara, 2006), *Cupressus*, and *Ceratonia* (Avital *et al.*, 2022), and could influence the  
728 succession dynamics of plant communities, particularly if carbon transfer from secondary hosts  
729 is sufficient to produce fruit bodies or if resources are transferred via exudates or common  
730 mycelial networks. As plant host ranges shift due to a changing climate and human  
731 introductions, understanding ECM partner specificity is key to predicting the establishment of  
732 these interactions in novel ecological contexts.

733

734

#### 735 **IV.ii: Competition**

736 Like all ECM fungal species, members of the genus *Suillus* compete to colonize host root tips  
737 as their main source of carbon in natural settings (Baldrian, 2009). Prior to molecular  
738 identification of ECM root tips, the relatively high annual production of sporocarps by most  
739 *Suillus* species suggested this genus was a dominant part of the ECM fungal community in  
740 many conifer forests (Dahlberg & Finlay, 1999). However, as molecular-based root tip surveys  
741 proliferated in the 1990s and 2000s, it became apparent that most *Suillus* species colonize only  
742 a limited number of root tips compared to other ECM genera such as *Tomentella* and *Russula*  
743 (Gardes & Bruns, 1996; Jonsson *et al.*, 1999; Peay *et al.*, 2007). However, in early successional  
744 forests, non-native environments, and post-fire environments *Suillus* often deviate from this  
745 pattern and dominate root tip communities (Visser, 1995; Chapela *et al.*, 2001; Hayward *et al.*,  
746 2015; Miyamoto *et al.*, 2021; Thompson *et al.*, 2022). This pattern of root-tip dominance can  
747 also be seen in some specialized habitats such as hummocks present in *Larix* systems  
748 (accumulations of organic matter that exist directly above the waterline), where *Suillus* can  
749 occupy a significantly higher proportion of the ECM fungal community (Kennedy *et al.*, 2018).  
750 In general, however, *Suillus* is rarely the most dominant genus colonizing ECM root tips in  
751 conifer forest soils where it is endemic.

752

753 Given the aforementioned pattern, one might speculate that *Suillus* species are relatively weak  
754 competitors for host root tips or easily excluded by other ECM fungi during fungal community  
755 succession. Support for a limited competitive ability has been observed in multiple seedling  
756 bioassay studies involving *Suillus* and other ECM fungi. For example, Kennedy *et al.* (2011)  
757 found that *S. pungens* was outcompeted by two species of *Rhizopogon* when colonizing *Pinus*  
758 *muricata* seedlings from mycelia. Similar results were found by Moeller and Peay (2016) who  
759 demonstrated that *S. pungens* was also outcompeted by both *Thelephora terrestris* and *R.*  
760 *occidentalis* when colonizing *P. muricata* seedlings from spore. Collectively, these  
761 experimental findings, in conjunction with root tip abundance patterns in field studies, suggest  
762 that *Suillus* species may indeed be relatively weak competitors when faced with either spore-  
763 or mycelial-based competition from other ECM fungi. It should be noted, however, that  
764 *Rhizopogon* and *Thelephora* are aggressive colonizers of seedlings, and studies across a much  
765 wider range of *Suillus* and competitor species are needed to have greater confidence in this  
766 pattern. The pattern of low root tip abundance, increased plant health, and large and abundant

767 fruit bodies suggests an effective carbon trading balance where only a limited number of root-  
768 tips are needed to achieve advantageous outcomes for both hosts and symbionts.

769  
770 Perhaps to compensate for limited interspecific competitive ability, members of *Suillus* appear  
771 to have specialized in reproductive traits that favor rapid and abundant dispersal (Policelli *et*  
772 *al.*, 2019), which likely favors their ability to colonize root tips that may have few other ECM  
773 fungi present. This has been best evidenced in the quantitative estimates of ECM fungal spore  
774 dispersal at increasing distances from established forests, which typically reveal that *Suillus*  
775 represent an increasing proportion of ECM spores at the farthest distances (Peay *et al.*, 2012;  
776 Thompson *et al.*, 2022). Similar tradeoffs in competitive versus colonization ability have been  
777 routinely observed in plant communities (Tilman, 1994) and *Suillus* is arguably one of the best  
778 documented examples of this phenomenon within ECM fungi (Peay *et al.*, 2007).

779  
780 Studies focused on congeneric competition among *Suillus* species have also revealed interesting  
781 ecological trends. Working with a set of species that are commonly associated with *Pinus*  
782 *strobus*, Kennedy *et al.* (2020) found a clear competitive hierarchy of *S. americanus* > *S.*  
783 *subaureus* > *S. spraguei*. The outcome of the competitive interactions appeared to be largely  
784 the result of differences in spore germination timing, with *S. americanus* being the first and  
785 most consistent to colonize *P. strobus* seedling root tips, followed by *S. subaureus*, and then *S.*  
786 *spraguei*. The early colonization by one species precluding the establishment of subsequent  
787 colonizers, also known as a priority effect, has been observed consistently in a variety of other  
788 ECM study systems as well (Kennedy, 2010; Bogar & Kennedy, 2013). Interestingly, *S.*  
789 *subaureus*, is a much rarer species than either *S. americanus* or *S. spraguei*, which suggests that  
790 other factors besides competitive ability are also likely important in shaping abundance in ECM  
791 fungal communities.

792  
793 To investigate the extent to which ECM host plants may influence ECM fungal competitive  
794 outcomes, Bogar *et al.* (2019) conducted a pair of seedling bioassays using the *Larix*-associated  
795 species; *S. clintonianus*, *S. grisellus*, and *S. spectabilis*. The authors used a split root design in  
796 which competing *Suillus* species were spatially isolated when colonizing a shared host  
797 (eliminating the role of direct fungal-fungal interactions). They found that colonization by two  
798 of the three species was not changed by a second species being present on a different portion  
799 of the root system, but that the colonization by *S. spectabilis* was significantly lower if a second  
800 *Suillus* species was present. This suggested that plants can differentiate amongst different  
801 *Suillus* species and can potentially mediate root tip colonization to privilege preferred species.  
802 Further evidence for differential plant investment was found in the second bioassay, in which  
803 seedlings allocated greater amounts of carbon to the portion of the split-root system that  
804 provided the greatest amount of nitrogen. Intriguingly, seedlings independently colonized by *S.*  
805 *spectabilis* were not significantly lower in needle nitrogen content than those colonized by *S.*  
806 *grisellus* or *S. clintonianus*, so exactly why the *Larix* host plants were preferentially colonized  
807 by the latter two species was not clear. In natural habitats, *S. spectabilis* can only be found with  
808 mature host plants, whereas the other two species can be found with younger hosts, suggesting  
809 a host age-dependent context of interactions. This study, along with additional studies  
810 manipulating soil nitrogen levels (Bogar *et al.*, 2022), highlight that competitive outcomes  
811 among ECM fungi may be influenced by host plant tendencies toward specific taxa.

812

#### 813 **IV.iii: Soil nutrient cycling**

814 Similar to other ECM fungi, *Suillus* are thought to have a limited capacity to decompose  
815 complex plant cell wall polymers (Kohler *et al.*, 2015). As the vast majority of ECM fungi  
816 evolved from saprotrophic ancestors (Tedersoo *et al.*, 2010) many have retained the ability to  
817 decompose less complex polymers such as protein and chitin, which could account for observed  
818 patterns of organic matter turnover (up to 30%) in introduced *Pinus* plantations (Chapela *et al.*,  
819 2001). Protein decomposition appears to be dependent on forest age, with *Suillus* species in  
820 younger forests having lower protein decomposition activity (Rineau *et al.*, 2016), which could  
821 be advantageous given that more mature forests contain higher levels of organic nitrogen.

822 Genomic and functional assays have demonstrated that *S. luteus* also uses endochitinases to  
823 decompose complex chitin polymers (Maillard *et al.*, 2023). Initial work suggests that in low  
824 carbon soils, *S. salmonicolor* (*cothurnatus*), like some other ECM species, can prime  
825 decomposition by saprotrophic fungi (Bhatnagar *et al.*, 2021). These pieces of evidence indicate  
826 that although limited in their capabilities to decompose complex plant cell wall polymers,  
827 *Suillus* can substantially contribute to decomposition of soil organic matter, and thus nutrient  
828 cycling, especially nitrogen cycling in soil.

829  
830 *Suillus* themselves can be consumed, decomposed, and returned to the soil as labile sources of  
831 carbon and nutrients. Members of this this genus often produce large aggregations of fruit  
832 bodies, estimated to be upward of 63.9kg ha<sup>-1</sup> year<sup>-1</sup> in dry weight for *S. variegatus* (Ohenoja  
833 & Koistinen, 1984), to one metric ton ha<sup>-1</sup> year<sup>-1</sup> dry weight for *S. luteus* (Hedger, 1986; Chapela  
834 *et al.*, 2001). Notably, these estimates did not include hyphae and rhizomorphs, which likely  
835 represent a substantial additional pool of tissue available for decomposition. These high nutrient  
836 resources are rapidly consumed by insects, in particular phorid and mycetophilid flies (Bruns,  
837 1984). The fruit bodies (Gohar *et al.*, 2022) and mycelium (Brabcová *et al.*, 2016) can also be  
838 decomposed by bacteria and fungi that appear to have specificity towards these fungal-derived  
839 resources (Nguyen, 2023). Enabled by the ready abundance of fruit bodies and the easy culture  
840 of *Suillus* mycelium Fernandez *et al.* (2019) and Maillard *et al.* (2022) used *S. grisellus* to  
841 conduct manipulative experiments characterizing the contributions of *Suillus* necromass to soil  
842 nutrient cycling as well as identifying the microbial members involved in this process. This  
843 area of research is currently experiencing a renaissance as modern tools and techniques provide  
844 exciting opportunities to dissect the molecular mechanisms and community interactions that  
845 facilitate the degradation of fungal necromass (Fernandez & Kennedy, 2018; Kennedy &  
846 Maillard, 2023) and will be influential in elucidating the contributions of *Suillus* (and other  
847 high-biomass producing fungal species) to soil nutrient cycling.

848  
849  
850

## 851 **V. Environmental (abiotic) adaptation**

852  
853 Studies in the genus *Suillus* have enabled significant advances in our knowledge of fungal  
854 abiotic adaptation. In this section, we outline recent progress in our understanding of how the  
855 genus impacts, and is impacted, by environmental variation. Specifically, we highlight studies  
856 assessing climate change, ionic radiation, metal pollution, and enzymatic activity and  
857 degradation. The availability of public protocols and genomic resources are expected to  
858 catalyze further research in these areas and assist in the development of new strategies to  
859 remediate and recover degraded habitats.

860

### 861 **V.i: Climate change**

862 Climate change, characterized by elevated temperatures and shifts in patterns of precipitation,  
863 is increasingly impacting forest ecosystems, and influencing symbiotic relationships between  
864 fungi and their host plants (Baldrian *et al.*, 2023). Recent research has identified *Suillus* as a  
865 key player in enhancing host plant resilience to environmental stress, and advanced our  
866 understanding of how ECM fungi respond and adapt to changing environmental conditions  
867 (Malcolm *et al.*, 2008; Branco *et al.*, 2017; Li *et al.*, 2021; Qi & Yin, 2022; Hou *et al.*, 2022;  
868 Erlandson *et al.*, 2022).

869

870 As a result of shifting climatic conditions, there has been a global increase in the frequency and  
871 severity of drought in recent years (Dai, 2012), posing a significant threat to the resilience of  
872 forest ecosystems (Brodribb *et al.*, 2020). *Suillus* exhibit considerable drought tolerance, which  
873 potentially sustain symbiotic relationships and enhance plant stress tolerance under drought  
874 conditions. The long-distance rhizomorphs of *Suillus* are well known to aid water transport  
875 (Duddridge *et al.*, 1980), which may help host plants to maintain positive water relations during

876 times of abiotic stress. Support for this hypothesis was recently demonstrated by Castaño et al.  
877 (2023), who found that *Pinus pinaster* seedlings exposed to experimental drought in Spain were  
878 enriched in ECM fungal taxa with long-distance rhizomorphs. In particular, they found that *S.*  
879 *variegatus* was an indicator species of drought (i.e. was more relatively abundant under the  
880 drought treatment), suggesting that some species of *Suillus* may be better able to tolerate low  
881 water availability than other ECM taxa. At the same time, this study also reported that the  
882 abundance of *S. bovinus* declined significantly in the drought treatment, indicating that not all  
883 *Suillus* species are drought resistant. This result is consistent with the findings of Erlandson et  
884 al. (2022), who showed that long-term water reduction dramatically decreased root tip  
885 colonization by *S. pungens* on *Pinus muricata* seedlings. This study found that while short-term  
886 water reduction led to major changes in the expression of functional genes associated with water  
887 acquisition, gene expression under long-term water stress was very similar to that of well-  
888 watered controls. This suggests that ECM fungi may be capable of significant acclimation under  
889 prolonged stress, which may aid in maintaining symbiotic functioning under challenging  
890 environmental conditions (Palumbi et al., 2014). Erlandson et al. (2022) further found that  
891 drought-exposed *S. pungens* was able to maintain cellular integrity by up-regulating genes  
892 related to fungal cell wall synthesis.

893

894 Significantly, *Suillus* may pass drought tolerance benefits onto their host trees. Several studies  
895 have found evidence for this effect and have proposed several potential mechanisms including  
896 osmoregulation, oxidative protection, and the preferential relocation of plant resources. For  
897 example, osmoprotective compounds such as proline and trehalose have been shown to  
898 contribute to plant osmoregulation by providing protection against drought-induced damage,  
899 and appear to be upregulated in *Suillus* during drought stress (Li et al., 2021; Erlandson et al.,  
900 2022). Osmoregulation may also occur by increasing host water uptake via the induction of  
901 water-channel proteins such as aquaporins (Lee et al., 2010; Xu & Zwiazek, 2020). While  
902 multiple studies have indicated that *S. tomentosus* can improve host tolerance to salt exposure  
903 via aquaporin-mediated water transport (Lee et al., 2010; Calvo-Polanco & Zwiazek, 2011),  
904 researchers have been unable to provide direct evidence that *Suillus* regulates host-water  
905 movement under drought conditions. As such, future research is needed to confirm the  
906 induction of water-channel proteins as a drought tolerance mechanism in the genus.

907

908 A second proposed mechanism for *Suillus* mediated drought tolerance is via oxidative defense.  
909 Li et al. (2021) found that roots colonized by *S. placidus* appeared to trigger antioxidant defense  
910 against drought-induced oxidative stress through the activation of antioxidant enzymes. These  
911 results suggest that some species of *Suillus* may be capable of counteracting the negative  
912 impacts of drought-induced oxidative damage, thereby promoting plant health and survival.

913

914 Finally, *Suillus* may mediate drought responses by altering the physiology of their hosts. For  
915 example, Qi and Yin (2023) found that under drought conditions *S. luteus* altered host root traits  
916 including diameter, length, surface area, and volume, improving water and nutrient uptake  
917 efficiency. A similar pattern was detected in *S. pungens*-inoculated mycorrhizal roots which  
918 responded to drought by up-regulating genes involved in energy production, signal  
919 transduction, metabolism, and the transport of amino acids, lipids, and carbohydrates  
920 (Erlandson et al., 2022). Despite the inhibitory effects of drought stress on photosynthesis, plant  
921 development, and growth, *Suillus* appears to assist in mitigating these effects by enhancing  
922 photosynthetic performance and increasing stomatal area and density (Li et al., 2021; Qi & Yin,  
923 2022). Taken together, these results suggest that *Suillus* may confer drought tolerance to host  
924 plants by increasing the production of photosynthetically-fixed carbon that is then preferentially  
925 reallocated to belowground structures, which in turn support mycorrhizal root growth and  
926 hyphal water uptake.

927

928

929 **V.ii: Ionizing Radiation**

930 High levels of radiation are a major concern for human health and ecosystem function. *Suillus*  
931 has provided important insights into how ionizing radiation affects the fungal kingdom.  
932 Irradiation results from radionuclides, unstable isotopes that release radiation when they break  
933 down to more stable elements. While these elements occur naturally, they are primarily  
934 concerning when produced artificially and subsequently enter the environment in high  
935 concentrations, typically via industrial waste or the detonation of nuclear weapons (Hu *et al.*,  
936 2010). Exposure to high levels of ionizing radiation has wide ranging negative effects, including  
937 genetic and epigenetic changes, decreased reproductive ability, abnormal morphology, and  
938 mortality (Hinton *et al.*, 2007).

939

940 The devastating accidents that occurred in Chernobyl (Ukraine) in 1986 and in Fukushima  
941 (Japan) in 2011 heavily impacted vast areas with radio-active fallout. These events have been  
942 widely studied to determine how chronic irradiation affects species and ecosystems. The genus  
943 *Pinus* is extremely sensitive to irradiation (Geras'kin *et al.*, 2008), and the nuclear accidents  
944 that occurred in Japan and Ukraine affected large swaths of pine-dominated forest (Vinichuk &  
945 Johanson, 2003; Pumpanen *et al.*, 2016; Bondarenko *et al.*, 2023). Studies from these areas  
946 suggest that some fungi, including *Suillus*, are relatively tolerant towards ionizing radiation  
947 (Dighton *et al.*, 2008). These fungi act as <sup>137</sup>Cs radionuclide sinks, effectively removing them  
948 from the soil. This process may influence the cycling of radionucleotides across soil  
949 compartments, as both *S. luteus* and *S. variegatus* accumulate high levels of <sup>137</sup>Cs in fruit body  
950 tissue (Nikolova *et al.*, 1997; Ronda *et al.*, 2022). Similarly, Vinichuk and Johanson (2003)  
951 found that an astounding 19% of total soil <sup>137</sup>Cs were transported into *S. variegatus* mycelium.  
952 Importantly, these effects appeared to be species-specific, as not all *Suillus* species accumulated  
953 <sup>137</sup>Cs to the same extent. The ability of some *Suillus* species to act in the bioaccumulation of  
954 <sup>137</sup>Cs raises the question of whether *Suillus* impacts <sup>137</sup>Cs uptake in host trees. In plants, <sup>137</sup>Cs  
955 uptake is mediated by K transporters and K channels (Zhu & Smolders, 2000). In *Suillus*  
956 however, the concentration of <sup>137</sup>Cs is not correlated with K concentration (Dighton *et al.*, 2008;  
957 Vinichuk *et al.*, 2011), suggesting that in fungi, <sup>137</sup>Cs uptake may be independent of K pathways.  
958 Interestingly, there seem to be differences in irradiation effects and molecular responses among  
959 fungal species that differently impact growth, melanin production, and ROS scavenging  
960 enzymes. For example, Kothamasi (2019) found that while multiple *S. luteus* isolates displayed  
961 reduced biomass upon irradiation exposure, *S. bovinus* was unaffected. Similarly, while all  
962 *Suillus* isolates showed melanin accumulation in response to irradiation, the response of ROS  
963 decomposing enzymes varied both between and within species.

964

965 Precisely how irradiation affects ECM fungi, if ECM fungi are involved in plant tolerance to  
966 ionizing radiation, and if so, by what mechanisms, are active areas of research (Ladeyn *et al.*,  
967 2008). Given the extensive areas affected by irradiation and the abundance and relevance of  
968 *Suillus* species in these ecosystems, *Suillus* is an ideal model to investigate the impacts of  
969 radiation, and a prime candidate for the development of bioremediation systems. To this end,  
970 members of the International *Suillus* Consortium are currently building a *Suillus* culture  
971 collection from Fukushima radionuclide-contaminated soil to investigate both the effects of  
972 irradiation in ECM systems and develop urgently needed bioremediation strategies.

973

974 **V.iii: Metal tolerance**

975 Soil metal contamination through anthropogenic activity can be extremely detrimental to  
976 ecosystems, negatively impacting metabolism, growth and reproduction (Branco *et al.*, 2022).  
977 Metal tolerance in *Suillus* is a widely documented phenomenon and several species are  
978 routinely found in metal contaminated soils, including sites near mining and metal

979 manufacturing facilities, decommissioned smelters and mills, and sites with heavy ore  
980 enrichment (Colpaert & van Assche, 1987; Leski *et al.*, 1995; Blaudez *et al.*, 2000; Adriaensen  
981 *et al.*, 2005). This finding has driven multiple studies focused on characterizing metal tolerant  
982 isolates with the goal of assessing whether the presence of metal contamination has selected for  
983 metal tolerant traits and led to local adaptation (Colpaert & van Assche, 1987; Bazzicalupo *et*  
984 *al.*, 2020).

985  
986 Early studies demonstrated that metal tolerance in *Suillus* not only varies across species, but  
987 also across isolates within the same species. For example, *S. luteus*, *S. variegatus* and *S.*  
988 *granulatus* display interspecific variation in tolerance to Cd, Pb, Sb and Zn (Hartley *et al.*,  
989 1997), while both *S. luteus* and *S. bovinus* display intraspecific variation in tolerance to Cd, Cu,  
990 Ni and Zn (Colpaert & Van Assche, 1992; Blaudez *et al.*, 2000; Adriaensen *et al.*, 2005).  
991 Interestingly, although *Suillus* isolated from metal contaminated sites generally display higher  
992 levels of metal tolerance, metal sensitive isolates have also been collected from metal  
993 contaminated soils (Blaudez *et al.*, 2000; Fomina *et al.*, 2005; Adriaensen *et al.*, 2005). Fomina  
994 *et al.* (2005) proposed that this observation could be explained by the heterogeneous nature of  
995 soil, which could lead to uneven metal concentrations across a single soil patch, allowing metal  
996 sensitive isolates to persist. However, metal tolerant *Suillus* have also been discovered in non-  
997 contaminated areas (Leski *et al.*, 1995; Hartley *et al.*, 1997; Colpaert *et al.*, 2004), suggesting  
998 that the fitness costs to metal tolerance may be inconsequential, or that there may be benefits  
999 to retaining metal tolerance traits apart from metal exposure. While Colpaert (2004) suggested  
1000 that gene flow could occasionally bring tolerant genes into sensitive populations, genetic  
1001 analysis of *S. luteus* from contaminated and non-contaminated sites did not support the presence  
1002 of separate populations (Bazzicalupo *et al.*, 2020).

1003  
1004 Metal tolerance in *Suillus* appears to result from the ability to maintain metal homeostasis  
1005 through a combination of metal immobilization, exclusion, and detoxification. These  
1006 mechanisms rely on a large number of genes, including transmembrane transporters (which  
1007 move metal into, out of and around the cell), chelators (that bind and immobilize ions both  
1008 within and outside the cell), and antioxidants that counteract the effects of metal toxicity  
1009 (Bazzicalupo *et al.*, 2020; Branco *et al.*, 2022). Analysis of *S. luteus* demonstrated that metal  
1010 tolerance is likely achieved through genetic variants arising from standing genetic variation  
1011 (Bazzicalupo *et al.*, 2020). In a recent study, Smith *et al.* (2023) conducted a transcriptomic  
1012 analysis of Zn-tolerant and Zn-sensitive *S. luteus* isolates exposed to high Zn concentrations  
1013 and confirmed that Zn tolerance in this species is polygenic. This study showed that different  
1014 isolates achieve tolerance through distinct mechanisms and that Zn-tolerant isolates are largely  
1015 constitutively tolerant regardless of metal exposure. However, the authors also identified two  
1016 Zn related genes for which expression was affected by Zn concentration, suggesting that Zn  
1017 tolerance in *S. luteus* also includes an environmental component.

1018  
1019 Studies investigating the genetics and physiology of *Suillus* metal tolerance have demonstrated  
1020 that metal accumulation in cells is a limiting factor for tolerance (Colpaert & van Assche, 1987).  
1021 For example, Colpaert *et al.* (2005) found that Zn uptake is negatively correlated with tolerance  
1022 in *S. luteus*, and that the exclusion of Zn from cells allows tolerant isolates to persist when  
1023 exposed to high Zn concentrations. Cadmium tolerance in *S. luteus* may similarly rely on a  
1024 metal exclusion mechanism, as tolerant isolates have been shown to take up less metal into the  
1025 mycelium than sensitive isolates (Krznaric *et al.*, 2009). Cadmium tolerant *S. luteus* isolates  
1026 have also been shown to express antioxidant transcripts at much lower levels than Cd sensitive  
1027 isolates, suggesting Cd tolerance is grounded in exclusion, and that Cd tolerant *Suillus* likely  
1028 avoid cellular damage by preventing Cd uptake in the first place (Ruytinx *et al.*, 2011).

1029  
1030 There has been substantial progress in illuminating the mechanisms of metal exclusion and  
1031 metal binding in *Suillus*. Multiple transmembrane transporters have been identified including  
1032 Cation Diffusion Facilitator (CDF) family transporters and Zrt-, Irt-Like Protein (ZIP)  
1033 transporters. Whereas CDF transporters are membrane bound proteins that transport a variety

1034 of metals (Kolaj-Robin *et al.*, 2015), ZIP transporters are Zn pumping proteins that are typically  
1035 localized to the plasma membrane. These transporters act to bring external Zn into the cells and  
1036 to draw Zn from vacuolar stores to prevent toxicity (Coninx *et al.*, 2017), and have been  
1037 characterized in *S. bovinus* (Ruytinx *et al.*, 2013). In *S. luteus* there are four CDF proteins  
1038 predicted to be Zn transporters. Two of these (SIZnT1 and SIZnT2), have been functionally  
1039 characterized, and SIZnT1 has been shown to confer Zn tolerance using heterologous  
1040 expression in yeast (Ruytinx *et al.*, 2017). *S. luteus* also has four predicted ZIP transporters,  
1041 two of which have been described. SIZRT1 is a plasma membrane transporter heavily involved  
1042 in maintaining cellular Zn homeostasis (Coninx *et al.*, 2017) and SIZRT2 is localized to the  
1043 plasma membrane and the perinuclear region and also appears to play a role in Zn uptake  
1044 (Coninx *et al.*, 2019).

1045  
1046 Metal binding is also an important mechanism of metal tolerance in *Suillus*. In *S. luteus*, there  
1047 are two known metallothioneins (small proteins that can bind and immobilize metals) involved  
1048 in Cu homeostasis (SIMTa and SIMTb). These genes are upregulated in excess Cu, but not in  
1049 excess Zn or Cd. Heterologous expression of these genes in a Cu sensitive yeast mutant has  
1050 been shown to restore tolerance (Nguyen *et al.*, 2017). Metallothionein homologues with a role  
1051 in Cu homeostasis have also been identified in *S. himalayensis*. These genes were shown to be  
1052 induced by excess Cu, but not by Cd and heterologous expression in yeast confirmed a role in  
1053 Cu tolerance, along with possible roles in Cd and Zn homeostasis (Kalsotra *et al.*, 2018).

1054  
1055 Notably, *Suillus* metal tolerance appears to have significant downstream effects on host plant  
1056 fitness (Adriaensen *et al.*, 2005). When associated with plant roots exposed to high metal  
1057 concentrations, some *Suillus* isolates can reduce metal transfer and accumulation in host plant  
1058 tissues while still efficiently transferring nutrients to the host (Adriaensen *et al.*, 2006; Colpaert,  
1059 2008; Krznanic *et al.*, 2010; Colpaert *et al.*, 2011; Branco *et al.*, 2022). While metal-tolerant  
1060 and metal-sensitive *Suillus* seem to be equally effective mycorrhizal partners regarding nutrient  
1061 uptake and transfer, there are clear differences in plant nutrient uptake and nutrient status when  
1062 exposed to high metal concentrations (Adriaensen *et al.*, 2004; Krznanic *et al.*, 2010). For  
1063 example, Andriaensen *et al.* (2005) demonstrated that inoculation with Cu-tolerant isolates of  
1064 *S. luteus* significantly improved plant phosphorus and nitrogen uptake in Cu-contaminated soil,  
1065 while Cu-sensitive isolates reduced nutrient uptake and transfer. *Suillus* colonization can also  
1066 facilitate metal tolerance indirectly, for example, Liu *et al.* (2020, 2021) demonstrated that *S.*  
1067 *luteus* activates multiple plant stress pathways, enhancing plant fitness during metal exposure.  
1068 Similarly, metal exclusion mechanisms in tolerant *Suillus* isolates have been shown to reduce  
1069 metal transfer to the host (Krupa & Kozdrój, 2004; Zhang *et al.*, 2021). For example, when  
1070 associated with *Pinus* roots under high external Zn, *S. luteus* accumulates Zn in the hyphal  
1071 mantle, preventing Zn accumulation in host tissues (Zhang *et al.*, 2021).

1072  
1073  
1074

## 1075 **VI. Biological introductions and invasions**

1076

1077 The strong ecological linkages between *Suillus* and the Pinaceae help explain the key role of  
1078 the genus in the spread and establishment of Pinaceae into novel habitats. While this ability has  
1079 long been beneficial for the forestry industry, enabling the wide-spread establishment and  
1080 success of tree plantations, it has also facilitated the invasion of introduced host taxa into native  
1081 plant communities around the world, displacing species and disrupting ecosystems (Read,  
1082 1998; Policelli *et al.*, 2019). While many *Suillus* host species have been introduced across the  
1083 world, not all introductions have resulted in biological invasions. The variables determining  
1084 invasion capacity, along with the ecological impacts of *Suillus*-associated host-tree  
1085 establishment in non-native environments are important areas of research.

1086  
1087

### **VI.i: Introduction via forestry**

1088 The genus *Pinus* is native to the northern hemisphere, but has been planted globally for shade,  
1089 shelter and the establishment of softwood plantations. Exotic *Pinus* plantation forests currently  
1090 cover over five million hectares, and their global effects on primary productivity,  
1091 biogeochemistry, and biodiversity represent some of the most widespread and impactful  
1092 introductions on Earth (Simberloff & Von Holle, 1999; Hoeksema *et al.*, 2020).

1093  
1094 Most early attempts to introduce conifers into the southern hemisphere failed due to the lack of  
1095 availability (and knowledge) of suitable ECM fungi (Mikola, 1970). As foresters became aware  
1096 of ECM and their role in forest nutrition, inoculating seedlings with ECM fungi became  
1097 common practice. Historically, the most commonly used inoculum source was transplanted soil  
1098 from established *Pinus* plantations (Marx, 1992; Marx *et al.*, 2002). This practice led to the  
1099 successful establishment of conifer plantations across both the northern and southern  
1100 hemispheres, along with the widespread dissemination of conifer-associated ECM fungi. Early  
1101 on, it was recognized that the ECM species that established themselves in exotic conifer  
1102 plantations represented only a fraction of native ECM biodiversity, and overwhelmingly  
1103 consisted of early colonizers such as *Suillus* and *Rhizopogon* (Theodorou, 1967; Theodorou &  
1104 Bowen, 1973). In areas where conifers have been introduced, this process of ecological filtering  
1105 has resulted in as few as 1-5 species of ECM fungi in New Zealand *Pinus* plantations (Chu-  
1106 Chou & Grace, 1988), and as few as one species (*S. luteus*) in *Pinus* plantations in Chile  
1107 (Hayward *et al.*, 2015). This reduction in diversity is also evident for other introduced members  
1108 of the Pinaceae. For example, *Larix* and *Pseudotsuga* have been widely planted in the southern  
1109 hemisphere, where they exhibit strong partner specificity with *Suillus*, including *S. cavipes* with  
1110 introduced *Larix* and *S. lakei* with *Pseudotsuga* (McNabb, 1968; Moeller *et al.*, 2015; Pietras  
1111 *et al.*, 2018).

1112  
1113  
1114

#### VI.ii: *Suillus*-*Pinaceae* co-invasions

1115 Successful afforestation and introduction of Pineaceae across the southern hemisphere has  
1116 inevitably resulted in the escape of introduced trees that interact with and sometimes displace  
1117 native plant communities (Richardson *et al.*, 1994; Simberloff & Von Holle, 1999; Nuñez *et*  
1118 *al.*, 2009). Burdon and Chilvers (1977) were the first to report the process of escape and  
1119 invasion by wilding conifers. The study detailed *P. radiata* invasion from plantations into  
1120 adjacent *Eucalyptus* forests near Canberra, Australia, resulting in the formation of a new type  
1121 of hybrid forest community. Follow up studies monitoring these forests over the subsequent 14  
1122 years, found that the growth of *P. radiata* was strongly affected by the presence of the other  
1123 plant species (Chilvers & Burdon, 1983; Burdon & Chilvers, 1994), leading to the eventual  
1124 coexistence between *Pinus* and native vegetation. Within Australia, the impact of *Pinus*  
1125 invasions seems to be relatively small with most wilding tree escapes occurring after periodic  
1126 disturbance or fire (Williams & Wardle, 2007; van Etten *et al.*, 2020). This is in contrast to  
1127 *Pinus* introductions on other continents, which have resulted in active invasion fronts,  
1128 tremendous loss of native biodiversity, and habitat alteration. The negative impacts of these  
1129 invasions have been greatest in regions where host trees are able to grow and shade out other  
1130 plant species, such as in the Cape Region of South Africa where *P. radiata* now threatens native  
1131 Fynbos vegetation (Richardson *et al.*, 1994), and in Brazil, Argentina, and Chile, where  
1132 invasive *Pinus* have escaped into native grasslands (Nuñez *et al.*, 2009). To date, efforts to  
1133 eradicate invasive Pineaceae hosts and restore these ecosystems have achieved only limited  
1134 success.

1135 The concept that ECM fungi had a role in facilitating host tree invasions was first suggested by  
1136 Richardson *et al.* (1994) and Read *et al.* (1998), and later confirmed using molecular approaches  
1137 (Nuñez *et al.*, 2009; Dickie *et al.*, 2010; Hynson *et al.*, 2013; Moeller *et al.*, 2015; Horton,  
1138 2017). This pattern of invasion has been termed “Linked Plant-Fungal Co-invasion” (Dickie  
1139 *et al.*, 2017). Today, plantation trees are inoculated with ECM fungi as a matter of course,  
1140 typically using soil-free spore-based methods or mycelial slurries where strain origin can be  
1141 controlled (Marx *et al.*, 2002). Contemporary commercial inoculum is produced using many

1142 ECM taxa, including *Pisolithus tinctorius*, *Laccaria* spp., *Rhizopogon* spp. *Hebeloma* spp., and  
1143 *Suillus* spp. (Marx *et al.*, 2002; Charya & Garg, 2019). Whether due to innate trait-based effects,  
1144 or legacy effects from soil-based inoculation, *Suillus* seem to have a disproportionate invasion  
1145 potential: in almost every study conducted to date on linked plant-fungal co-invasions, *Suillus*  
1146 species have been identified as keystone community members. While tree plantations are the  
1147 best studied source of *Suillus* introductions into non-native ranges, it is important to note that  
1148 ECM fungi can be translocated along with any compatible host tree or introduced intentionally  
1149 or inadvertently with the movement of soil. This underscores the importance of knowing  
1150 mycorrhizal status and the identity of the ECM fungi associated with introduced trees for  
1151 applications such as forest restoration, shelterbelts (windbreaks), and ornamental plantings.

1152 To generate an up to date assessment of *Suillus* introductions, we used the Global Database of  
1153 Alien Macrofungi (Monteiro *et al.*, 2020). This database includes 770 ECM distribution records  
1154 originally assembled by Vellinga *et al.* (Vellinga *et al.*, 2009), but is extended to include  
1155 multiple guilds, and studies completed since 2009, for a total of 1,966 observations. We pruned  
1156 this dataset to exclude taxa with uncertain species-level assignments, and those annotated as  
1157 having established from uncertain geographic origins. To identify ECM species, guild was  
1158 assigned across the database using FUNGuild (Nguyen *et al.*, 2016a), retaining only species  
1159 that could be confidently identified as ECM and excluding mixed-guild types, resulting in 946  
1160 observations, spanning 55 genera and 241 species. Out of these 55 ECM genera, *Suillus* made  
1161 up the vast majority of observed introductions, with a total of 193 records, followed by  
1162 *Scleroderma* at 91, and *Rhizopogon* at 72 (Fig. 5A). While the dominance of *Suillus* is striking,  
1163 as noted by Vellinga *et al.* (Vellinga *et al.*, 2009), occurrence records of species such as  
1164 *Scleroderma* and *Rhizopogon* are likely underestimated due to the relative difficulty of locating  
1165 their fruitbodies. These results broaden our understanding of genus-level ECM invasion  
1166 frequency, and emphasize the ongoing prevalence of *Suillus* in ECM introductions as reported  
1167 15 years ago in Vellinga *et al.* (Vellinga *et al.*, 2009). In total, there were 24 species of *Suillus*  
1168 in introduced ranges (Fig. 5B). Largely in agreement with previous studies (Read, 1998;  
1169 Dunstan *et al.*, 1998; Policelli *et al.*, 2019), the most common species were the European *Pinus*  
1170 associated taxa *S. luteus* (44 observations) and *S. granulatus* (34 observations), followed by the  
1171 larch associate *S. grevillei* (18) and Douglas Fir associate *S. lakei* (15). It should be noted that  
1172 the western north American species *S. quiescens* (typified in 2010) is also an important  
1173 introduced species (Policelli *et al.*, 2019) but was absent from the database (likely combined  
1174 with the morphologically similar *S. brevipes*).

1175 The dominance of *Suillus* in ECM introductions is likely the result of both the historical  
1176 frequency of using soil-based inoculations, and the confluence of several traits exhibited by the  
1177 genus. Policelli *et al.* (2019) identified several ecological traits which help to make *Suillus*  
1178 strong facilitators of host tree invasions. These include 1) long-distance dispersal capacity, 2)  
1179 the ability to generate a long-lived resistant spore bank, 3) the establishment of positive biotic  
1180 interactions with mammals, 4) the rapid colonization of host roots, and 5) the long-distance  
1181 exploration type of *Suillus* extraradical mycelium.

1182 Within their exotic range, and in the absence of competition from other fungi, *Suillus* species  
1183 often fruit especially prolifically, further contributing to their proliferation and spread in these  
1184 habitats. Hedger (1986) and Chapela (2001) estimated that *S. luteus* fruit body production in  
1185 introduced Ecuadoran *P. radiata* forests, could amount to over one metric ton of fungal tissue  
1186 per hectare (dry weight): an impressive yield by any measure. These fruit bodies produce  
1187 enormous numbers of basidiospores which are widely dispersed well beyond the forest  
1188 boundary. Once dispersed, *Suillus* spores become part of a long-lived drought and fire-resistant  
1189 soil spore-bank, and are able to wait many years for a wilding *Pinus* seed to land beside them  
1190 (Bruns *et al.*, 2010).

1191 In addition to wind dispersal, mycophagy is recognized as an important dispersal mechanism  
1192 and likely contributes to the establishment of *Suillus* in invasive ranges (Wood *et al.*, 2015;

1193 Caiafa *et al.*, 2021; Elliott *et al.*, 2022). Numerous studies have documented fruit body  
1194 consumption by both native and non-native animals including rodents, marsupials, deer, pigs,  
1195 rabbits and potentially birds, which can translocate spores over great distances and disperse  
1196 them through their excrement (Ashkannejhad & Horton, 2006; Nuñez *et al.*, 2013; Wood *et al.*,  
1197 2015; Horton, 2017; Aguirre *et al.*, 2021; Caiafa *et al.*, 2021; Policelli *et al.*, 2022).

1198 Perhaps the strongest factor linking *Suillus* to host tree invasions is the fact that *Suillus* species  
1199 colonize young host seedlings so readily. Once established, *Suillus* can rapidly spread to nearby  
1200 root systems by the production of the abundant rhizomorphs that are a hallmark of this clade.  
1201 This type of extramatrical mycelium, which Agerer (2001) termed the “long-distance  
1202 exploration-type”, not only enables mycelial-based colonization of neighboring host trees, but  
1203 likely helps to facilitate the success of *Pinus* across exotic ranges by enabling better water  
1204 withdrawal, increased access to resources patches, and increased acquisition of organic nitrogen  
1205 (Hobbie & Agerer, 2009; Koide *et al.*, 2014). Although *Suillus* are often thought of as weak  
1206 competitors, new plantation environments are typically depauperate in ECM symbionts capable  
1207 of colonizing the introduced host trees, resulting in reduced competition (Hedger, 1986;  
1208 Chapela *et al.*, 2001). In addition, novel environments offer a high likelihood of ‘enemy escape’  
1209 from co-evolved pathogens, which may also contribute to establishment across introduced  
1210 ranges (Dickie *et al.*, 2010).

1211

#### 1212 **VI.iii: Downstream effects and unresolved questions**

1213 The strong obligate symbiosis between *Suillus* and the Pinaceae is an example of a positive  
1214 plant-soil feedback. Plant-soil feedbacks are often associated with biological invasions, and  
1215 have numerous downstream effects on ecosystems (Simberloff & Von Holle, 1999; Simberloff,  
1216 2006). *Pinus* plantations are known to greatly alter soil biogeochemistry by increasing soil  
1217 acidity and litter production, and depleting soil carbon (Chapela *et al.*, 2001). This feedback is  
1218 thought to be one of the primary factors preventing effective ecological restoration (Hoeksema  
1219 *et al.*, 2020). Even after host removal, the enzymatic function of soil can be slow to recover  
1220 (Sapsford & Dickie, 2023). As primary colonizers of introduced Pinaceae forests, *Suillus* spp.  
1221 likely contribute to the biogeochemical changes associated with plantation forestry. The precise  
1222 role that *Suillus* fungi play in altering soil chemistry and the mechanisms involved in this  
1223 phenomenon are active areas of research.

1224

1225 The combined impacts of simultaneous co-invasion can have disastrous consequences on native  
1226 biodiversity, a process which has been termed “invasional meltdown” by Simberloff and Von  
1227 Holle (1999). Whether facilitated directly by the fungus, or via host-mediated mechanisms,  
1228 early evidence points toward the capacity of *Suillus* (and also likely *Rhizopogon*) to displace  
1229 native fungal communities at least when the density of introduced hosts is high (Sapsford *et al.*,  
1230 2022; Mujic *et al.*, 2023). In linked plant-fungal co-invasions, it is difficult to uncouple the  
1231 invasive capacity of the fungus from that of the host, complicating the question of whether  
1232 *Suillus* should be treated as invasive species in their own right. Generally, ECM species are  
1233 primarily constrained to their introduced hosts (Vellinga *et al.*, 2009; Policelli *et al.*, 2020).  
1234 While the high partner specificity exhibited by *Suillus* should reduce overall invasion potential  
1235 compared to host generalist species that are more likely to establish on novel hosts after  
1236 introduction, this benefit seems to be overwhelmed by the influence of the early-successional  
1237 life history traits exhibited by *Suillus* (Vlk *et al.*, 2020).

1238

1239 Previous work has shown no phylogenetic signal for informing which species of *Suillus* are  
1240 likely to be introduced (Vellinga *et al.*, 2009), and while phylogenetically diverse *Suillus*  
1241 species are capable of establishing in introduced ranges, occurrence records are dominated by  
1242 a small number of species. Interesting questions remain as to what traits and qualities make  
1243 these species invasive, while the majority of *Suillus* species have never been associated with  
1244 introduction or linked plant-fungal co-invasions.

1245

1246

1247

## VII. Conclusions and future directions

1248

1249

1250

1251

1252

1253

1254

1255

1256

1257

1258

1259

Here, we have shown that *Suillus* fungi play essential roles in natural and managed ecosystems, have greatly added to our understanding of fungal evolution, the ecology of fungal-host and fungal-fungal interactions, the stress responses of ECM fungi to both biotic and abiotic factors, as well as to population genetics, comparative genomics, and invasion biology. *Suillus* holds immense potential to serve as a model genus to advance our understanding of the ecology and evolution of ECM systems. Additionally, the combination of experimental tractability, the availability of experimental protocols, the flexibility of a well-defined genus-level system, a large set of publicly available genomes and respective annotations, and the cultures associated with these genomes, make *Suillus* one of the few well-established ECM fungal models for addressing questions that can scale from genes to ecosystems.

1260

### VII.i: Outstanding questions and research priorities

1261

1262

1263

1264

1265

1266

1267

1268

1269

1270

1271

1272

1273

1274

1275

Research leveraging *Suillus* over the last decade has helped to define and prioritize outstanding questions in the field. First, phylogenetic analysis to define and place taxa from under sampled areas will help to fully delineate taxonomic boundaries and clarify the evolutionary history of host switching and diversification in the genus. While the strength of the *Suillus* system is based at the genus level, selecting target species for deep investigation will allow researchers to leverage both intra- and interspecific comparisons to derive the true impacts of fungal trait variation. Currently, the most well-studied species of *Suillus* is *S. luteus*, driven largely by the fact that it was the first species in the genus to undergo whole genome sequencing (Kohler *et al.*, 2015). However, current efforts to develop other species including *S. brevipes*, *S. quiescens*, *S. salmonicolor*, *S. pungens*, and *S. tomentosus* will soon provide more insight into trait and genome variation and provide a framework for testing specific hypotheses that have so far been difficult to address. These efforts include both population-level sampling, and chromosome-level reference assemblies, enabling a deeper understanding of genomic diversity, and the influence of genome structure on ecological traits.

1276

1277

1278

1279

1280

1281

Another research priority is to define the mechanisms facilitating partner specificity and partner switching in *Suillus*. This line of research will not only enhance our fundamental understanding of these ecological phenomena but also has practical implications for effective ecosystem management and conservation, particularly in the face of climate change-induced shifts in host tree distributions.

1282

1283

1284

1285

1286

1287

1288

1289

Ongoing research efforts using *Suillus* should aim to quantify key aspects of the ECM symbiosis such as realized nutrient trading dynamics. For instance, employing emerging techniques such as quantum dots and new advances in stable isotope tracing will help to address a multitude of both ecological and mechanistic questions related to nutrient cycling. In addition to quantifying carbon and nitrogen trading across species and environmental variables, defining the role and importance of non-canonically traded macro and micro-nutrients should be a priority for gaining a comprehensive understanding of metabolic interactions.

1290

1291

1292

1293

1294

1295

1296

Clarifying the roles and mechanisms of non-nutritional partner benefits, such as stress tolerance to abiotic factors like drought, metal exposure, and ionizing radiation will help researchers to understand how *Suillus* and their hosts will adapt to a rapidly changing world. Understanding these variables will also help to identify the characteristics that contribute to the invasive potential of certain *Suillus* species, thereby enabling researchers to safeguard native ecosystems while supporting forestry practices.

1297

1298

Finally, research on the interactions between *Suillus* and other community members beyond the plant-fungal partnership is also vital for understanding the broader ecological context in which

1299 the ECM symbiosis operates. Exploring the consequences of these interactions, including those  
1300 involving other fungi, bacteria, non-host plants, macro- and micro-fauna, will provide insights  
1301 into the mechanisms of coexistence, competition, and facilitation.

1302

### 1303 **VII.ii: Limitations**

1304 While the *Suillus* system presents numerous advantages, it is important to also acknowledge its  
1305 limitations within specific research contexts. In fact, some of the very traits that make *Suillus*  
1306 an ideal model for certain research objectives render it unsuitable for others. For example, the  
1307 close association between *Suillus* and the Pinaceae, makes the genus valuable for studying  
1308 partner specificity and diversification events associated with host switching, but also confines  
1309 its applicability primarily to host associations within the Pinaceae (although associations of  
1310 certain *Suillus* species with angiosperms may bring different perspectives). Similarly, because  
1311 *Suillus* is naturally found only in the northern hemisphere where Pinaceae hosts are native, and  
1312 in the southern hemisphere only as introduced species, it serves as an excellent system for  
1313 studying reproductive bottlenecks, dispersal, and invasion biology across introduced ranges.  
1314 These benefits do not negate the fact that *Suillus* is non-native on half the planet, limiting its  
1315 applicability as a model for understanding many systems across the global south, such as  
1316 understudied tropical ECM forests.

1317

1318 Finally, we emphasize that the advancement of the *Suillus* system as a model does not preclude  
1319 the progress of other systems. On the contrary, we hope that formalizing the development of  
1320 the *Suillus* model will catalyze researchers to establish similar collaborative working groups on  
1321 other promising ECM model genera such as *Laccaria*, *Russula*, and *Pisolithus*, which may be  
1322 better suited for specific research objectives. The ECM lifestyle is thought to have evolved  
1323 independently at least 78 times over, and represents at least 251 distinct genera, with diverse  
1324 traits, host-associations, and geographic distributions (Tedersoo & Smith, 2013). With  
1325 recognition that no single set of taxa can hope to represent the entirety of ECM diversity,  
1326 concentrating efforts around the development of carefully chosen groups will enable the rapid  
1327 development of resources needed to address the most pressing questions in ECM ecology and  
1328 evolution.

1329

### 1330 **VII.iii: Technical challenges and future directions**

1331 Fully developing *Suillus* as a model genus presents some technical challenges that the  
1332 community is currently working to address. Resolving these issues will soon open up a  
1333 multitude of possibilities to further use *Suillus* as a means to understand ECM ecology, biology,  
1334 and evolution.

1335

1336 First, using *Suillus* to fully address the mechanisms underlying ECM metabolism, requires the  
1337 development of systems capable of tracking nutrient dynamics under field conditions, with  
1338 adult trees, and in complex communities. Bolstered by the availability of a wide array of  
1339 publicly available *Suillus* genomes, continual improvements in functional -omics and meta-  
1340 omics approaches promise to facilitate comprehensive links between genomes and functional  
1341 traits. These advancements are expected to have significant implications across both laboratory  
1342 and field settings, facilitating a better understanding of ECM biodiversity across biological  
1343 scales: linking genes to communities and ecosystems.

1344

1345 Another significant challenge is the complexity of working with heterokaryotic dikaryons; a  
1346 biological consideration for most ECM basidiomycetes. Optimized protocols to dependably  
1347 monokaryotize and dikaryotize *Suillus* strains in culture, along with efficient assays to confirm  
1348 karyotic state, would go far to facilitate studies in ECM fungal genetics, gene regulation, and  
1349 nuclear control.

1350

1351 An additional challenge common to most ECM fungi is the inability to dependably produce  
1352 fruit bodies under laboratory conditions. In theory, this limitation is related to the size of the  
1353 carbon stores necessary to support the development of reproductive structures. Seedling-based

1354 bioassays are favored in ECM studies because of the impracticality of manipulating large trees  
1355 in the laboratory. This is thought to limit total photosynthetic capacity and the amount of carbon  
1356 transferred to the fungus, particularly over limited time scales. Circumnavigating this limitation  
1357 would require a system which increases carbon transfer from the host without disrupting the  
1358 symbiosis, or a system for triggering fruit body development apart from the host entirely. The  
1359 development of such a system would facilitate significant advances in our understanding of  
1360 ECM genetics, enabling gene association studies, and studies in morphogenesis and histology.

1361  
1362 Finally, the development of gene editing systems for *Suillus* will radically improve our ability  
1363 to manipulatively address the genetic basis of adaptation, evolution, and the role of specific  
1364 genes in important ecological and biological processes. The International *Suillus* Consortium  
1365 considers the development of gene editing tools to be of the highest priority and has formed a  
1366 working group to address this challenge, making use of multiple gene editing platforms and  
1367 targeting multiple species in the genus.

1368  
1369

#### 1370 **VII.iv: Conclusion**

1371 Studies in *Suillus* have already made significant contributions to our understanding of the  
1372 biology and ecology and evolution of the ECM symbiosis. The availability of tools and  
1373 protocols optimized for *Suillus*, including specific approaches for *in vitro* culturing, plant  
1374 partner bioassays, high quality DNA and RNA extraction included in Section VIII, along with  
1375 the large number of publicly available reference genomes will catalyze future research to  
1376 address long-standing questions in the field. We expect that continued community development  
1377 of the *Suillus* system will solidify the genus as a model for understanding the ecology and  
1378 evolution of plant-fungal mutualisms. We invite anyone interested in collaborating with The  
1379 International *Suillus* Consortium to please contact the authors.

1380  
1381  
1382

## 1383 **VIII. Resources and protocols**

1384

1385 Despite the importance of ECM fungi to both natural and managed forests, manipulative work  
1386 has largely lagged behind other experimental systems due in part to the technical challenges of  
1387 working with host-associated, dikaryotic species. In an effort to standardize protocols and  
1388 stimulate future improvements that will build off of these resources, we present a series of  
1389 protocols for working with *Suillus* that will allow researchers to circumnavigate many of these  
1390 challenges. These protocols are available as a series of supplemental files on topics ranging  
1391 from axenic isolation and long-term storage, to mycorrhizal synthesis and the application of  
1392 DNA based approaches. Here, we provide instruction for navigating these resources, outline  
1393 specific use cases, limitations, and recommendations.

1394  
1395

#### 1396 **VIII.i: Fruit body and spore collection (Supplemental file S1)**

1397 One of the benefits of the *Suillus* system is that fruit bodies of the genus are conspicuous,  
1398 frequently abundant, and relatively easy to identify. Both amateur and professional mycologists  
1399 alike have long targeted the collection of charismatic *Suillus* mushrooms, with over 46,000  
1400 vouchered specimens currently digitized in the MyCoPortal collections database  
1401 (<https://www.mycportal.org/>) and over 62,000 observations of *Suillus* currently in iNaturalist  
1402 (<https://www.inaturalist.org/>). Vouchered collections have facilitated phylogenetic  
1403 reclassifications across the genus (Nguyen *et al.*, 2016b), illuminated the association between  
1404 species diversification and host shifts (Zhang *et al.*, 2022), clarified species rarity (Kennedy *et al.*,  
1405 2020), and helped to identify the biotic and abiotic factors facilitating range shifts and  
1406 invasion potential (Pietras *et al.*, 2018). The collection of *Suillus* fruit bodies is the first step to  
1407 obtaining pure cultures, spores for use in bioassays, and preserved specimens for morphological

1408 studies or new species descriptions. Supplemental file S1 details the process of finding,  
1409 identifying, and collecting *Suillus* fruit bodies for down-stream applications, and the processing  
1410 of fruit bodies for spore collection and preservation.  
1411

#### 1412 **VIII.ii: Isolation (Supplemental file S2)**

1413 Unlike many groups of ECM fungi, *Suillus* can be readily isolated and grown in pure culture.  
1414 Cultures can be isolated from fruit bodies, ECM rootlets, or basidiospores, to flexibly obtain  
1415 dikaryotic or monokaryotic strains. Supplemental file S2 gives an overview of best practices  
1416 and specific instructions for obtaining pure cultures of *Suillus* from all three of these sources.  
1417

#### 1418 **VIII.iii: Culture conditions and culture storage (Supplemental file S3)**

1419 *Suillus* can accommodate growth on a variety of media types, but are most commonly cultured  
1420 on Fries Media, Hagens Agar, Modified Melin-Norkrans (MMN) or Pachlewski's Media (Px).  
1421 Recipes for all four media types, slightly modified from their original formulations for  
1422 optimized *Suillus* growth, are available in Supplemental file S3, along with notes on optimized  
1423 growth conditions and recommendations for culture storage and long-term culture preservation.  
1424  
1425  
1426

#### 1427 **VIII.v: Sequencing (Supplemental files S4-S6)**

1428 Whole genome and next-generation sequencing using genomic, transcriptomic, and  
1429 metatranscriptomic approaches have revolutionized our ability to identify and characterize  
1430 ECM fungal communities (Bork *et al.*, 2015; Thompson *et al.*, 2017; Quince *et al.*, 2017; Lu &  
1431 Salzberg, 2020), and helped to shed light on fungal ecology and evolution (Miyachi *et al.*,  
1432 2020; Feurtey *et al.*, 2023; Ali *et al.*, 2023). In *Suillus*, these technologies have enabled the  
1433 characterization of high resolution phylogenies, revealed the extent of intragenomic genome  
1434 diversity, and helped to illuminate the functional mechanisms of important traits like stress  
1435 tolerance and partner specificity (Liao *et al.*, 2016; Bazzicalupo *et al.*, 2020; Lofgren *et al.*,  
1436 2021; Erlandson *et al.*, 2022). However, compared to other ECM groups, *Suillus* spp. present  
1437 unique technical challenges to sample preparation for -omics analysis. The genus is a notorious  
1438 producer of pigments and extraction inhibitors that can complicate quality control  
1439 measurements, interfere with PCR amplification, and confound DNA/cDNA library  
1440 construction. DNA and RNA extraction from mixed biological samples such as mycorrhizal  
1441 root tips is a critical but often difficult step in preparing material for molecular work. In general,  
1442 ectomycorrhizae (which are a mix of plant and fungal tissue) contain polysaccharides,  
1443 hydroxybenzenes, esters, and other secondary metabolites that can confound extraction  
1444 protocols. Compared to other ECM groups, *Suillus* also has a greatly expanded secondary  
1445 metabolite repertoire (Lofgren *et al.*, 2021), that likely further complicates DNA extraction.  
1446 Additionally, because *Suillus* associates almost exclusively with plants in the family Pinaceae,  
1447 which famously produce a wide array of secondary metabolites (Bashalkhanov & Rajora,  
1448 2008), the confounding effect of extraction inhibitors are likely compounded across the genus.  
1449

1450 To minimize the impact of inhibitors, we have developed and optimized three methods for  
1451 nucleic acid extraction that cover a broad range of applications including fresh and dried  
1452 mushrooms, root-tips, cultures, and soil, and include them here as three separate Supplementary  
1453 protocols. The NaOH extraction method yields good DNA for fast genotyping of cultures or  
1454 root-tips (Supplementary file S4). The CTAB extraction method is generally used for whole  
1455 genome sequencing and can be performed either for DNA alone, or as a DNA/RNA co-  
1456 extraction (Supplementary file S5). The high-molecular weight DNA extraction method is  
1457 generally good for long-read sequencing (Supplementary file S6).

1458

#### 1459 **VIII.vi: Host-*Suillus* bioassays (Supplemental file S7)**

1460 The tractability of *Suillus* spp. to *in vitro* mycorrhization via both spore inoculation and  
1461 mycelial extension is one of the primary factors making this genus an excellent experimental  
1462 system for ecological and evolutionary research. While spore-based inoculation methods are  
1463 high-throughput and easily incorporate fungal genetic variation into mycorrhizal synthesis  
1464 studies, mycelial-based methods provide a mechanism to control strain provenance and can be  
1465 used to construct either mid-throughput pot-based assays, or low-throughput but highly flexible  
1466 mesocosms that are otherwise axenic, facilitating the ability to control and manipulate the rest  
1467 of the microbial community. All *Suillus* species we have examined to date are amenable to  
1468 laboratory mycorrhization using spore and mycelial methods, achieving a high level of  
1469 colonization (up to 100% of root tips) when paired with the appropriate host (Table S1). In  
1470 Supplementary file S7, we provide an overview of these procedures and specific protocols for  
1471 inoculating *Suillus* onto host trees to conduct bioassays using spore-based, and mycelial-based  
1472 mycorrhizal synthesis.

1473  
1474

#### 1475 **VIII.vii: *Suillus*-host associations (Supplemental Table S1)**

1476 Understanding the specificity patterns between different *Suillus* species and their host trees in  
1477 natural environments is fundamental to successful experimental design, interrupting high  
1478 throughput sequence data, and to locating and identifying *Suillus* fruitbodies. In Table S1, we  
1479 provide putative host associations for all currently recognized species of *Suillus* and as well as  
1480 several additional taxa that are yet to be formally typified, but clearly represent independent  
1481 species based on molecular analysis. Citations are provided for all putative host associations.  
1482 Species that have one or more associated whole genome sequencing projects are noted.

1483  
1484  
1485

## 1486 **Author contributions**

1487

1488 SB, LL, and NHN developed the concept and outline for this manuscript. SB and LL  
1489 coordinated all author contributions. LL synthesized and edited the initial and final drafts. SB  
1490 and LL wrote section I, NHN wrote section II with contributions from LL. LL wrote section  
1491 III with contributions from NHN and SB. PK and EP-P wrote section IV with contributions  
1492 from NHN. SB, HW, AS, JF, JR, and KZ wrote section V. RV and LL wrote section VI. LL,  
1493 SB, and NHN wrote section VII. LL wrote section VIII with contributions from H-LL, HW,  
1494 HVTC, KMH, EE, and NHN.

1495  
1496

## 1497 **Acknowledgements**

1498

1499 LL is supported by funding from the National Institutes of Health grant no. T32-AI052080 via  
1500 the Tri-I MMPTP Fellowship. SB and H-LL are supported by NSF IOS-PBI (2029168). NHN  
1501 is supported by the U.S. Department of Energy, Office of Science, Office of Biological and  
1502 Environmental Research, Genomic Science Program, under award numbers DE-SC0020163  
1503 and DE-SC0023106. Sequencing of *Suillus* genomes and transcriptomes are provided by the  
1504 Joint Genome Institute grants CSP 502931 to NHN and FICUS CSP 49514 to H-LL, JR is  
1505 supported by Research Foundation Flanders (FWO G082621N and K226022N). Thank you to  
1506 all of the members of The International *Suillus* Consortium past and present and to the larger  
1507 *Suillus* and ECM research communities who drive and inspire the work reviewed here. We  
1508 would like to thank Ian Dickie and two anonymous reviewers for their helpful comments on a  
1509 previous version of this manuscript.

1510  
1511

## 1512 Author's note

1513

1514 *Suillus* fungi have long captivated the attention of the mycological community. The formalized  
1515 efforts of the International *Suillus* Consortium over the last decade represent only the most  
1516 recent expression of this ongoing research trajectory. The literature dedicated to *Suillus* is  
1517 extensive, spanning over a century and encompassing a breadth of knowledge that far exceeds  
1518 the scope of this text. Considering this context, the authors would like to acknowledge and  
1519 appreciate the pioneering work of the many experts whose profound contributions paved the  
1520 way for contemporary researchers and continue to inspire future advancements in the ecology  
1521 and ecology of this enigmatic yet enchanting genus.

1522

1523

## 1524 Figure Legends

1525

1526 **Figure 1: Morphological and phenotypic variation across *Suillus*.** A-H) Representative members of the major  
1527 clades within the *Suillus* phylogeny: Larch associate species A) *S. ampliporus* showing the radiating pores  
1528 characteristic most members of the genus. B) *S. clintonianus* displaying the copious viscus slime often produced by  
1529 *Suillus* fruit bodies. C) *S. grisellus* highlighting morphological variation present just within the *Larix*-associated  
1530 species. D) *Pseudotsuga*-associated *S. lakei* (Rocky Mountain form) highlighting the significant carbon allocation  
1531 needed to form *Suillus* fruit bodies, which here exceed ~15 cm in diameter. *Pinus* associated species: E) *S.*  
1532 *subalutaceus* representing species with longer stipes. F) *S. luteus*, the type species of the genus, displaying a  
1533 prominent veil, a diagnostic feature for some species. G) *S. spraguei*, displaying distinctive red scales on the cap  
1534 cuticle. H) *S. tomentosus* with robust fruiting bodies and variable bluing reactions of the trama tissue. I-P) Other  
1535 diagnostic and morphologically important characteristics of *Suillus* I) A young specimen of *S. weaverae* showing  
1536 milky droplets and clearly punctate stipe as a result of glandular cells. J) An old specimen of *S. caerulescens*, strongly  
1537 staining blue, with insect larvae tunnels. K) Insect larvae quickly consume *Suillus* fruit bodies and can quickly ruin  
1538 an attempt at making spore prints. L) A spore print of *S. lakei* on aluminum foil for spore collection. M) A portion  
1539 of a *S. lakei* fairy ring with large fruit bodies which require substantial carbon allocations for the host. N) A  
1540 mycorrhizal root of *S. paluster* with rhizomorphs at the base, typically found in *Suillus* mycorrhizae. O) The  
1541 mycoparasitic *Chroogomphus vinicolor* (left) fruiting next to its host, a senescing *S. pungens* (right) P) Clusters of  
1542 hyphae of *C. vinicolor*, stained dark violet with Melzer's reagent, within a mycorrhizal root of *S. pungens*.

1543

1544 **Figure 2: Representatives of the *Suillus* genome-strain culture collection on four media types.** Cultures were  
1545 grown in 9 cm petri plates on four media types including (A, E, I, M) Modified Melin-Norkrans (MMN), (B, F, J,  
1546 N) Modified Fries Media (Fries), (C, G, K, O) Modified Hagem's Agar (Hagem's), and (D, H, L, P) Pachlewski's  
1547 Media (Pachlewski's). See supplemental material S3 for *Suillus* optimized media recipes. All media types were  
1548 prepared at their full respective carbon concentrations and adjusted to pH 6 prior to autoclaving. Cultures were grown  
1549 for 28 days, at room temperature, in the dark, prior to being photographed. Strains represented here are (A-D) *S.*  
1550 *weaverae* FC27, (E-H) *S. pungens* FC27, (I-L) *S. fuscotomentosus* FC203, and (M-P) the monokaryotic strain *S.*  
1551 *hirtellus* EM16. These four strains are part of the *Suillus* Genome Strain Culture Collection, and the *Suillus*  
1552 phenotype database, SuilluScope. To access phenotype information and NRRL accession numbers for the full strain  
1553 collection, please visit [www.SuilluScope.com](http://www.SuilluScope.com).

1554

1555 **Figure 3: Total colony area over time for representatives of the *Suillus* genome-strain culture collection.**  
1556 Cultures were started by placing 3mm agar plugs on Modified Melin-Norkrans (MMN) media, adjusted to pH 6  
1557 prior to autoclaving, and grown in temperature adjustable incubators, in the dark, for a total of 33 days (at n=4  
1558 replicates per species per temperature treatment). Starting on day 8, colony area was recorded twice per week over  
1559 the course of the assay by marking the colony margin on the back of each petri dish with a fine-tip sharpie. After  
1560 33 days of growth, the back of the petri plates were imaged using a flatbed scanner, and colony area was calculated  
1561 for each time point using the program imageJ. The data presented here is for replicates grown at room temperature  
1562 (20°C). To access growth data at other temperatures, and for the full *Suillus* Genome Strain Culture Collection  
1563 please visit [www.SuilluScope.com](http://www.SuilluScope.com).

1564

1565 **Figure 4: Associations between *Suillus* and their Pinaceae hosts.** Time calibrated phylogenies of proposed *Suillus*  
1566 subgenera and Pinaceae genera and subgenera, displaying distinct topologies and no indication of cospeciation.  
1567 Molecular clock dating supports the earliest divergence of *Suillus* between 40.2 and 71.1 MYA depending on the  
1568 method of fossil calibration used. Dashed lines represent host associations, and ancestral host-switching events,  
1569 highlighting ancestral associations with *Larix* and the host switch of *Suillus* subgenus *Suillus* from the host subgenera  
1570 *Strobos* to the subgenus *Pinus*, which is thought to have occurred at least four times independently. Figure redrawn

1571 with permission from Zhang *et al.* (Zhang *et al.*, 2022).

1572

1573 **Figure 5: Biological introduction records for A) all ECM fungi and B) Suillus.** Records from the Global Database  
1574 of Alien Macrofungi (Monteiro *et al.*, 2020) were pruned to exclude taxa with uncertain species-level assignments,  
1575 and those annotated as having established from uncertain geographic origins. Guild was assigned across the database  
1576 using FUNGuild retaining only species that could be confidently identified as ECM, resulting in 946 observations.  
1577 A) Out of 55 genera of ECM fungi with introduction records, *Suillus* accounted for the majority of observed  
1578 introductions, totaling 193 records. B) In total, 24 species of *Suillus* have introduction records, with the most  
1579 common species identified as *S. luteus* (44 observations) and *S. granulatus* (34 observations), but was absent from  
1580 the database (likely combined with the morphologically similar *S. brevipes*).

1581

1582

## 1583 References

1584

- 1585 **Adriaensen K, Van Der Lelie D, Van Laere A, Vangronsveld J, Colpaert JV. 2004.** A  
1586 zinc-adapted fungus protects pines from zinc stress. *The New phytologist* **161**: 549–555.
- 1587 **Adriaensen K, Vangronsveld J, Colpaert JV. 2006.** Zinc-tolerant *Suillus bovinus* improves  
1588 growth of Zn-exposed *Pinus sylvestris* seedlings. *Mycorrhiza* **16**: 553–558.
- 1589 **Adriaensen K, Vrålstad T, Noben J-P, Vangronsveld J, Colpaert JV. 2005.** Copper-  
1590 adapted *Suillus luteus*, a symbiotic solution for pines colonizing Cu mine spoils. *Applied and*  
1591 *environmental microbiology* **71**: 7279–7284.
- 1592 **Agerer R. 2001.** Exploration types of ectomycorrhizae. *Mycorrhiza* **11**: 107–114.
- 1593 **Aguirre F, Nouhra E, Urcelay C. 2021.** Native and non-native mammals disperse exotic  
1594 ectomycorrhizal fungi at long distances from pine plantations. *Fungal ecology* **49**: 101012.
- 1595 **Ali S, Gladieux P, Ravel S, Adreit H, Meusnier I, Milazzo J, Cros-Arteil S, Bonnot F, Jin**  
1596 **B, Dumartinet T, et al. 2023.** Evolution of the rice blast pathogen on spatially structured rice  
1597 landraces maintains multiple generalist fungal lineages. *Molecular ecology* **32**: 2519–2533.
- 1598 **Arraiano-Castilho R, Bidartondo MI, Niskanen T, Clarkson JJ, Brunner I,**  
1599 **Zimmermann S, Senn-Irlet B, Frey B, Peintner U, Mrak T, et al. 2021.** Habitat  
1600 specialisation controls ectomycorrhizal fungi above the treeline in the European Alps. *The*  
1601 *New phytologist* **229**: 2901–2916.
- 1602 **Ashkannejhad S, Horton TR. 2006.** Ectomycorrhizal ecology under primary succession on  
1603 coastal sand dunes: interactions involving *Pinus contorta*, suilloid fungi and deer. *The New*  
1604 *phytologist* **169**: 345–354.
- 1605 **Avital S, Rog I, Livne-Luzon S, Cahanovitch R, Klein T. 2022.** Asymmetric belowground  
1606 carbon transfer in a diverse tree community. *Molecular ecology* **31**: 3481–3495.
- 1607 **Baldrian P. 2009.** Ectomycorrhizal fungi and their enzymes in soils: is there enough evidence  
1608 for their role as facultative soil saprotrophs? *Oecologia* **161**: 657–660.
- 1609 **Baldrian P, López-Mondéjar R, Kohout P. 2023.** Forest microbiome and global change.  
1610 *Nature reviews. Microbiology* **21**: 487–501.
- 1611 **Bashalkhanov S, Rajora OP. 2008.** Protocol: A high-throughput DNA extraction system  
1612 suitable for conifers. *Plant methods* **4**: 20.
- 1613 **Baura G, Szaro TM, Bruns TD. 1992.** *Gastrosuillus laricinus* is a Recent Derivative of  
1614 *Suillus grevillei*: Molecular Evidence. *Mycologia* **84**: 592–597.
- 1615 **Bazzicalupo AL, Ruytinx J, Ke Y-H, Coninx L, Colpaert JV, Nguyen NH, Vilgalys R,**  
1616 **Branco S. 2020.** Fungal heavy metal adaptation through single nucleotide polymorphisms  
1617 and copy-number variation. *Molecular ecology* **29**: 4157–4169.
- 1618 **Besl H, Bresinsky A. 1997.** Chemosystematics of Suillaceae and Gomphidiaceae (suborder  
1619 Suillineae). *Plant systematics and evolution = Entwicklungsgeschichte und Systematik der*  
1620 *Pflanzen* **206**: 223–242.
- 1621 **Bhatnagar J, Vilgalys R, Averill C, Brzostek E, Liao H-L, Chen K-H, Nash J, Policelli N,**  
1622 **Tappero R. 2021.** Roles of ectomycorrhizal fungi in plant and soil function under simulated  
1623 global change. In: B11D-04.
- 1624 **Bidartondo MI, Bruns TD. 2002.** Fine-level mycorrhizal specificity in the Monotropoideae  
1625 (Ericaceae): specificity for fungal species groups. *Molecular ecology* **11**: 557–569.

1626 **Binder M, Hibbett DS. 2006.** Molecular systematics and biological diversification of  
1627 Boletales. *Mycologia* **98**: 971–981.

1628 **Blaudez D, Jacob C, Turnau K, Colpaert JV, Ahonen-Jonnarth U, Finlay R, Botton B,**  
1629 **Chalot M. 2000.** Differential responses of ectomycorrhizal fungi to heavy metals *in vitro*.  
1630 *Mycological research* **104**: 1366–1371.

1631 **Bogar LM, Kennedy PG. 2013.** New wrinkles in an old paradigm: neighborhood effects can  
1632 modify the structure and specificity of *Alnus*-associated ectomycorrhizal fungal communities.  
1633 *FEMS microbiology ecology* **83**: 767–777.

1634 **Bogar L, Peay K, Kornfeld A, Huggins J, Hortal S, Anderson I, Kennedy P. 2019.** Plant-  
1635 mediated partner discrimination in ectomycorrhizal mutualisms. *Mycorrhiza* **29**: 97–111.

1636 **Bogar LM, Tavasieff OS, Raab TK, Peay KG. 2022.** Does resource exchange in  
1637 ectomycorrhizal symbiosis vary with competitive context and nitrogen addition? *The New*  
1638 *phytologist* **233**: 1331–1344.

1639 **Bondarenko V, Geras'kin S, Bondarenko E, Yoschenko V, Bondarenko S, Khanova A,**  
1640 **Garbaruk D, Nanba K. 2023.** Comparative analysis of epigenetic variability in two pine  
1641 species exposed to chronic radiation in the Chernobyl and Fukushima affected zones.  
1642 *Environmental pollution* **330**: 121799.

1643 **Bonello P, Bruns TD, Gardes M. 1998.** Genetic structure of a natural population of the  
1644 ectomycorrhizal fungus *Suillus pungens*. *The New phytologist* **138**: 533–542.

1645 **Bork P, Bowler C, de Vargas C, Gorsky G, Karsenti E, Wincker P. 2015.** Tara Oceans.  
1646 Tara Oceans studies plankton at planetary scale. Introduction. *Science* **348**: 873.

1647 **Brabcová V, Nováková M, Davidová A, Baldrian P. 2016.** Dead fungal mycelium in forest  
1648 soil represents a decomposition hotspot and a habitat for a specific microbial community. *The*  
1649 *New phytologist* **210**: 1369–1381.

1650 **Bradshaw AJ, Autumn KC, Rickart EA, Dentinger BTM. 2022.** On the origin of feces:  
1651 Fungal diversity, distribution, and conservation implications from feces of small mammals.  
1652 *Environmental DNA (Hoboken, N.J.)* **4**: 608–626.

1653 **Branco S, Bi K, Liao H-L, Gladioux P, Badouin H, Ellison CE, Nguyen NH, Vilgalys R,**  
1654 **Peay KG, Taylor JW, et al. 2017.** Continental-level population differentiation and  
1655 environmental adaptation in the mushroom *Suillus brevipes*. *Molecular ecology* **26**: 2063–  
1656 2076.

1657 **Branco S, Gladioux P, Ellison CE, Kuo A, LaButti K, Lipzen A, Grigoriev IV, Liao H-L,**  
1658 **Vilgalys R, Peay KG, et al. 2015.** Genetic isolation between two recently diverged  
1659 populations of a symbiotic fungus. *Molecular ecology* **24**: 2747–2758.

1660 **Branco S, Schauster A, Liao H-L, Ruytinx J. 2022.** Mechanisms of stress tolerance and  
1661 their effects on the ecology and evolution of mycorrhizal fungi. *The New phytologist* **235**:  
1662 2158–2175.

1663 **Brodribb TJ, Powers J, Cochard H, Choat B. 2020.** Hanging by a thread? Forests and  
1664 drought. *Science* **368**: 261–266.

1665 **Brundrett M, Bougher N, Dell B, Grove T, Malajczuk N. 1996.** *Working with mycorrhizas*  
1666 *in forestry and agriculture*. Australian Centre for International Agricultural Research.

1667 **Bruns TD. 1984.** Insect mycophagy in the Boletales : Fungivore diversity and the mushroom  
1668 habitat. *Fungus-Insect relationships-Perspectives in Ecology and Evolution*.

1669 **Bruns TD, Bidartondo MI, Taylor DL. 2002.** Host specificity in ectomycorrhizal  
1670 communities: what do the exceptions tell us? *Integrative and comparative biology* **42**: 352–  
1671 359.

1672 **Bruns TD, Fogel R, White TJ, Palmer JD. 1989.** Accelerated evolution of a false-truffle  
1673 from a mushroom ancestor. *Nature* **339**: 140–142.

1674 **Bruns TD, Grubisha LC, Trappe JM, Kerekes JF, Vellinga EC. 2010.** *Suillus quiescens*, a  
1675 new species commonly found in the spore bank in California and Oregon. *Mycologia* **102**:  
1676 438–446.

1677 **Bruns TD, Hale ML, Nguyen NH. 2019.** *Rhizopogon olivaceotinctus* increases its inoculum  
1678 potential in heated soil independent of competitive release from other ectomycorrhizal fungi.  
1679 *Mycologia* **111**: 936–941.

1680 **Bubriski R, Kennedy P. 2014.** A molecular and morphological analysis of the genus

1681 *Rhizopogon* subgenus *Villosuli* section *Villosuli* as a preface to ecological monitoring.  
1682 *Mycologia* **106**: 353–361.

1683 **Burchhardt KM, Rivera Y, Baldwin T, Vanearden D, Kretzer AM. 2011.** Analysis of  
1684 genet size and local gene flow in the ectomycorrhizal basidiomycete *Suillus spraguei*  
1685 (synonym *S. pictus*). *Mycologia* **103**: 722–730.

1686 **Burdon JJ, Chilvers GA. 1977.** Preliminary studies on a native Australian eucalypt forest  
1687 invaded by exotic pines. *Oecologia* **31**: 1–12.

1688 **Burdon JJ, Chilvers GA. 1994.** Demographic changes and the development of competition  
1689 in a native Australian eucalypt forest invaded by exotic pines. *Oecologia* **97**: 419–423.

1690 **Caiafa MV, Jusino MA, Wilkie AC, Díaz IA, Sieving KE, Smith ME. 2021.** Discovering  
1691 the role of Patagonian birds in the dispersal of truffles and other mycorrhizal fungi. *Current*  
1692 *biology: CB* **31**: 5558-5570.e3.

1693 **Cairney JWG, Chambers SM. 1997.** Interactions between *Pisolithus tinctorius* and its  
1694 hosts: a review of current knowledge. *Mycorrhiza* **7**: 117–131.

1695 **Calvo-Polanco M, Zwiazek JJ. 2011.** Role of osmotic stress in ion accumulation and  
1696 physiological responses of mycorrhizal white spruce (*Picea glauca*) and jack pine (*Pinus*  
1697 *banksiana*) to soil fluoride and NaCl. *Acta physiologiae plantarum / Polish Academy of*  
1698 *Sciences, Committee of Plant Physiology Genetics and Breeding* **33**: 1365–1373.

1699 **Castaño C, Suarez-Vidal E, Zas R, Bonet JA, Oliva J, Sampedro L. 2023.**  
1700 Ectomycorrhizal fungi with hydrophobic mycelia and rhizomorphs dominate in young pine  
1701 trees surviving experimental drought stress. *Soil biology & biochemistry* **178**: 108932.

1702 **Chapela IH, Osher LJ, Horton TR, Henn MR. 2001.** Ectomycorrhizal fungi introduced  
1703 with exotic pine plantations induce soil carbon depletion. *Soil biology & biochemistry* **33**:  
1704 1733–1740.

1705 **Chapman WK, Paul L. 2012.** Evidence that Northern Pioneering Pines with Tuberculate  
1706 Mycorrhizae are Unaffected by Varying Soil Nitrogen Levels. *Microbial ecology* **64**: 964–  
1707 972.

1708 **Charya LS, Garg S. 2019.** Advances in methods and practices of ectomycorrhizal research.  
1709 In: Meena SN, Naik MM, eds. *Advances in Biological Science Research*. Academic Press,  
1710 303–325.

1711 **Chilvers GA, Burdon JJ. 1983.** Further studies on a native Australian eucalypt forest  
1712 invaded by exotic pines. *Oecologia* **59**: 239–245.

1713 **Cho HJ, Park KH, Park MS, Cho Y, Kim JS, Seo CW, Oh S-Y, Lim YW. 2021.**  
1714 Determination of Diversity, Distribution and Host Specificity of Korean *Laccaria* Using Four  
1715 Approaches. *Mycobiology* **49**: 461–468.

1716 **Chu-Chou M, Grace LJ. 1988.** Mycorrhizal fungi of radiata pine in different forests of the  
1717 north and south islands in New Zealand. *Soil biology & biochemistry* **20**: 883–886.

1718 **Colpaert JV. 2008.** Chapter 11 Heavy metal pollution and genetic adaptations in  
1719 ectomycorrhizal fungi. In: Avery SV, Stratford M, Van West P, eds. *British Mycological*  
1720 *Society Symposia Series*. Academic Press, 157–173.

1721 **Colpaert JV, Adriaensen K, Muller LAH, Lambaerts M, Faes C, Carleer R,**  
1722 **Vangronsveld J. 2005.** Element profiles and growth in Zn-sensitive and Zn-resistant Suilloid  
1723 fungi. *Mycorrhiza* **15**: 628–634.

1724 **Colpaert JV, van Assche JA. 1987.** Heavy Metal Tolerance in Some Ectomycorrhizal Fungi.  
1725 *Functional ecology* **1**: 415–421.

1726 **Colpaert JV, Muller LAH, Lambaerts M, Adriaensen K, Vangronsveld J. 2004.**  
1727 Evolutionary adaptation to Zn toxicity in populations of Suilloid fungi. *The New phytologist*  
1728 **162**: 549–559.

1729 **Colpaert JV, Van Assche JA. 1992.** Zinc toxicity in ectomycorrhizal *Pinus sylvestris*. *Plant*  
1730 *and soil* **143**: 201–211.

1731 **Colpaert JV, Wevers JHL, Krznaric E, Adriaensen K. 2011.** How metal-tolerant ecotypes  
1732 of ectomycorrhizal fungi protect plants from heavy metal pollution. *Annals of forest science*  
1733 **68**: 17–24.

1734 **Coninx L, Smisdom N, Kohler A, Arnauts N, Ameloot M, Rineau F, Colpaert JV,**  
1735 **Ruytinx J. 2019.** SIZRT2 Encodes a ZIP Family Zn Transporter With Dual Localization in

1736 the Ectomycorrhizal Fungus *Suillus luteus*. *Frontiers in microbiology* **10**: 2251.

1737 **Coninx L, Thoonen A, Slenders E, Morin E, Arnauts N, Op De Beeck M, Kohler A,**

1738 **Ruytinx J, Colpaert JV. 2017.** The SIZRT1 Gene Encodes a Plasma Membrane-Located ZIP

1739 (Zrt-, Irt-Like Protein) Transporter in the Ectomycorrhizal Fungus *Suillus luteus*. *Frontiers in*

1740 *microbiology* **8**: 2320.

1741 **Cripps C, R. A. 2011.** Native Ectomycorrhizal Fungi of Limber and Whitebark Pine:

1742 Necessary for Forest Sustainability? The future of high-elevation, five-needle white pines in

1743 Western North America: proceedings of the High Five Symposium. Fort Collins (CO): USDA

1744 Forest Service, Rocky Mountain Research Station. *USDA Forest Service Proceedings*

1745 **RMRS-P-63**: 37-44).

1746 **Dahlberg A, Finlay RD. 1999.** *Suillus*. In: Cairney JWG, Chambers SM, eds.

1747 Ectomycorrhizal Fungi Key Genera in Profile. Berlin, Heidelberg: Springer Berlin

1748 Heidelberg, 33–64.

1749 **Dahlberg A, Stenlid J. 1990.** Population structure and dynamics in *Suillus bovinus* as

1750 indicated by spatial distribution of fungal clones. *The New phytologist* **115**: 487–493.

1751 **Dahlberg A, Stenlid J. 1994.** Size, distribution and biomass of genets in populations of

1752 *Suillus bovinus* (L.: Fr.) Roussel revealed by somatic incompatibility. *The New*

1753 *phytologist* **128**: 225–234.

1754 **Dai A. 2012.** Increasing drought under global warming in observations and models. *Nature*

1755 *climate change* **3**: 52–58.

1756 **Dauphin B, Peter M. 2024.** Tracking signatures of selection in natural populations of

1757 ectomycorrhizal fungi - progress, challenges, and prospects. *The New phytologist*.

1758 **Den Bakker HC, Zuccarello GC, Kuypers TW, Noordeloos ME. 2004.** Evolution and host

1759 specificity in the ectomycorrhizal genus *Leccinum*. *The New phytologist* **163**: 201–215.

1760 **Dickie IA, Bolstridge N, Cooper JA, Peltzer DA. 2010.** Co-invasion by *Pinus* and its

1761 mycorrhizal fungi. *The New phytologist* **187**: 475–484.

1762 **Dickie IA, Bufford JL, Cobb RC, Desprez-Loustau M-L, Grelet G, Hulme PE,**

1763 **Klironomos J, Makiola A, Nuñez MA, Pringle A, et al. 2017.** The emerging science of

1764 linked plant-fungal invasions. *The New phytologist* **215**: 1314–1332.

1765 **Dighton J, Tugay T, Zhdanova N. 2008.** Fungi and ionizing radiation from radionuclides.

1766 *FEMS microbiology letters* **281**: 109–120.

1767 **Doudrick RL, Stewart EL, Alm AA. 2011.** Survey and ecological aspects of presumed

1768 ectomycorrhizal fungi associated with black spruce in northern Minnesota. *Canadian journal*

1769 *of botany. Journal canadien de botanique*.

1770 **Dowie NJ, Hemenway JJ, Miller SL. 2012.** Identity, genetic lineages and putative hybrids of

1771 an obligate mycobiont associated with the mycoheterotrophic plant *Pterospora andromedea*

1772 in the south-central Rocky Mountains. *Fungal ecology* **5**: 137–146.

1773 **Duddridge JA, Malibari A, Read DJ. 1980.** Structure and function of mycorrhizal

1774 rhizomorphs with special reference to their role in water transport. *Nature* **287**: 834–836.

1775 **Dunstan WA, Dell B, Malajczuk N. 1998.** The diversity of ectomycorrhizal fungi associated

1776 with introduced *Pinus* spp. in the Southern Hemisphere, with particular reference to Western

1777 Australia. *Mycorrhiza* **8**: 71–79.

1778 **Elliott TF, Truong C, Jackson SM, Zúñiga CL, Trappe JM, Vernes K. 2022.** Mammalian

1779 mycophagy: A global review of ecosystem interactions between mammals and fungi. *Fungal*

1780 *systematics and evolution* **9**: 99–159.

1781 **Erlandson SR, Margis R, Ramirez A, Nguyen N, Lofgren LA, Liao H-L, Vilgalys R,**

1782 **Kennedy PG, Peay KG. 2022.** Transcriptional acclimation and spatial differentiation

1783 characterize drought response by the ectomycorrhizal fungus *Suillus pungens*. *The New*

1784 *phytologist*.

1785 **van Etten EJB, Belen CA, Calviño-Cancela M. 2020.** Invasion patterns of *Pinus pinaster* in

1786 south-west Australia in relation to fire, vegetation type and plantation management. *Forest*

1787 *ecology and management* **463**: 118042.

1788 **Fernandez CW, Heckman K, Kolka R, Kennedy PG. 2019.** Melanin mitigates the

1789 accelerated decay of mycorrhizal necromass with peatland warming. *Ecology letters* **22**: 498–

1790 505.

1791 **Fernandez CW, Kennedy PG. 2018.** Melanization of mycorrhizal fungal necromass  
1792 structures microbial decomposer communities. *The Journal of ecology* **106**: 468–479.

1793 **Feurtey A, Lorrain C, McDonald MC, Milgate A, Solomon PS, Warren R, Puccetti G,**  
1794 **Scalliet G, Torriani SFF, Gout L, et al. 2023.** A thousand-genome panel retraces the global  
1795 spread and adaptation of a major fungal crop pathogen. *Nature communications* **14**: 1059.

1796 **Fomina MA, Alexander IJ, Colpaert JV, Gadd GM. 2005.** Solubilization of toxic metal  
1797 minerals and metal tolerance of mycorrhizal fungi. *Soil biology & biochemistry* **37**: 851–866.

1798 **Fries N. 1994.** Sexual incompatibility in *Suillus variegatus*. *Mycological research* **98**: 545–  
1799 546.

1800 **Fries Elias. 1821-[1832].** *Systema mycologicum : sistens fungorum ordines, genera et*  
1801 *species, huc usque cognitatas, quas ad normam methodi naturalis determinavit.* Lundae, Ex  
1802 Officina Berlingiana, 1821-[1832].

1803 **Fries N, Neumann W. 1990.** Sexual incompatibility in *Suillus luteus* and *S. granulatus*.  
1804 *Mycological research* **94**: 64–70.

1805 **Fries N, Sun Y-P. 1992.** The mating system of *Suillus bovinus*. *Mycological research* **96**:  
1806 237–238.

1807 **Gardes M, Bruns TD. 1996.** Community structure of ectomycorrhizal fungi in a *Pinus*  
1808 *muricata* forest: above- and below-ground views. *Canadian journal of botany. Journal*  
1809 *canadien de botanique* **74**: 1572–1583.

1810 **Geras'kin SA, Fesenko SV, Alexakhin RM. 2008.** Effects of non-human species irradiation  
1811 after the Chernobyl NPP accident. *Environment international* **34**: 880–897.

1812 **Gernandt DS, Holman G, Campbell C, Parks M, Mathews S, Raubeson LA, Liston A,**  
1813 **Stockey RA, Rothwell GW. 2016.** Phylogenetics of extant and fossil Pinaceae: methods for  
1814 increasing topological stability. *Botany* **94**: 863–884.

1815 **Gohar D, Pöldmaa K, Tedersoo L, Aslani F, Furneaux B, Henkel TW, Saar I, Smith ME,**  
1816 **Bahram M. 2022.** Global diversity and distribution of mushroom-inhabiting bacteria.  
1817 *Environmental microbiology reports* **14**: 254–264.

1818 **Grigoriev IV, Nikitin R, Haridas S, Kuo A, Ohm R, Otilar R, Riley R, Salamov A, Zhao**  
1819 **X, Korzeniewski F, et al. 2014.** MycoCosm portal: gearing up for 1000 fungal genomes.  
1820 *Nucleic acids research* **42**: D699-704.

1821 **Grubisha LC, Dowie NJ, Miller SL, Hazard C, Klooster MR. 2014.** *Rhizopogon kretzeriae*  
1822 sp. nov.: The rare fungal symbiont in the tripartite system with *Pterospora andromedea* and  
1823 *Pinus strobus*. *Botany* **92**.

1824 **Grubisha LC, Trappe JM, Molina R, Spatafora JW. 2002.** Biology of the ectomycorrhizal  
1825 genus *Rhizopogon*. VI. Re-examination of infrageneric relationships inferred from  
1826 phylogenetic analyses of ITS sequences. *Mycologia* **94**: 607–619.

1827 **Hartley J, Cairney JWG, Sanders FE, Meharg AA. 1997.** Toxic interactions of metal ions  
1828 ( $\text{Cd}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Zn}^{2+}$  and  $\text{Sb}^{3-}$ ) on *in vitro* biomass production of ectomycorrhizal fungi. *The*  
1829 *New phytologist* **137**: 551–562.

1830 **Hayward J, Horton TR, Pauchard A, Nuñez MA. 2015.** A single ectomycorrhizal fungal  
1831 species can enable a *Pinus* invasion. *Ecology* **96**: 1438–1444.

1832 **Hedger J. 1986.** *Suillus luteus* on the equator. *Bulletin of the British Mycological Society* **20**:  
1833 53–54.

1834 **Heitman J, Sun S, James TY. 2013.** Evolution of fungal sexual reproduction. *Mycologia*  
1835 **105**: 1–27.

1836 **Hinton TG, Alexakhin R, Balonov M, Gentner N, Hendry J, Prister B, Strand P,**  
1837 **Woodhead D. 2007.** Radiation-induced effects on plants and animals: findings of the United  
1838 Nations Chernobyl Forum. *Health physics* **93**: 427–440.

1839 **Hobbie E a., Agerer R. 2009.** Nitrogen isotopes in ectomycorrhizal sporocarps correspond to  
1840 belowground exploration types. *Plant and soil* **327**: 71–83.

1841 **Hoeksema JD, Averill C, Bhatnagar JM, Brzostek E, Buscardo E, Chen K-H, Liao H-L,**  
1842 **Nagy L, Policelli N, Ridgeway J, et al. 2020.** Ectomycorrhizal Plant-Fungal Co-invasions as  
1843 Natural Experiments for Connecting Plant and Fungal Traits to Their Ecosystem  
1844 Consequences. *Frontiers in Forests and Global Change* **3**.

1845 **Hoffman CS, Wood V, Fantes PA. 2015.** An Ancient Yeast for Young Geneticists: A Primer

1846 on the *Schizosaccharomyces pombe* Model System. *Genetics* **201**: 403–423.

1847 **Horton TR. 2006.** The number of nuclei in basidiospores of 63 species of ectomycorrhizal

1848 Homobasidiomycetes. *Mycologia* **98**: 233–238.

1849 **Horton TR. 2017.** Spore Dispersal in Ectomycorrhizal Fungi at Fine and Regional Scales. In:

1850 Tedersoo L, ed. Biogeography of Mycorrhizal Symbiosis. Cham: Springer International

1851 Publishing, 61–78.

1852 **Hou Z, Wei Y, Sun L, Xia R, Xu H, Li Y, Feng Y, Fan W, Xin G. 2022.** Effects of drying

1853 temperature on umami taste and aroma profiles of mushrooms (*Suillus granulatus*). *Journal of*

1854 *food science* **87**: 1983–1998.

1855 **Hu Q-H, Weng J-Q, Wang J-S. 2010.** Sources of anthropogenic radionuclides in the

1856 environment: a review. *Journal of environmental radioactivity* **101**: 426–437.

1857 **Hynson NA, Merckx VSFT, Perry BA, Treseder KK. 2013.** Identities and distributions of

1858 the co-invading ectomycorrhizal fungal symbionts of exotic pines in the Hawaiian Islands.

1859 *Biological invasions* **15**: 2373–2385.

1860 **Izumi H, Anderson IC, Alexander IJ, Killham K, Moore ERB. 2006.** Endobacteria in

1861 some ectomycorrhiza of Scots pine (*Pinus sylvestris*). *FEMS microbiology ecology* **56**: 34–43.

1862 **Jacobson KM, Miller OK. 1994.** Postmeiotic mitosis in the basidia of *Suillus granulatus*:

1863 Implications for population structure and dispersal biology. *Mycologia* **86**: 511–516.

1864 **Jenkins ML, Cripps CL, Gains-Germain L. 2018.** Scorched Earth: *Suillus* colonization of

1865 *Pinus albicaulis* seedlings planted in wildfire-impacted soil affects seedling biomass, foliar

1866 nutrient content, and isotope signatures. *Plant and soil* **425**: 113–131.

1867 **Jonsson L, Dahlberg A, Nilsson M-C, Kårén O, Zackrisson O. 1999.** Continuity of

1868 ectomycorrhizal fungi in self-regenerating boreal *Pinus sylvestris* forests studied by

1869 comparing mycobiont diversity on seedlings and mature trees. *The New phytologist* **142**: 151–

1870 162.

1871 **Kabir MA, Hussain MA, Ahmad Z. 2012.** *Candida albicans*: A Model Organism for

1872 Studying Fungal Pathogens. *International Scholarly Research Notices* **2012**.

1873 **Kalsotra T, Khullar S, Agnihotri R, Reddy MS. 2018.** Metal induction of two

1874 metallothionein genes in the ectomycorrhizal fungus *Suillus himalayensis* and their role in

1875 metal tolerance. *Microbiology* **164**: 868–876.

1876 **Kamada T, Sano H, Nakazawa T, Nakahori K. 2010.** Regulation of fruiting body

1877 photomorphogenesis in *Coprinopsis cinerea*. *Fungal genetics and biology: FG & B* **47**: 917–

1878 921.

1879 **Karathia H, Vilaprinyo E, Sorribas A, Alves R. 2011.** *Saccharomyces cerevisiae* as a model

1880 organism: a comparative study. *PLoS one* **6**: e16015.

1881 **Karlsen-Ayala E, Smith ME, Askey BC, Gazis R. 2022.** Native ectomycorrhizal fungi from

1882 the endangered pine rocklands are superior symbionts to commercial inoculum for slash pine

1883 seedlings. *Mycorrhiza* **32**: 465–480.

1884 **Kawai M, Yamahara M, Ohta A. 2008.** Bipolar incompatibility system of an

1885 ectomycorrhizal basidiomycete, *Rhizopogon rubescens*. *Mycorrhiza* **18**: 205–210.

1886 **Ke Y-H, Branco S, Bazzicalupo AL, Nguyen NH, Liao H-L, Kennedy P, Bruns TD, Kuo**

1887 **A, LaButti K, Barry K, et al. 2023.** Genomic determination of breeding systems and trans-

1888 specific evolution of HD MAT genes in suilloid fungi. *Genetics* **224**.

1889 **Kennedy P. 2010.** Ectomycorrhizal fungi and interspecific competition: species interactions,

1890 community structure, coexistence mechanisms, and future research directions. *The New*

1891 *phytologist* **187**: 895–910.

1892 **Kennedy PG, Gagne J, Perez-Pazos E, Lofgren LA, Nguyen NH. 2020.** Does fungal

1893 competitive ability explain host specificity or rarity in ectomycorrhizal symbioses? *PLoS one*

1894 **15**: e0234099.

1895 **Kennedy PG, Higgins LM, Rogers RH, Weber MG. 2011.** Colonization-competition

1896 tradeoffs as a mechanism driving successional dynamics in ectomycorrhizal fungal

1897 communities. *PLoS one* **6**: e25126.

1898 **Kennedy PG, Maillard F. 2023.** Knowns and unknowns of the soil fungal necrobiome.

1899 *Trends in microbiology* **31**: 173–180.

1900 **Kennedy PG, Mielke LA, Nguyen NH. 2018.** Ecological responses to forest age, habitat,

1901 and host vary by mycorrhizal type in boreal peatlands. *Mycorrhiza* **28**: 315–328.

1902 **Kennedy PG, Peay KG. 2007.** Different soil moisture conditions change the outcome of the

1903 ectomycorrhizal symbiosis between *Rhizopogon* species and *Pinus muricata*. *Plant and soil*

1904 **291**: 155–165.

1905 **Kennedy PG, Smith DP, Horton TR, Molina RJ. 2012.** *Arbutus menziesii* (Ericaceae)

1906 facilitates regeneration dynamics in mixed evergreen forests by promoting mycorrhizal fungal

1907 diversity and host connectivity. *American journal of botany* **99**: 1691–1701.

1908 **Kohler A, Kuo A, Nagy LG, Morin E, Barry KW, Buscot F, Canbäck B, Choi C,**

1909 **Cichocki N, Clum A, et al. 2015.** Convergent losses of decay mechanisms and rapid turnover

1910 of symbiosis genes in mycorrhizal mutualists. *Nature genetics* **47**: 410–415.

1911 **Koide RT, Fernandez C, Malcolm G. 2014.** Determining place and process: functional traits

1912 of ectomycorrhizal fungi that affect both community structure and ecosystem function. *The*

1913 *New phytologist* **201**: 433–439.

1914 **Kolaj-Robin O, Russell D, Hayes KA, Pembroke JT, Soulimane T. 2015.** Cation Diffusion

1915 Facilitator family: Structure and function. *FEBS letters* **589**: 1283–1295.

1916 **Kothamasi D, Wannijn J, Van Hees M, Nauts R, Van Gompel A, Vanhoudt N,**

1917 **Vandenhove H. 2019.** Exposure to ionizing radiation affects the growth of ectomycorrhizal

1918 fungi and induces increased melanin production and increased capacities of reactive oxygen

1919 species scavenging enzymes. *Journal of environmental radioactivity* **197**: 16–22.

1920 **Koukol O, Valda S, Gaisler J, Kunca V, Dowie NJ. 2022.** *Rhizopogon confusus* sp. nov., a

1921 correct name for a fungus previously recorded in Central Europe as the North American

1922 *Rhizopogon salebrosus*. *Mycological progress* **21**: 49.

1923 **Kretzer A, Bruns TD. 1997.** Molecular revisitation of the genus *Gastrospora*. *Mycologia*

1924 **89**: 586–589.

1925 **Kretzer A, Li Y, Szaro T, Bruns TD. 1996.** Internal transcribed spacer sequences from 38

1926 recognized species of *Suillus* sensu lato: Phylogenetic and taxonomic implications. *Mycologia*

1927 **88**: 776–785.

1928 **Krupa P, Kozdrój J. 2004.** Accumulation of Heavy Metals by Ectomycorrhizal Fungi

1929 Colonizing Birch Trees Growing in an Industrial Desert Soil. *World journal of microbiology*

1930 *& biotechnology* **20**: 427–430.

1931 **Krznaric E, Verbruggen N, Wevers JHL, Carleer R, Vangronsveld J, Colpaert JV. 2009.**

1932 Cd-tolerant *Suillus luteus*: a fungal insurance for pines exposed to Cd. *Environmental*

1933 *pollution* **157**: 1581–1588.

1934 **Krznaric E, Wevers JHL, Cloquet C, Vangronsveld J, Vanhaecke F, Colpaert JV. 2010.**

1935 Zn pollution counteracts Cd toxicity in metal-tolerant ectomycorrhizal fungi and their host

1936 plant, *Pinus sylvestris*. *Environmental microbiology* **12**: 2133–2141.

1937 **Ladeyn I, Plassard C, Staunton S. 2008.** Mycorrhizal association of maritime pine, *Pinus*

1938 *pinaster*, with *Rhizopogon roseolus* has contrasting effects on the uptake from soil and root-

1939 to-shoot transfer of <sup>137</sup>Cs, <sup>85</sup>Sr and <sup>95m</sup>Tc. *Journal of environmental radioactivity* **99**: 853–863.

1940 **Lee SH, Calvo-Polanco M, Chung GC, Zwiazek JJ. 2010.** Role of aquaporins in root water

1941 transport of ectomycorrhizal jack pine (*Pinus banksiana*) seedlings exposed to NaCl and

1942 fluoride. *Plant, cell & environment* **33**: 769–780.

1943 **Leopold P, Renaud PE, Ambrose WG, Berge J. 2019.** High Arctic *Mytilus* spp.:

1944 occurrence, distribution and history of dispersal. *Polar biology* **42**: 237–244.

1945 **Lepage B, Currah R, Stockey R, Rothwell G. 1997.** Fossil ectomycorrhizae from the

1946 Middle Eocene. *American journal of botany* **84**: 410.

1947 **Leski T, Rudawska M, Kieliszewska-Rokicka B. 1995.** Intraspecific aluminium response in

1948 *Suillus luteus* (L.) s.f. gray., an ectomycorrhizal symbiont of scots pine. *Acta Societatis*

1949 *Botanicorum Poloniae* **64**: 97–105.

1950 **Li M, Wang H, Zhao X, Lu Z, Sun X, Ding G. 2021.** Role of *Suillus placidus* in Improving

1951 the Drought Tolerance of Masson Pine (*Pinus massoniana* Lamb.) Seedlings. *Forests, Trees*

1952 *and Livelihoods* **12**: 332.

1953 **Liao H-L, Chen Y, Vilgalys R. 2016.** Metatranscriptomic Study of Common and Host-

1954 Specific Patterns of Gene Expression between Pines and Their Symbiotic Ectomycorrhizal

1955 Fungi in the Genus *Suillus*. *PLoS genetics* **12**: e1006348.

- 1956 **van der Linde S, Suz LM, Orme CDL, Cox F, Andreae H, Asi E, Atkinson B, Benham S,**  
1957 **Carroll C, Cools N, et al. 2018.** Environment and host as large-scale controls of  
1958 ectomycorrhizal fungi. *Nature* **558**: 243–248.
- 1959 **Liu H, Chen H, Ding G, Li K, Ren Q. 2020.** Identification of candidate genes conferring  
1960 tolerance to aluminum stress in *Pinus massoniana* inoculated with ectomycorrhizal fungus.  
1961 *BMC plant biology* **20**: 521.
- 1962 **Liu H, Chen H, Ding G, Li K, Wang Y. 2021.** Proteomic Insight into the Symbiotic  
1963 Relationship of *Pinus massoniana* Lamb and *Suillus luteus* towards Developing Al-Stress  
1964 Resistance. *Life* **11**.
- 1965 **Lofgren L, Nguyen NH, Kennedy PG. 2018.** Ectomycorrhizal host specificity in a changing  
1966 world: can legacy effects explain anomalous current associations? *The New phytologist* **220**:  
1967 1273–1284.
- 1968 **Lofgren LA, Nguyen NH, Vilgalys R, Ruytinx J, Liao H-L, Branco S, Kuo A, LaButti K,**  
1969 **Lipzen A, Andreopoulos W, et al. 2021.** Comparative genomics reveals dynamic genome  
1970 evolution in host specialist ectomycorrhizal fungi. *The New phytologist* **230**: 774–792.
- 1971 **Lofgren LA, Uehling JK, Branco S, Bruns TD, Martin F, Kennedy PG. 2019.** Genome-  
1972 based estimates of fungal rDNA copy number variation across phylogenetic scales and  
1973 ecological lifestyles. *Molecular ecology* **28**: 721–730.
- 1974 **Looney BP, Meidl P, Piatek MJ, Miettinen O, Martin FM, Matheny PB, Labbé JL.**  
1975 **2018.** Russulaceae: a new genomic dataset to study ecosystem function and evolutionary  
1976 diversification of ectomycorrhizal fungi with their tree associates. *The New phytologist* **218**:  
1977 54–65.
- 1978 **Lu J, Salzberg SL. 2020.** Ultrafast and accurate 16S rRNA microbial community analysis  
1979 using Kraken 2. *Microbiome* **8**: 124.
- 1980 **Maillard F, Fernandez CW, Mundra S, Heckman KA, Kolka RK, Kauserud H,**  
1981 **Kennedy PG. 2022.** Warming drives a “hummockification” of microbial communities  
1982 associated with decomposing mycorrhizal fungal necromass in peatlands. *The New*  
1983 *phytologist* **234**: 2032–2043.
- 1984 **Maillard F, Kohler A, Morin E, Hossann C, Miyauchi S, Ziegler-Devin I, Gérant D,**  
1985 **Angeli N, Lipzen A, Keymanesh K, et al. 2023.** Functional genomics gives new insights into  
1986 the ectomycorrhizal degradation of chitin. *The New phytologist* **238**: 845–858.
- 1987 **Malcolm GM, López-Gutiérrez JC, Koide RT, Eissenstat DM. 2008.** Acclimation to  
1988 temperature and temperature sensitivity of metabolism by ectomycorrhizal fungi. *Global*  
1989 *change biology* **14**: 1169–1180.
- 1990 **Marmeisse R, Guidot A, Gay G, Lambilliotte R, Sentenac H, Combier J-P, Melayah D,**  
1991 **Fraissinet-Tachet L, Debaud JC. 2004.** *Hebeloma cylindrosporum*- a model species to study  
1992 ectomycorrhizal symbiosis from gene to ecosystem. *The New phytologist* **163**: 481–498.
- 1993 **Martin F, Aerts A, Ahrén D, Brun A, Danchin EGJ, Duchaussoy F, Gibon J, Kohler A,**  
1994 **Lindquist E, Pereda V, et al. 2008.** The genome of *Laccaria bicolor* provides insights into  
1995 mycorrhizal symbiosis. *Nature* **452**: 88–92.
- 1996 **Marx DH. 1992.** The practical significance of ectomycorrhizae in forest establishment. In: 27.  
1997 **Marx DH, Marrs LF, Cordell CE. 2002.** Practical use of the mycorrhizal fungal technology  
1998 in forestry, reclamation, arboriculture, agriculture, and horticulture. *Dendrobiology* **47**: 27–40.
- 1999 **Massicotte HB, Molina R, Luoma DL, Smith JE. 1994.** Biology of the ectomycorrhizal  
2000 genus, *Rhizopogon*. II. Patterns of host-fungus specificity following spore inoculation of  
2001 diverse hosts grown in monoculture and dual culture. *The New phytologist* **126**: 677–690.
- 2002 **Mateos E, Olaizola J, Pajares JA, Pando V, Diez JJ. 2017.** Influence of *Suillus luteus* on  
2003 *Fusarium* damping-off in pine seedlings. *African Journal of Biotechnology* **16**: 268–273.
- 2004 **McNabb RFR. 1968.** The Boletaceae of New Zealand. *New Zealand journal of botany* **6**:  
2005 137–176.
- 2006 **Mikola P. 1970.** Mycorrhizal Inoculation in Afforestation. In: Romberger JA, Mikola P, eds.  
2007 International Review of Forestry Research. Elsevier, 123–196.
- 2008 **Miller-Fleming L, Giorgini F, Outeiro TF. 2008.** Yeast as a model for studying human  
2009 neurodegenerative disorders. *Biotechnology journal* **3**: 325–338.
- 2010 **Miyamoto Y, Danilov AV, Bryanin SV. 2021.** The dominance of *Suillus* species in

2011 ectomycorrhizal fungal communities on *Larix gmelinii* in a post-fire forest in the Russian Far  
2012 East. *Mycorrhiza* **31**: 55–66.

2013 **Miyamoto Y, Maximov TC, Sugimoto A, Nara K. 2019.** Discovery of *Rhizopogon*  
2014 associated with *Larix* from northeastern Siberia: Insights into host shift of ectomycorrhizal  
2015 fungi. *Mycoscience* **60**: 274–280.

2016 **Miyauchi S, Kiss E, Kuo A, Drula E, Kohler A, Sánchez-García M, Morin E,**  
2017 **Andreopoulos B, Barry KW, Bonito G, et al. 2020.** Large-scale genome sequencing of  
2018 mycorrhizal fungi provides insights into the early evolution of symbiotic traits. *Nature*  
2019 *communications* **11**: 5125.

2020 **Moeller HV, Dickie IA, Peltzer DA, Fukami T. 2015.** Mycorrhizal co-invasion and novel  
2021 interactions depend on neighborhood context. *Ecology* **96**: 2336–2347.

2022 **Moeller HV, Peay KG. 2016.** Competition-function tradeoffs in ectomycorrhizal fungi.  
2023 *PeerJ* **4**: e2270.

2024 **Molina R, Massicotte H, Trappe JM. 1992.** Specificity phenomena in mycorrhizal  
2025 symbioses: community-ecological consequences and practical applications. In: Chapman and  
2026 Hall, 357.

2027 **Molina R, Smith JE, McKay D, Melville LH. 1997.** Biology of the ectomycorrhizal genus,  
2028 *Rhizopogon*: III. Influence of co-cultured conifer species on mycorrhizal specificity with the  
2029 arbutoid hosts *Arctostaphylos uva-ursi* and *Arbutus menziesii*. *The New phytologist* **137**: 519–  
2030 528.

2031 **Monteiro M, Reino L, Schertler A, Essl F, Figueira R, Ferreira MT, Capinha C. 2020.** A  
2032 database of the global distribution of alien macrofungi. *Biodiversity data journal* **8**: e51459.

2033 **Mudbhari S, Lofgren L, Appidi MR, Vilgalys R, Hettich RL, Abraham P. 2023.**  
2034 Decoding the chemical language of *Suillus* fungi: genome mining and untargeted  
2035 metabolomics uncover terpene chemical diversity. *bioRxiv*: 2023.11.20.567897.

2036 **Mujic AB, Kuo A, Tritt A, Lipzen A, Chen C, Johnson J, Sharma A, Barry K, Grigoriev**  
2037 **IV, Spatafora JW. 2017.** Comparative Genomics of the Ectomycorrhizal Sister Species  
2038 *Rhizopogon vinicolor* and *Rhizopogon vesiculosus* (Basidiomycota: Boletales) Reveals a  
2039 Divergence of the Mating Type B Locus. *G3* **7**: 1775–1789.

2040 **Mujic AB, Policelli N, Nuñez MA, Truong C, Smith ME. 2023.** Co-invasive  
2041 ectomycorrhizal fungi alter native soil fungal communities. *Plant and soil* **484**: 547–567.

2042 **Muller LAH, Vangronsveld J, Colpaert JV. 2007.** Genetic structure of *Suillus luteus*  
2043 populations in heavy metal polluted and nonpolluted habitats. *Molecular ecology* **16**: 4728–  
2044 4737.

2045 **Murakami C, Kaeberlein M. 2009.** Quantifying yeast chronological life span by outgrowth  
2046 of aged cells. *Journal of visualized experiments: JoVE*.

2047 **Murrill WA. 1909.** The Boletaceae of North America—I. *Mycologia* **1**: 4–18.

2048 **Nara K. 2006.** Pioneer dwarf willow may facilitate tree succession by providing late  
2049 colonizers with compatible ectomycorrhizal fungi in a primary successional volcanic desert.  
2050 *The New phytologist* **171**: 187–197.

2051 **Nguyen NH. 2023.** Fungal Hyphosphere Microbiomes Are Distinct from Surrounding  
2052 Substrates and Show Consistent Association Patterns. *Microbiology spectrum* **11**: e0470822.

2053 **Nguyen NH, Hynson NA, Bruns TD. 2012.** Stayin’ alive: survival of mycorrhizal fungal  
2054 propagules from 6-yr-old forest soil. *Fungal ecology* **5**: 741–746.

2055 **Nguyen H, Rineau F, Vangronsveld J, Cuypers A, Colpaert JV, Ruytinx J. 2017.** A novel,  
2056 highly conserved metallothionein family in basidiomycete fungi and characterization of two  
2057 representative *SIMTa* and *SIMTb* genes in the ectomycorrhizal fungus *Suillus luteus*.  
2058 *Environmental microbiology* **19**: 2577–2587.

2059 **Nguyen NH, Song Z, Bates ST, Branco S, Tedersoo L, Menke J, Schilling JS, Kennedy**  
2060 **PG. 2016a.** FUNGuild: An open annotation tool for parsing fungal community datasets by  
2061 ecological guild. *Fungal ecology* **20**: 241–248.

2062 **Nguyen NH, Vellinga EC, Bruns TD, Kennedy PG. 2016b.** Phylogenetic assessment of  
2063 global *Suillus* ITS sequences supports morphologically defined species and reveals  
2064 synonymous and undescribed taxa. *Mycologia* **108**: 1216–1228.

2065 **Nieuwenhuis BPS, Billiard S, Vuilleumier S, Petit E, Hood ME, Giraud T. 2013.**

2066 Evolution of uni- and bifactorial sexual compatibility systems in fungi. *Heredity* **111**: 445–  
2067 455.

2068 **Nikolova I, Johanson KJ, Dahlberg A. 1997.** Radiocaesium in fruitbodies and mycorrhizae  
2069 in ectomycorrhizal fungi. *Journal of environmental radioactivity* **37**: 115–125.

2070 **Nuñez MA, Hayward J, Horton TR, Amico GC, Dimarco RD, Barrios-Garcia MN,**  
2071 **Simberloff D. 2013.** Exotic mammals disperse exotic fungi that promote invasion by exotic  
2072 trees. *PloS one* **8**: e66832.

2073 **Nuñez MA, Horton TR, Simberloff D. 2009.** Lack of belowground mutualisms hinders  
2074 Pinaceae invasions. *Ecology* **90**: 2352–2359.

2075 **Nygren CMR, Edqvist J, Elfstrand M, Heller G, Taylor AFS. 2007.** Detection of  
2076 extracellular protease activity in different species and genera of ectomycorrhizal fungi.  
2077 *Mycorrhiza* **17**: 241–248.

2078 **Ohenoja E, Koistinen R. 1984.** Fruit body production of larger fungi in Finland. 2: Edible  
2079 fungi in northern Finland 1976—1978. *Annales botanici Fennici* **21**: 357–366.

2080 **Olsson PA, Münzenberger B, Mahmood S, Erland S. 2000.** Molecular and anatomical  
2081 evidence for a three-way association between *Pinus sylvestris* and the ectomycorrhizal fungi  
2082 *Suillus bovinus* and *Gomphidius roseus*. *Mycological research* **104**: 1372–1378.

2083 **Palmer JG. 1971.** *Techniques and procedures for culturing ectomycorrhizal fungi*  
2084 *Mycorrhizae: Proceedings of the First North American Conference on Mycorrhizae*. USDA,  
2085 U.S. Forest Service.

2086 **Palumbi SR, Barshis DJ, Traylor-Knowles N, Bay RA. 2014.** Mechanisms of reef coral  
2087 resistance to future climate change. *Science* **344**: 895–898.

2088 **Paul LR, Chapman BK, Chanway CP. 2007.** Nitrogen fixation associated with *Suillus*  
2089 *tomentosus* tuberculate ectomycorrhizae on *Pinus contorta* var. *latifolia*. *Annals of botany* **99**:  
2090 1101–1109.

2091 **Paul LR, Chapman W K, Chanway CP. 2012.** Diazotrophic bacteria reside inside *Suillus*  
2092 *tomentosus* / *Pinus contorta* tuberculate ectomycorrhizae. *Botany*.

2093 **Peay KG. 2016.** The Mutualistic Niche: Mycorrhizal Symbiosis and Community Dynamics.  
2094 *Annual review of ecology, evolution, and systematics* **47**: 143–164.

2095 **Peay KG, Bruns TD, Kennedy PG, Bergemann SE, Garbelotto M. 2007.** A strong species-  
2096 area relationship for eukaryotic soil microbes: island size matters for ectomycorrhizal fungi.  
2097 *Ecology letters* **10**: 470–480.

2098 **Peay KG, Russo SE, McGuire KL, Lim Z, Chan JP, Tan S, Davies SJ. 2015.** Lack of host  
2099 specificity leads to independent assortment of dipterocarps and ectomycorrhizal fungi across a  
2100 soil fertility gradient (J Chave, Ed.). *Ecology letters*.

2101 **Peay KG, Schubert MG, Nguyen NH, Bruns TD. 2012.** Measuring ectomycorrhizal fungal  
2102 dispersal: macroecological patterns driven by microscopic propagules. *Molecular ecology* **21**:  
2103 4122–4136.

2104 **Pérez-Pazos E, Certano A, Gagne J, Lebeuf R, Siegel N, Nguyen N, Kennedy PG. 2021.**  
2105 The slippery nature of ectomycorrhizal host specificity: *Suillus* fungi associated with novel  
2106 pinoid (*Picea*) and abietoid (*Abies*) hosts. *Mycologia* **113**: 891–901.

2107 **Pietras M, Litkowiec M, Gołębiewska J. 2018.** Current and potential distribution of the  
2108 ectomycorrhizal fungus *Suillus lakei* ((Murrill) A.H. Sm. & Thiers) in its invasion range.  
2109 *Mycorrhiza* **28**: 467–475.

2110 **Pildain MB, Marchelli P, Azpilicueta MM, Starik C, Barroetaveña C. 2021.**  
2111 Understanding introduction history: Genetic structure and diversity of the edible  
2112 ectomycorrhizal fungus, *Suillus luteus*, in Patagonia (Argentina). *Mycologia* **113**: 715–724.

2113 **Plaza DF, Lin C-W, van der Velden NSJ, Aebi M, Künzler M. 2014.** Comparative  
2114 transcriptomics of the model mushroom *Coprinopsis cinerea* reveals tissue-specific armories  
2115 and a conserved circuitry for sexual development. *BMC genomics* **15**: 492.

2116 **Plett JM, Kohler A, Martin F. 2024.** Masters of Manipulation: How Our Molecular  
2117 Understanding of Model Symbiotic Fungi and Their Hosts Is Changing the Face of  
2118 “Mutualism.” In: Hsueh Y-P, Blackwell M, eds. *Fungal Associations*. Cham: Springer  
2119 International Publishing, 249–272.

2120 **Policelli N, Bruns TD, Vilgalys R, Nuñez MA. 2019.** Suilloid fungi as global drivers of pine

2121 invasions. *The New phytologist* **222**: 714–725.

2122 **Policelli N, Horton TR, García RA, Naour M, Pauchard A, Nuñez MA. 2020.** Native and  
 2123 non-native trees can find compatible, mycorrhizal partners in each other's dominated areas.  
 2124 *Plant and soil* **454**: 285–297.

2125 **Policelli N, Horton TR, Kitzberger T, Nuñez MA. 2022.** Invasive ectomycorrhizal fungi  
 2126 can disperse in the absence of their known vectors. *Fungal ecology* **55**: 101124.

2127 **Pumpanen J, Ohashi M, Endo I, Hari P, Bäck J, Kulmala M, Ohte N. 2016.** <sup>137</sup>Cs  
 2128 distributions in soil and trees in forest ecosystems after the radioactive fallout - Comparison  
 2129 study between southern Finland and Fukushima, Japan. *Journal of environmental*  
 2130 *radioactivity* **161**: 73–81.

2131 **Qi J, Yin D. 2022.** Effects of *Suillus luteus* on the Growth, Photosynthesis, Stomata, and  
 2132 Root System of *Pinus tabulaeformis* Under Drought Stress. *Journal of plant growth*  
 2133 *regulation*: 1–12.

2134 **Qi J, Yin D. 2023.** Effects of *Suillus luteus* on the Growth, Photosynthesis, Stomata, and Root  
 2135 System of *Pinus tabulaeformis* Under Drought Stress. *Journal of plant growth regulation* **42**:  
 2136 3486–3497.

2137 **Quince C, Walker AW, Simpson JT, Loman NJ, Segata N. 2017.** Shotgun metagenomics,  
 2138 from sampling to analysis. *Nature biotechnology* **35**: 833–844.

2139 **Randall BL, Grand LF. 1985.** Morphology and possible mycobiont (*Suillus pictus*) of a  
 2140 tuberculate ectomycorrhiza on *Pinus strobus*. *Canadian journal of botany. Journal canadien*  
 2141 *de botanique*.

2142 **Read DJ. 1998.** The Mycorrhizal Status of *Pinus*. In: Richardson D, ed. Ecology and  
 2143 Biogeography of *Pinus*. Cambridge University Press, 324–340.

2144 **Ribeiro AM, Canchaya CA, Penalzoza F, Galindo J, da Fonseca RR. 2019.** Population  
 2145 genomic footprints of environmental pollution pressure in natural populations of the  
 2146 Mediterranean mussel. *Marine genomics* **45**: 11–15.

2147 **Richardson DM, Williams PA, Hobbs RJ. 1994.** Pine Invasions in the Southern  
 2148 Hemisphere: Determinants of Spread and Invasibility. *Journal of biogeography* **21**: 511–527.

2149 **Rineau F, Stas J, Nguyen NH, Kuyper TW, Carleer R, Vangronsveld J, Colpaert JV,**  
 2150 **Kennedy PG. 2016.** Ectomycorrhizal Fungal Protein Degradation Ability Predicted by Soil  
 2151 Organic Nitrogen Availability. *Applied and environmental microbiology* **82**: 1391–1400.

2152 **Roche CM, Loros JJ, McCluskey K, Glass NL. 2014.** *Neurospora crassa*: looking back and  
 2153 looking forward at a model microbe. *American journal of botany* **101**: 2022–2035.

2154 **Rochet J, Moreau P-A, Manzi S, Gardes M. 2011.** Comparative phylogenies and host  
 2155 specialization in the alder ectomycorrhizal fungi *Alnicola*, *Alpova* and *Lactarius*  
 2156 (Basidiomycota) in Europe. *BMC evolutionary biology* **11**: 40.

2157 **Ronda O, Grządka E, Ostolska I, Orzel J, Cieślak BM. 2022.** Accumulation of  
 2158 radioisotopes and heavy metals in selected species of mushrooms. *Food chemistry* **367**:  
 2159 130670.

2160 **Roy M, Dubois M-P, Proffit M, Vincenot L, Desmarais E, Selosse M-A. 2008.** Evidence  
 2161 from population genetics that the ectomycorrhizal basidiomycete *Laccaria amethystina* is an  
 2162 actual multihost symbiont. *Molecular ecology* **17**: 2825–2838.

2163 **Ruytinx J, Coninx L, Nguyen H, Smisdorn N, Morin E, Kohler A, Cuypers A, Colpaert**  
 2164 **JV. 2017.** Identification, evolution and functional characterization of two Zn CDF-family  
 2165 transporters of the ectomycorrhizal fungus *Suillus luteus*. *Environmental microbiology reports*  
 2166 **9**: 419–427.

2167 **Ruytinx J, Craciun AR, Verstraelen K, Vangronsveld J, Colpaert JV, Verbruggen N.**  
 2168 **2011.** Transcriptome analysis by cDNA-AFLP of *Suillus luteus* Cd-tolerant and Cd-sensitive  
 2169 isolates. *Mycorrhiza* **21**: 145–154.

2170 **Ruytinx J, Nguyen H, Van Hees M, Op De Beeck M, Vangronsveld J, Carleer R,**  
 2171 **Colpaert JV, Adriaensen K. 2013.** Zinc export results in adaptive zinc tolerance in the  
 2172 ectomycorrhizal basidiomycete *Suillus bovinus*. *Metallomics: integrated biometal science* **5**:  
 2173 1225–1233.

2174 **Sánchez-García M, Ryberg M, Khan FK, Varga T, Nagy LG, Hibbett DS. 2020.** Fruiting  
 2175 body form, not nutritional mode, is the major driver of diversification in mushroom-forming

2176 fungi. *Proceedings of the National Academy of Sciences of the United States of America* **117**:  
2177 32528–32534.

2178 **Sanger TJ, Kircher BK. 2017.** Model Clades Versus Model Species: Anolis Lizards as an  
2179 Integrative Model of Anatomical Evolution. In: Sheng G, ed. *Avian and Reptilian*  
2180 *Developmental Biology: Methods and Protocols*. New York, NY: Springer New York, 285–  
2181 297.

2182 **Sapsford SJ, Dickie IA. 2023.** Slow soil enzyme recovery following invasive tree removal  
2183 through gradual changes in bacterial and fungal communities. *The Journal of ecology*.  
2184 **Sapsford SJ, Wakelin A, Peltzer DA, Dickie IA. 2022.** Pine invasion drives loss of soil  
2185 fungal diversity. *Biological invasions* **24**: 401–414.

2186 **Sato H, Toju H. 2019.** Timing of evolutionary innovation: scenarios of evolutionary  
2187 diversification in a species-rich fungal clade, Boletales. *The New phytologist* **222**: 1924–1935.

2188 **Sato H, Yumoto T, Murakami N. 2007.** Cryptic species and host specificity in the  
2189 ectomycorrhizal genus *Strobilomyces* (Strobilomycetaceae). *American journal of botany* **94**:  
2190 1630–1641.

2191 **Selker EU, Cambareri EB, Jensen BC, Haack KR. 1987.** Rearrangement of duplicated  
2192 DNA in specialized cells of *Neurospora*. *Cell* **51**: 741–752.

2193 **Shemesh H, Bruns TD, Peay KG, Kennedy PG, Nguyen NH. 2023.** Changing balance  
2194 between dormancy and mortality determines the trajectory of ectomycorrhizal fungal spore  
2195 longevity over a 15-yr burial experiment. *The New phytologist* **238**: 11–15.

2196 **Simberloff D. 2006.** Invasional meltdown 6 years later: important phenomenon, unfortunate  
2197 metaphor, or both? *Ecology letters* **9**: 912–919.

2198 **Simberloff D, Von Holle B. 1999.** Positive Interactions of Nonindigenous Species:  
2199 Invasional Meltdown? *Biological invasions* **1**: 21–32.

2200 **Smith ME, Castellano MA, Frank JL. 2018.** *Hymenogaster macmurphyi* and  
2201 *Splanchnomyces behrii* are sequester species of *Xerocomellus* from the western United  
2202 States. *Mycologia* **110**: 605–617.

2203 **Smith ME, Douhan GW, Fremier AK, Rizzo DM. 2009.** Are true multihost fungi the  
2204 exception or the rule? Dominant ectomycorrhizal fungi on *Pinus sabiniana* differ from those  
2205 on co-occurring *Quercus* species. *The New phytologist* **182**: 295–299.

2206 **Smith SE, Read DJ. 2010.** *Mycorrhizal Symbiosis*. Academic Press.

2207 **Tang N, Lebreton A, Xu W, Dai Y, Yu F, Martin FM. 2021.** Transcriptome Profiling  
2208 Reveals Differential Gene Expression of Secreted Proteases and Highly Specific Gene  
2209 Repertoires Involved in *Lactarius-Pinus* Symbioses. *Frontiers in plant science* **12**: 714393.

2210 **Tedersoo L, May TW, Smith ME. 2010.** Ectomycorrhizal lifestyle in fungi: global diversity,  
2211 distribution, and evolution of phylogenetic lineages. *Mycorrhiza* **20**: 217–263.

2212 **Tedersoo L, Smith ME. 2013.** Lineages of ectomycorrhizal fungi revisited: Foraging  
2213 strategies and novel lineages revealed by sequences from belowground. *Fungal biology*  
2214 *reviews* **27**: 83–99.

2215 **Tedersoo L, Suvi T, Jairus T, Ostonen I, Pölme S. 2009.** Revisiting ectomycorrhizal fungi  
2216 of the genus *Alnus*: differential host specificity, diversity and determinants of the fungal  
2217 community. *The New phytologist* **182**: 727–735.

2218 **Theodorou C. 1967.** Inoculation with Pure Cultures of Mycorrhizal Fungi of Radiata Pine  
2219 Growing in Partially Sterilized Soil. *Australian Forestry* **31**: 303–309.

2220 **Theodorou C, Bowen GD. 1973.** Inoculation of seeds and soil with basidiospores of  
2221 mycorrhizal fungi. *Soil biology & biochemistry* **5**: 765–771.

2222 **Thompson L, Swift SOI, Egan CP, Yogi D, Chapin T, Hynson NA. 2022.** Traits and  
2223 tradeoffs among non-native ectomycorrhizal fungal symbionts affect pine seedling  
2224 establishment in a Hawaiian coinvasion landscape. *Molecular ecology* **31**: 4176–4187.

2225 **Thompson LR, The Earth Microbiome Project Consortium, Sanders JG, McDonald D,**  
2226 **Amir A, Ladau J, Locey KJ, Prill RJ, Tripathi A, Gibbons SM, et al. 2017.** A communal  
2227 catalogue reveals Earth’s multiscale microbial diversity. *Nature* **551**: 457–463.

2228 **Tilman D. 1994.** Competition and biodiversity in spatially structured habitats. *Ecology* **75**: 2–  
2229 16.

2230 **Timonen S, Hurek T. 2011.** Characterization of culturable bacterial populations associating

2231 with *Pinus sylvestris* – *Suillus bovinus* mycorrhizospheres. *Canadian journal of microbiology*.

2232 **Torres P, Honrubia M. 1994.** Inoculation of containerized *Pinus halepensis* (Miller)

2233 seedlings with basidiospores of *Pisolithus arhizus* (Pers) Rauschert, *Rhizopogon roseolus*

2234 (Corda) Th M Fr and *Suillus collinitus* (Fr) O Kuntze. *Annales des Sciences Forestières* **51**:

2235 521–528.

2236 **Twyford AD, Streisfeld MA, Lowry DB, Friedman J. 2015.** Genomic studies on the nature

2237 of species: adaptation and speciation in *Mimulus*. *Molecular ecology* **24**: 2601–2609.

2238 **Vellinga EC, Wolfe BE, Pringle A. 2009.** Global patterns of ectomycorrhizal introductions.

2239 *The New phytologist* **181**: 960–973.

2240 **Vinichuk MM, Johanson KJ. 2003.** Accumulation of <sup>137</sup>Cs by fungal mycelium in forest

2241 ecosystems of Ukraine. *Journal of environmental radioactivity* **64**: 27–43.

2242 **Vinichuk M, Rosén K, Johanson KJ, Dahlberg A. 2011.** Correlations between potassium,

2243 rubidium and cesium <sup>133</sup>Cs and <sup>37</sup>Cs in sporocarps of *Suillus variegatus* in a Swedish boreal

2244 forest. *Journal of environmental radioactivity* **102**: 386–392.

2245 **Visser S. 1995.** Ectomycorrhizal fungal succession in jack pine stands following wildfire. *The*

2246 *New phytologist* **129**: 389–401.

2247 **Vlk L, Tedersoo L, Antl T, Větrovský T, Abarenkov K, Pergl J, Albrechtová J, Vosátka**

2248 **M, Baldrian P, Pyšek P, et al. 2020.** Early successional ectomycorrhizal fungi are more

2249 likely to naturalize outside their native range than other ectomycorrhizal fungi. *The New*

2250 *phytologist* **227**: 1289–1293.

2251 **Wang B, Mitchell-Olds T. 2017.** Balancing selection and trans-specific polymorphisms.

2252 *Genome biology* **18**: 231.

2253 **Wang J, Zhang H, Gao J, Zhang Y, Liu Y, Tang M. 2021.** Effects of ectomycorrhizal fungi

2254 (*Suillus variegatus*) on the growth, hydraulic function, and non-structural carbohydrates of

2255 *Pinus tabulaeformis* under drought stress. *BMC plant biology* **21**: 171.

2256 **Williams MC, Wardle GM. 2007.** *Pinus radiata* invasion in Australia: Identifying key

2257 knowledge gaps and research directions. *Austral ecology* **32**: 721–739.

2258 **Wills C. 2007.** *Principles of Population Genetics, 4th edition*. Oxford Academic.

2259 **Wilson AW, May TW, Mueller GM. 2017.** Biogeography of the Ectomycorrhizal Mushroom

2260 Genus *Laccaria*. In: Tedersoo L, ed. *Biogeography of Mycorrhizal Symbiosis*. Cham:

2261 Springer International Publishing, 273–297.

2262 **Wood JR, Dickie IA, Moeller HV, Peltzer DA, Bonner KI, Rattray G, Wilmshurst JM.**

2263 **2015.** Novel interactions between non-native mammals and fungi facilitate establishment of

2264 invasive pines. *The Journal of ecology* **103**: 121–129.

2265 **Wu CA, Lowry DB, Cooley AM, Wright KM, Lee YW, Willis JH. 2008.** *Mimulus* is an

2266 emerging model system for the integration of ecological and genomic studies. *Heredity* **100**:

2267 220–230.

2268 **Wu G, Miyauchi S, Morin E, Kuo A, Drula E, Varga T, Kohler A, Feng B, Cao Y, Lipzen**

2269 **A, et al. 2022.** Evolutionary innovations through gain and loss of genes in the

2270 ectomycorrhizal Boletales. *The New phytologist* **233**: 1383–1400.

2271 **Wu G, Wu K, Qi L-L, Morozova OV, Alexandrova AV, Gorbunova IA, Li Y, Liu J-W,**

2272 **Yang ZL. 2020.** *Psiloboletinus* is an independent genus sister to *Suillus*. *Mycologia* **112**: 185–

2273 196.

2274 **Xu H, Zwiazek JJ. 2020.** Fungal Aquaporins in Ectomycorrhizal Root Water Transport.

2275 *Frontiers in plant science* **11**: 302.

2276 **Zhang R, Shi X-F, Liu P-G, Wilson AW, Mueller GM. 2022.** Host shift speciation of the

2277 ectomycorrhizal genus *Suillus* (suillineae, Boletales) and biogeographic comparison with its

2278 host Pinaceae. *Frontiers in microbiology* **13**: 831450.

2279 **Zhang K, Tappero R, Ruytinx J, Branco S, Liao H-L. 2021.** Disentangling the role of

2280 ectomycorrhizal fungi in plant nutrient acquisition along a Zn gradient using X-ray imaging.

2281 *The Science of the total environment* **801**: 149481.

2282 **Zhou Z, Miwa M, Hogetsu T. 1999.** Analysis of Genetic Structure of a *Suillus grevillei*

2283 Population in a *Larix kaempferi* Stand by Polymorphism of Inter-Simple Sequence Repeat

2284 (ISSR). *The New phytologist* **144**: 55–63.

2285 **Zhou Z, Miwa M, Hogetsu T. 2001.** Polymorphism of simple sequence repeats reveals gene

2286 flow within and between ectomycorrhizal *Suillus grevillei* populations. *The New phytologist*  
2287 **149**: 339–348.  
2288 **Zhu YG, Smolders E. 2000.** Plant uptake of radiocaesium: a review of mechanisms,  
2289 regulation and application. *Journal of experimental botany* **51**: 1635–1645.  
2290 **Zou Y-N, Wu Q-S, Kuča K. 2021.** Unravelling the role of arbuscular mycorrhizal fungi in  
2291 mitigating the oxidative burst of plants under drought stress. *Plant biology* **23 Suppl 1**: 50–  
2292 57.